

<u>Supplementary Figure 1.</u> *iRegulon (Janky et al. 2014) predicts the key regulons for the SASP cells.* (a) The motif with the highest enrichment for the SASP genes, according to iRegulon, was Factorbook-NFKB1. (b) The most important transcription factors with 92, 28 and 34 targets within SenMayo were BCL3, RXRA and NFIC, respectively. The first transcription factor controlled (c) BCL3, NFKB1 and -2, RELA and IKZF1, thus confirming the RNA-Seq predictions in the young and old dataset (Fig. 1B). (d) The three transcription factors control a majority (95/125) of SenMayo genes. Source data are provided as a Source Data file.

а



b

<u>Supplementary Figure 2.</u> The SenMayo gene set predicts aging in two mRNA-seq data sets. Out of the 50 available genes in the R-HSA-2559582 gene set, two were significantly enriched in the aging cohort (a), while 13 out of 125 of the SenMayo genes were enriched (b). Canonical markers of the SASP such as *CCL24*, *SEMA3F*, *FGF2*, and *IGFBP7* were upregulated with aging in the RNA-seq of human bone/bone marrow samples in (c) cohort A ((CCL24: and IGFBP7 two-sided unpaired t-test, SEMA3F and FGF2: Kolmogorov-Smirnov test, *CCL24*: p=0.062, *SEMA3F*: p=0.001, *FGF2*: p=0.0034, *IGFBP7*: p=0.0022), and (d) cohort B (two-sided unpaired t-test, *CCL24*: p=0.060, *SEMA3F*: p<0.0001, *FGF2*: p=0.0028, *IGFBP7*: p=0.0001). Moreover, the senescence markers, *CDKN1A/p21^{CIP1}* and SEMA3F, correlate with each other in cohort A (spearman-correlation, p<0.0001, e) and cohort B (spearman-correlation, p=0.0148, f), demonstrating a potential circumvention of high interindividual variability by combining more than one gene. *p<0.05, **p<0.01, ***p<0.001, ****p<0.0001. Cohort A (blue): n=38 (19 young, 19 old, all female), Cohort B (purple): n=30 (15 young, 15 old, all female). Depicted are mean ± SEM. Source data are provided as a Source Data file.



<u>Supplementary Figure 3.</u> The senescent phenotype of SASP cells in human hematopoietic bone marrow. (a) Dotplot depicting the canonical marker genes per cluster along with the reference for each marker and cell type. (b) 46% of CD16⁺ monocytic cells had (high) SASP expression pattern; (spearman-correlation, p<0.0001, c) Bivariate correlation plots with Spearman correlation for genAge (R=0.43) and (spearman-correlation, p<0.0001, d) CellAge (R=0.35) show reliable correlations with SenMayo; e) The SASP cell cluster displayed a shift towards the G1 cell cycle phase, suggesting reduced replicative potential (Chi-square test, p<0.0001), f) The enriched terms of the SASP cluster, depicted in a Manhattan plot, show the high expression of cell cycle arrest (GO: 0007050), apoptosis (WP: WP254), and negative proliferation patterns (GO: 0008285) (multiple t-test with Benjamini-Hochberg adjustment); (g) SASP cells emerged in the final phases of cellular differentiation and increased *MIF* expression (yellow on the left, late phase) at their late developmental phase as revealed by pseudotime analysis. ****p<0.0001, n= 22 (10 male, 12 female). Source data are provided as a



<u>Supplementary Figure 4.</u> *MIF and PECAM pathways in human hematopoietic bone marrow cell types.* (a) The MIF pathway and its key members show a highly heterogeneous expression pattern among all cell clusters. While CD10⁺ B cells show a high expression of *MIF*, *CD74* and *CXCR4*, the expression of *CD44* is low. An overall high expression of all MIF members is evident in CD8⁺ effector T cells and conventional dendritic cells and SASP cells. (b) The PECAM pathway shows an expression of PECAM1 in CD16⁺ monocytes, plasma cells and SASP cells. Source data are provided as a Source Data file.



Supplementary Figure 5. Communication patterns of the SASP cluster in human hematopoietic bone marrow cells. (a) The SASP cluster has an overall high outgoing and moderate incoming interaction strength; (b) The SASP cells exerted various signaling functions (sender, receiver, mediator, and influencer) in the MIF pathway and (c) the PECAM1 pathway, which was used mostly by CD16+ monocytes, plasma cells, and SASP cells (color-code in D); (d) The outgoing signaling pattern revealed the relevance of the MIF pathway among all other pathways, while the relative incoming signaling pattern was likewise substantial; (e) A direct MIF-driven interaction (*via* CD74/CD44 or CD74/CXCR4) from the SASP cells was detected among the majority of other cell types, especially plasmacytoid dendritic cells and B cells, while the PECAM1 pathway mostly targeted the abovementioned three cell types. *p*-values computed from one-sided permutation test, n= 22 (10 male, 12 female). Source data are provided as a Source Data file.





Supplementary Figure 6. Key marker genes and correlation plots for genAge and CellAge in the murine dataset. (a) A dotplot indicating cell cluster marker genes along with the references for each cell type. (b) A bivariate correlation plots with Spearman correlation indicate the reliable correlation of SenMayo with genAge (Spearman-correlation, p<0.0001, R=0.61) and (c) CellAge (Spearmancorrelation, p<0.0001, R=0.67). Source data are provided as a Source Data file.



<u>Supplementary Figure 7.</u> Murine SASP cells in mesenchymal cells from bone and bone marrow are mainly of osteolineage origin and communicate via MIF. (a) SASP cells were mostly recruited from osteolineage cells (OLC), and leptin-positive (Lep⁺) mesenchymal stem cells (MSCs), while (b) 24% of OLC 1 and 18% of OLC2 cells were SASP cell members; (c) Mesenchymal SASP cells in murine bone and bone marrow significantly changed their replicative state from G2M to G1, indicating a replicative stop (Chi-square test, p<0.0001); (d) A Manhattar plot depicts an enrichment of genes associated with cellular senescence (KEGG 04218), negative regulation of proliferation (GO0008285), and cytokine-receptor interaction (KEGG 04060) within the SASP cluster (multiple t-test with Benjamini-Hochberg adjustment). (e) SASP cells function as both senders and influencers within the *MIF* network, and mostly as influencers in the PECAM1 network; (f) The outgoing interaction strength of the SASP cells was high, while they simultaneously showed a substantial incoming signaling strength; (g) Direct cell-cell interactions in the *MIF* pathway from the SASP cells is predominantly directed to hypertrophic chondrocytes, chondrocytic progenitors, and mineralizing osteocytes, while the Pecam1 pathway is directed to chondrocytes, endothelial cells, mast cells, and the SASP cells themselves. *p*-values computed from one-sided permutation test. ****p<0.0001, n= 8 (4 bone, 4 bone marrow, all male). Source data are provided as a Source Data file.



Supplementary Figure 8. SCENIC (Aibar et al. 2017) predicts the key regulons for the SASP cells within the human single cell dataset. (a) A regulon-based tSNE is constructed, where the SASP cells contribute substantially to the upper-middle continent (purple). (b) The predicted regulons, BCL3 and RXRA, indeed control the SASP cells containing continent. Source data are provided as a Source Data file.



<u>Supplementary Figure 9.</u> Trajectory interference using velocity (La Manno et al. 2018). (a) The overall trajectory interference shows that SASP cells are mostly developing from OLC1, OLC2 and Lepr⁺ MSCs. (b) A focus on these four cell types reveals the OLC1 and Lepr⁺ MSCs as main origin of the upper-left continent, and the OLC2 and Lepr⁺ MSCs as the origin of a different SASP cell population in the bottom-right continent. Source data are provided as a Source Data file.



<u>Supplementary Figure 10.</u> The murine SASP cluster showed an enrichment of senescent pathways and contained distinct expression modules. (a) Within the SASP cluster, different expression modules were mathematically predicted. In this module, *Pappa* and *Fgf7* are present, which can be visualized spatially in (b) tSNE, having a similar kernel-weighed density. Other predicted co-expressional patterns were demonstrated by the pairs *Dkk1-Cdkn2a* and *Bmp2-Cdkn1a*. n= 8 (4 bone, 4 bone marrow, all male). Source data are provided as a Source Data file.

Supplementary References

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