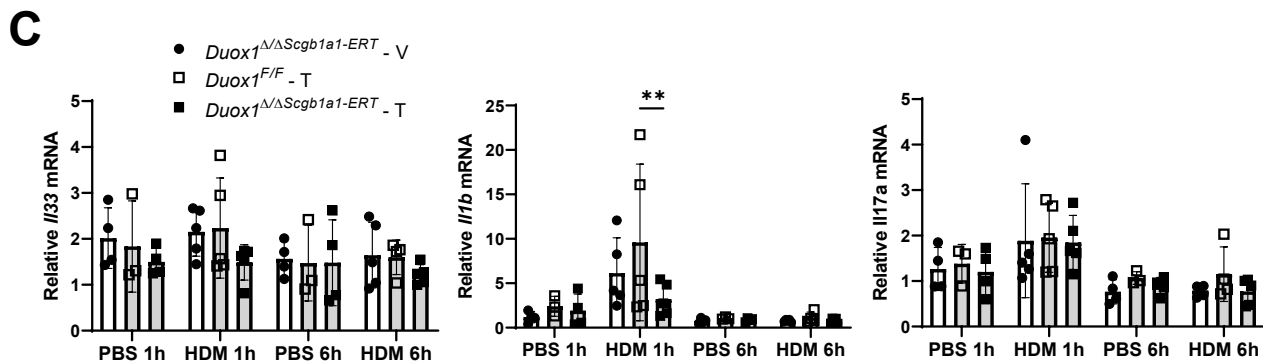
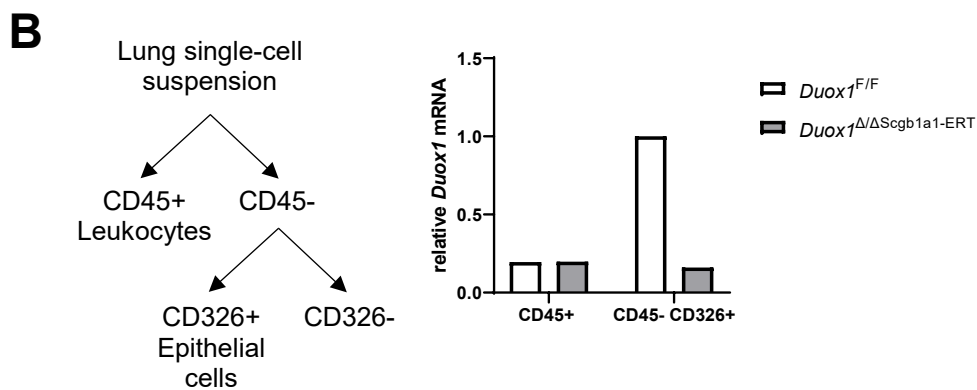
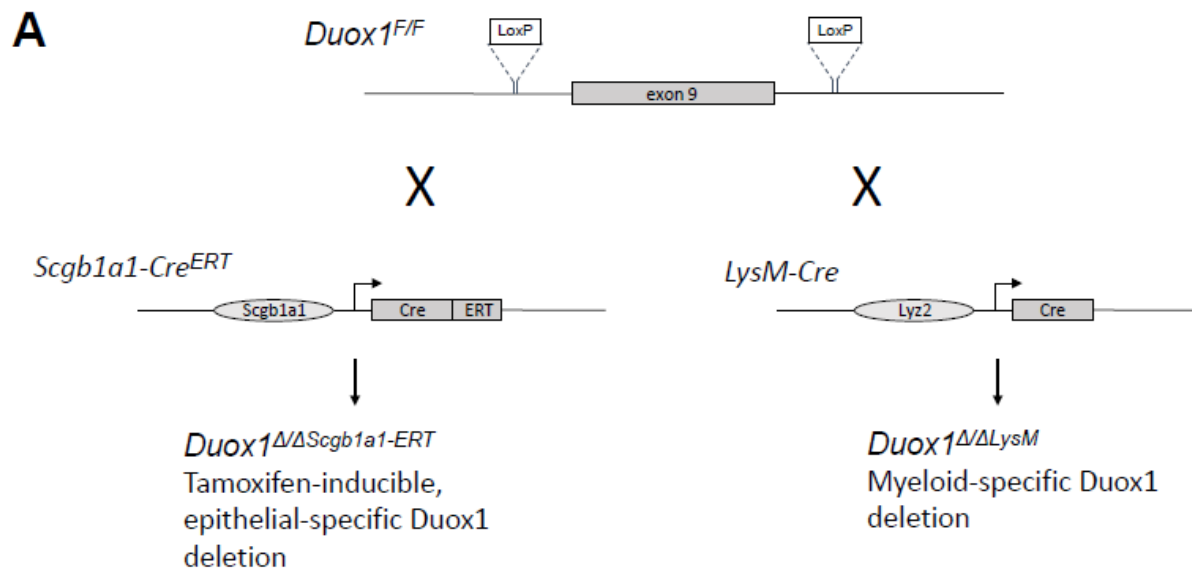


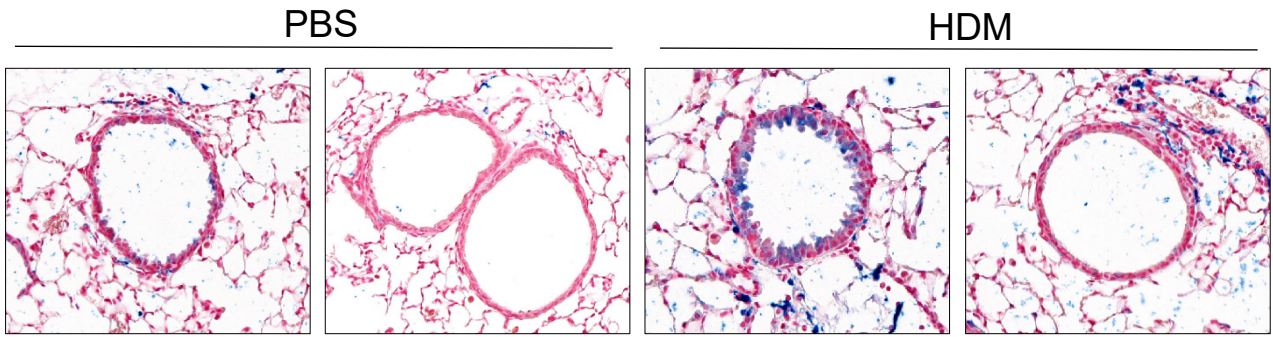
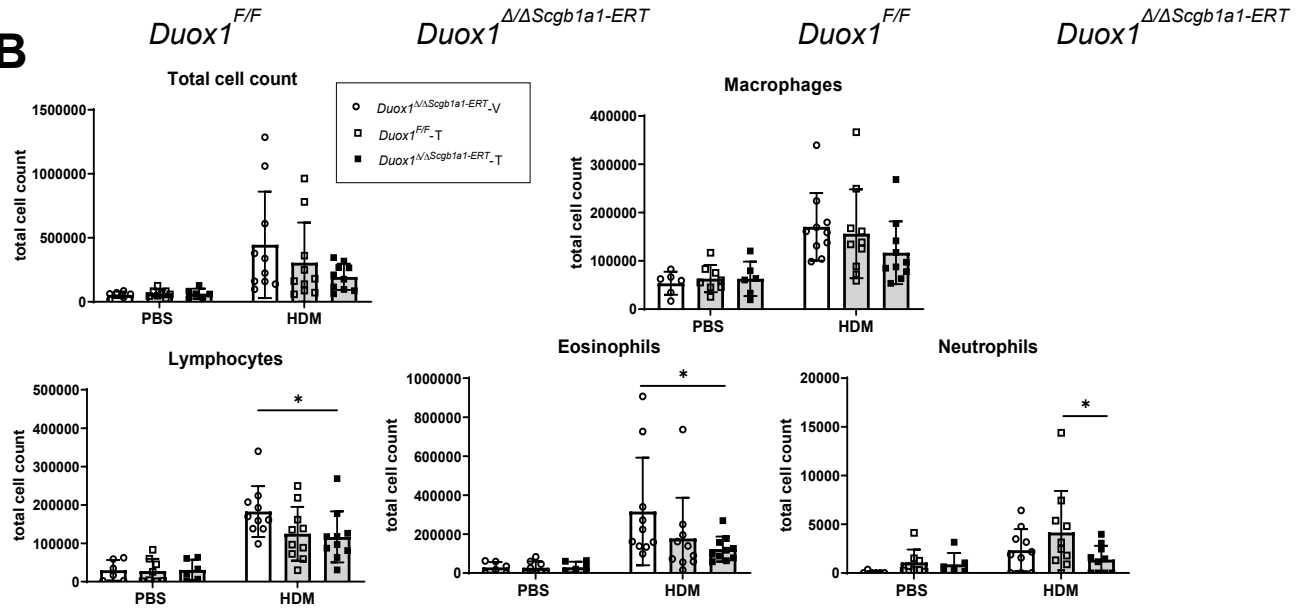
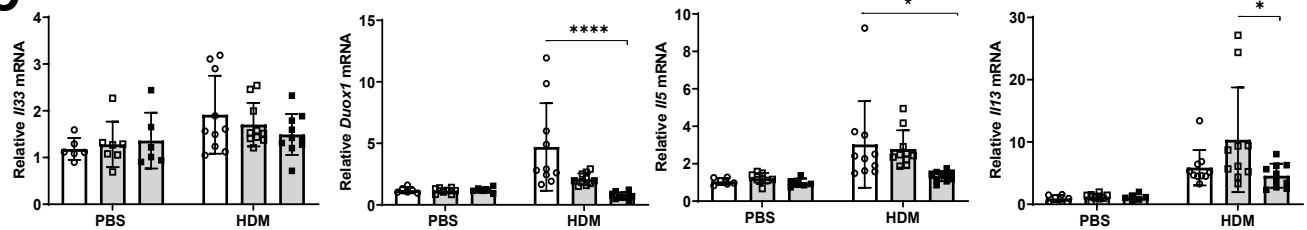
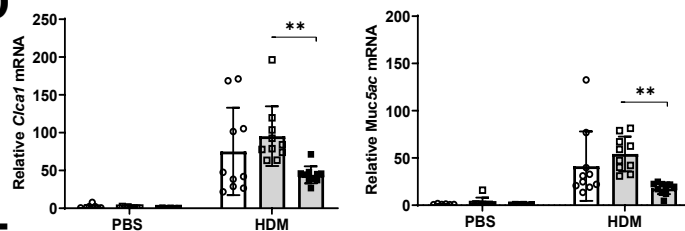
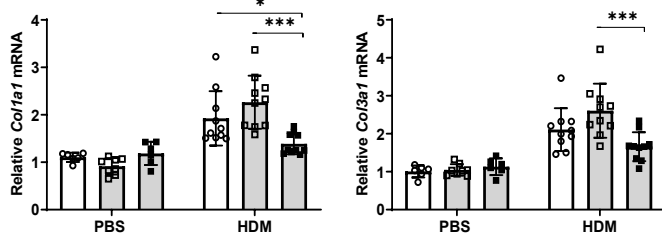
Supplementary Figures

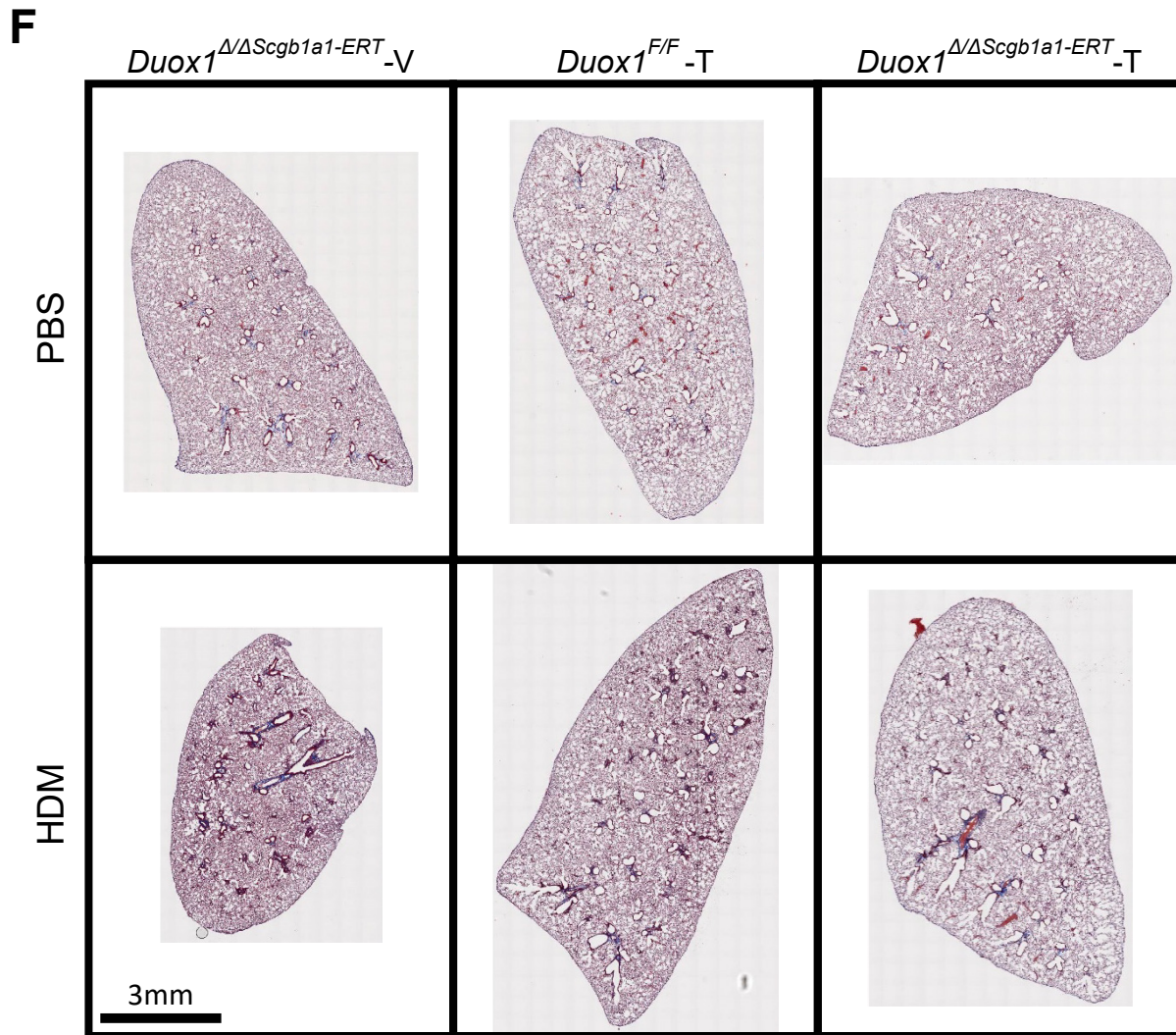
Macrophage-intrinsic DUOX1 contributes to type 2 inflammation and mucus metaplasia during allergic airway disease

Carolyn R. Morris, Aida Habibovic, Christopher M. Dustin, Caspar Schiffers, Miao-Chong Lin, Jennifer L. Ather, Yvonne M.W. Janssen-Heininger, Matthew E. Poynter, Olaf Utermohlen, Martin Krönke, and Albert van der Vliet

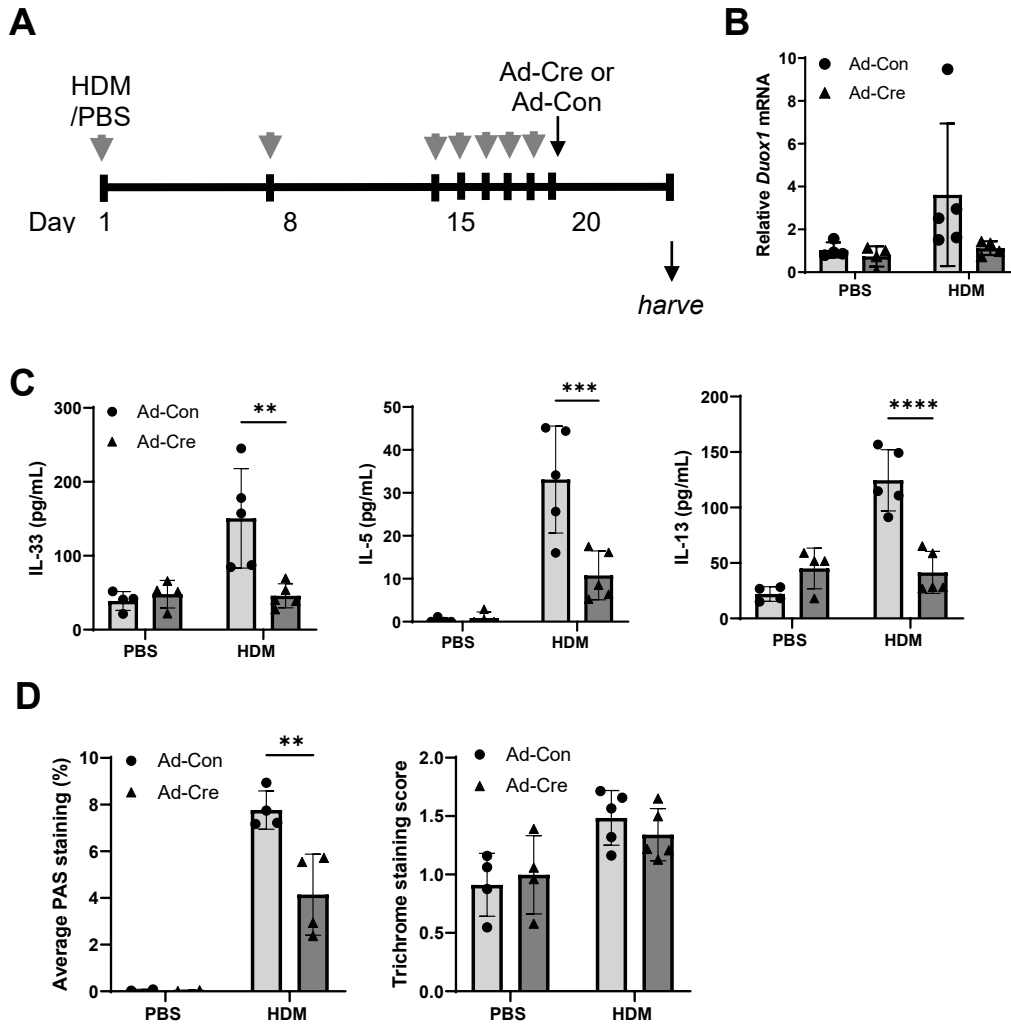


Supplementary Figure 1: (A) Schematic illustration of mouse lines used in this study. (B) Confirmation of selective *Duox1* ablation in CD326+ epithelial cells from tamoxifen-treated *Duox1^{ΔΔScgb1a1-ERT}* mice. Average results of duplicate analyses are shown. (C) RT-PCR analysis of *Il1b*, *Il33*, and *Il17a* mRNA in lung tissues from *Duox1^{F/F}* or *Duox1^{ΔΔScgb1a1-ERT}* mice, pretreated with either tamoxifen (T) or vehicle control (V), following acute intranasal HDM challenge. **: $p < 0.01$.

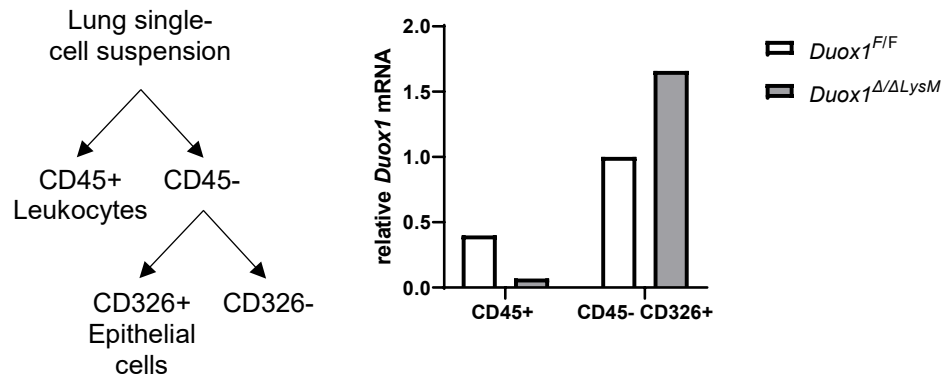
A**B****C****D****E**



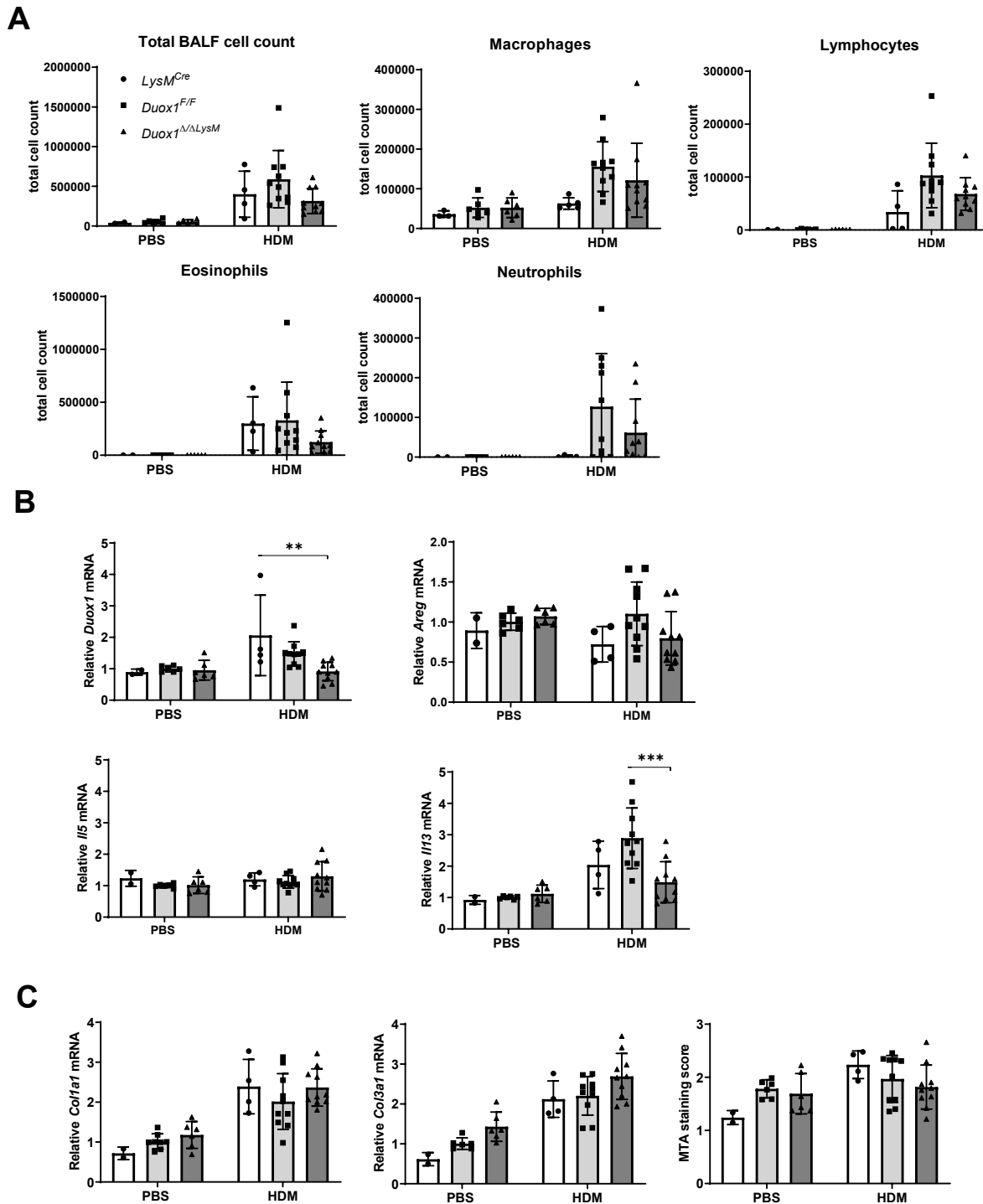
Supplementary Figure 2: Epithelial DUOX1 ablation attenuates type 2 inflammation and airway remodeling during repeated HDM challenge. *Duox1*^{F/F} or *Duox1*^{Δ/ΔScgb1a1-ERT} mice pretreated with either tamoxifen (T) or vehicle control (V) were subjected to repeated intranasal HDM or PBS. **(A)** Immunohistochemical analysis of DUOX1 protein in tamoxifen-treated *Duox1*^{F/F} or *Duox1*^{Δ/ΔScgb1a1-ERT} mice following repeated PBS or HDM challenge, revealing selective ablation of DUOX1 within the epithelium. **(B)** Quantification of BAL inflammatory cell types. **(C-E)** Lung tissue analysis of mRNA levels of genes related to type 2 inflammation **(C)**, mucus metaplasia **(D)**, and collagen production **(E)**. **(F)** Whole slide images of representative full lung sections following Masson's trichrome staining. *: p<0.05; **: p<0.01; ***: p<0.001; ****: p<0.0001.



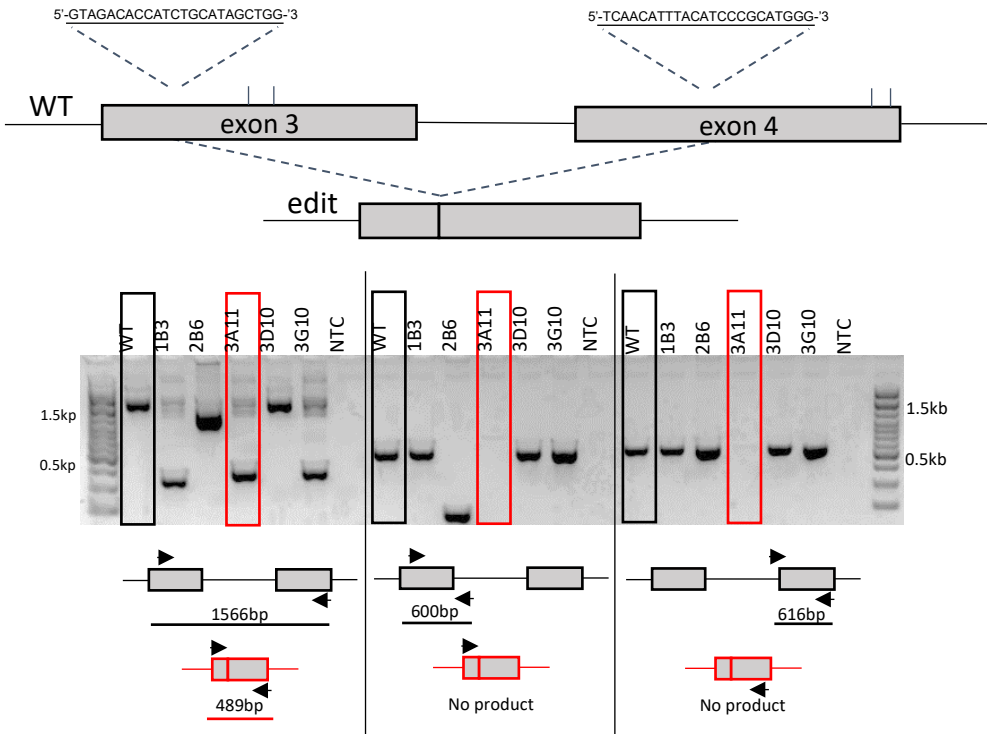
Supplementary Figure 3: DUOX1 ablation during ongoing allergic airway inflammation promotes resolution of inflammation and remodeling. (A) Schematic of experimental design and timeline of HDM and Adeno-Cre administration. **(B)** Analysis of lung tissue *Duox1* mRNA expression. **(C)** ELISA analysis of type cytokines in BAL fluids. **(D)** Quantification of PAS staining and Masson's trichrome staining. **: $p < 0.01$; ***: $p < 0.001$; ****: $p < 0.0001$.



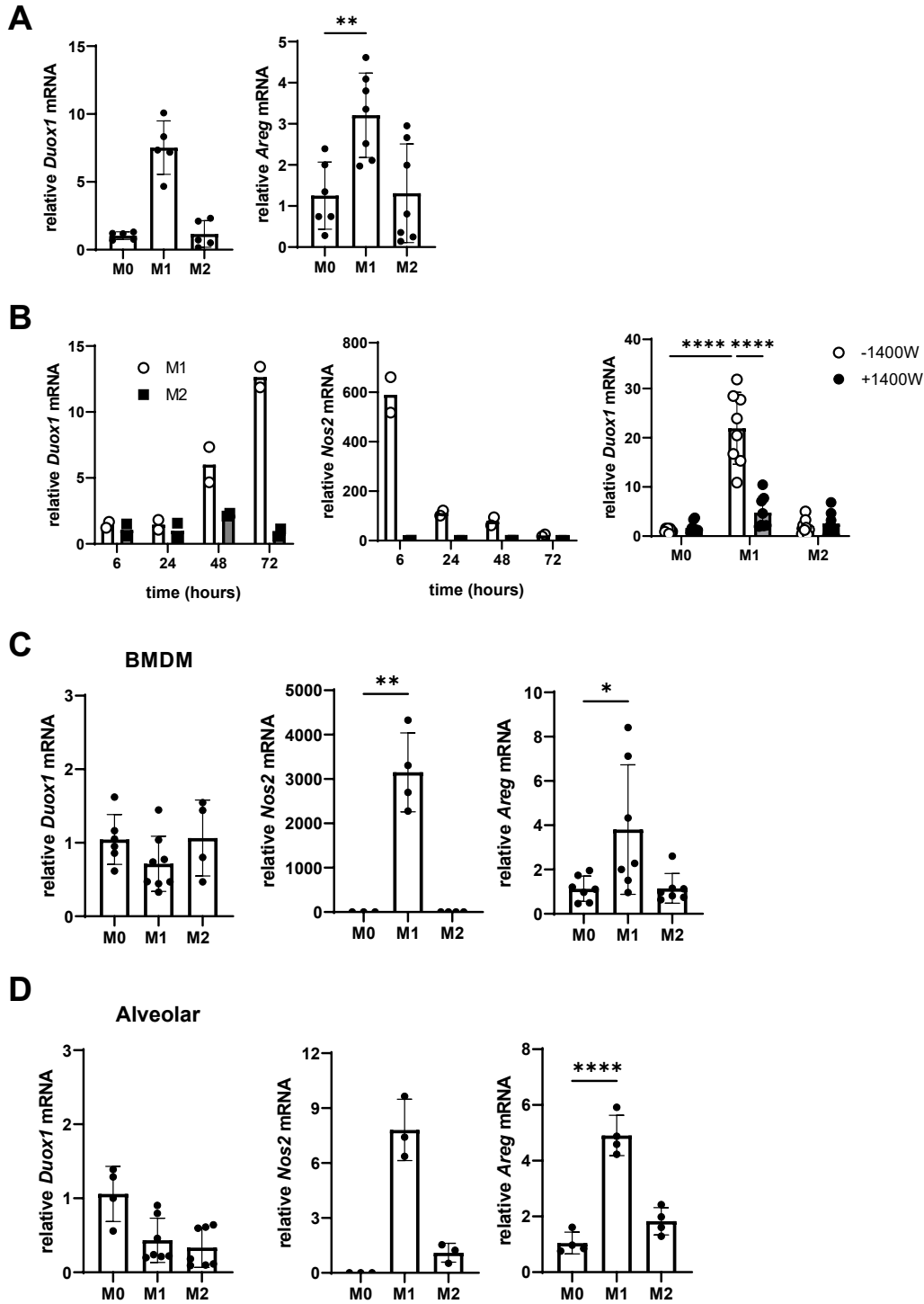
Supplementary Figure 4: Analysis of selective *Duox1* ablation in CD45+ cells but not CD326+ epithelial cells from *Duox1*^{Δ/ΔLysM} mice. Average results of duplicate analyses are shown.



Supplementary Figure 5: Selective ablation of myeloid DUOX1 attenuates type 2 inflammation and mucus metaplasia during chronic HDM challenge. *Duox1*^{Δ/ΔLysM} mice and *LysM*^{Cre+/-} or *Duox1*^{F/F} as controls, were subjected to repeated challenge with HDM or PBS, and BAL fluid were analysed for inflammatory cell counts (**A**), and lung tissues were analyzed for mRNA levels of genes related to type 2 inflammation (**B**). (**C**) Lung tissue analysis of mRNA levels of collagen genes or Masson's trichrome staining. **: p<0.01; ***: p<0.001.



Supplementary Figure 6: Schematic of *Duox1* editing and screening in MH-S cell clones.



Supplementary Figure 7: Impact of M1/M2 polarization on macrophage *Duox1* expression. MH-S cells were subjected to M1 or M2 polarization for 48 hrs (A) or indicated times (B), and analyzed for mRNA expression of *Duox1* or M1 markers *Nos2* or *Areg*. (B, right panel) Impact of NOS2 inhibition by 1400W on *Duox1* mRNA during 72-hr M1 polarization. (C,D) Impact of 48-hr M1/M2 polarization of mouse bone marrow-derived macrophages (BMDM) or alveolar macrophages on *Duox1*, *Nos2* or *Areg*. *: $p < 0.05$; **: $p < 0.01$; ****: $p < 0.0001$.