

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

- | | |
|-------------------------------------|--|
| n/a | Confirmed |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> The statistical test(s) used AND whether they are one- or two-sided
<i>Only common tests should be described solely by name; describe more complex techniques in the Methods section.</i> |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> A description of all covariates tested |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals) |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
<i>Give P values as exact values whenever suitable.</i> |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated |

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

Data analysis

1. Drift correction: MotionCor version 2.1.2.3
2. CTF estimation: GCTF version 1.06,
3. 2D and 3D Reconstruction, 3D refinement, post-processing: Relion Versions (3.06, 3.1) cisTEM 1.0.0 beta, DeepEMhancer
4. Local resolution estimation: Resmap version 1.1.5
5. Pore profile calculation: HOLE version 3.0, Channel annotation package 0.9.1
6. Model visualization: PyMOL version 2.0.4
7. 3D volume visualization: Chimera version 1.11.2, ChimeraX 0.93
8. MRC to MTZ conversion: CCP4I version 7.0.057
9. Manual model building: Coot 0.8.9.1
10. Structure refinement: Phenix version 1.17rc5-3630
11. Electrostatic potential calculation: APBS version 1.5
12. Protein surface area and interfaces analysis: PDBePISA server v1.48
13. MD simulations: Gromacs version 2018, 2020
14. Figure generation: CorelDraw version 20.1.0.708
15. Electrophysiology data analysis: Clampfit version 10.7.0.3
16. Electrophysiology traces: OriginLab Version b9.5.0.193
17. Cavity prediction: fpocket 2.0
18. Electrophysiology statistical analysis: Graphpad Prism 7.04

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

The atomic coordinates and cryo-EM maps for the GlyR-THC, GlyR-0.1gly-THC, and GlyR-0.1gly structures have been deposited in the Protein Data Bank and Electron Microscopy Data Bank with accession codes PDB-7M6M (EMD-23700), PDB-7M6O (EMD-23702) and PDB-7M6N (EMD-23701), respectively. The atomic coordinates and cryo-EM maps for the GlyR-1gly-THC-State1, GlyR-1gly-THC-State2, GlyR-1gly-THC-State3 and GlyR-1gly structures have been deposited in the Protein Data Bank and Electron Microscopy Data Bank with accession codes PDB-7M6Q (EMD-23704), PDB-7M6R (EMD-23705), PDB-7M6S (EMD-23706) and PDB-7M6P (EMD-23703), respectively.

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

- Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	For electrophysiological recordings, the wild type and mutant currents were recorded from independently injected oocytes. Ligand activation and potentiation were observed in multiple oocytes (sample size indicated in the figure legend). No sample size calculation was made and the sample sizes were considered to be sufficient based on studies that used similar methodology.
Data exclusions	For electrophysiological recordings, currents from unhealthy or leaky oocytes were not included in the analysis.
Replication	Electrophysiology experiments were performed on independent oocytes, from multiple different surgeries. Simulations were set up in triplicates. Number of replicates for experimental and simulation data are indicated in the legend. All attempts at replication were successful. Structure determination was not carried out in replicates (standard practice).
Randomization	Randomization is not applicable for structural studies. For electrophysiology experiments, oocytes were randomly used for various types of ligand-application experiments.
Blinding	Blinding was not used since it is not applicable to structural studies or reported functional measurements.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

Methods

- | n/a | Included in the study |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Antibodies |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Eukaryotic cell lines |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Palaeontology and archaeology |
| <input type="checkbox"/> | <input checked="" type="checkbox"/> Animals and other organisms |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Human research participants |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Clinical data |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Dual use research of concern |

- | n/a | Included in the study |
|-------------------------------------|---|
| <input checked="" type="checkbox"/> | <input type="checkbox"/> ChIP-seq |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> Flow cytometry |
| <input checked="" type="checkbox"/> | <input type="checkbox"/> MRI-based neuroimaging |

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)	Sf9 cell purchased from Invitrogen.
Authentication	We have only used commercially available cell lines. The Sf9 cells used were not authenticated by us.
Mycoplasma contamination	The cell lines were not tested for mycoplasma contamination
Commonly misidentified lines (See ICLAC register)	No commonly misidentified lines were used in the study

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals	Female <i>Xenopus laevis</i> (extra large adult, oocyte positive) were purchased from Nasco.
Wild animals	No wild animals were used in this study
Field-collected samples	No field-collected samples were used in this study
Ethics oversight	Dr. W. F. Boron kindly provided oocytes used in this study. We complied with all relevant ethical regulations for animal testing and research. Animal experimental procedures were approved by Institutional Animal Care and Use Committee (IACUC) of Case Western Reserve University

Note that full information on the approval of the study protocol must also be provided in the manuscript.