Structural insights into human PA28-20S proteasome enabled by efficient tagging

and purification of endogenous proteins

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Supplementary Figures

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ML557 TAATACGACTCACTATAG -> **T7** Gene Specific **Constant Stem** Promoter Sequence Loop Region В PSMB2 (B4) PSMB7 (B2) PSMB6 (B1) Control Cells PSMB2 KI PSMB6 KI PSMB7 KI No Knock-In mNG211-StrepII mNG211-StrepII mNG211-StrepII 293F Suspension Cells mNG2 NG2 FSC-A ssc-A FSC-H 10 mNG2+ All Events Cell Events PSME1 (PA28a) С PSME1 KI Control Cells sfCherry₂11-ALFA PSMB2 mNG211 KI in PSMB2 KI Line 293F Suspension Cells with PSMB2 mNG211 KI Cherry FSC-A FSC-A

Supplementary Figure 1. gRNA IVT Template Design and FACS Sorting of KIs. (A) PCR schematic for the IVT template to make Cas9 gRNA. The gene specific primers (provided in the Methods section) anneal to ML611 containing the constant stem loop region. ML557 and ML558 are used to amplify the gene-specific template created by the gene-specific oligo and ML611. (B) FACS sorting of PSMB knock-in Expi293F cells. Shown are control cells used to set gating and PSMB2, PSMB6, and PSMB7 KIs containing the mNG211-StrepII donor. Cells in the gate were collected for downstream structural studies. (C) FACS sorting of PSME1 knock-in. This knock-in was performed in the sorted PSMB2 KI cell line. Shown are control cells used to set gating and PSME1 knock-in was performed in the softed results.

FSC-H

All Events

Cell Events

sfCherrv2+



Supplementary Figure 2. Negative stain EM of β 4-tagged proteasomal complexes. 2D classification of the images show a diverse population of different proteasomal complexes formed between 20S, 19S, PA28, and PA200. Scale bar, 50 Å.





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ALFA-PSME1





Supplementary Figure 3. Cryo-EM image processing workflow. 2D and 3D classification for PSMB2-StrepII, ALFA-PSME1, and PSMB2-StrepII + MG132 samples.





Supplementary Figure 4. Cryo-EM map statistics. (**A**) Fourier shell correlation (FSC) and 1D directional FSC (dFSC) plots. (**B**) Visualization of 3D dFSC. (**C**) Plots showing coverage of Fourier space. (**D**) Plots showing distribution of viewing angles from final refined datasets. (**E**) Corresponding local resolution map with color scale.



Supplementary Figure 5. Mass spectrometry (left) of $20S(\beta4-StrepII)$ pull-down shows co-purification with PA28 α (PSME1), PA28 β (PSME2), and PA28 γ (PSME3). Cryo-EM density map of PA28-20S(β 4-StrepII) was modeled with both PA28 $\alpha\beta$ and PA28 γ (right). The density fits PA28 γ sidechains in some regions (F34, L29) while PA28 β sidechains are better fit in other regions (Y33, H192, H218), indicating that the map is an average of PA28 $\alpha\beta$ and PA28 γ .







Supplementary Figure 6. PA28 model fit to map density. For each chain in PA28, PA28a (PSME1) and PA28b (PSME2) were modeled into the cryo-EM density map of single capped PA28-20S proteasome. Key differences in the model-to-map fit are highlighted and support a stoichiometry of {a,b,a,b,a,b,b} for chains {c,d,e,f,g,h,i}, respectively. Scale bar, 1 Angstrom.



Supplementary Figure 7. (**A**) Cryo-EM map of double capped PA28-20S-PA28 shows ALFA peptide density in both antechambers. (**B**) The walls of the 20S proteasome inner chambers are made up of both hydrophilic and hydrophobic regions.



Supplementary Figure 8. Fit of MG132 into the PA28-20S cryo-EM density at the proteolytic sites of β 1, β 2, and β 5.



Supplementary Figure 9. PA28 C-terminal tail model and map density. Density is observed for the C-terminal tails of PA28 subunits for 6 out of 7 binding sites.

Supplementary Table 1. Cryo-EM imaging parameters

PSMB2-StrepII pulldown			
Microscope	Polara (300 kV), UCSF		
Camera (software)	Gatan K2 Summit (SerialEM, 1 shot/hole)		
Magnification (at detector)	40.984X		
Exposure rate (total)	8 e/pixel/s (43 e/Å^2)		
Frames	30		
Pixel size	1.22 Å/pixel		
Defocus range (um)	1.2 - 3.1		
Micrographs (# initial particles)	3761 (1,912,794)		
# of 3D classes (# particles)	3 (550,396)		
Class 1 (# particles), codes	20S (499,629), EMD-24275, PDB 7NAN		
Resolution (0.143/0.5)	2.8/3.8 A (mask), 3.5/4.1 A (no mask)		
Local resolution (min-max)	2.7 – 29 Å		
3DFSC spread (0.143)	2.5 – 2.8A		
Sharpening B-factor	0		
Class 2 (# particles), codes	PA200-20S (50,767), EMD-24278, PDB 7NAQ		
Resolution (0.143/0.5)	3.2/4.4 A (mask), 4.2/7.6 A (no mask)		
Local resolution (min-max)	2.8 – 50 A		
3DFSC spread (0.143)	3.0 – 3.6 A		
Sharpening B-factor	0		
Class 3 (# particles)	19S-20S (did not refine)		
ALFA-PSME1 pulldown			
Microscope	Polara (300 kV), UCSF		
Camera (software)	Gatan K2 Summit (SerialEM, 1 shot/hole)		
Magnification (at detector)	40,984X		
Exposure rate (total)	8 e/pixel/s (43 e/Å^2)		
Frames	30		
Pixel size	1.22 Å/pixel		
Defocus range (um)	1.3 – 2.0		
Micrographs (# initial particles)	5994 (2,540,092)		
# of 3D classes (# particles)	2 (158,883)		
Class 1 (# particles), codes	PA28-20S (135,937), EMD-24276, PDB 7NAO		
Resolution (0.143/0.5)	2.9/3.9 A (mask), 3.8/5.3 A (no mask)		
Local resolution (min-max)	2.7 – 43 A		
3DFSC spread (0.143)	2.6 – 3.2 A		
Sharpening B-factor	0		
Class 2 (# particles), codes	PA28-20S-PA28 (22,946), EMD-24277, PDB 7NAP		
Resolution (0.143/0.5)	3.2/4.5 A (mask), 6.1/8.5 A (no mask)		
Local resolution (min-max)	2.7 – 52 A		
3DFSC spread (0.143)	3.0 – 3.4 A		
Sharpening B-factor	0		
PSMB2-StrepII pulldown + MG132			
Microscope	Titan Krios (300 kV), SBP		
Camera (software)	Gatan K3 (SerialEM, 4-shots/hole)		

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Magnification (at detector)	47,170X		
Exposure rate (total)	25 e/pixel/s (30 e/Å^2)		
Frames	20		
Pixel size	1.06 Å/pixel		
Defocus range (um)	1.3 – 2.3		
Micrographs (# initial particles)	8128 (3.537.033)		
# of 3D classes	7 (729,026)		
Class 1 (# particles), codes	20S (214,790), EMD-27013, PDB 8CVR		
Resolution (0.143/0.5)	2.7/3.1 A (masked), 3.3/4.1 A (unmasked)		
Local resolution (min-max)	2.5 – 45 A		
3DFSC spread (0.143)	2.5 – 2.8 A		
Sharpening B-factor	0		
Class 2 (# particles), codes	PA200-20S (70,412), EMD-27015, PDB 8CVS		
Resolution (0.143/0.5)	3.1/4.3 A (masked), 4.1/8.0 (unmasked)		
Local resolution (min-max)	2.4 – 40 A		
3DFSC spread (0.143)	2.2 – 3.6 A		
Sharpening B-factor	0		
Class 3 (# particles), codes	PA28-20S (136,392), EMD-27014		
Resolution (0.143/0.5)	2.8/3.4 A (masked), 4.1/7.8 A (unmasked)		
Local resolution (min-max)	2.6 – 45 A		
3DFSC spread (0.143)	2.3 – 3.0 A		
Sharpening B-factor	0		
Class 4 (# particles), codes	19S-20S, S _A /E _A (65,390), EMD-27016		
Resolution (0.143/0.5)	3.3/4.1 A (masked), 7.1/9.4 A (unmasked)		
Local resolution (min-max)	3.1 – 57 A		
3DFSC spread (0.143)	3.0 – 4.0 A		
Sharpening B-factor	0		
Class 5 (# particles), codes	19S-20S, S _{D1} (45,734), EMD-27017		
Resolution (0.143/0.5)	3.4/4.2 A (masked), 7.3/11 A (unmasked)		
Local resolution (min-max)	3.1 – 57 A		
3DFSC spread (0.143)	3.2 – 3.9 A		
Sharpening B-factor			
Class 6 (# particles), codes	19S-20S, S _{D2} /E _{D2} (147,696), EMD-27018, PDB 8CV1		
Resolution (0.143/0.5)	3.1/3.8 A (masked), 4.7/8.9 A (unmasked)		
Local resolution (min-max)	2.8 – 51 A		
Sharponing R factor	2.2 – 4.0 A		
Sharpening B-factor			
Liass / (# particles), codes	$195-205$, $5_{D3}/E_{C2}$ (48,612), EMD-2/019		
Resolution (0.143/0.5)	3.3/4.1 A (MASKEO), 7.2/11 A (UNMASKEO)		
Local resolution (min-max)	3.1 - 3/ A		
SUFSC spread (0.143)	3.1 – 3.0 A		
Snarpening B-factor	U		

Supplementary Table 2. Comparing PA28(4a3b) and PA28(3a4b) models

Model composition	20S-PA28(4a3b)	20S-PA28(3a4b)
PDB code	8CXB	7NAO
Atomic modeling package	Coot	Coot
CCvolume/CCmask	0.85/0.85	0.85/0.85
B-factor (masked/unmasked)	70/75	70/75
RMSD bond lengths, angles	0.007/1.281	0.006/1.090
Molprobity score	1.41	1.04
Clashscore	4.59	2.51
Rotamers outliers (%)	1.71	0
C-beta outliers (%)	0.25	0
Ramachandrans (Favored, outliers) %	98.57/0.09	98.63/0.07
CaBLAM outliers (%)	1.39	1.41
EMRinger score	3.05	3.09