

1 **Supporting Information**

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3 **⁶⁸Ga-labeled maleimide for blood pool and lymph PET**
4 **imaging through covalent bonding to serum albumin *in vivo***

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6 *Lixia Feng¹, Jianyang Fang¹, Xinying Zeng¹, Huanhuan Liu¹, Jingru Zhang¹, Lumei*
7 *Huang¹, Zhide Guo^{1*}, Rongqiang Zhuang^{1*}, Xianzhong Zhang^{1*}*

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9 ¹State Key Laboratory of Molecular Vaccinology and Molecular Diagnostics & Center
10 for Molecular Imaging and Translational Medicine, School of Public Health, Xiamen
11 University, 4221-116 Xiang'An South Road, Xiamen 361102, China

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13 ***Correspondence to:**

14 Xianzhong Zhang, E-mail: zhangxzh@xmu.edu.cn;

15 Rongqiang Zhuang, E-mail: zhrq@xmu.edu.cn;

16 Zhide Guo, E-mail: gzd666888@xmu.edu.cn

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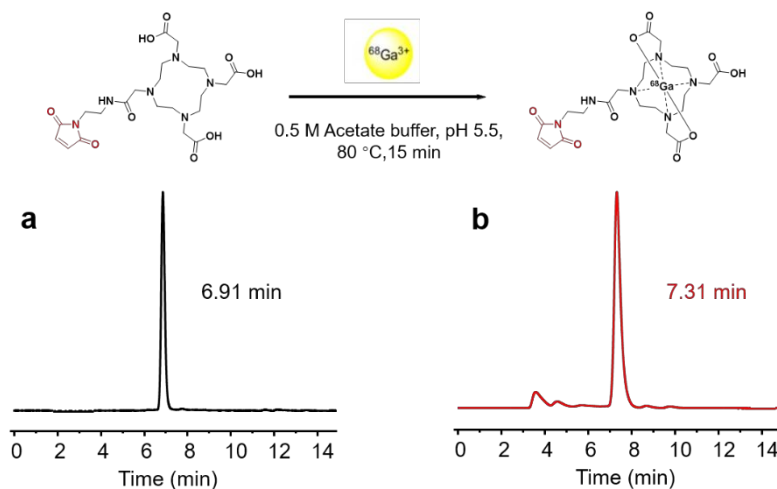
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1 **1. Methods of HPLC**

2 As the UV retention time of the precursor was very close to that of the labeled
3 product, High-Performance Liquid Chromatography (HPLC) conditions and the
4 retention time of the precursor were explored to distinguish the radioactive product
5 from others.

6 **Method 1:** mobile phases were acetonitrile with 0.1% trifluoroacetic acid (A) and
7 water with 0.1% trifluoroacetic acid (B), 5% A, 95% B for 0-10 min, 10% A, 90% B
8 for 10-20 min, flow rate:1 mL/min, UV monitoring wavelength = 220 nm. Hypersil
9 GOLD C18 selectivity HPLC column (25005-254630, 250×4.6 mm, 5 μm, Thermo
10 Scientific, USA) was used.

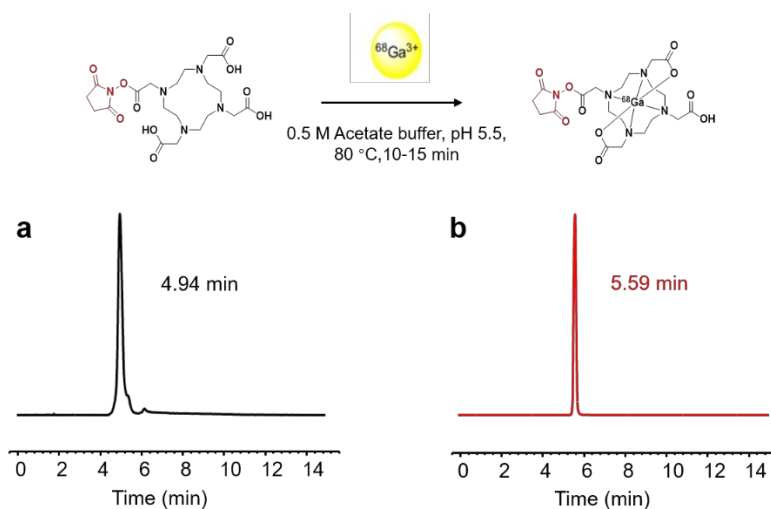
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13 **Figure S1. (a) UV-HPLC analysis of precursor DM and (b) radio-HPLC analysis**
14 **of $[^{68}\text{Ga}]\text{Ga-DM}$**

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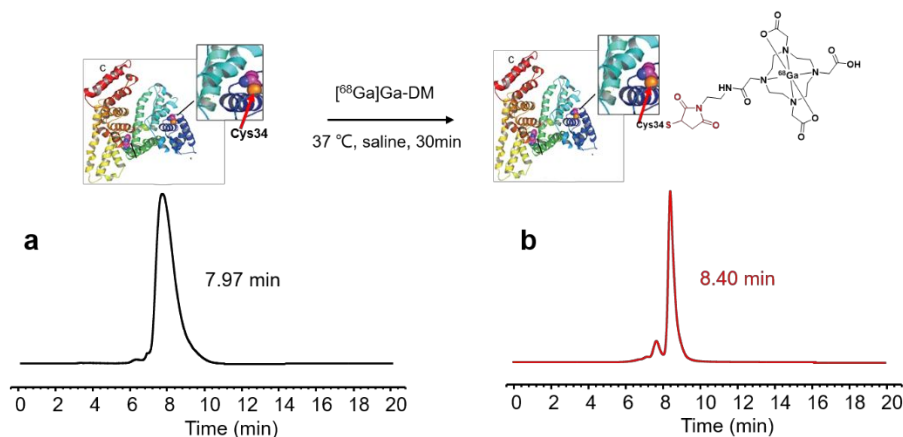
2 **Figure S2. (a) UV-HPLC analysis of precursor DOTA-NHS and (b) radio-HPLC**
 3 **analysis of $[^{68}\text{Ga}]\text{Ga-DOTA-NHS}$**

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5 About 37 MBq/50 μL of purified tracer $[^{68}\text{Ga}]\text{Ga-DM}$ was added to the albumin
 6 solution (HSA & BSA), co-incubated at 37 °C for 30 min, and transferred to an
 7 ultrafiltration centrifuge tube of 0.5 mL/30 kDa for separation and purification,
 8 centrifuged at 14,000 rpm, 4 °C for 15 min to obtain an albumin-bound tracer.

9 **Method 2:** mobile phase was 1 \times PBS solution with a pH of 7.4 (C), 100% C for
 10 20 min, flow rate:1 mL/min, and UV monitoring wavelength = 278 nm. Acclaim SEC-
 11 300 size exclusion chromatography HPLC column (0797233, 300 \times 4.6 mm, 5 μm ,
 12 Thermo Scientific, USA) was used.

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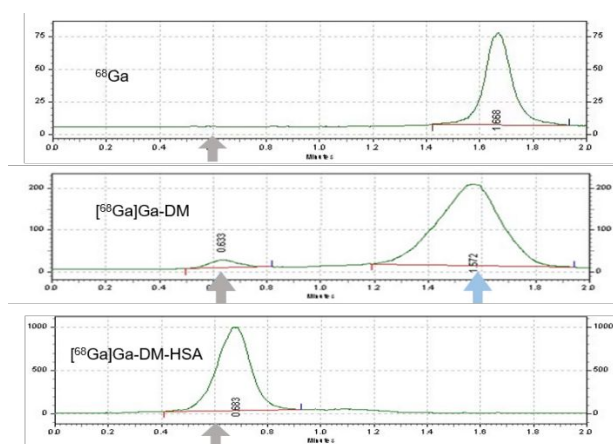


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2 **Figure S3. (a) UV-HPLC analysis of precursor HSA and (b) radio-HPLC**
3 **analysis of [⁶⁸Ga]Ga-DM-HSA**

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5 **2. Methods of iTLC**

6 The R_f value of the albumin-bound tracer was 0–0.1, which was lower than the
7 unbound [⁶⁸Ga]Ga-DM (R_f value was 0.7–0.9) and the free ⁶⁸Ga ion (R_f value was 0.9–
8 1.0), so, the iTLC method was established to judge the stability of the tracer binding
9 with albumin in saline and serum.

10 **Method 3:** the samples were loaded on an iTLC-SG strip (5×70mm) and 0.1 M
11 sodium citrate solution (pH = 5.0) as the developing agent. The iTLC-SG strip was
12 detected by a MiniScanTLC scanner.



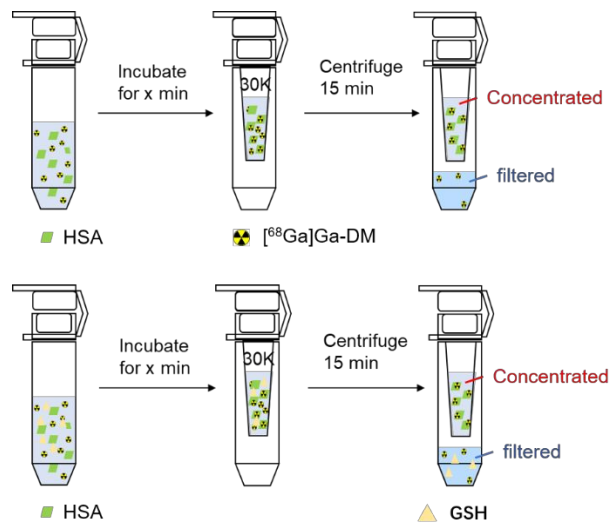
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15 **Figure S4. iTLC analysis of ⁶⁸Ga, [⁶⁸Ga]Ga-DM, and [⁶⁸Ga]Ga-DM-HSA**

1 **3. Methods of purification**

2 **Method 4:** Tracer was co-incubated with HSA for several minutes. Samples were
3 transferred to an ultrafiltration centrifuge tube of 0.5 mL/30 kDa for separation and
4 purification, centrifuged at 14,000 rpm, 4 °C for 15 min to obtain the concentrated
5 solution, and filtered solution represented the albumin-bound tracers, and unbound
6 tracers, respectively. The radioactivity counts of the two parts were detected by the γ -
7 counter and the radioactivity bound fraction was calculated as follows:

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$$\text{Bound fraction} = \frac{\text{CPM (concentrated)}}{\text{CPM (concentrated)} + \text{CPM (filtered)}}$$

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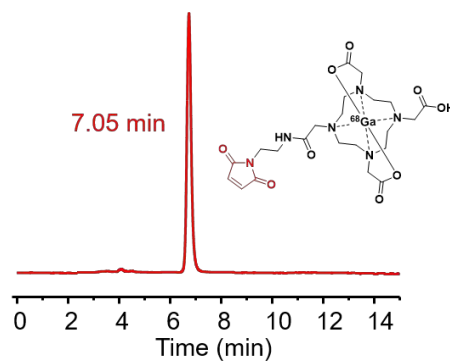
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Figure S5. Diagram of purification

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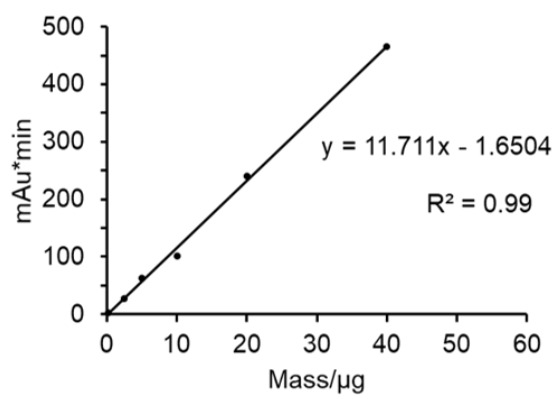


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Figure S6. Radio-HPLC analysis of [⁶⁸Ga]Ga-DM injection

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Figure S7. Ultraviolet standard curve for DM with a detection wavelength of 220

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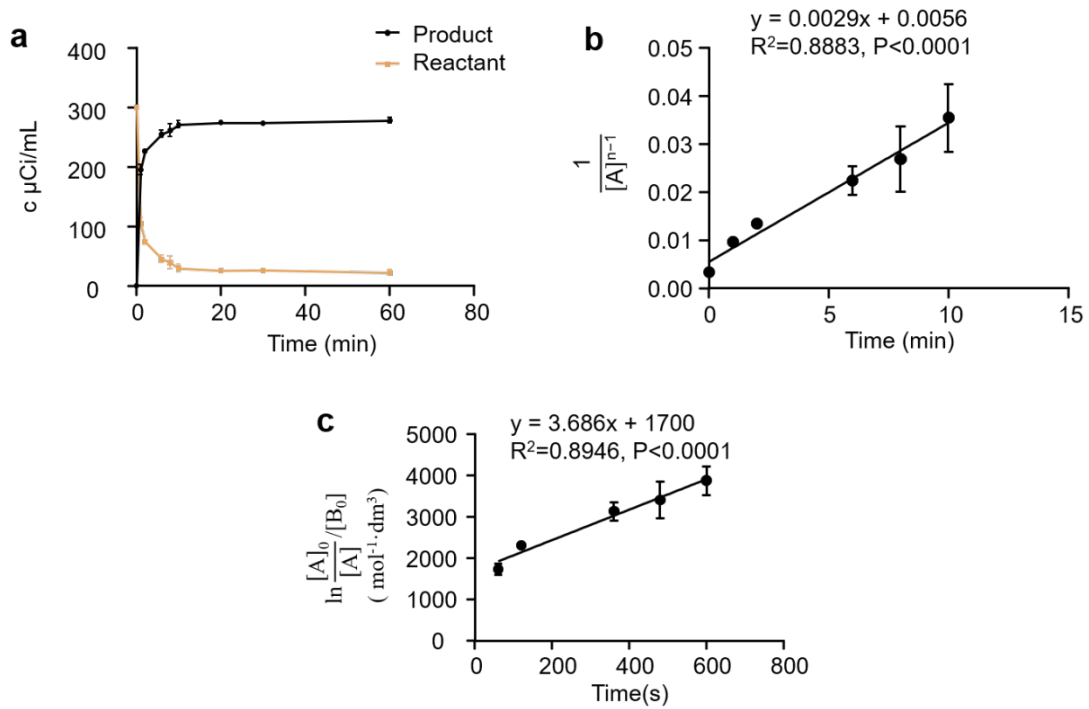
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1 **4. Calculation of the covalent binding reaction rate**

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4 **Figure S8. (a) Curves of reactant and product concentrations over time; (b)**
 5 **$\frac{1}{[A]^{n-1}}$ -time diagram when the reaction order is 2; (c) $\ln \frac{[A]_0}{[B]_0} / [A]$ -time chart**

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7 Consider the second-order reaction:



11 The dose of reactant B is much higher than that of reactant A, so, $[B] \approx [B]^0$, the rate
 12 equation corresponds to:

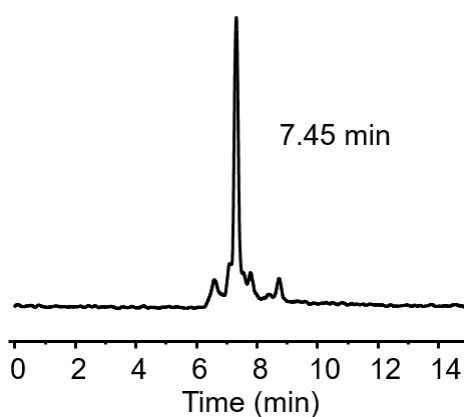
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$$k = \frac{1}{t} \ln \frac{[A]^0}{[A]} / [B]^0 \quad \text{mol}^{-1} \cdot \text{dm}^3 \cdot \text{s}^{-1}$$

14 Serve $\ln \frac{[A]^0}{[A]} / [B]^0$ as y and t as x to obtain the linear ship as shown in Figure S8c,

15 therefore, the covalent binding reaction rate K is about $3.69 \text{ mol}^{-1} \cdot \text{dm}^3 \cdot \text{s}^{-1}$.

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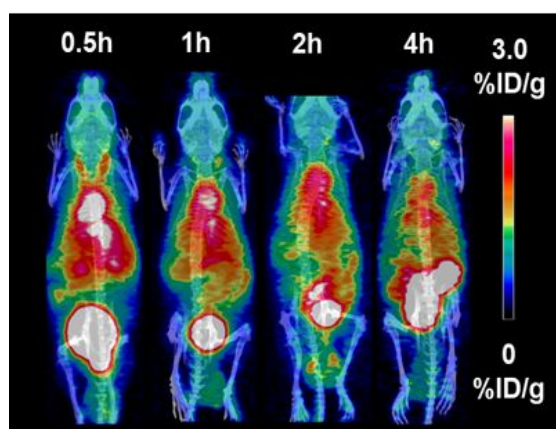
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3 **Figure S9. Radio-HPLC analysis of urine at 5 min after administration**

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6 **Figure S10. Representative PET-CT images of $[^{68}\text{Ga}]\text{Ga-DM-BSA}$**

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Table S1. Uptake ratio of blood/organs at selected time points

Time (min)	Blood /organ (Mean \pm SEM, n=4)			
	Heart	Liver	Lung	Kidney
2	2.72 \pm 0.15	3.77 \pm 0.10	1.16 \pm 0.09	0.50 \pm 0.10
5	3.72 \pm 0.23	4.67 \pm 0.28	1.72 \pm 0.07	0.97 \pm 0.27
30	4.28 \pm 0.13	3.84 \pm 0.67	1.78 \pm 0.12	1.54 \pm 0.32
60	2.64 \pm 0.68	3.22 \pm 0.78	1.51 \pm 0.42	1.63 \pm 0.44
120	3.16 \pm 0.38	3.70 \pm 0.32	1.65 \pm 0.17	1.97 \pm 0.12
180	3.35 \pm 0.21	3.19 \pm 0.23	1.88 \pm 0.17	1.71 \pm 0.03 *

* n=3

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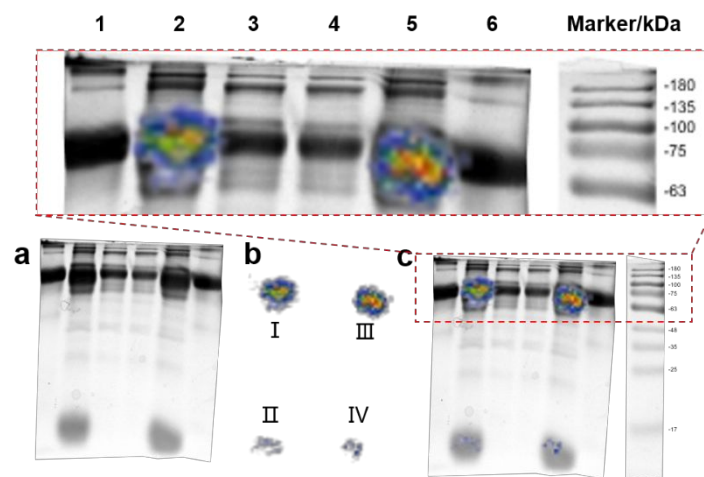
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1 **5. Gel electrophoresis and autoradiography analysis of [⁶⁸Ga]Ga-DM cultured**
2 **plasma**

3 Plasma protein concentration was measured at 79 mg/mL via bicinchoninic acid
4 assay (BCA)¹, 4.9 MBq of [⁶⁸Ga]Ga-DM was incubated with mouse whole blood with
5 anticoagulant for 30 min, then, centrifuged at 14000 rpm for 10 min to obtain a clear
6 plasma sample. The sample was diluted 20 times, then, 10 μL of it was taken for protein
7 electrophoresis, and commercially obtained BSA and mouse serum protein were used
8 as controls. After electrophoresis, the radioactive signal was localized by
9 autoradiography of the gel. Gel electrophoresis was carried out at 80 V for 30 min, and
10 120 V for 1.5 h. Exposure duration was 30 min for autoradiography.

11 The results of gel electrophoresis were shown in **Figure S11a**, each protein can be
12 well separated. Albumin in plasma samples can be localized by the electrophoretic band
13 of BSA (1, 6), where the molecular weight was between 75 kDa and 63 kDa. Compared
14 to the electrophoretic band of mouse serum protein (3, 4), [⁶⁸Ga]Ga-DM co-incubation
15 with blood did not change the properties of plasma proteins. The results of gel
16 autoradiography were shown in **Figure S11b**. There were only two radioactive signals,
17 one with molecular weights between 75 kDa and 63 kDa (I, III), and another with
18 molecular weights less than 17 kDa (II, IV). The quantitative analysis data were shown
19 in **Table S2**. Fusion of coomassie brilliant blue staining and autoradiography (**Figure**
20 **S11c**) showed that 75% of the agent was bound to albumin.

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Figure S11. (a) Coomassie brilliant blue staining after gel electrophoresis; (b) Autoradiography after gel electrophoresis; (c) Merge of coomassie brilliant blue staining and autoradiography. Lanes 1, 6 represent commercially obtained BSA; Lanes 2, 5 represent diluted plasma samples; Lanes 3,4 represent mouse serum protein.

Table S2. Quantitative analysis data of gel autoradiography

No.	DLU /mm2	Percentage (%)
□	61622.83 ± 2755.40	75.16 ± 1.08
□	20356.20 ± 861.71	24.84 ± 1.08
□	76008.97 ± 3949.14	75.73 ± 0.36
□	24347.23 ± 841.43	24.27 ± 0.36

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References

(1) Arslan A, Duman H, Kaplan M, Uzkuç H, Bayraktar A, Ertürk M, Alkan M, Frese SA, Duar RM, Henrick BM, Karav S. Determining Total Protein and Bioactive Protein Concentrations in Bovine Colostrum. J Vis Exp. 2021 Dec 10;(178). doi: 10.3791/63001.