## Supporting Information

## Supramolecular Complex of Photochromic Diarylethene and Cucurbit[7]uril: Fluorescent Photoswitching System for Biolabeling and Imaging

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## Abbreviations

ACN [acetonitrile], ap [antiparallel conformer], Ar [argon], calcd [calculated], CB7 [cucurbit[7]uril], CD<sub>3</sub>OD [deuterated methanol], CF [closed form], d [day], DAE [diarylethene], DAIB [diacetoxyiodosobenzene], DCM [dichloromethane], DFT [density functional theory], DIPEA [*N*,*N*-diisopropyl ethyl amine], DMF [dimethylformamide], EA [ethyl acetate], ESI [electrospray ionization], EtOH [ethanol], equiv. [equivalents], FP [fluorescent protein], GFP [green fluorescent protein], h [hour], HPLC [high performance liquid chromatography], HRMS [high-resolution mass spectrometry], IRF [instrumental response function], K<sub>2</sub>CO<sub>3</sub> [photassium carbonate], MeOH [methanol], min [minute], Na<sub>2</sub>SO<sub>4</sub> [sodium sulfate], n-Hex [n-hexane], NHS [*N*-hydroxysuccinimidyl], NMR [nuclear magnetic resonance], OF [open form], p [parallel conformer], PBS [phosphate buffered saline], Pd(dba)<sub>2</sub> [bis(dibenzylideneacetone)palladium(0)], PL [photoluminescence], RESOLFT [reversible saturable optical fluorescence transitions], RP [reversed phase], rsFP [reversibly photoswitchable fluorescent protein], r.t. [room temperature], SPhos [2-dicyclohexylphosphino-2',6'-dimethoxybiphenyl], TBAC [tetrabutylammonium chloride], TEMPO [2,2,6,6-tetramethylpiperidine 1-oxyl], TFA [trifluoroacetic acid], THF [tetrahydrofuran], TLC [thin layer chromatography], TSTU [*N*,*N*,*N*',*N*'-tetramethyl-*O*-succinimidyluronium tetrafluoroborate], UV/Vis [ultraviolet/visible light], 3D [three-dimensional].

Synthesis of DAE



Compound **b** was prepared according to the literature procedure.<sup>[S1]</sup> To a DCM solution (15 mL) containing compound **b** (200 mg, 0.25 mmol) was added iodomethane (1.42 g, 10 mmol, 40 equiv.). The reaction mixture was stirred at r.t. for 3 d. The precipitate was filtered and was washed with DCM. The solid product was dissolved in MeOH (10 mL), and a 10 mL MeOH solution containing TBAC (1.39 g, 5.0 mmol, 20 equiv.) was added dropwise. The solution was further stirred for 1 d. The reaction mixture was concentrated in vacuo, and the residue recrystallized from a mixture of EtOH and n-Hex. Yield 135 mg (60%) of **DAE** as an orange-yellow powder. ap:p = 5:3.

<sup>1</sup>H-NMR (400 MHz, CD<sub>3</sub>OD): δ (ppm) = 8.22 (d, J = 1.6 Hz, 1.25H, ap), 8.14 – 8.12 (m, 1.25H, ap), 8.11 (d, J = 1.8 Hz, 0.75H, p), 8.10 – 8.06 (m, 2.75H, ap/p), 8.05 – 7.97 (m, 4.5H, ap/p), 7.95 – 7.89 (m, 1.5H, ap/p), 7.65 (d, J = 8.0 Hz, 0.75H, p), 7.58 (d, J = 8.0 Hz, 1.25H, ap), 3.74 (s, 11.25H, ap), 3.71 (s, 6.75H, p), 2.79 – 2.46 (m, 4H, ap/p), 1.40 (t, J = 7.6 Hz, 2.25H), 1.06 (t, J = 7.6 Hz, 3.75H). <sup>13</sup>C NMR (101 MHz, CD<sub>3</sub>OD): δ (ppm) = 150.7, 150.5, 148.7, 148.6, 143.1, 143.0, 141.5, 141.4, 138.1, 137.9, 134.3, 134.0, 130.3, 130.2, 123.0, 125.6, 125.4, 124.3, 122.2, 121.9, 121.8, 57.8, 20.4, 20.3, 12.4 12.1. <sup>19</sup>F NMR (367 MHz, CD<sub>3</sub>OD): δ (ppm) = -111.1 (m, 4F, ap/p), -132.9 (m, 2F, ap/p). HRMS (ESI+, *m*/z): M<sup>2+</sup> calcd for C<sub>43</sub>H<sub>42</sub>F<sub>6</sub>N<sub>2</sub>O<sub>4</sub>S<sub>2</sub><sup>2+</sup>, 414.1240; found, 414.1250.

Synthesis of compound d



Compound **c** was prepared according to the literature procedure.<sup>[S2]</sup> A flask was charged with compound **c** (150 mg, 0.18 mmol), 4-(dimethylamino)phenylboronic acid (72 mg, 0.44 mmol, 2.5 equiv.), Pd(dba)<sub>2</sub> (25 mg, 0.044 mmol, 0.25 equiv.), and SPhos (18 mg, 0.044 mmol, 0.25 equiv.). THF (20 mL) and saturated aqueous  $K_2CO_3$  (20 mL) were added, and the mixture was degassed by Ar bubbling for 10 min. The solution was stirred at 80 °C for 1 h, while monitoring the progress of the reaction by TLC. After the reaction was complete, the mixture was poured into brine and extracted with DCM. The organic solution was dried over Na<sub>2</sub>SO<sub>4</sub> and concentrated under reduced pressure. The crude material

was subjected to chromatography on silica gel with gradient elution (EA/n-Hex:  $0/100 \rightarrow 50/50$ ) to give 112 mg (76%) of **d** as an orange powder. ap:p = 5:3.

<sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.29 (dd, *J* = 3.7, 1.6 Hz, 1.25H, ap), 8.17 (d, *J* = 1.7 Hz, 0.75H, p), 8.11 (ddd, *J* = 8.2, 2.8, 1.8 Hz, 1.25H, ap), 7.91 (ddd, *J* = 7.8, 5.9, 1.8 Hz, 0.75H, p), 7.82 – 7.71 (m, 3.25H, ap/p), 7.71 – 7.63 (m, 2.75H, ap/p), 6.90 – 6.84 (m, 2.5H, ap), 6.80 (d, *J* = 8.5 Hz, 1.5H, p), 4.55 (t, *J* = 5.1 Hz, 0.37H, p), 4.47 (t, *J* = 5.1 Hz, 0.63H, ap), 3.59 (q, *J* = 6.2 Hz, 0.75H, p), 3.38 (ddt, *J* = 21.6, 10.9, 5.4 Hz, 1.25H, ap), 3.03 (s, 7.5H, ap), 2.98 (s, 4.5H, p), 2.81 – 2.47 (m, 4H, ap/p), 2.06 – 1.34 (m, 5.12H, ap/p), 1.00 (t, *J* = 7.6 Hz, 1.88H, p). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 152.34, 152.31, 152.25, 148.7, 148.4, 147.8, 147.6, 145.4, 145.2, 145.1, 137.6, 137.50, 137.47, 131.9, 131.8, 131.41, 131.38, 128.94, 128.89, 128.84, 127.0, 126.9, 126.8, 126.7, 125.7, 125.64, 125.55, 125.50, 124.7, 124.4, 120.03, 119.97, 119.89, 113.59, 113.57, 113.51, 62.0, 61.9, 40.60, 40.56, 33.8, 33.7 26.6, 26.6, 25.5, 25.3, 20.1, 19.8, 12.9, 12.5. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = -109.8 (m, 4F, ap/p), -131.2 (m, 2F, ap/p).

HRMS (ESI+, *m/z*): [M+2H]<sup>2+</sup> calcd for C<sub>43</sub>H<sub>42</sub>F<sub>6</sub>N<sub>2</sub>O<sub>5</sub>S<sub>2</sub><sup>2+</sup>, 422.1214; found, 422.1215.

Synthesis of compound e



To a dioxane solution (2 mL) containing compound **d** (54 mg, 64 µmol), was added iodomethane (2 mL, 32 mmol, 500 equiv.). The mixture was heated to 80 °C and stirred for 24 h. The precipitate was filtered off and subjected to preparative HPLC with gradient elution (0.1% aq. TFA / ACN, 85/15  $\rightarrow$  10/90). Lyophilization yielded 52 mg (73%) of compound **e** as a yellow powder. ap:p = 7:3.

<sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.57 (dd, J = 5.9, 1.6 Hz, 1.4H, ap), 8.47 (d, J = 1.6 Hz, 0.6H, p), 8.39 – 8.32 (m, 4H, ap/p), 8.30 – 8.21 (m, 4H, ap/p), 8.18 (dq, J = 6.3, 2.1 Hz, 0.6H, p), 8.16 – 8.11 (m, 1.4H, ap), 8.01 – 7.97 (m, 0.6H, p), 7.90 (t, J = 8.5 Hz, 1.4H, ap), 3.96 (s, 12.6H, ap), 3.91 (s, 5.4H, p), 3.61 (t, J = 6.2 Hz, 0.6H, p), 3.41 (t, J = 6.2 Hz, 1.4H, ap), 2.89 – 2.54 (m, 4H, ap/p), 2.08 – 1.34 (m, 4.9H, ap/p), 1.03 (t, J = 7.6 Hz, 2.1H, ap). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 159.8, 159.5, 150.3, 149.5, 149.12, 149.09, 149.05, 142.8, 142.7, 142.5, 140.4, 140.3, 140.2, 137.6, 137.5, 137.4, 134.60, 134.58, 134.2, 134.1, 130.0, 129.94, 129.87, 129.8, 129.7, 129.6, 126.0, 125.94, 125.85, 124.3, 124.0, 122.68, 122.65, 122.59, 122.12, 122.08, 122.0, 120.0, 117.1, 62.0, 61.9, 57.74, 57.69, 33.7, 33.6, 26.7, 26.6, 25.4, 25.2, 20.3, 20.0, 12.8, 12.4. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = -109.5 (m, 4F,

ap/p), -130.7 (m, 2F, ap/p). HRMS (ESI+, m/z): M<sup>2+</sup> calcd for C<sub>45</sub>H<sub>46</sub>F<sub>6</sub>N<sub>2</sub>O<sub>5</sub>S<sub>2</sub><sup>2+</sup>, 436.1371; found, 436.1372.

Synthesis of compound DAE-COOH



Compound e (30 mg, 27 µmol) was dissolved in a mixture of ACN (400 µL) and water (400 µL), and the solution was cooled to -10 °C. To this solution were added TEMPO (21 mg, 140 µmol, 5 equiv.) and DAIB (88 mg, 270 µmol, 10 equiv.), and the reaction mixture was stirred for 2 h, while monitoring the progress of the reaction by analytical HPLC. Purification was carried out by preparative HPLC with gradient elution (0.1% aq. TFA / ACN, 95/5  $\rightarrow$  50/50). Lyophilization yielded 24 mg (79%) of compound **DAE-COOH** as a yellow powder. ap:p = 7:3.

<sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.57 (dd, J = 18.2, 1.6 Hz, 1.4H, ap), 8.48 – 8.40 (m, 0.6H, p), 8.38 – 8.12 (m, 10H, ap/p), 7.98 – 7.89 (m, 2H, ap/p), 3.96 (d, J = 1.6 Hz, 12.6H, ap), 3.91 (d, J = 2.4 Hz, 5.4H, p), 2.88 – 2.55 (m, 4H, ap/p), 2.50 – 1.61 (m, 4H, ap/p), 1.42 (t, J = 7.5 Hz, 0.9H, p), 1.01 (t, J = 7.5 Hz, 2.1H, ap). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 174.83, 174.77, 149.13, 149.06, 142.9, 142.8, 140.5, 140.3, 137.5, 134.6, 134.5, 130.02, 129.97, 129.90, 129.86, 129.7, 129.6, 126.1, 125.9, 124.8, 124.7, 123.98, 123.97, 122.74, 122.68, 122.62, 122.59, 122.1, 57.75, 57.70, 31.2, 25.75, 25.70, 23.96, 23.90, 19.99, 19.96, 12.8, 12.2. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = –109.1 (m, 4F, ap/p), –130.5 (m, 2F, ap/p). HRMS (ESI+, *m/z*): M<sup>2+</sup> calcd for C<sub>45</sub>H<sub>44</sub>F<sub>6</sub>N<sub>2</sub>O<sub>6</sub>S<sub>2</sub><sup>2+</sup>, 443.1267; found, 443.1266.

Synthesis of compound DAE-Male1



To an ACN solution (1.5 mL) containing compound **DAE-COOH** (32 mg, 28 µmol) and TSTU (17 mg, 57 µmol, 2 equiv.), was added DIPEA (15 mg, 114 µmol, 4 equiv.). The reaction mixture was stirred at r.t. for 10 min. 1-(2-Aminoethyl)maleimide hydrochloride (20 mg, 114 µmol, 4 equiv.) and DIPEA (15 mg, 114 µmol, 4 equiv.) were added to the reaction mixture which was then stirred for 30 min at r.t.

Purification was carried out by preparative HPLC with gradient elution (0.1% aq. TFA / ACN, 95/5 → 50/50). Lyophilization yielded 22 mg (61%) of compound **DAE-Male1** as a yellow powder. ap:p = 7:3. <sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.55 (dd, *J* = 14.4, 1.6 Hz, 1.4H, ap), 8.46 (t, *J* = 1.7 Hz, 0.6H, p), 8.38 – 8.31 (m, 4H, ap/p), 8.31 – 8.22 (m, 4H, ap/p), 8.21– 7.86 (m, 4H, ap/p), 7.02 (s, 0.6H, p), 7.00 (s, 1.4H, ap), 3.96 (s, 12.6H, ap), 3.91 (d, *J* = 0.8 Hz, 5.4H, p), 3.66 – 3.22 (m, 4H, ap/p), 2.91 – 2.59 (m, 4H, ap/p), 2.42 – 1.59 (m, 4H, ap/p), 1.42 (t, *J* = 7.5 Hz, 0.9H), 1.07 (t, *J* = 7.6 Hz, 2.1H). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 172.8, 172.7, 172.32, 172.28, 149.11, 149.08, 149.06, 142.8, 142.7, 142.5, 140.5, 140.3, 140.2, 137.6, 137.5, 137.4, 135.6, 134.7, 134.6, 130.07, 129.95, 129.88, 129.86, 129.75, 129.6, 125.9, 125.8, 124.5, 124.0, 122.7, 122.6, 122.1, 57.8, 26.2, 24.14, 24.11, 20.3, 20.1, 12.8, 12.4. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = –109.2 (m, 4F, ap/p), –130.4 (m, 2F, ap/p). HRMS (ESI+, *m/z*): M<sup>2+</sup> calcd for C<sub>51</sub>H<sub>50</sub>F<sub>6</sub>N<sub>4</sub>O<sub>7</sub>S<sub>2</sub><sup>2+</sup>, 504.1507; found, 504.1520.

Synthesis of compound DAE-Male2



To an ACN solution (1 mL) containing compound **DAE-COOH** (25 mg, 22 µmol) and TSTU (14 mg, 45 µmol, 2 equiv.), was added DIPEA (12 mg, 90 µmol, 4 equiv.). The reaction mixture was stirred at r.t. for 10 min. N-(5-Aminopenthyl)maleimide hydrochloride (20 mg, 90 µmol, 4 equiv.) and DIPEA (12 mg, 90 µmol, 4 equiv.) were added to the reaction mixture, which was then stirred for 40 min at r.t. Purification was carried out by preparative HPLC with gradient elution (0.1% aq. TFA / ACN, 95/5  $\rightarrow$  50/50). Lyophilization yielded 14.4 mg (50%) of compound **DAE-Male2** as a yellow powder. ap:p = 7:3.

<sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.56 (dd, J = 14.7, 1.7 Hz, 1.4H, ap), 8.47 (d, J = 1.7 Hz, 0.6H, p), 8.38 – 8.31 (m, 4H, ap/p), 8.31 – 8.22 (m, 4H, ap/p), 8.22 – 7.87 (m, 4H), 7.03 (s, 0.6H, p), 7.02 (s, 1.4H, ap), 3.96 (d, J = 1.3 Hz, 12.6H, ap), 3.91 (d, J = 2.1 Hz, 5.4H, p), 3.53 – 3.03 (m, 4H, ap/p), 2.90 – 2.56 (m, 4H, ap/p), 2.46 – 1.22 (m, 10.9H, ap/p), 1.05 (t, J = 7.6 Hz, 2.1H, p). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 172.30, 172.28, 172.2, 159.9, 159.5, 150.3, 149.11, 149.08, 149.05, 142.8, 142.7, 142.54, 142.48, 140.5, 140.3, 140.23, 140.20, 137.54, 137.48, 137.46, 137.4, 135.6, 134.7, 134.59, 130.04, 129.95, 129.88, 129.85, 129.75, 129.6, 126.0, 125.8, 124.6, 122.7, 122.6, 122.1, 122.03, 121.99, 57.8, 57.74, 57.69, 39.80, 39.77, 38.32, 38.31, 26.2, 26.1, 25.1, 25.0, 24.7, 24.5, 20.0, 12.8, 12.4. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = -109.2 (m, 4F, ap/p), -130.5 (m, 2F, ap/p).

HRMS (ESI+, *m/z*): M<sup>2+</sup> calcd for C<sub>54</sub>H<sub>56</sub>F<sub>6</sub>N<sub>4</sub>O<sub>7</sub>S<sub>2</sub><sup>2+</sup>, 525.1742; found, 525.1754.

Synthesis of compound SI-a



NH2-PEG2-COOH (100 mg, 0.56 mmol) was dissolved in 3 mL anhydrous MeOH, and the solution was stirred at 0 °C for 5 min. Di-tert-butyl dicarbonate (123 mg, 0.56 mmol, 1 equiv.) was added to the reaction mixture, which was then stirred for 30 min at 0 °C. The temperature was changed to r.t., and then the mixture was stirred for 90 min. The mixture was poured into DCM and brine, and extracted with DCM. The combined organic solutions were dried over  $Na_2SO_4$  and evaporated. Lyophilization with 1,4-dioxane yielded 112 mg (72%) of compound **SI-a** as a pale yellow oil.

<sup>1</sup>H-NMR (400 MHz, chloroform-*d*):  $\delta$  (ppm) = 5.07 (s, 1H), 3.77 (t, *J* = 6.1 Hz, 2H), 3.65 – 3.56 (m, 4H), 3.56 – 3.47 (m, 2H), 3.34 – 3.21 (m, 2H), 2.79 – 2.52 (m, 2H), 1.44 (s, 9H). HRMS (ESI+, *m/z*): [M+Na]<sup>+</sup> calcd for C<sub>12</sub>H<sub>23</sub>NNaO<sub>6</sub><sup>+</sup>, 300.1418; found, 300.1420.

Synthesis of compound SI-b



To a DMF solution (1.5 mL) containing compound SI-a (25 mg, 90 µmol) and TSTU (41 mg, 140 µmol, 1.5 equiv.), was added DIPEA (41 mg, 320 µmol, 3.5 equiv.). The reaction mixture was stirred at r.t. for 10 min. N-(5-Aminopenthyl)maleimide hydrochloride (59 mg, 270 µmol, 3 equiv.) and DIPEA (12 mg, 320 µmol, 3.5 equiv.) were added to the reaction mixture, which was then stirred for 30 min at r.t. The crude material was subjected to chromatography on silica gel with gradient elution (MeOH/DCM: 0/100  $\rightarrow$  15/85) to give 36 mg (90%) of SI-b as a yellow oil.

<sup>1</sup>H-NMR (400 MHz, Chloroform-*d*):  $\delta$  (ppm) = 6.68 (s, 2H), 6.30 (s, 1H), 5.01 (s, 1H), 3.73 (t, *J* = 5.8 Hz, 2H), 3.62 (s, 4H), 3.57 – 3.48 (m, 4H), 3.35 – 3.26 (m, 2H), 3.22 (q, *J* = 6.9 Hz, 2H), 2.46 (t, *J* = 5.8 Hz, 2H), 1.64 – 1.48 (m, 4H), 1.44 (s, 9H), 1.35 – 1.28 (m, 2H).

HRMS (ESI+, m/z): [M+Na]<sup>+</sup> calcd for C<sub>21</sub>H<sub>35</sub>N<sub>3</sub>NaO<sub>7</sub><sup>+</sup>, 464.2367; found, 464.2364.

Synthesis of compound SI-c



To a DCM solution (1.2 mL) containing compound **SI-b** (30 mg), was added TFA (300  $\mu$ L). The reaction mixture was stirred at r.t. for 3 h. Evaporation and lyophilization with 1,4-dioxane yielded 30 mg (99%) of compound **SI-c** as a pale yellow oil.

<sup>1</sup>H-NMR (400 MHz, Chloroform-*d*): δ (ppm) = 8.05 (s, 2H), 6.70 (s, 2H), 6.45 (s, 1H), 3.83 – 3.75 (m, 4H), 3.70 – 3.61 (m, 4H), 3.54 – 3.49 (m, 2H), 3.27 – 3.18 (m, 4H), 2.47 (t, *J* = 5.5 Hz, 2H), 1.65 – 1.49 (m, 4H), 1.34 – 1.28 (m, 2H).

HRMS (ESI+, *m/z*): [M+Na]<sup>+</sup> calcd for C<sub>21</sub>H<sub>35</sub>N<sub>3</sub>NaO<sub>7</sub><sup>+</sup>, 464.2367; found, 464.2364.

Synthesis of compound DAE-Male3



To a DMF solution (600 µL) containing compound **DAE-COOH** (10 mg, 9.0 µmol) and TSTU (5.4 mg, 18 µmol, 2 equiv.), was added DIPEA (9.3 mg, 36 µmol, 4 equiv.). The reaction mixture was stirred at r.t. for 10 min. Compound **SI-c** (9.2 mg, 27 µmol, 3 equiv.) and DIPEA (9.3 mg, 36 µmol, 4 equiv.) were added to the reaction mixture, which was then stirred for 30 min at r.t. Purification was carried out by preparative HPLC with gradient elution (0.1% aq. TFA / ACN, 95/5  $\rightarrow$  70/30). Lyophilization yielded 8.8 mg (68%) of compound **DAE-Male3** as an orange yellow powder. ap:p = 7:3.

<sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.56 (dd, J = 13.5, 1.7 Hz, 1.4H, ap), 8.46 (d, J = 1.6 Hz, 0.6H, p), 8.37 – 8.32 (m, 4H, ap/p), 8.30 – 8.23 (m, 4H, ap/p), 8.22 – 7.79 (m, 4H, ap/p), 7.03 – 6.99 (m, 2H, ap/p), 3.96 (s, 12.6H, ap), 3.91 (d, J = 2.3 Hz, 5.4H, p), 3.73 – 3.07 (m, 14H, ap/p), 2.90 – 2.55 (m, 4H, ap/p), 2.47 – 1.23 (m, 12.9H, ap/p), 1.04 (t, J = 7.6 Hz, 2.1H, ap). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 172.5, 172.3, 171.33, 171.29, 149.10, 149.07, 149.04, 142.8, 142.7, 142.53, 142.47, 140.5, 140.3, 140.22, 140.19, 137.54, 137.48, 137.4, 135.6, 134.7, 134.6, 130.1, 130.0, 129.9, 129.8, 129.7, 129.6, 126.2, 125.9, 124.5, 123.9, 123.8, 122.7, 122.59, 122.57, 122.1, 122.0, 71.1, 71.0, 71.0, 70.9, 70.7, 70.5, 68.33, 68.30, 57.8, 57.74, 57.69, 40.0, 39.9, 39.7, 38.3, 37.62, 37.55, 29.1, 26.1, 25.0, 24.4, 20.0, 12.8, 12.4. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = -109.1 (m, 4F, ap/p), -130.3 (m, 2F, ap/p).

HRMS (ESI+, *m/z*): M<sup>2+</sup> calcd for C<sub>61</sub>H<sub>69</sub>F<sub>6</sub>N<sub>5</sub>O<sub>10</sub>S<sub>2</sub><sup>2+</sup>, 604.7190; found, 604.7173.

Synthesis of compound f



To an ACN solution (900 µL) containing compound **DAE-COOH** (25 mg, 22 µmol) and TSTU (14 mg, 45 µmol, 2 equiv.), was added DIPEA (12 mg, 90 µmol, 4 equiv.). The reaction mixture was stirred at r.t. for 10 min.  $\gamma$ -Aminobutyric acid (9.3 mg, 90 µmol, 4 equiv.) and DIPEA (12 mg, 90 µmol, 4 equiv.) were added to the reaction mixture which was then stirred for 40 min at r.t. Purification was carried out by preparative HPLC with gradient elution (0.1% aq. TFA / ACN, 95/5  $\rightarrow$  50/50). Lyophilization yielded 18.7 mg (70%) of compound **f** as a yellow powder. ap:p = 7:3.

<sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.55 (dd, *J* = 15.4, 1.6 Hz, 1.4H, ap), 8.47 (s, 0.6H, p), 8.38 – 8.31 (m, 4H, ap/p), 8.31 – 8.22 (m, 4H, ap/p), 8.22 – 7.86 (m, 4H, ap/p), 3.96 (d, *J* = 1.5 Hz, 12.6H, ap), 3.91 (d, *J* = 1.8 Hz, 5.4H, p), 3.28 – 3.08 (m, 2H, ap/p), 2.90 – 2.58 (m, 4H, ap/p), 2.48 – 1.62 (m, 8H, ap/p), 1.43 (t, *J* = 7.5 Hz, 0.9H, p), 1.05 (t, *J* = 7.5 Hz, 2.1H). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 175.3, 175.2, 172.4, 159.9, 159.6, 149.1, 149.0, 142.8, 142.7, 142.53, 142.47, 140.4, 140.3, 140.21, 140.19, 137.54, 137.46, 137.45, 137.4, 134.7, 134.6, 134.2, 134.1, 130.0, 129.94, 129.87, 129.8, 129.7, 129.64, 129.58, 126.0, 125.8, 124.5, 123.9, 122.7, 122.60, 122.57, 122.5, 122.1, 122.0, 57.73, 57.68, 39.4, 32.4, 32.3, 26.17, 26.15, 24.7, 24.4, 20.3, 20.0, 12.8, 12.4. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = -109.4 (m, 4F, ap/p), -130.5 (m, 2F, ap/p).

HRMS (ESI+, m/z): M<sup>2+</sup> calcd for C<sub>49</sub>H<sub>51</sub>F<sub>6</sub>N<sub>3</sub>O<sub>7</sub>S<sub>2</sub><sup>2+</sup>, 485.6531; found, 485.6539.

Synthesis of compound DAE-NHS



To an ACN solution (1 mL) containing compound **f** (20 mg, 17  $\mu$ mol) and TSTU (7.5 mg, 25  $\mu$ mol, 1.5 equiv.), was added DIPEA (7.8 mg, 60  $\mu$ mol, 3.6 equiv.). The reaction mixture was stirred at r.t. for 30 min. Purification was carried out by preparative HPLC with gradient elution (0.1% aq. TFA / ACN,

 $95/5 \rightarrow 50/50$ ). Lyophilization yielded 12 mg (56%) of compound **DAE-NHS** as a yellow powder. ap:p = 7:3.

<sup>1</sup>H-NMR (400 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 8.55 (dd, *J* = 15.1, 1.7 Hz, 1.4H, ap), 8.46 (s, 0.6H, p), 8.38 – 8.31 (m, 4H), 8.30 – 8.22 (m, 4H), 8.21 – 7.86 (m, 4H), 3.95 (d, *J* = 2.5 Hz, 12.6H, ap), 3.91 (d, *J* = 1.9 Hz, 5.4H, p), 3.37 – 3.19 (m, 2H, ap/p), 2.96 – 2.93 (m, 4H, ap/p), 2.89 – 2.54 (m, 5H, ap/p), 2.51 – 1.57 (m, 7H, ap/p), 1.43 (t, *J* = 7.5 Hz, 0.9H, p), 1.05 (t, *J* = 7.5 Hz, 2.1H, ap). <sup>13</sup>C NMR (101 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = 172.58, 172.55, 171.4, 171.3, 170.14, 170.12, 150.2, 149.1, 149.0, 142.73, 142.68, 140.4, 140.3, 137.5, 137.4, 134.6, 130.0, 129.92, 129.85, 129.82, 125.9, 125.8, 124.5, 123.9, 122.6, 122.5, 122.1, 57.72, 57.67, 39.0, 29.2, 26.7, 26.4, 26.1, 25.9, 25.8, 24.3, 20.0, 12.8, 12.4. <sup>19</sup>F NMR (367 MHz, DMF-d<sub>7</sub>):  $\delta$  (ppm) = -109.4 (m, 4F, ap/p), -130.5 (m, 2F, ap/p).

HRMS (ESI+, m/z): M<sup>2+</sup> calcd for C<sub>53</sub>H<sub>54</sub>F<sub>6</sub>N<sub>4</sub>O<sub>9</sub>S<sub>2</sub><sup>2+</sup>, 534.1613; found, 534.1613.



**Figure S1.** Absorption spectra (extinction coefficients) and normalized emission spectra of compounds **DAE** (A) and **DAE-COOH** (B) in aqueous solutions. The black and red lines are the spectra of open and closed isomers, respectively.



**Figure S2.** Absorption and emission changes of an aqueous solution of compound **DAE** ( $5.4 \mu$ M) under irradiation with 365 nm light (A-D) and 470 nm light (E-F).



**Figure S3.** Lifetime decay profiles of (a) **DAE** and (b) **DAE**@CB7 complex in aqueous solution. The excitation and emission wavelengths were 405 nm and 555 nm, respectively. The green, blue, and red lines represent lifetime decay, instrumental response functions (IRF), and fitted curve, respectively. The lower red line represents the residual.



**Figure S4.** Lifetime decay profiles of (a) **DAE** and (b) **DAE**@CB7 complex in aqueous solution after argon purging for 15 min. The excitation and emission wavelengths were 405 nm and 555 nm, respectively. The green, blue, and red lines represent lifetime decay, instrumental response functions (IRF), and fitted curve, respectively. The lower red line represents the residual.

**Table S1.** Fluorescence quantum yields ( $\Phi_{fl}$ ), fluorescence lifetimes ( $\tau_{fl}$ ), radiative ( $k_r$ ) and non-radiative ( $k_{nr}$ ) rate constants of free **DAE** and **DAE**@CB7 complex at the photostationary state upon 365 nm irradiation in aqueous solution before and after argon purging.

	DAE				DAE@CB7			
	$\Phi_{ m fl}$	$\tau_{\rm fl}(\rm ns)$	$k_{r} (s^{-1})$	$k_{nr} (s^{-1})$	$\Phi_{ m fl}$	$\tau_{\rm fl}(\rm ns)$	$k_{r} (s^{-1})$	$k_{nr} (s^{-1})$
H <sub>2</sub> O	0.40	1.83	2.19E8	3.28E8	0.63	2.33	2.70E8	1.59E8
H <sub>2</sub> O (Ar purged)	0.41	1.80	2.28E8	3.28E8	0.63	2.31	2.73E8	1.60E8



Figure S5. (a) UV/vis and (b) PL spectra of DAE  $(10^{-5} \text{ M})$  in water upon CB7 addition. (c) PL intensity changes upon CB7 addition.



Figure S6. The raw data of the isothermal titration calorimetry (ITC) measurements: thermographs of titration of 1000  $\mu$ M DAE into 100  $\mu$ M CB7 (black line) and titration of 1000  $\mu$ M DAE into pure water (red line). The ligand-target binding signal was subtracted by control signal, and used for Figure 2b in the main text.



**Figure S7.** (a) UV/vis and (b) PL spectra of the mixed solutions of **DAE** and CB7 in water,  $[DAE]+[CB7]=10^{-5}$  M; (c) the Job's plot for complexation of **DAE** with CB7, derived from PL spectra (b), PL intensity: emission area,  $\chi_{CB7}$ : molar fraction of CB7 in a mixture.



Figure S8. High resolution mass-spectrum (HRMS) of DAE@CB7; a peak with m/z = 1576.4698 supports 1:2 stoichiometry of the DAE@CB7 complex.



**Figure S9.** (a) <sup>1</sup>H NMR spectra of **DAE**@CB7 complex in D<sub>2</sub>O. CB7 titration induced changes in the resonance signals: (b) <sup>1</sup>H NMR aromatic region and (c) <sup>1</sup>H NMR aliphatic region of **DAE** in D<sub>2</sub>O. [DAE] = 2 mM. The changes of the chemical shifts were observed for 10 out of total 14 aromatic protons. Shielding = encapsulation by CB7. The chemical shifts of ethyl groups at C-2/C-2' did not change considerably; these groups are situated outside of CB7. One set of signals of ethyl groups in the complex indicate the presence of only one (anti-parallel) conformer.



Figure S10. Results of the DFT geometry optimization (B3LYP/6-31G) in water (CPCM) for the antiparallel conformer, parallel conformer, and the closed-ring isomer of DAE in the CB7 complex: DAE(ap)@CB7, DAE(p)@CB7, and DAE(c)@CB7. All energies are given in *kcal/mol*, relative to the closed-ring isomer, DAE(c)@CB7.



**Figure S11.** Results of the DFT geometry optimization (B3LYP/6-31G) in water (CPCM) for the antiparallel conformer of the open-ring isomer, parallel conformer of the open-ring isomer, and the closedring isomer of free **DAE**: **DAE(ap)**, **DAE(p)**, and **DAE(c)**. All energies are given in *kcal/mol* relative to the closed-ring isomer, **DAE(c)**.



**Figure S12.** Absorption and emission changes of an aqueous solution of compound **DAE** (5.4  $\mu$ M) containing CB7 (4 eq.) under irradiation with 365 nm light (A-D) and 470 nm light (E-F).



**Figure S13.** Absorption and emission changes of an aqueous solution of compound **DAE-COOH** (5.4  $\mu$ M) under irradiation with 365 nm light (A-D) and 470 nm light (E-F)).



**Figure S14.** Absorption and emission changes of an aqueous solution of compound **DAE-COOH** (5.4  $\mu$ M) containing CB7 (4 eq.) under irradiation with 365 nm light (A-D) and 470 nm light (E-F)).



**Figure S15.** 2D maps of LC-MS (HPLC with the molecular mass detection; positive mode ESI-MS) after repetitive photoswitching experiments: (a) free **DAE** and (b) **DAE**@CB7 were irradiated with 365 nm light in aqueous solutions. OF: open form, CF: closed form, MP: main byproduct, BP1: main byproduct 1, and BP2: main byproduct 2. The molecular masses of MP and BP1 (808), as well as BP2 (824), correspond to  $-2F+H_2O$ , as well as -2F+2OH transformations of the (CF<sub>2</sub>)<sub>3</sub> bridge.



**Figure S16.** (a) <sup>1</sup>H NMR spectra of **DAE-COOH**@CB7 complex in D<sub>2</sub>O. CB7 titration induced changes in the chemical shifts: (b) <sup>1</sup>H NMR aromatic region and (c) <sup>1</sup>H NMR aliphatic region of **DAE-COOH** in D<sub>2</sub>O. [DAE-COOH] = 2 mM. The signals of 10 out of total 14 aromatic protons are shifted (due to encapsulation by CB7). The positions of the signals of CH<sub>2</sub> groups (ethyl and carboxyalkyl groups) are not particularly affected, as these residues are located outside of CB7. The 1:2 complex **DAE-COOH**@CB7 displays one set of signals (of an anti-parallel conformer).



**Figure S17.** (a) <sup>1</sup>H NMR spectra of **DAE-Male1**@CB7 complex in D<sub>2</sub>O. CB7 titration induced changes in the resonance signals: (b) <sup>1</sup>H NMR aromatic region and (c) <sup>1</sup>H NMR aliphatic region of **DAE-Male1** in D<sub>2</sub>O. [DAE-Male1] = 2 mM. The signals of 10 out of total 14 aromatic protons are shifted (due to encapsulation by CB7). The positions of the signals of CH<sub>2</sub> groups (ethyl and amidoalkyl groups) are not particularly affected, as these residues are located outside of CB7. **DAE-Male1**@CB7 (1:2 complex) displays one set of signals(of an anti-parallel conformer).



**Figure S18.** (a) UV/vis and (b) PL spectra of **DAE-COOH** ( $10^{-5}$  M) in aqueous solutions upon CB7 addition. (c) PL intensity change upon CB7 addition.



**Figure S19.** (a) UV/vis and (b) PL spectra of the aqueous solutions containing **DAE-COOH** and CB7,  $[DAE-COOH]+[CB7]=10^{-5}$  M. (c) the Job's plot for complexation of **DAE-COOH** with CB7 derived from the PL spectra (b), PL intensity: emission area,  $\chi_{CB7}$ : molar fraction of CB7 in a mixture.



**Figure S20.** Lifetime decay profiles of (a) **DAE-COOH** and (b) **DAE-COOH**@CB7 complex in aqueous solution. The excitation and emission wavelengths were 405 nm and 555 nm, respectively. The green, blue, and red lines represent lifetime decay, IRF, and fitted curve, respectively. The lower red line represents the residual.



**Figure S21.** Reversible photoswitching cycles of free **DAE-COOH** (red) and **DAE-COOH**@CB7 (black) complex dissolved in pure water: (a) A zoom into the first 20 cycles, (b) Full cycles up to N=400.



**Figure S22.** (a) UV/vis and (b) PL spectra of **DAE-Male1** (10<sup>-5</sup> M) in aqueous solution upon addition of CB7. (c) the PL intensity changes upon CB7 addition.



**Figure S23.** (a) UV/vis and (b) PL spectra of the aqueous solutions: mixtures of **DAE-Male1** and CB7,  $[DAE-Male1]+[CB7]=10^{-5}$  M; the (c) Job's plot for complexation of **DAE-Male1** with CB7, derived from the PL spectra (b), PL intensity: emission area,  $\chi_{CB7}$ : molar fraction of CB7 in a mixture.



**Figure S24.** Absorption spectra of the prepared bioconjugates: secondary antibodies labeled with the indicated reactive DAEs. Spectra were normalized to the absorption at 280 nm.



**Figure S25.** Confocal images of fixed Cos7 cells, stained with secondary antibodies labeled with **DAE-NHS**, with addition of CB7 (2 mM) during the staining with the secondary. A schematic representation of the protocol is presented on the top. The confocal images were obtained by mounting in PBS (middle row). In the bottom row, images of other cells and other fields of view in the same sample are presented after the addition of CB7 (2 mM). The markers are switched-on before the recording of each pixel, with a short pulse of 355 nm light. We conclude from the experiment that the CB7-DAE complex is dissociated during the washing steps with PBS, probably due to the removal of CB7 during the washing step and/or the presence of single-charged cations (Na<sup>+</sup> and K<sup>+</sup>) in the PBS buffer. Scale bar: 10  $\mu$ m



Figure S26. Confocal images of fixed Cos7 cells, stained with secondary antibodies labeled with the indicated DAE before (top) and after (bottom) CB7 addition. The markers are switched-on before the recording of each pixel with a short pulse of 355 nm light. The images relate to Figure 5 in the main text: the same samples and imaging conditions. Scale bar:  $10 \mu m$ .



**Figure S27.** Confocal (A) and RESOLFT (B) images of fixed Cos7 cells, stained with secondary antibodies labeled with **DAE-Male3**; mounted in PBS *without CB7*. The image here corresponds to the image displayed in Figure 7: the same sample and imaging conditions. Scale bar: 2 µm.



**Figure S28.** Confocal (A) and RESOLFT (B) images of fixed Cos7 cells, stained with secondary antibodies labeled with **DAE-NHS**, mounted in PBS *without CB7*. The image correspond to the one displayed in Figure 7: the same sample and imaging conditions. Scale bar: 2 μm.



Figure S29. <sup>1</sup>H NMR spectrum (400 MHz, CD<sub>3</sub>OD) of DAE.



Figure S30. <sup>13</sup>C NMR spectrum (101 MHz, CD<sub>3</sub>OD) of DAE.







Figure S32. <sup>1</sup>H NMR spectrum (400 MHz, DMF-d<sub>7</sub>) of compound d.



Figure S33. <sup>13</sup>C NMR spectrum (101 MHz, DMF-d<sub>7</sub>) of compound d.



Figure S34. <sup>19</sup>F NMR spectrum (367 MHz, DMF-d<sub>7</sub>) of compound **d**.



Figure S35. <sup>1</sup>H NMR spectrum (400 MHz, DMF-d<sub>7</sub>) of compound e.





Figure S37. <sup>19</sup>F NMR spectrum (367 MHz, DMF-d<sub>7</sub>) of compound e.



Figure S38. <sup>1</sup>H NMR spectrum (400 MHz, DMF-d<sub>7</sub>) of compound DAE-COOH.



Figure S39. <sup>13</sup>C NMR spectrum (101 MHz, DMF-d<sub>7</sub>) of compound DAE-COOH.



Figure S40. <sup>19</sup>F NMR spectrum (367 MHz, DMF-d<sub>7</sub>) of compound DAE-COOH.



Figure S41. <sup>1</sup>H NMR spectrum (400 MHz, DMF-d<sub>7</sub>) of compound DAE-Male1.



Figure S42. <sup>13</sup>C NMR spectrum (101 MHz, DMF-d<sub>7</sub>) of compound DAE-Male1.



Figure S43. <sup>19</sup>F NMR spectrum (367 MHz, DMF-d<sub>7</sub>) of compound DAE-Male1.



Figure S44. <sup>1</sup>H NMR spectrum (400 MHz, DMF-d<sub>7</sub>) of compound DAE-Male2.



Figure S45. <sup>13</sup>C NMR spectrum (101 MHz, DMF-d<sub>7</sub>) of compound DAE-Male2.





SI-b\_1H --- 5.015 - 2.475 - 2.461 - 2.447 3.615 3.551 3.551 3.551 3.551 3.551 3.555 3.555 3.555 3.555 3.350 3.310 3.310 3.216 3.231 3.2512 3.2512 3.2512 3.25120 3.2512 3.2512 3.25120 3.2512000000000 684 - 6.301 8.731 8.731 8.717 450 2.475 2.461 2.447 1.443 1.342 1.329 1.312 1.312 283 283 3.527 3.510 3.492 3.336 3.336 3.320 3.310 3.295 3.746 3.248 3.231 3.216 1.640 1.621 1.604 485 SI-b 30 20 250 10 200 2.08 ę. 3.8 3.0 2.8 2.6 2.4 f1 (ppm) 3.6 3.4 3.2 2.2 2.0 1.8 1.6 1.2 1.4 150 100 50 0 0.78 2.08 0.787 3.394 9.534 2.384 12 0.71-2.00 4.32 4.32 5.0 4.5 4.0 f1 (ppm) 7.5 7.0 6.5 2.5 6.0 5.5 3.0 2.0 1.5 1.0 3.5

Figure S48. <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>) of compound SI-b.



Figure S49. <sup>1</sup>H NMR spectrum (400 MHz, CDCl<sub>3</sub>) of compound SI-c.



Figure S50. <sup>1</sup>H NMR spectrum (400 MHz, DMF-d<sub>7</sub>) of compound DAE-Male3.





Figure S52. <sup>19</sup>F NMR spectrum (367 MHz, DMF-d<sub>7</sub>) of compound DAE-Male3.



Compound g\_13C  $< \frac{175.323}{175.238}$   $\sim 172.386$ 30.333 30.124 20.116 26.167 26.167 26.148 24.665 24.665 24.665 24.347 22.023 20.023 12.802 57.679 39.414 36.074 35.057 35.657 35.657 35.657 35.657 35.657 35.657 35.657 35.657 35.657 35.657 35.657 35.657 35.657 35.657 31.167 100 163.44 0.749 - 90 - 50 175.323 175.238 175.238 163.441 163.150 [ 163.150 [ 163.150 [ 163.150 [ 163.592 159.592 159.592 149.102 134,080 139,073 129,937 129,937 129,836 129,640 125,959 129,640 125,595 124,508 124,508 124,508 124,508 122,570 122,57 142.705 142.530 142.530 142.469 140.429 140.313 140.214 140.214 137.450 137.450 137.450 137.450 137.450 137.457 134.677 - 80 40 - 70 30 - 60 20 - 50 10 40 175 170 165 160 155 150 145 140 135 130 125 120 fl (ppm) - 30 - 20 - 10 - 0 170 160 150 130 120 100 90 f1 (ppm) 50 40 180 140 110 10 80 . 70 60 30 20

Figure S54. <sup>13</sup>C NMR spectrum (101 MHz, DMF-d<sub>7</sub>) of compound **f**.



Figure S55. <sup>19</sup>F NMR spectrum (367 MHz, DMF-d<sub>7</sub>) of compound **f**.



Figure S56. <sup>1</sup>H NMR spectrum (400 MHz, DMF-d<sub>7</sub>) of compound DAE-NHS.





Figure S58. <sup>19</sup>F NMR spectrum (367 MHz, DMF-d<sub>7</sub>) of compound DAE-NHS.

## **Supplementary References**

[S1] Iwai, R.; Morimoto, M.; Irie, M., Turn-on mode fluorescent diarylethenes: effect of electrondonating and electron-withdrawing substituents on photoswitching performance. *Photochem. Photobiol. Sci.* **2020**, *19* (6), 783-789.

[S2] Uno, K.; Aktalay, A.; Bossi, M. L.; Irie, M.; Belov, V. N.; Hell, S. W., Turn-on mode diarylethenes for bioconjugation and fluorescence microscopy of cellular structures. *Proc. Natl. Acad. Sci. U. S. A.* 2021, *118*, e2100165118.