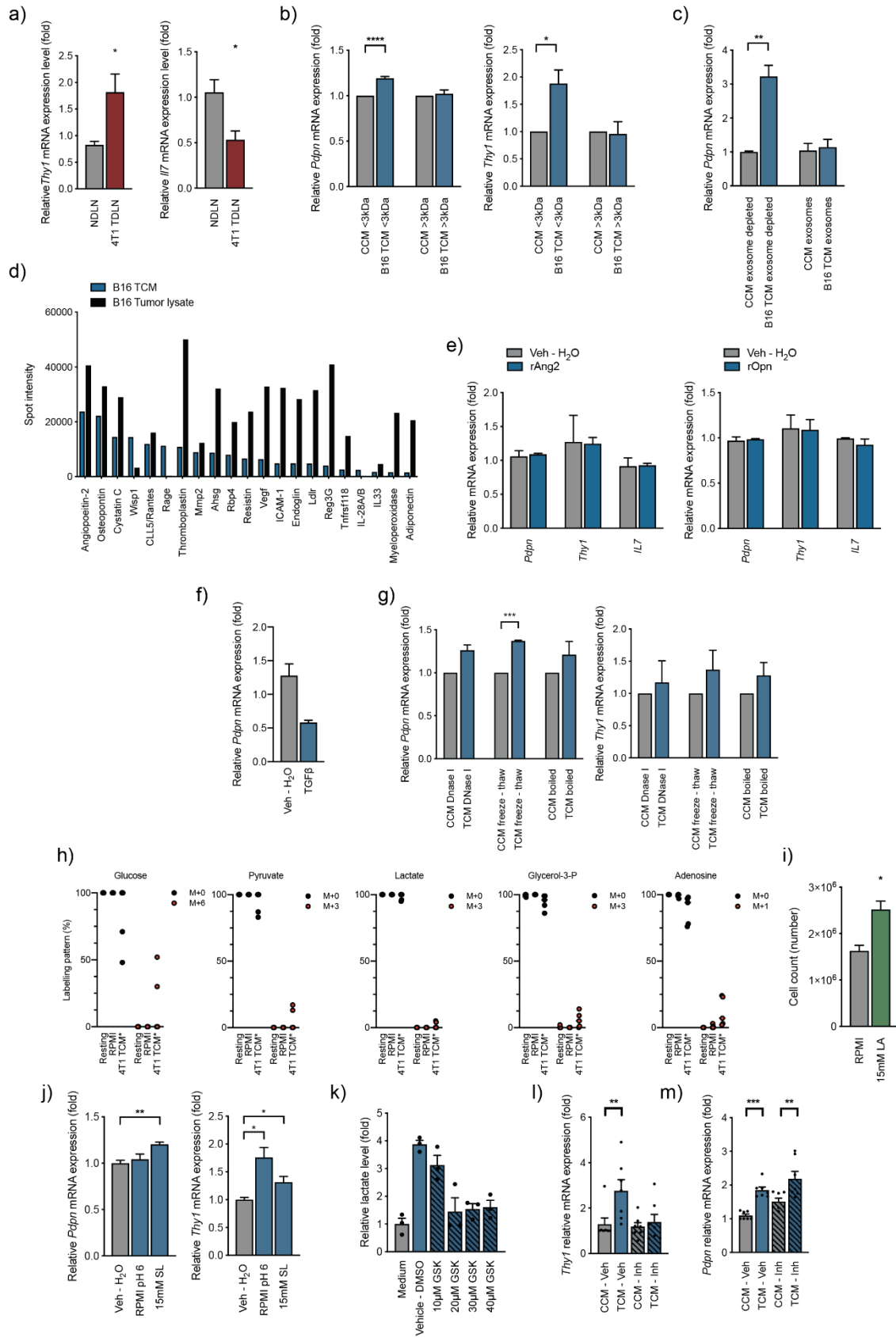


Supp Fig 1



Supplementary figure S1. Factor depletion, sodium lactate and low pH medium controls, and proliferation assay.

(A) Quantitative RT-PCR analysis of *Thy1* (left) and *Ii7* (right) in non-draining lymph nodes (NDLNs) and tumor-draining LNs (TDLNs) from 4T1 orthotopically injected, female Balb/c mice. n = 2 independent biological experiments with 2-3 replicates.

(B) Quantitative RT-PCR analysis of *Pdpm* (left) and *Thy1* (right) in FRCs cultured *in vitro* and treated for 4 days with control conditioned medium (CCM) or B16.F10 tumor conditioned medium (B16 TCM) with either medium at <3kDa or >3kDa. n = 4 independent biological experiments.

(C) Quantitative RT-PCR analysis of *Pdpm* in *in vitro* cultured FRCs treated for 4 days with exosome depleted CCM or TCM or with exosomes isolated from CCM or B16 TCM. n = 2-5 independent biological experiments.

(D) Cytokine array of B16 TCM and B16 tumor lysates showing the top 21 detected cytokines as measured by spot intensity. n = 1

(E) Quantitative RT-PCR analysis of *Pdpm*, *Thy1*, and *Ii7* in *in vitro* cultured FRCs treated for 4 days with 1 µg/ml recombinant murine angiopoietin 2 (rAng2; left) or 5 µg/ml recombinant murine osteopontin (rOPN; right). n = 2 independent biological experiments with 3 replicates.

(F) Quantitative RT-PCR analysis of *Pdpm* in *in vitro* cultured FRCs treated for 4 days with 2 ng/ml TGFβ. n = 3 independent biological experiments.

(G) Quantitative RT-PCR analysis of *Pdpm* (left) and *Thy1* (right) in *in vitro* cultured FRCs treated for 4 days with CCM or TCM subjected to DNase I treatment, freeze-thaw cycles or boiling. n = 2 independent biological experiments.

(H) Labeling pattern of metabolites of whole tissues of brachial LNs. TCM of ¹³C-glucose labeled 4T1 tumor cells (4T1 TCM*) was injected subcutaneously every day for 11 days at the shoulder in lieu of a tumor *in vivo*. Metabolites were measured using LS-MS. 'M+n' is the molecular mass plus the number of incorporated heavy carbons. n = 1 experiment with 5 LNs.

(I) Total LN cells in draining LNs from female C57Bl/6 mice assessed by flow cytometry injected over 11 days subcutaneously each day with 15mM LA in RPMI. Mean with SD. n = 1 experiment with 3 LNs.

(J) Quantitative RT-PCR analysis of *Pdpm* (left) and *Thy1* (right) in FRCs cultured *in vitro* and treated with Vehicle (Veh – H₂O), RPMI at pH 6 or 15mM sodium lactate (SL) for 4 days. n = 3 independent biological experiments.

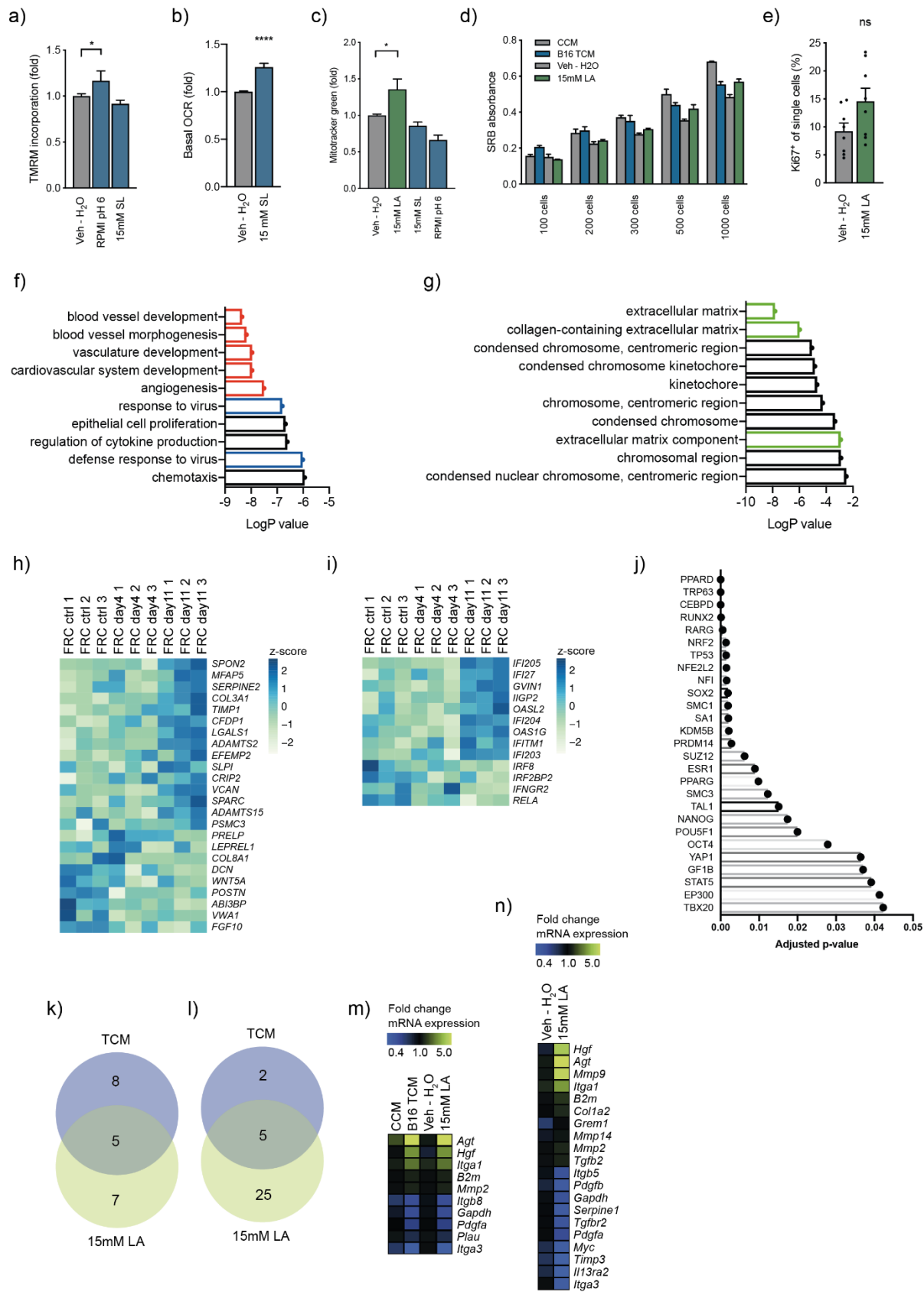
(K) Relative lactate levels in TCM of B16.F10 treated for 24h with a *Ldha*/*Ldhd* inhibitor (GSK 2837808A). n = 3 independent biological experiments.

(L) Quantitative RT-PCR analysis of *Thy1* in FRCs cultured *in vitro* and cultivated with TCM of B16.F10 tumor cells treated with either vehicle (DMSO, Veh) or 40µM Inhibitor (Inh) for 4 days. n = 4 independent biological experiments with 1-2 replicates.

(M) Same as in (L), but Quantitative RT-PCR analysis of *Pdpm*.

All data are means with SEM from biological independent replicates (unless stated differently). Significance (**P*<0.05, ***P*<0.01, ****P*<0.001 and *****P*<0.0001) was determined by unpaired two-tailed *t*-test.

Supp Fig 2



Supplementary figure S2. Transcriptional analyses of FRCs.

(A) *In vitro* FRCs treated for 4 days as in (I), stained with TMRM and analyzed by flow cytometry. n = 3 independent biological experiments with 1-3 replicates.

(B) Baseline OCR from vehicle (Veh – H₂O) and 15mM sodium lactate (SL) treated FRCs. n = 3-4 independent biological experiments with each 5-replicates.

(C) *In vitro* FRCs treated for 4 days with vehicle (Veh – H₂O), 15mM LA, 15mM sodium lactate (SL), or RPMI at pH 6, stained with Mitotracker green and analyzed by flow cytometry. n = 3 independent biological experiments with 1-2 replicates.

(D) SRB proliferation assay of *in vitro* cultured FRCs treated for 4 days with CCM, B16 TCM, vehicle (Veh – H₂O) or 15mM LA and seeded in a 96-well plate at different starting cell numbers. n = 1 with 3 replicates. Mean with SD.

(E) *In vitro* FRCs treated for 4 days with vehicle (Veh – H₂O) or 15mM LA, stained for Ki67 and analyzed by flow cytometry. n = 3 independent biological experiments with 2-3 replicates.

(F) Top 10 most significant gene ontology (GO) terms for 'biological process'. All blood vessel development processes marked in red, all response to virus in blue.

(G) Top 10 most significant gene ontology (GO) terms for 'cellular component'. All extracellular matrix processes marked in green.

(H) Extracellular matrix genes deregulated in *ex vivo* sorted FRCs of tumor-draining lymph nodes (TDLNs; Riedel et al., 2016)

(I) Interferon related genes deregulated in *ex vivo* sorted FRCs of TDLNs (Riedel et al., 2016)

(J) Transcription factor (TF) analysis displaying TFs with adjusted $P < 0.05$ (Enrichr, ChEA 2016) using all up and down regulated genes in 15mM LA to Veh – H₂O.

(K) Overlap of genes significantly ($P < 0.05$) upregulated covered by the PCR array (PAMM-120ZG, Mouse Fibrosis, QIAGEN) in comparisons between B16 TCM to CCM or 15mM LA to vehicle (Veh – H₂O). FRCs were treated for 4 days and each sample was run 5 times from 5 independent biological replicates.

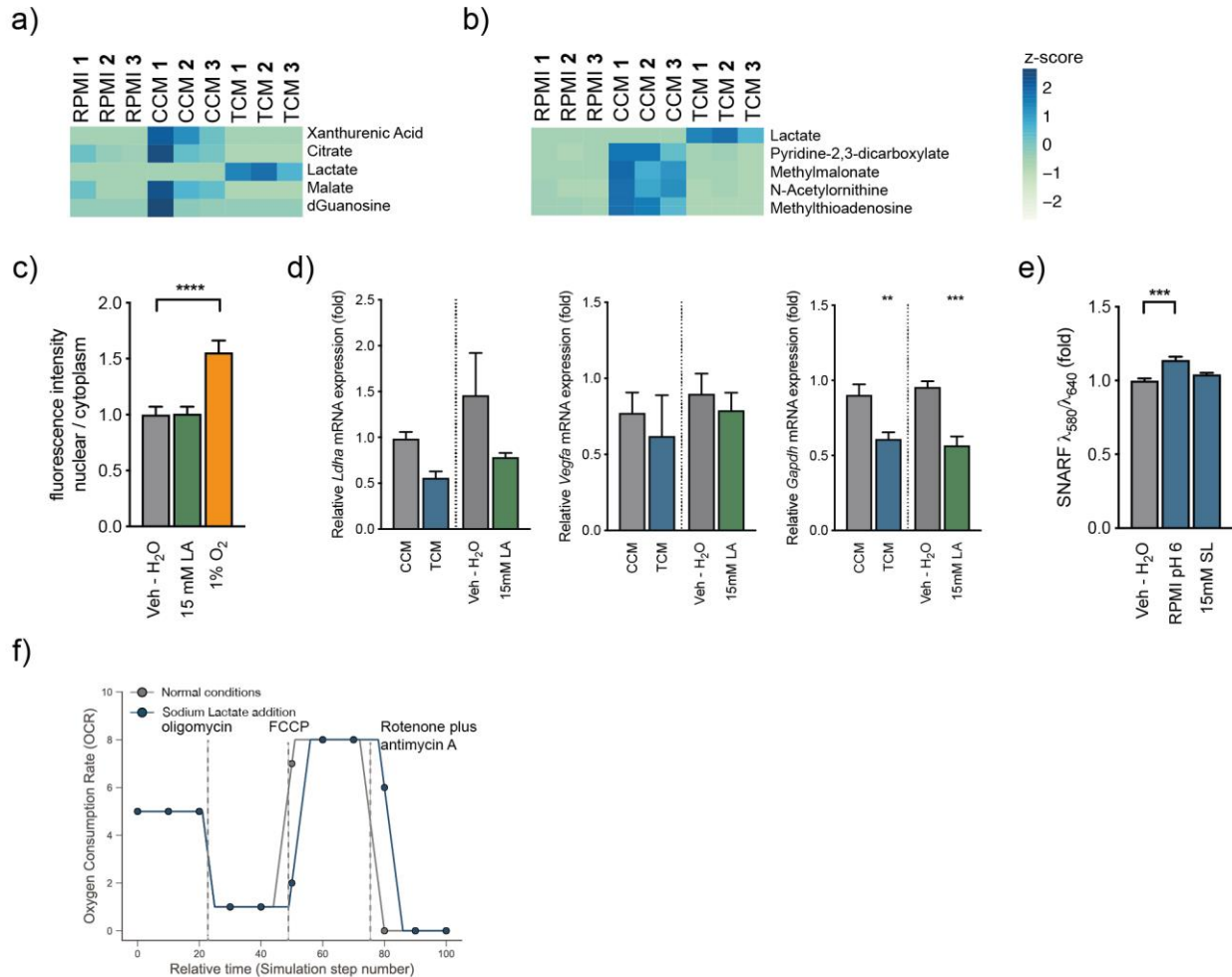
(L) Overlap of genes significantly ($P < 0.05$) downregulated in comparisons as in (A).

(M) Heatmap depicting the 10 commonly deregulated genes between B16 TCM to CCM and 15mM LA to vehicle (Veh – H₂O) as average of fold changes of mRNA expression.

(N) Heatmap depicting the top 10 up- and downregulated genes between 15mM LA to vehicle (Veh – H₂O) as average of fold changes of mRNA expression.

All data are means with SEM from biological independent replicates (unless stated differently). Significance ($*P < 0.05$, $**P < 0.01$, $***P < 0.001$ and $****P < 0.0001$) was determined by unpaired two-tailed *t*-test.

Supp Fig 3



Supplementary figure S3. pH regulates metabolic shift.

(A) Metabolites in same order as in (Fig 4c), but measured in the respective conditioned medium (CCM or TCM) measured by LC-MS.

(B) Significant ($P < 0.05$) metabolites in TCM vs CCM measured by LC-MS.

(C) Quantification of confocal fluorescent images stained for Hif1a and nuclei. Fluorescent intensity was calculated as nuclear / cytoplasm and was normalized on cell number. $n = 3$ independent biological experiments with each 8-10 fields of view.

(D) Quantitative RT-PCR analysis of *Ldha* (left), *Gapdh* (middle) and *Vegfa* (right) in *in vitro* cultured FRCs treated for 4 days with CCM, B16 TCM, vehicle (Veh - H₂O) or 15mM LA. $n = 2-5$ independent biological experiments.

(E) Intracellular pH of FRCs treated for 4 days with vehicle (Veh - H₂O), RPMI at a pH of 6 (RPMI pH 6), or 15mM sodium lactate (SL) and stained with SNARF. $n = 2-6$ independent biological experiments.

(F) Metabolic analysis of mitochondrial function in the computational model. Shown is the OCR for mitochondria exposed to sodium lactate compared to control cells.

All data are means with SEM from biological independent replicates (unless stated differently). Significance ($*P < 0.05$, $**P < 0.01$, $***P < 0.001$ and $****P < 0.0001$) was determined by unpaired two-tailed *t*-test.