

# Supplementary Materials for

## Predicting the potential for zoonotic transmission and host associations for novel viruses

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## Supplementary Results

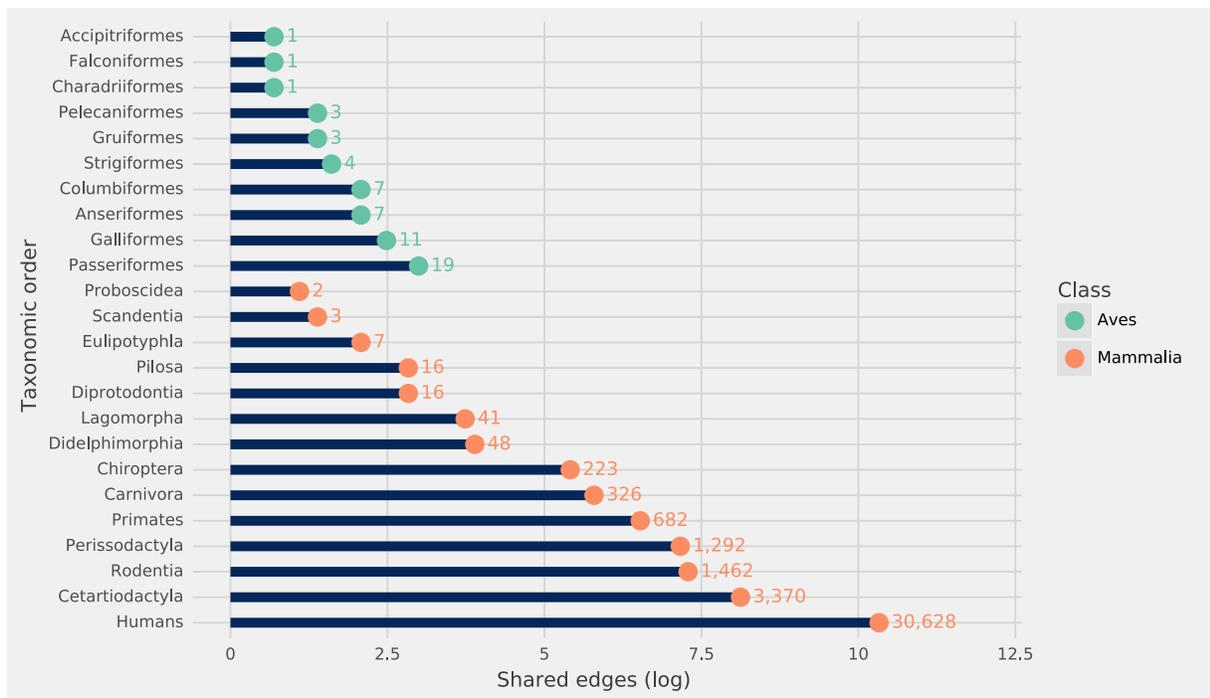
### Feature selection based on Pearson's correlation coefficient:

Resource allocation index, Jaccard coefficient, Adamic Adar, and preferential attachment coefficient were highly correlated with each other ( $R^2 > 0.95$ ), hence we decided to use only the Jaccard coefficient as a feature in training the model (Fig. S7).

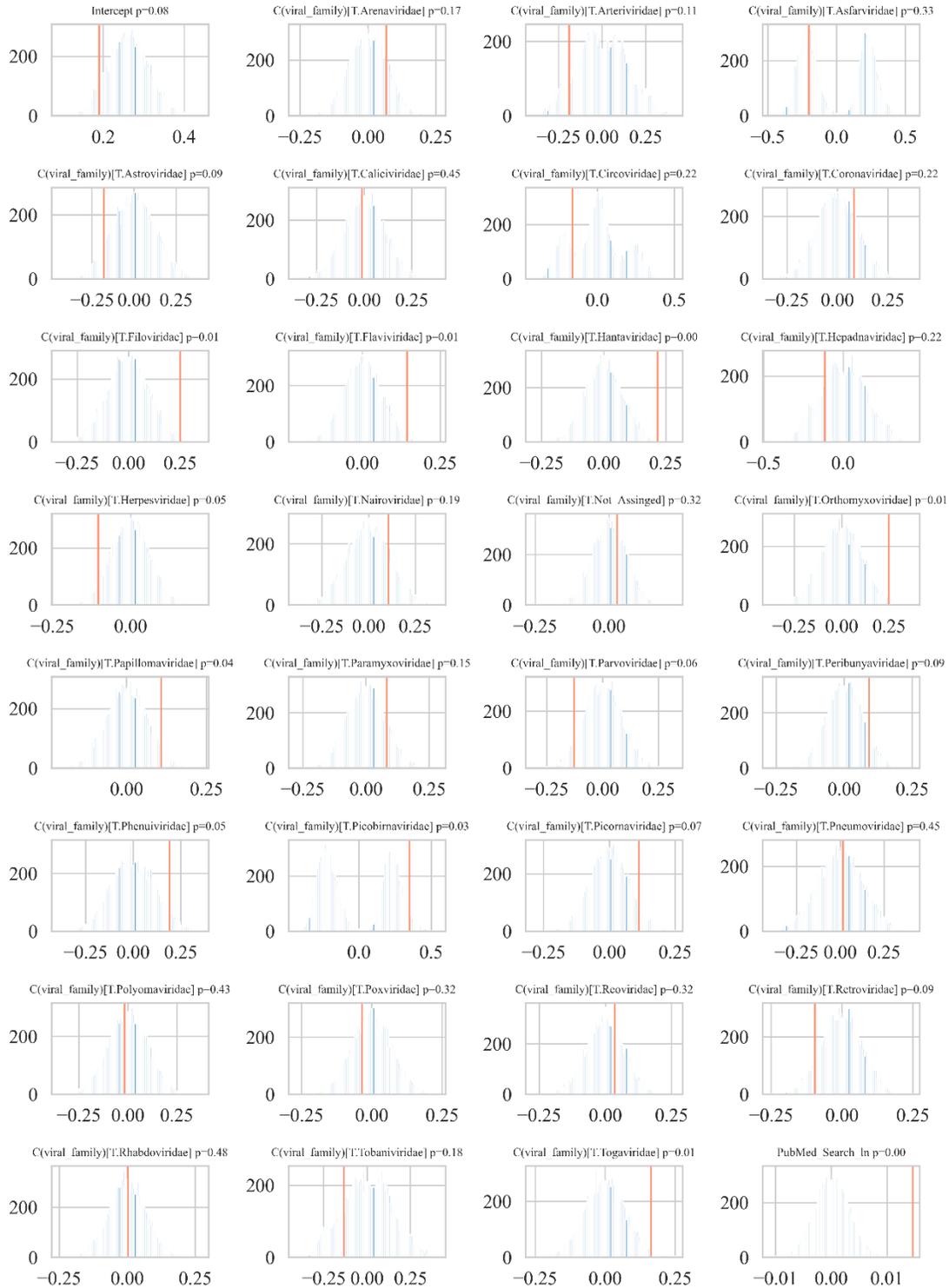
### Hypertuning binary model:

Grid search for tuning the best model parameters for the binary model resulted in the following parameters that performed the best for the XGBoost classifier.

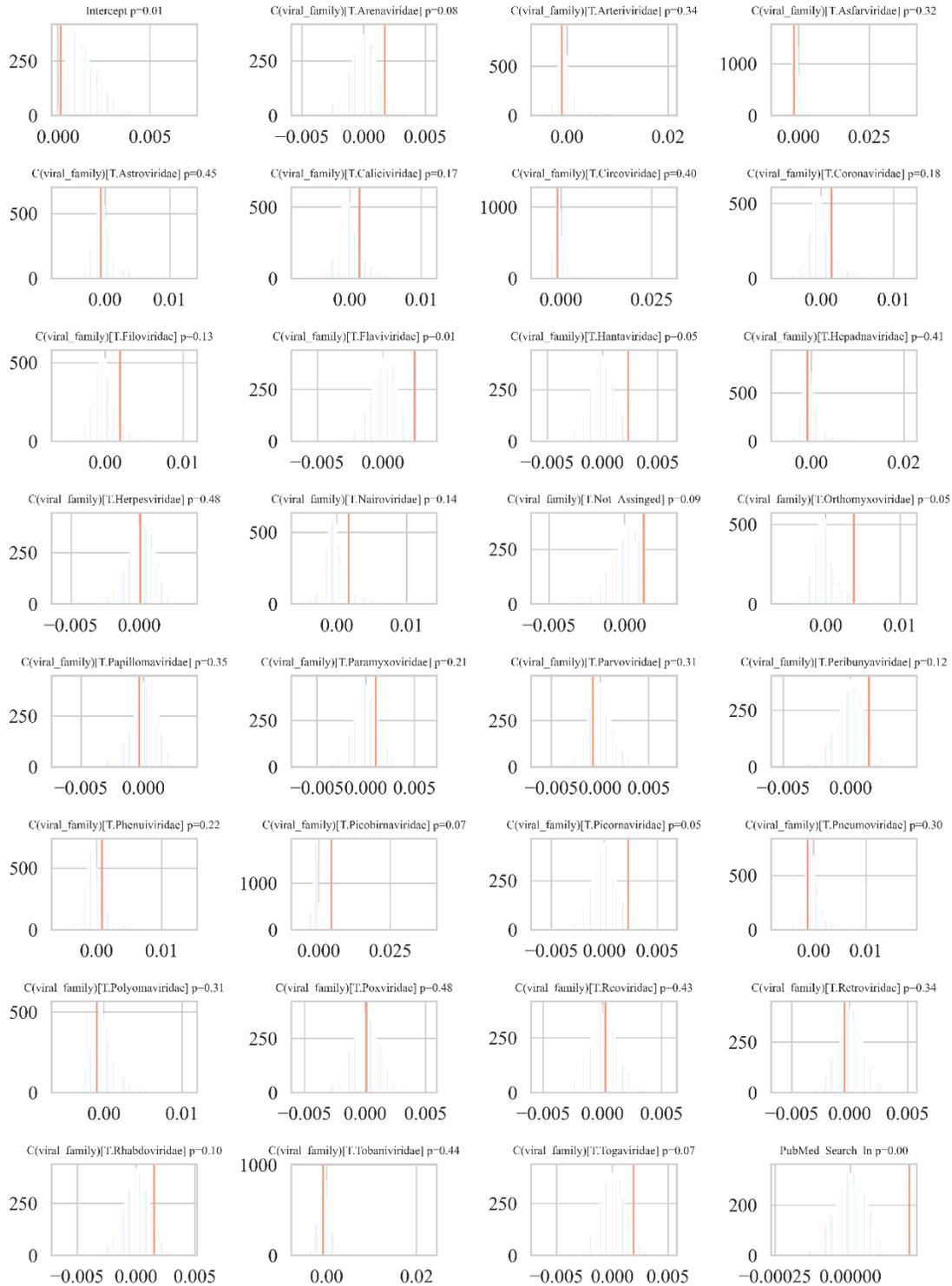
```
XGBClassifier(base_score=0.5, booster='gbtree', colsample_bylevel=1,  
              colsample_bytree=0.7, gamma=0.1, learning_rate=0.2,  
              max_delta_step=0, max_depth=12, min_child_weight=3, missing=None,  
              n_estimators=100, n_jobs=1, nthread=None,  
              objective='binary:logistic', random_state=0, reg_alpha=0,  
              reg_lambda=1, scale_pos_weight=1, seed=None, silent=True,  
              subsample=1)
```



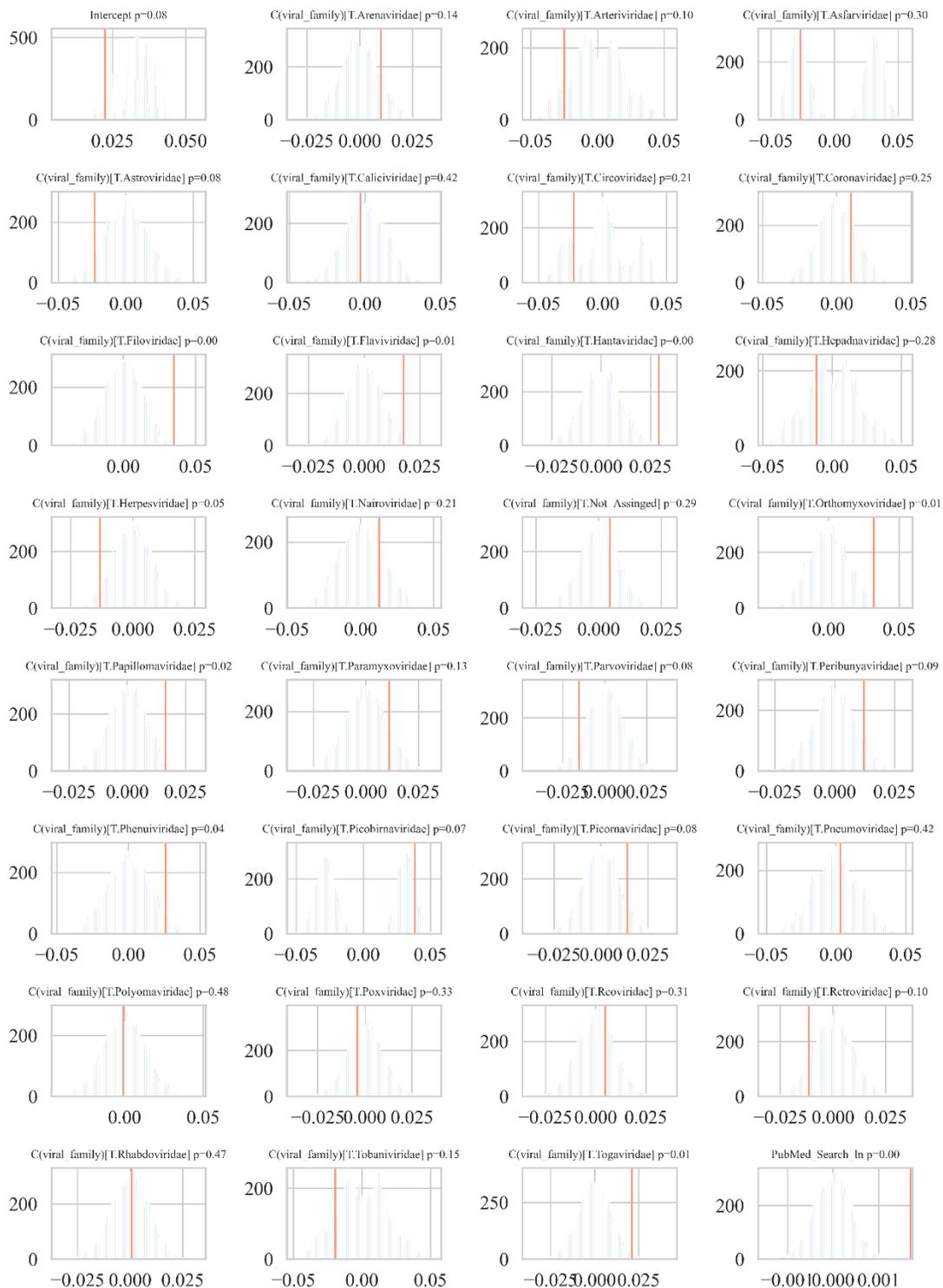
**Fig. S 1:** Distribution of edges by avian and mammalian taxonomic orders in the unipartite complete network ( $G_c$ ) generated by sharing of hosts by zoonotic and non-zoonotic viruses.



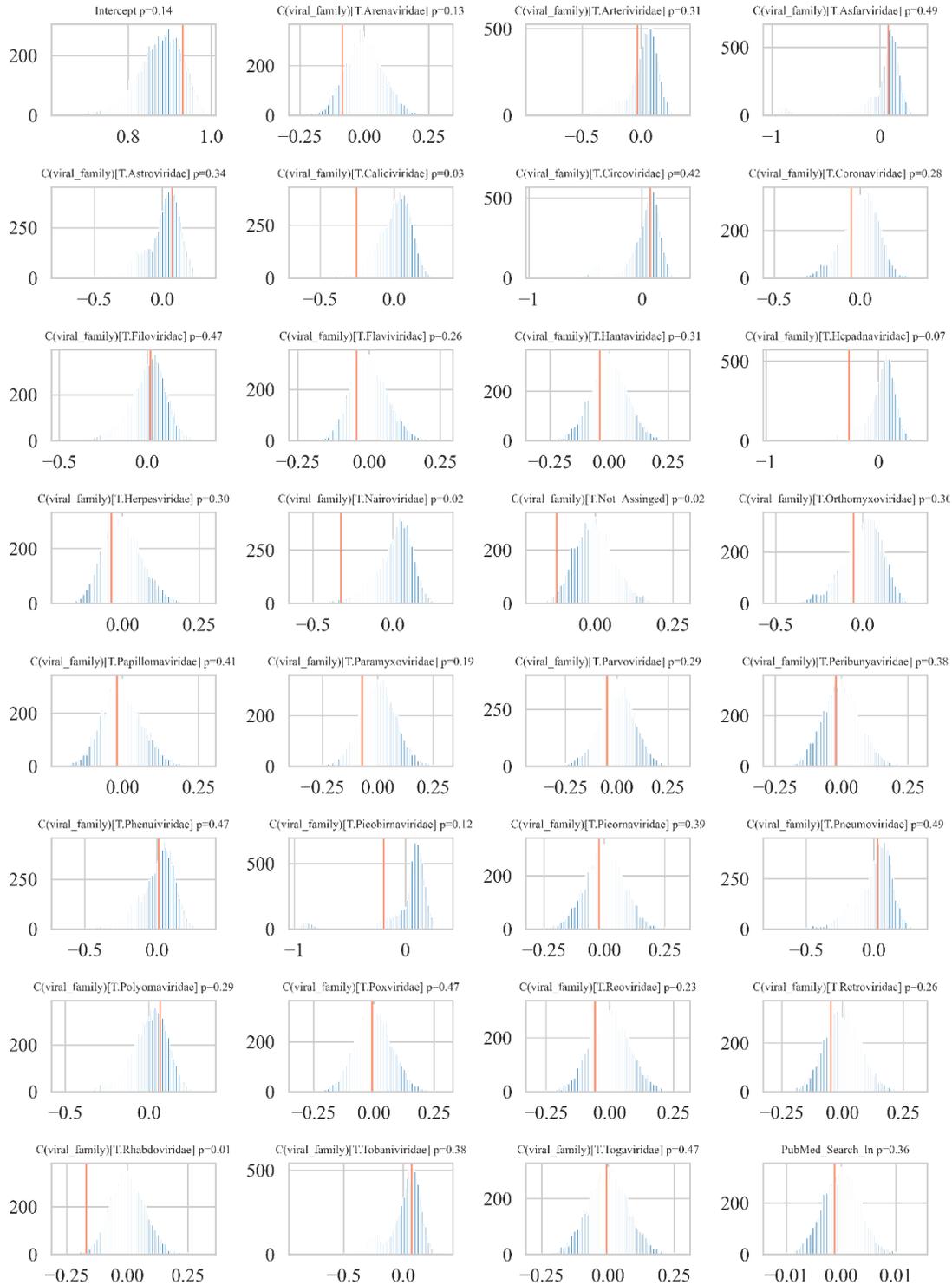
**Fig. S2.** Results of permutation-based regression analysis to understand the effects of virus family and citations on the degree centrality of viruses in the observed network. Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



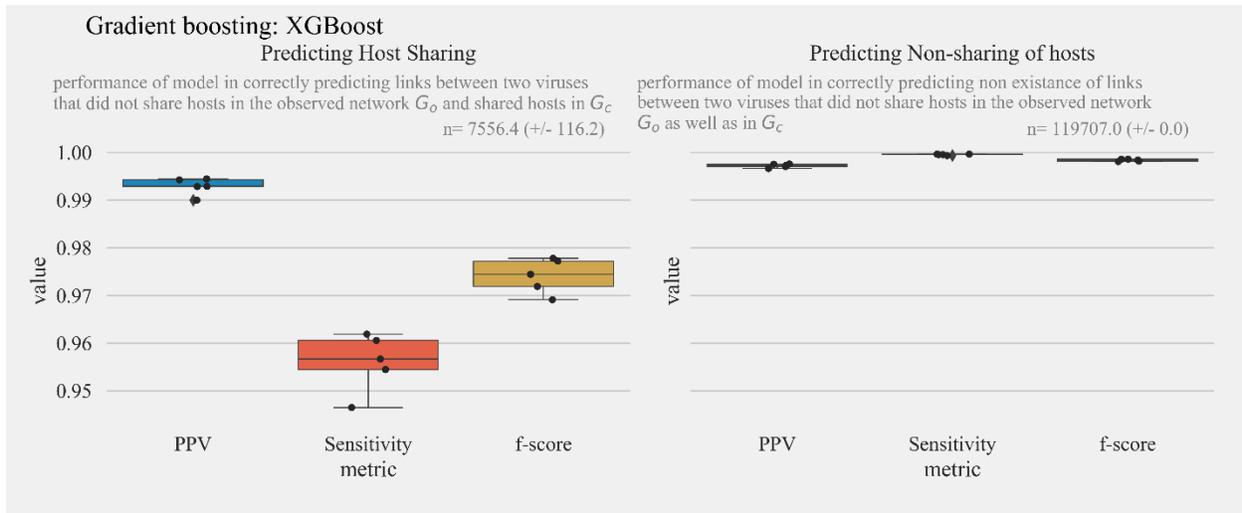
**Fig. S3.** Results of permutation-based regression analysis to understand the effects of viral family and citations on the betweenness centrality of viruses in the observed network. Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



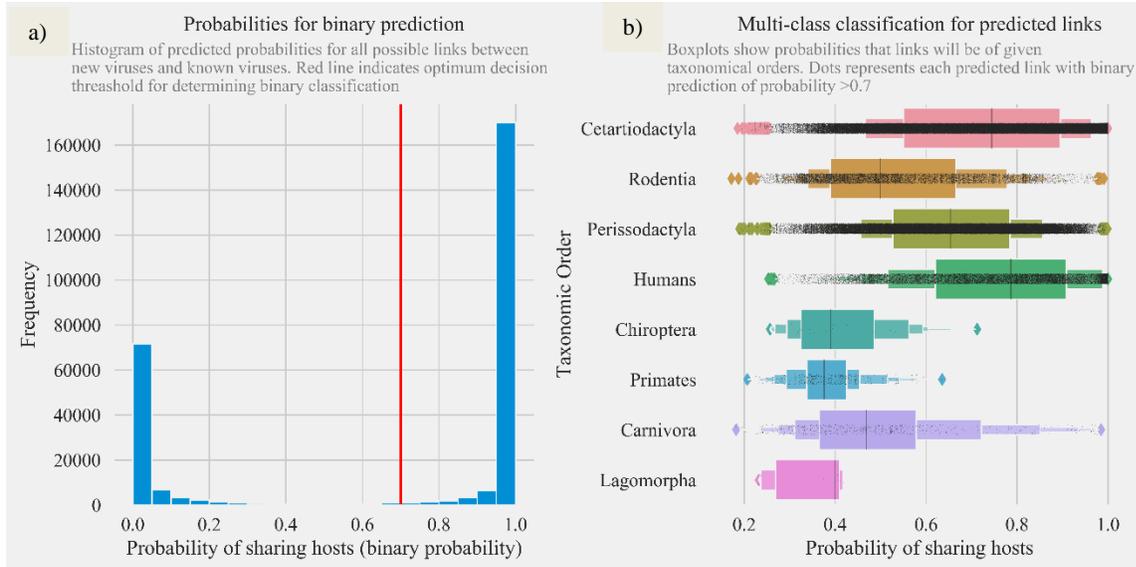
**Fig. S4.** Results of permutation-based regression analysis to understand the effects of viral family and citations on the eigenvector centrality of viruses in the observed network. Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



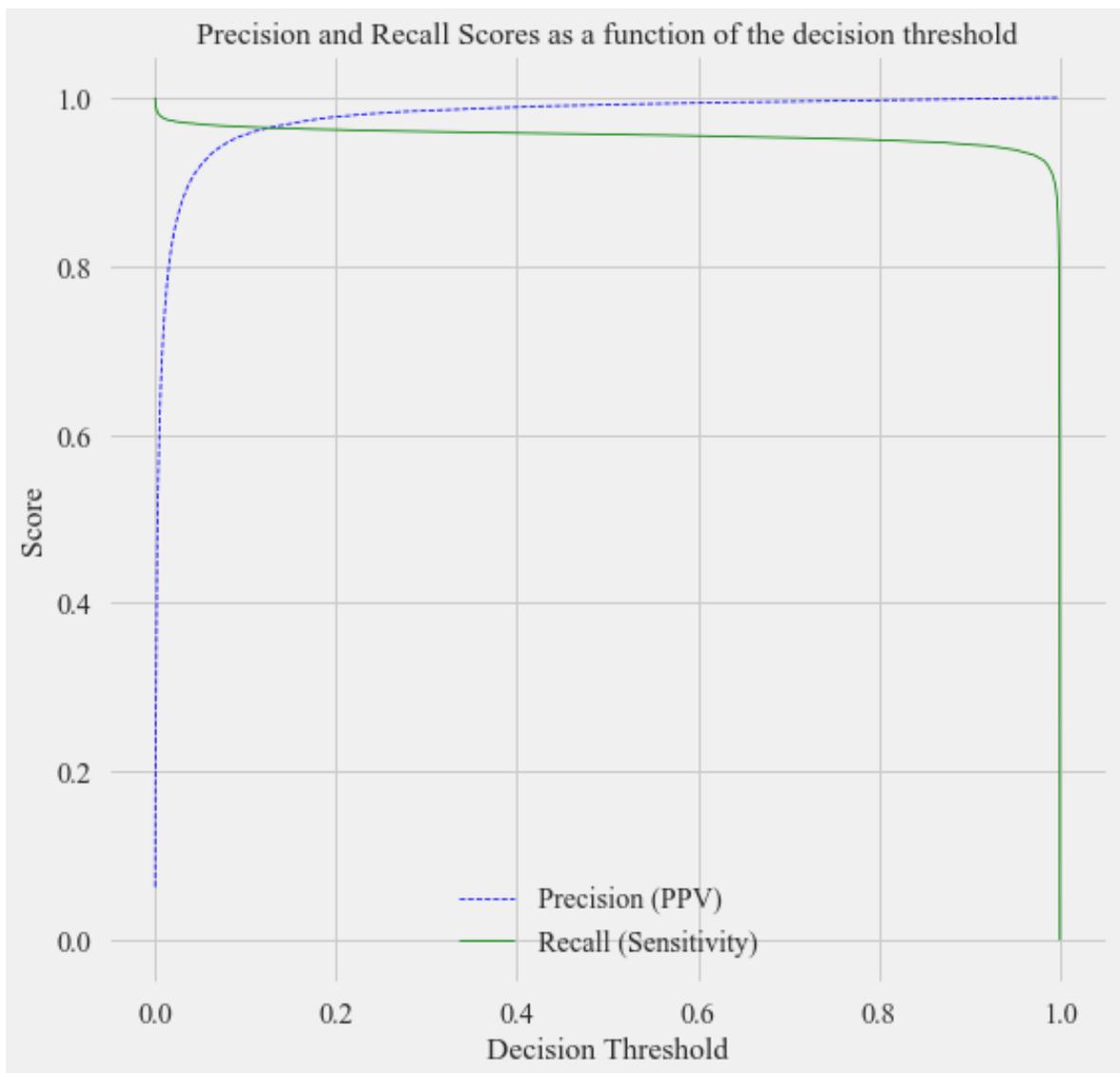
**Fig. S 5.** Results of permutation-based regression analysis to understand the effects of viral family and citations on the clustering coefficient of viruses in the observed network. Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



**Fig. S 6.** Sensitivity, positive predictive value, and f-score for correctly predicting the sharing of hosts in  $G_c$  by viral pairs that did not share hosts in  $G_o$  and non-sharing of hosts in  $G_c$  by virus pairs that did not share hosts in  $G_o$  for binary XGBoost model.



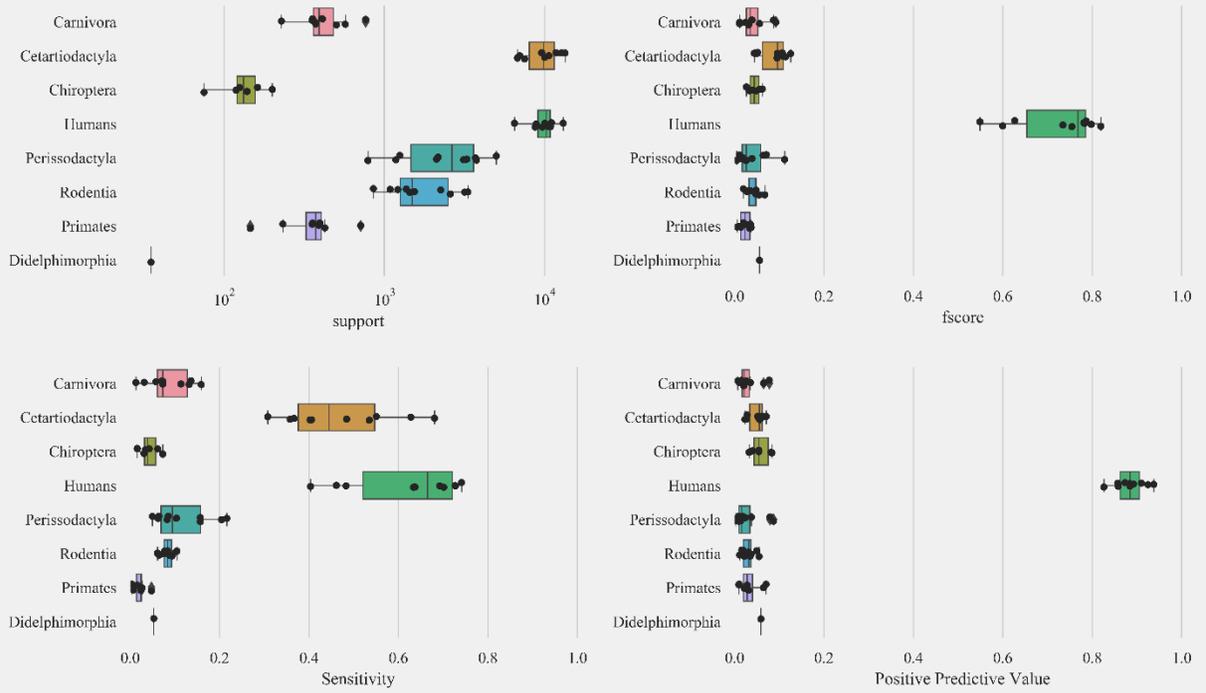
**Fig. S 7:** Predictions for PREDICT viruses. a) Histogram of predicted probabilities for the presence of a link between all possible pairs of PREDICT viruses and known zoonotic and non-zoonotic viruses (left panel). The red line indicates an optimum decision threshold for determining binary classification. On the right b), boxplots show predicted probabilities that the links predicted by the binary model between viruses are generated by the sharing of hosts from taxonomic order.



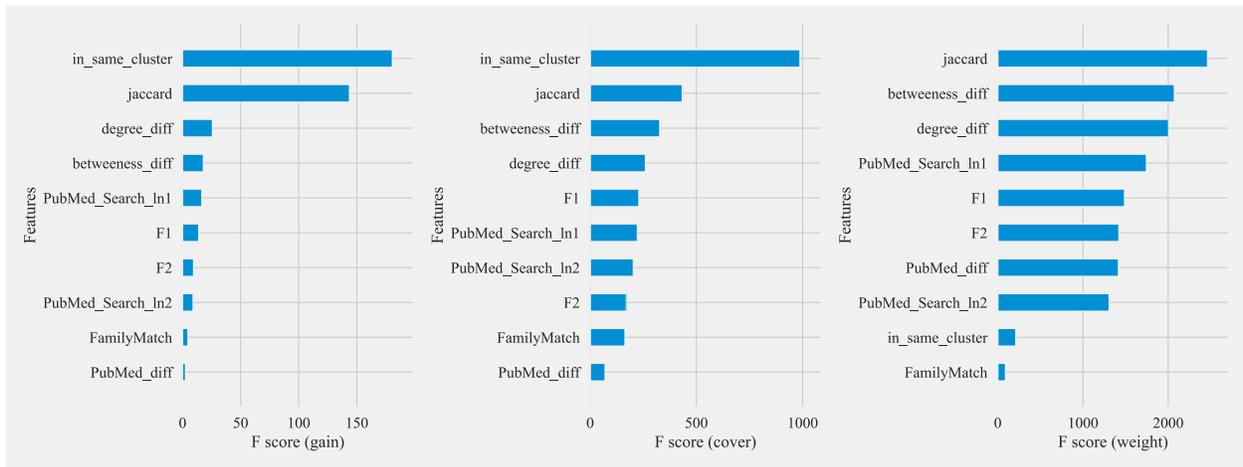
**Fig. S 8:** Precision recall scores as a function of the decision threshold shows a high score (beyond 0.15) model predicted probability for the binary model.

### Predicting Shared Host Order

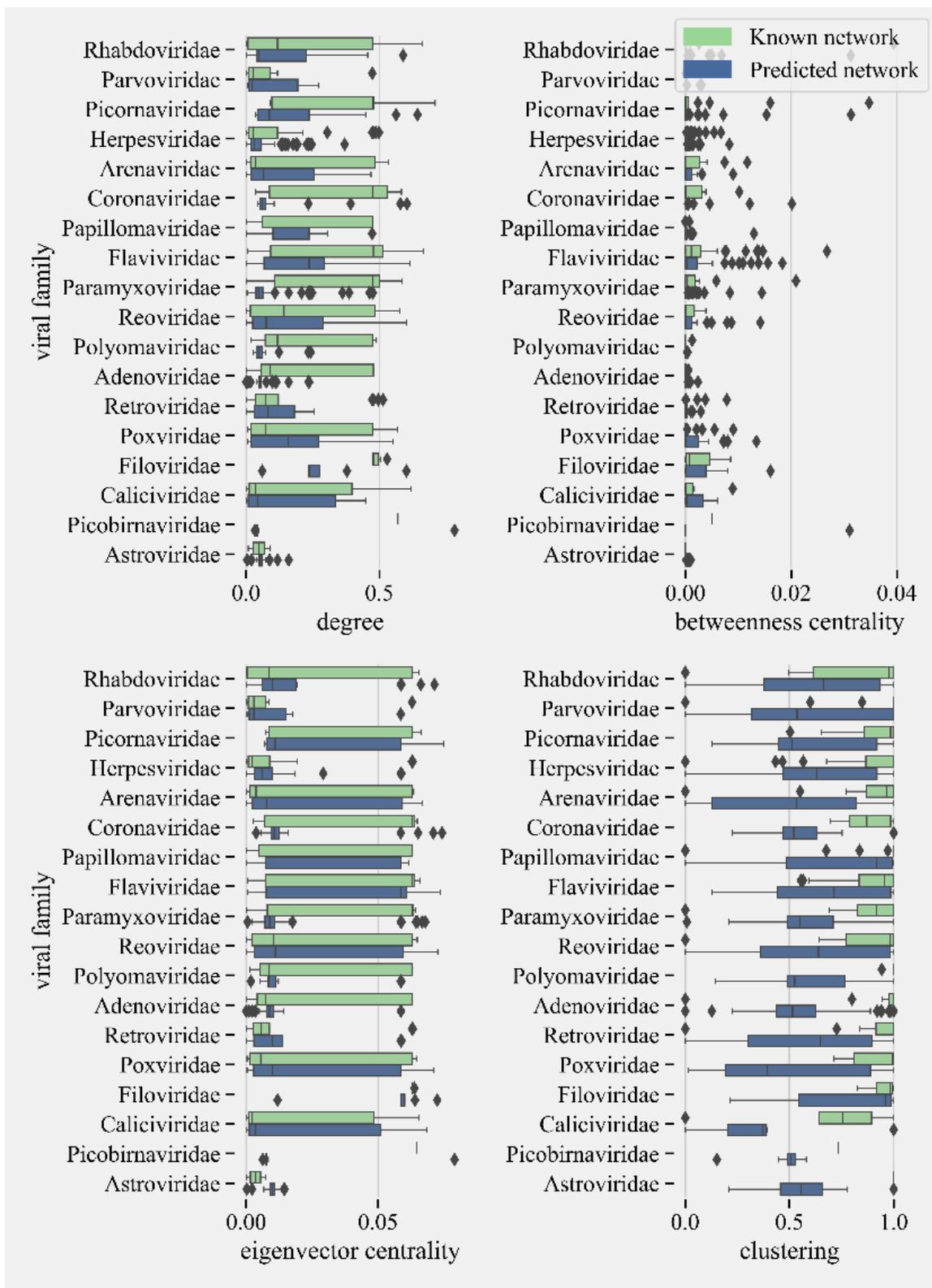
Sample size for validation of XGBoost model performance in correctly predicting the type of links (host order) between two viruses that did not share hosts in the observed network  $G_O$  and shared hosts in  $G_C$ .



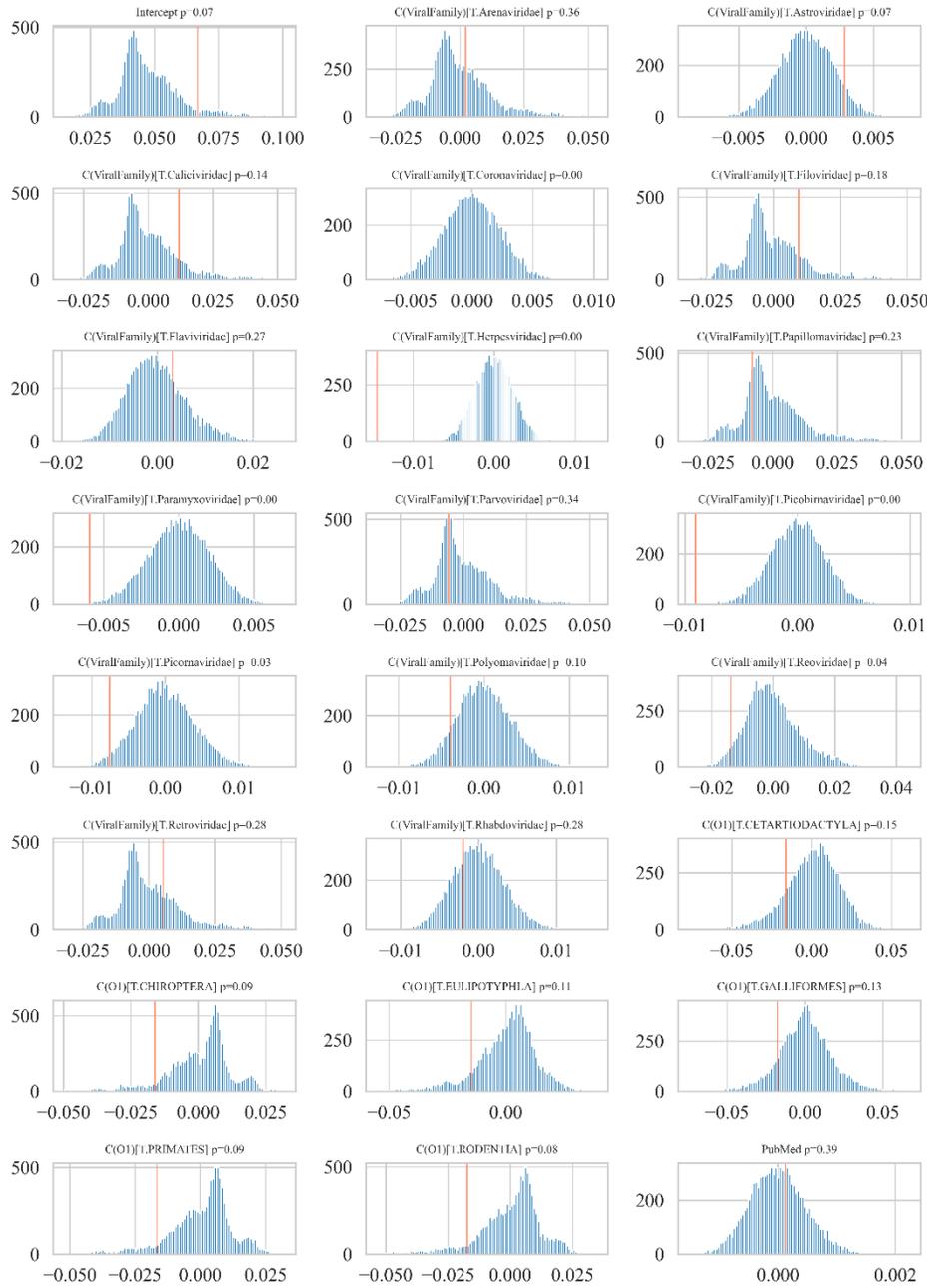
**Fig. S 9.** Sample size, f-score, sensitivity, and positive predictive value for validation of multiclass multioutput XGBoost model for correctly predicting the type of host sharing between viruses that did not share hosts in  $G_O$  and shared hosts in  $G_C$ .



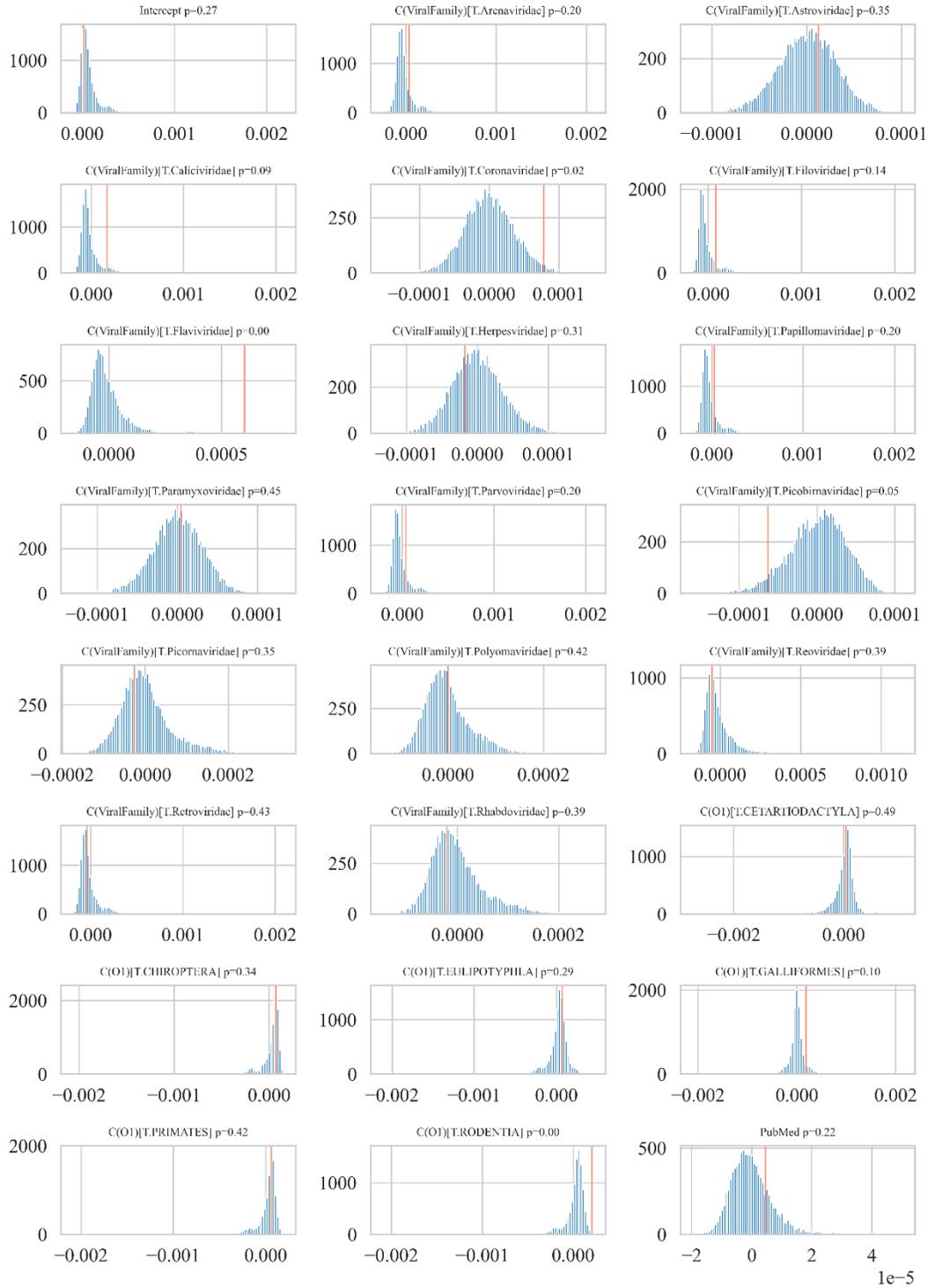
**Fig. S 10:** Feature importance of network topological characteristics and virological features used in training binary model based on gain, cover and weight.



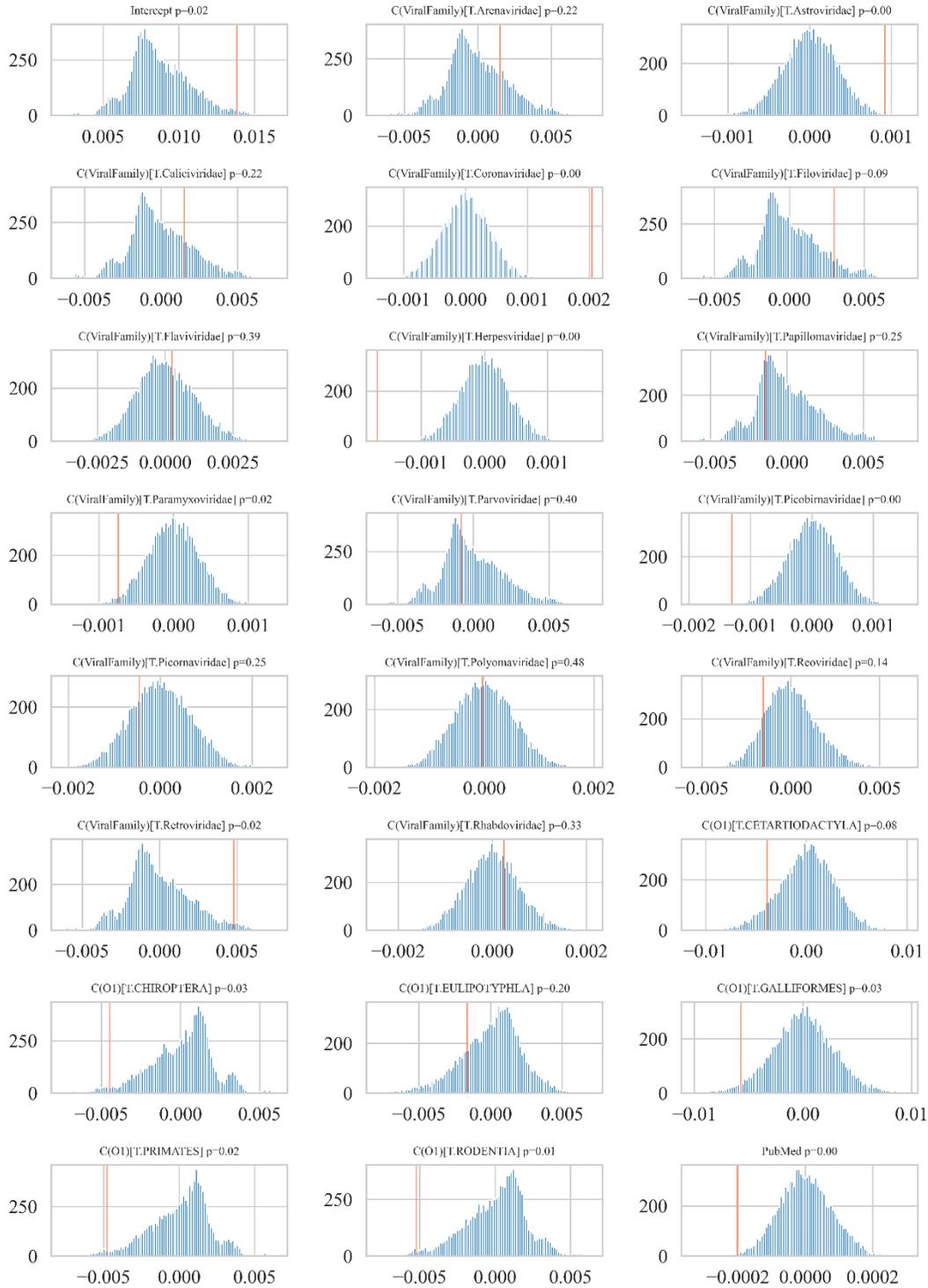
**Fig. S 11:** Comparison of centrality measures and clustering coefficient for selected viral families in predicted and known network.



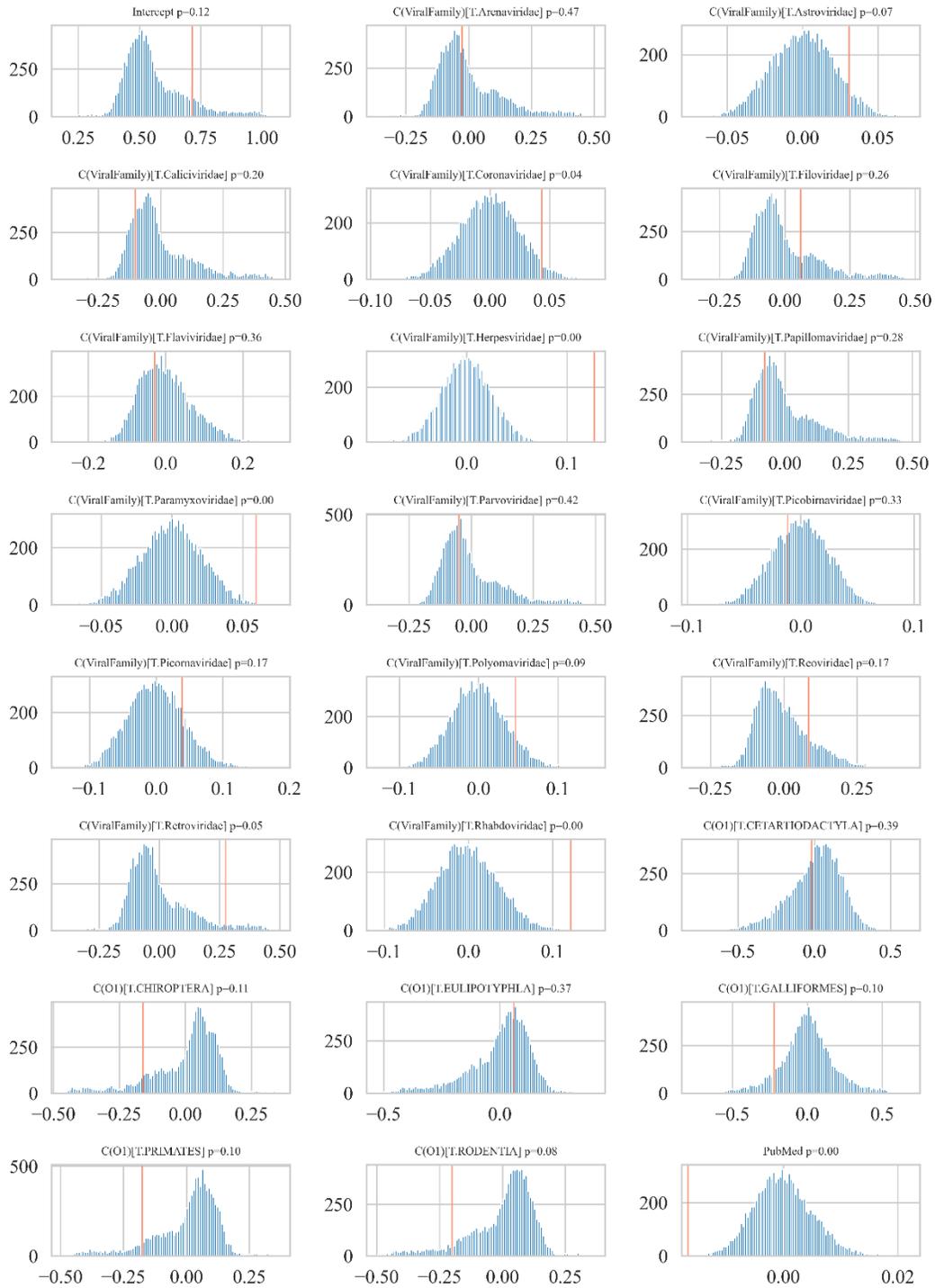
**Fig. S 12:** Results of permutation-based regression analysis to understand the effects of virus family, host taxonomic order and number of detections on the degree centrality of novel viruses in the predicted network ( $G_{predicted}$ ). Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



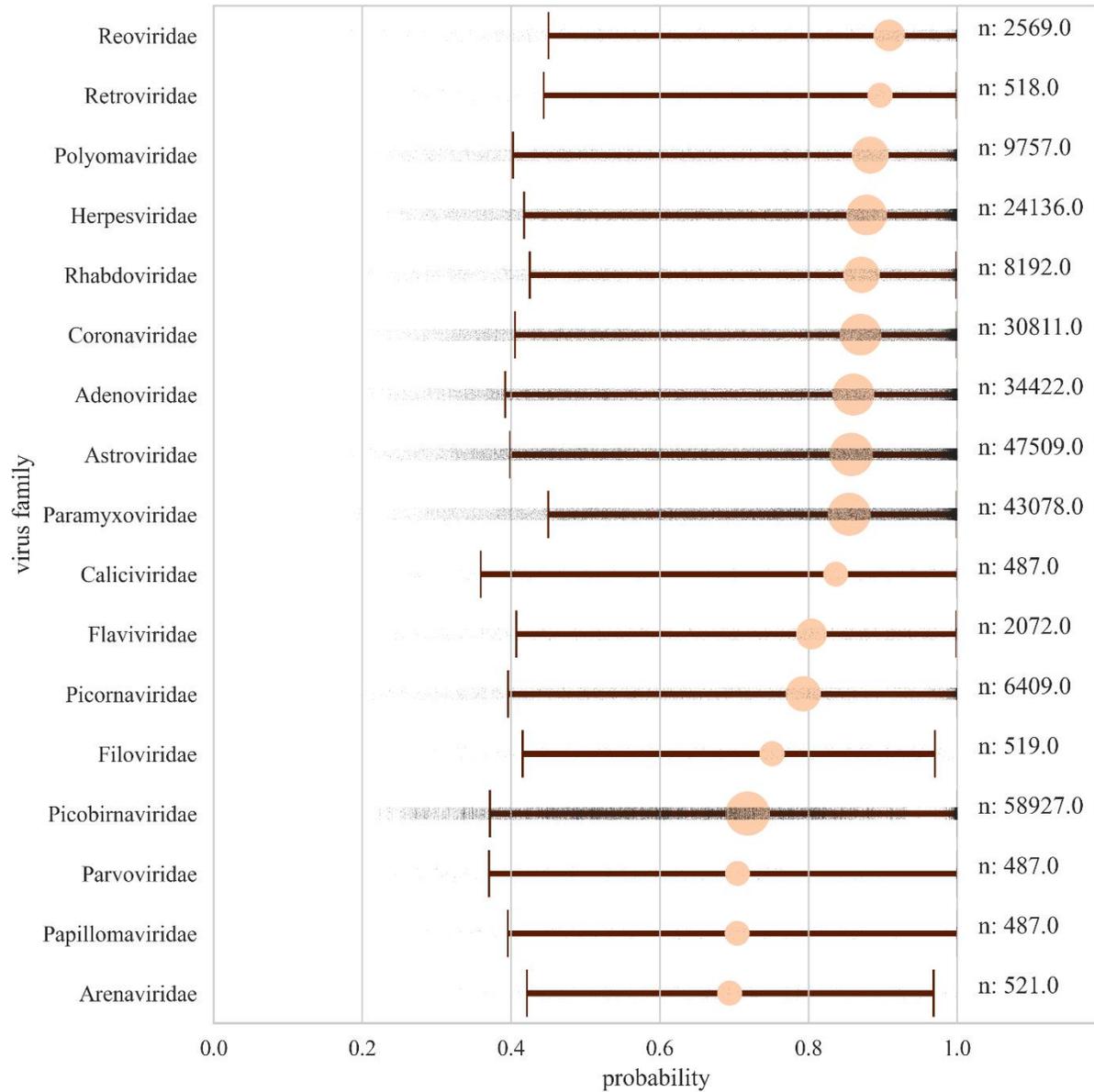
**Fig. S 13:** Results of permutation-based regression analysis to understand the effects of virus family, host taxonomic order, and number of detections on the betweenness centrality of novel viruses in the predicted network ( $G_{predicted}$ ). Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



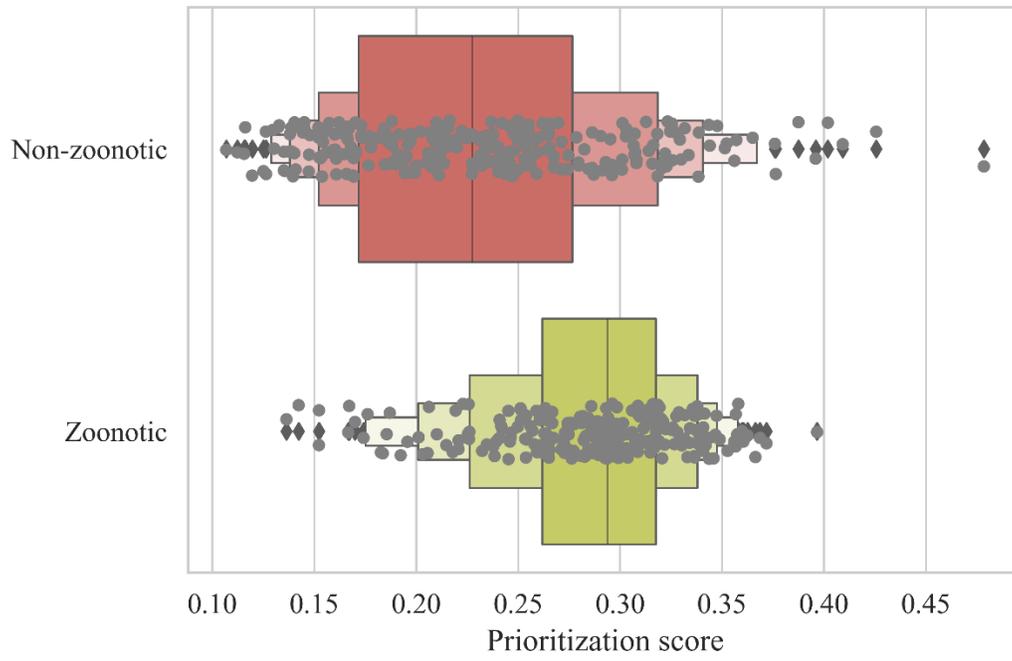
**Fig. S 14:** Results of permutation-based regression analysis to understand the effects of virus family, host taxonomic order and number of detections on the eigenvector centrality of novel viruses in the predicted network ( $G_{predicted}$ ). Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



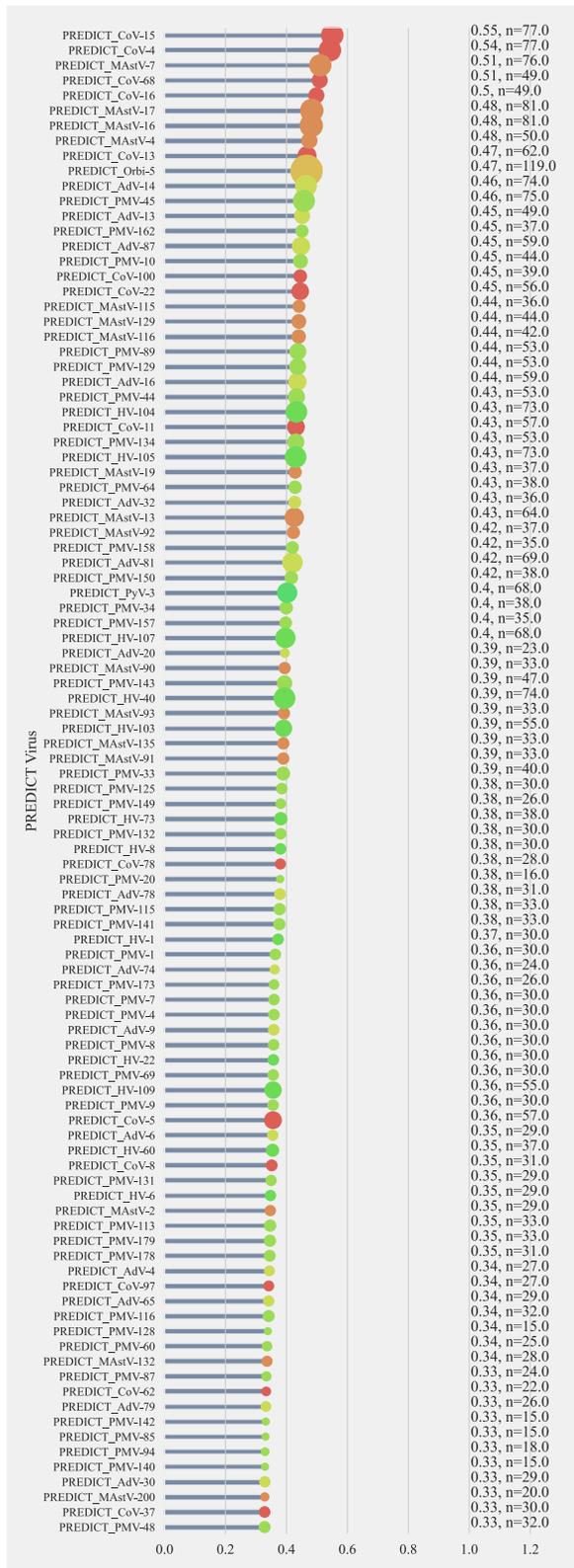
**Fig. S 15:** Results of permutation-based regression analysis to understand the effects of virus family, host taxonomic order and number of detections on the clustering coefficient of novel viruses in the predicted network ( $G_{predicted}$ ). Blue lines show the distribution of coefficients after 10,000 random permutations. Red lines show original coefficients.



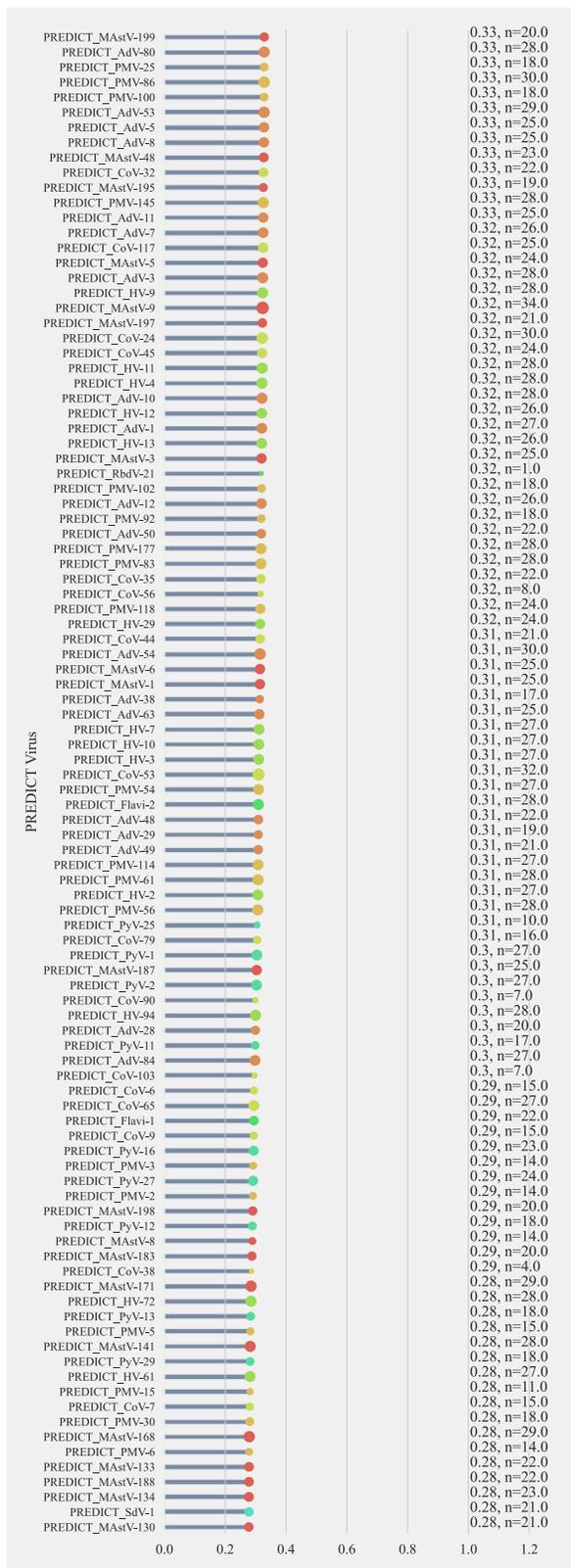
**Fig. S 16:** Predicted probabilities of human links between newly discovered viruses and known zoonotic and non-zoonotic viruses based on the multiclass model. Circles represent the median for all probabilities estimated for viruses within the virus family. The size of the circle is proportional to the number of human links predicted for newly discovered viruses within the virus family (represented with n). Error represents the 5th, and 95th percentile for the distribution, and grey shaded dots represent probabilities for each predicted human link.



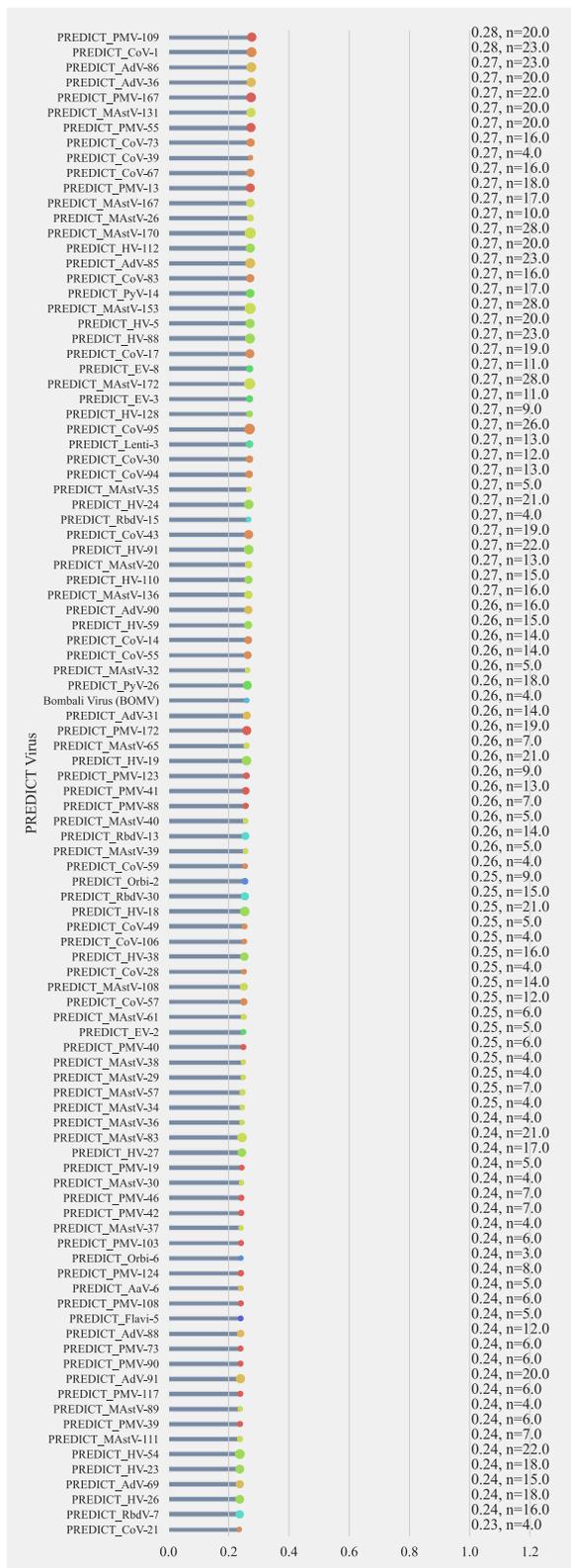
**Fig. S 17: Prioritization scores for known zoonotic and non-zoonotic viruses.**



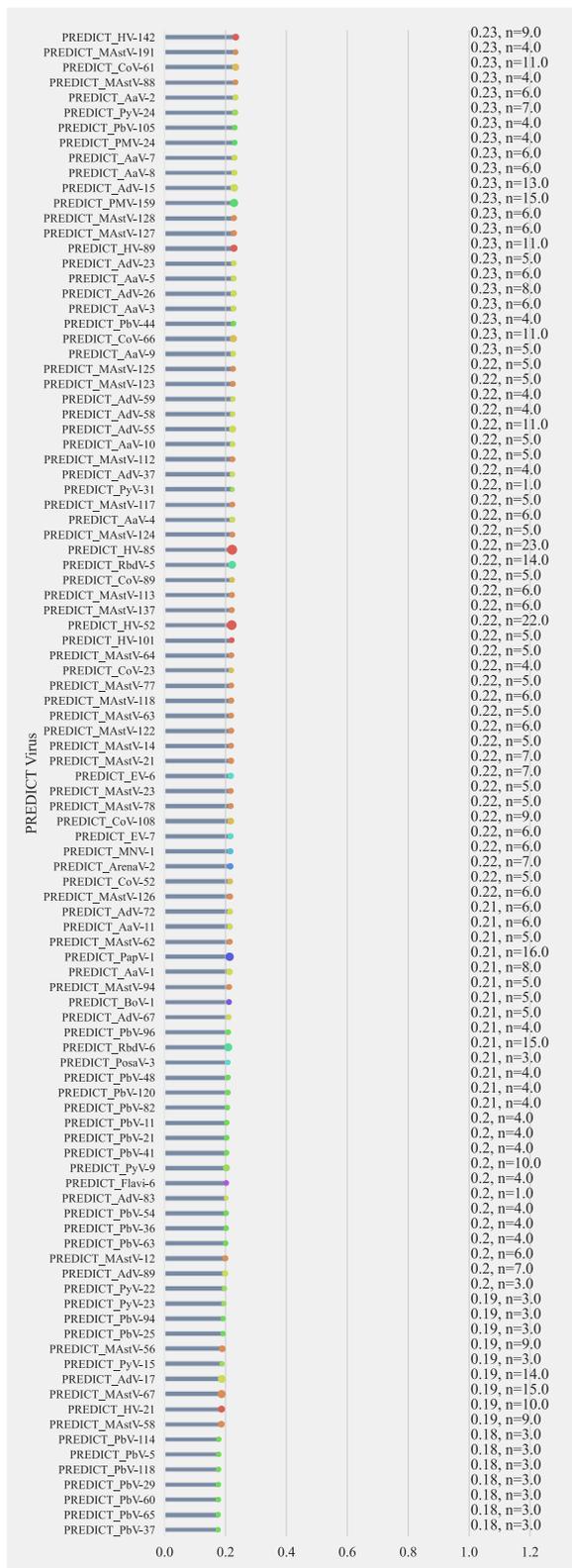
**Fig. S 18:** Prioritization metrics for novel viruses to understand zoonotic risk: Virus prioritization scores based on multiclass model predictions. Annotations show the score and support represented by number of human links predicted (Part 1).



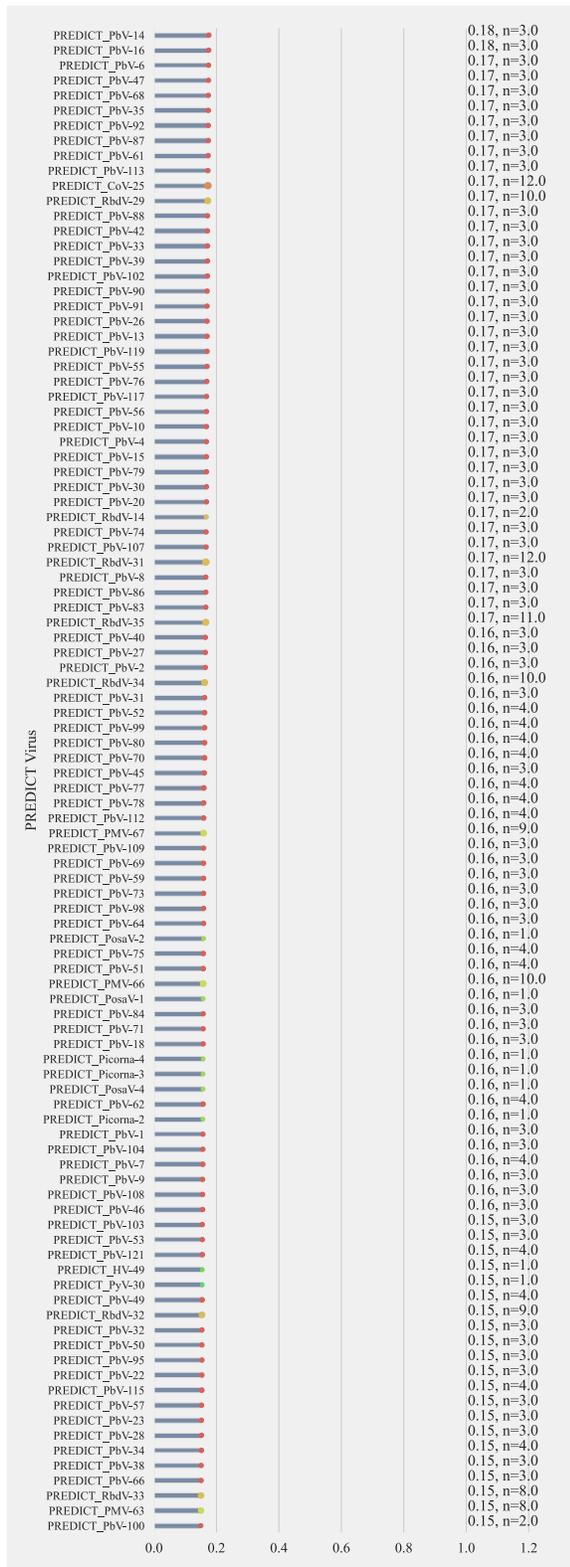
**Fig. S 19: Prioritization metrics for novel viruses to understand zoonotic risk:** Virus prioritization scores based on multiclass model predictions. Annotations show the score and support represented by number of human links predicted (Part 2).



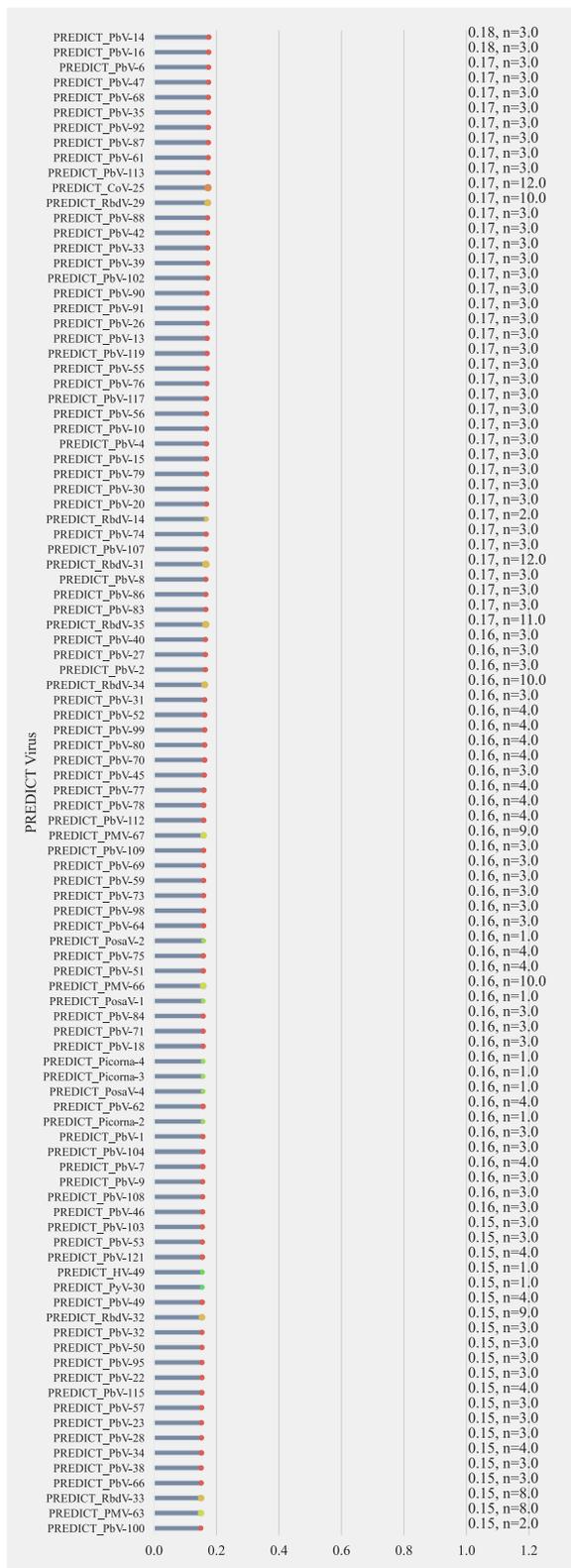
**Fig. S 20: Prioritization metrics for novel viruses to understand zoonotic risk:** Virus prioritization scores based on multiclass model predictions. Annotations show the score and support represented by number of human links predicted (Part 3).



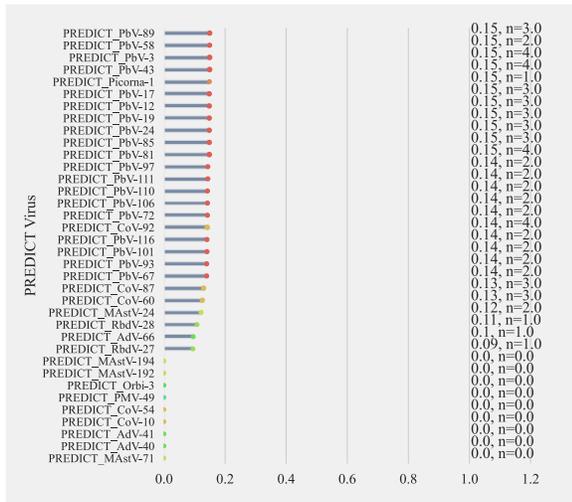
**Fig. S 21: Prioritization metrics for novel viruses to understand zoonotic risk:** Virus prioritization scores based on multiclass model predictions. Annotations show the score and support represented by number of human links predicted (Part 4).



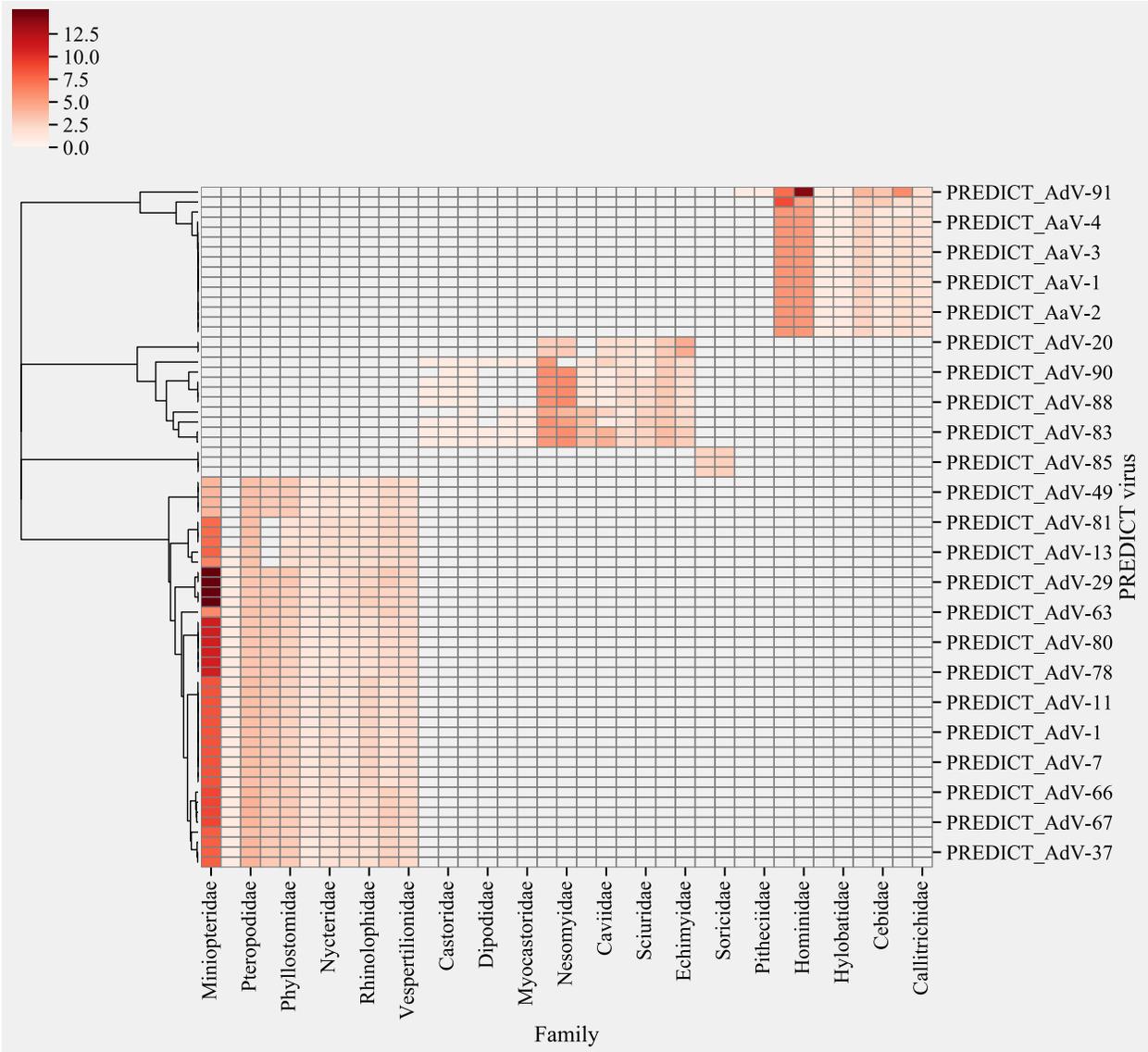
**Fig. S 22: Prioritization metrics for novel viruses to understand zoonotic risk: Virus prioritization scores based on multiclass model predictions. Annotations show the score and support represented by number of human links predicted (Part 5)**



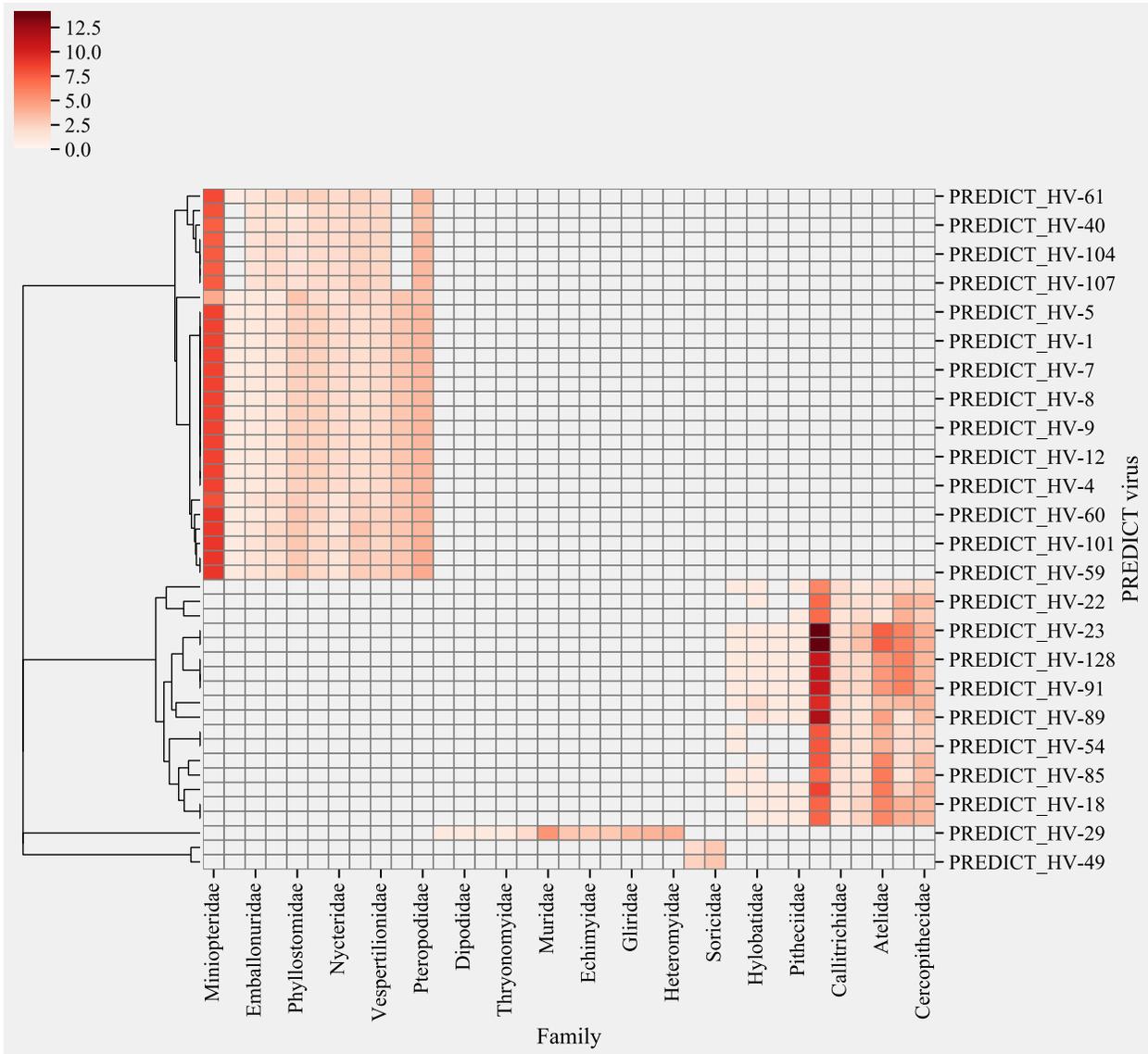
**Fig. S 23: Prioritization metrics for novel viruses to understand zoonotic risk:** Virus prioritization scores based on multiclass model predictions. Annotations show the score and support represented by number of human links predicted (Part 6)



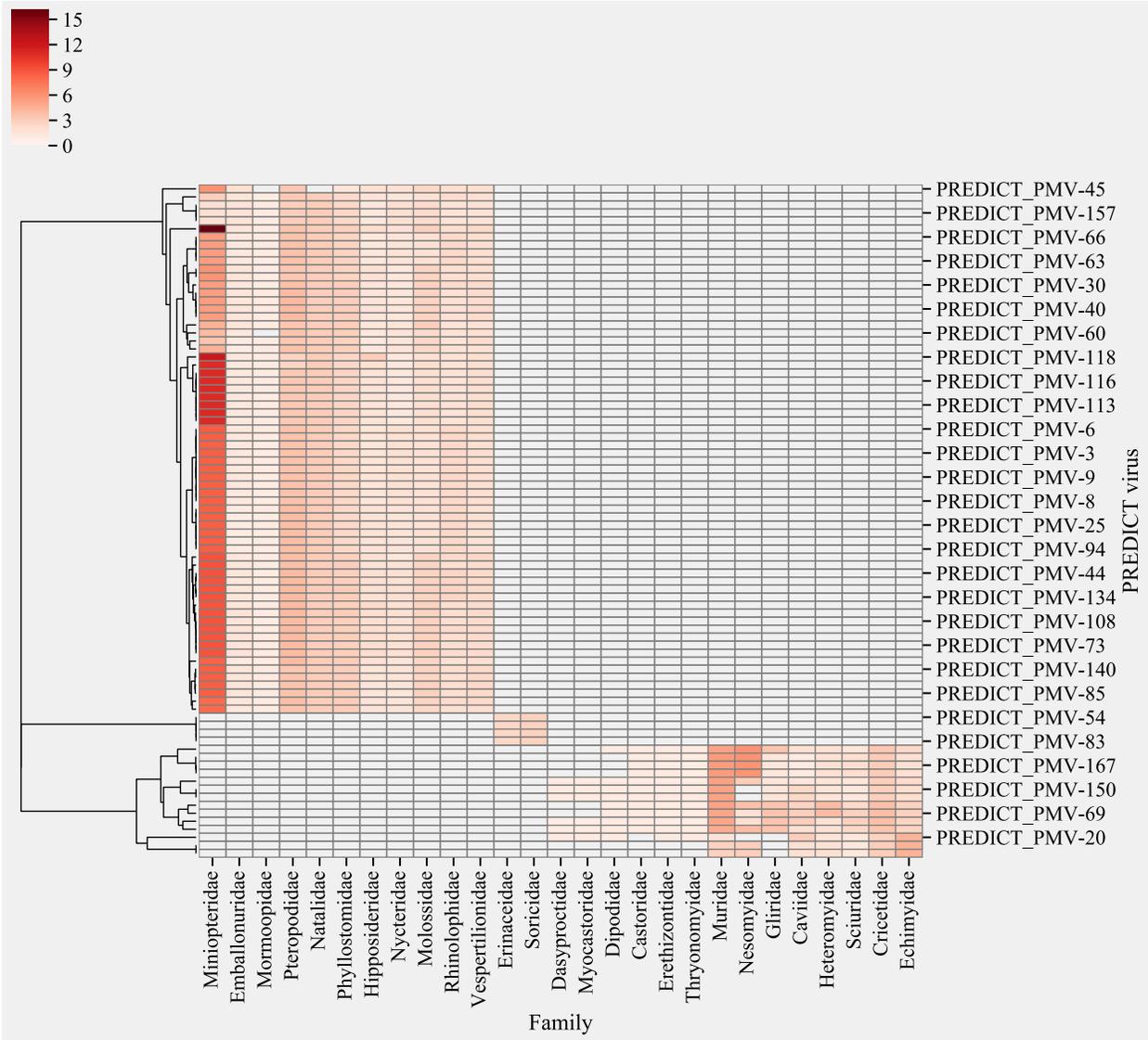
**Fig. S 24: : Prioritization metrics for novel viruses to understand zoonotic risk:** Virus prioritization scores based on multiclass model predictions. Annotations show the score and support represented by number of human links predicted (Part 7).



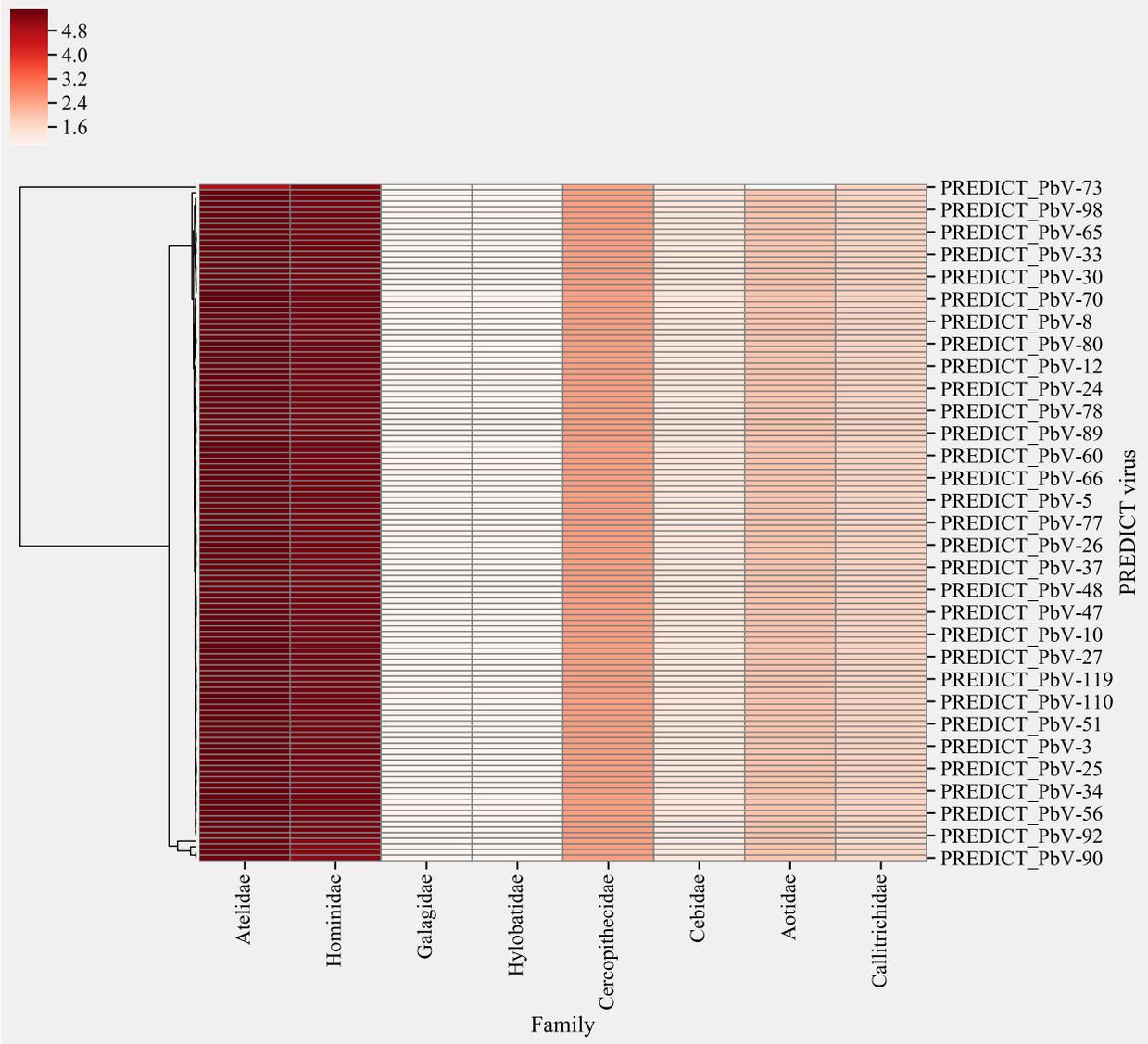
**Fig. S 25:** Surveillance targets for PREDICT adenoviruses based on predicted sharing of hosts with known viruses than their hosts. Red color represents the evidence towards species in the taxonomic family (frequency of links and probability). Clustering is based on Bray-curtis dissimilarity index.



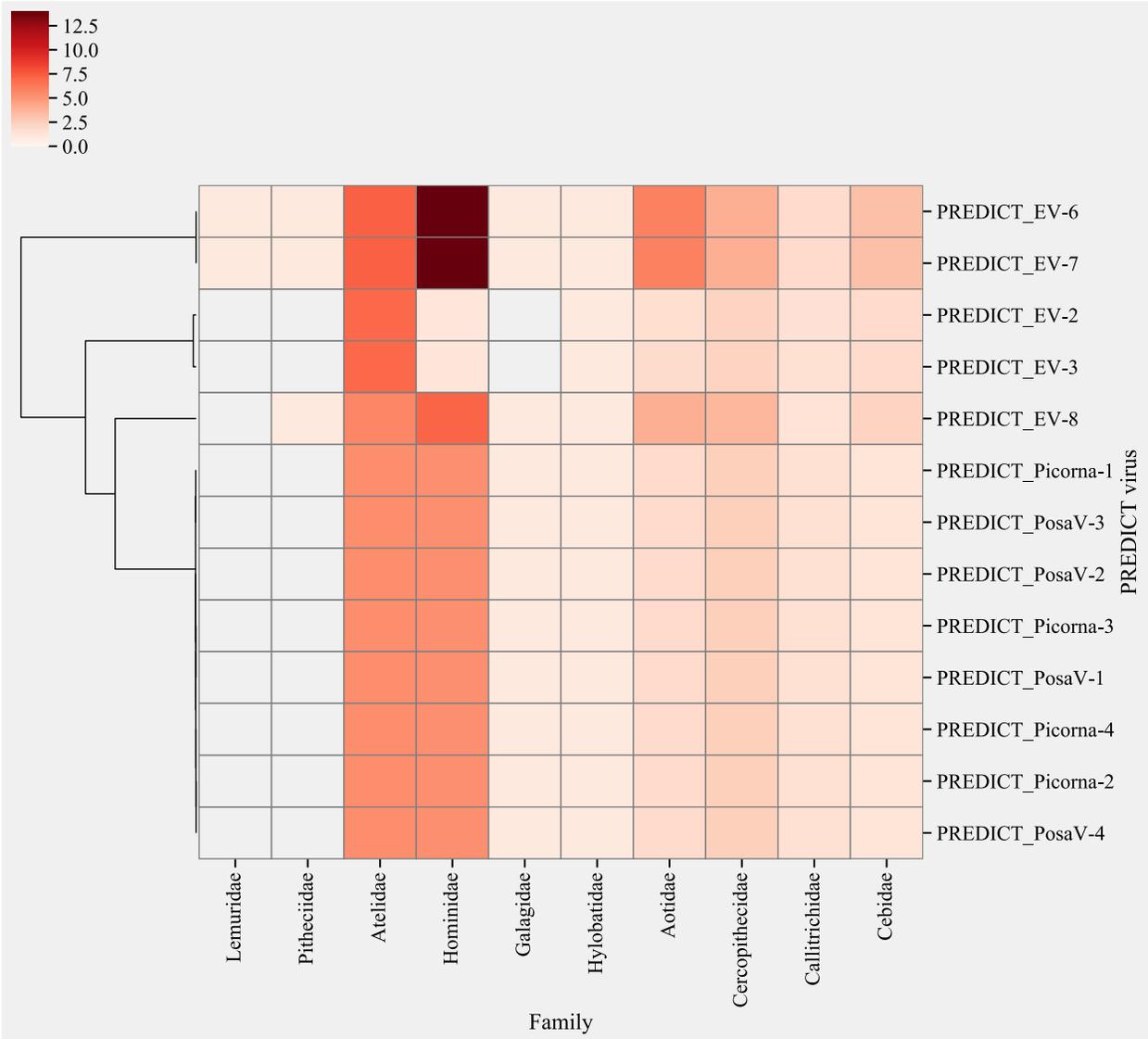
**Fig. S 26:** Surveillance targets for PREDICT herpesviruses based on predicted sharing of hosts with known viruses than their hosts. Red color represents the evidence towards species in the taxonomic family (frequency of links and probability). Clustering is based on Bray-curtis dissimilarity index.



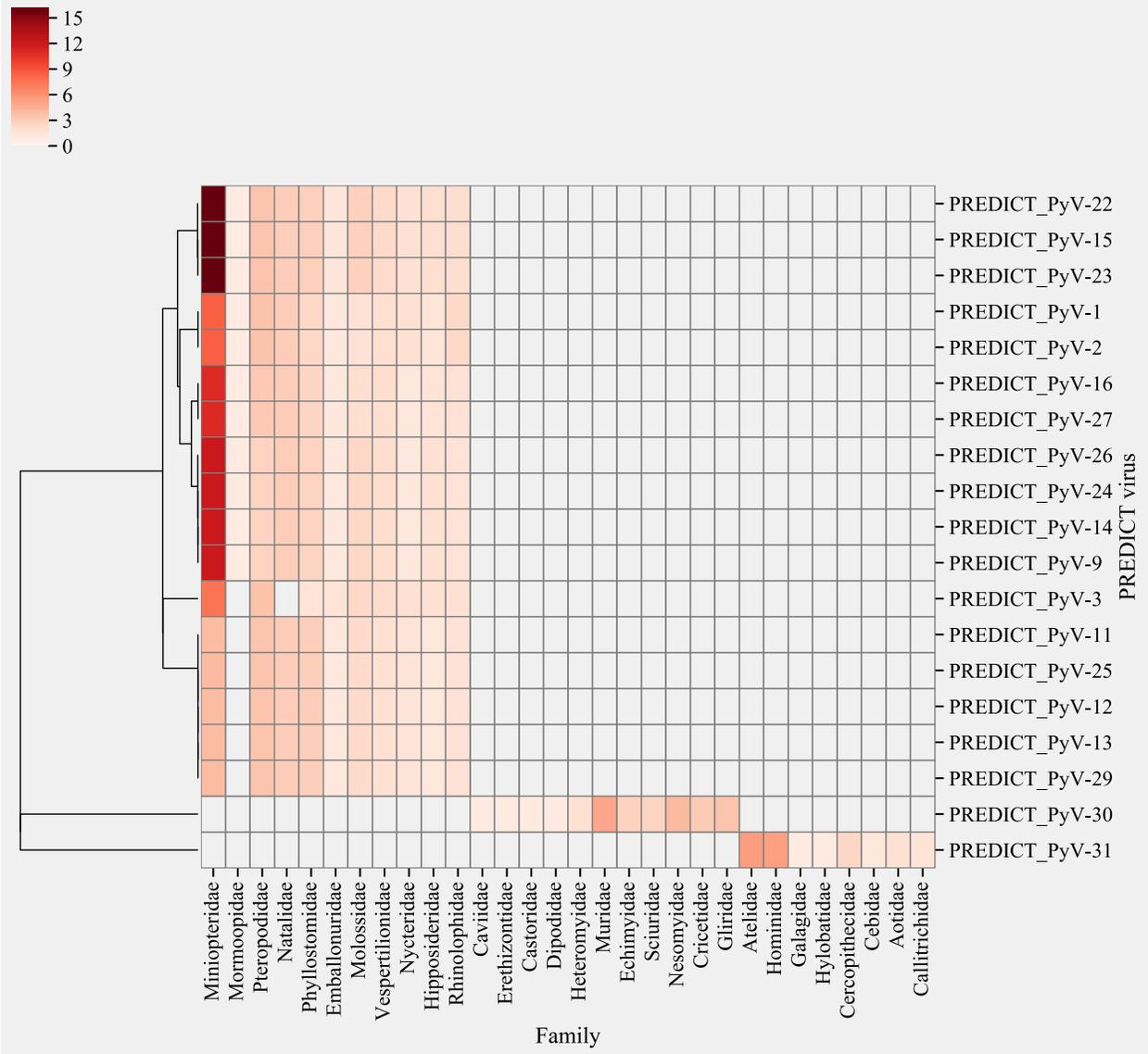
**Fig. S 27:** Surveillance targets for PREDICT paramyxoviruses based on predicted sharing of hosts with known viruses than their hosts. Red color represents the evidence towards species in the taxonomic family (frequency of links and probability). Clustering is based on Bray-curtis dissimilarity index.



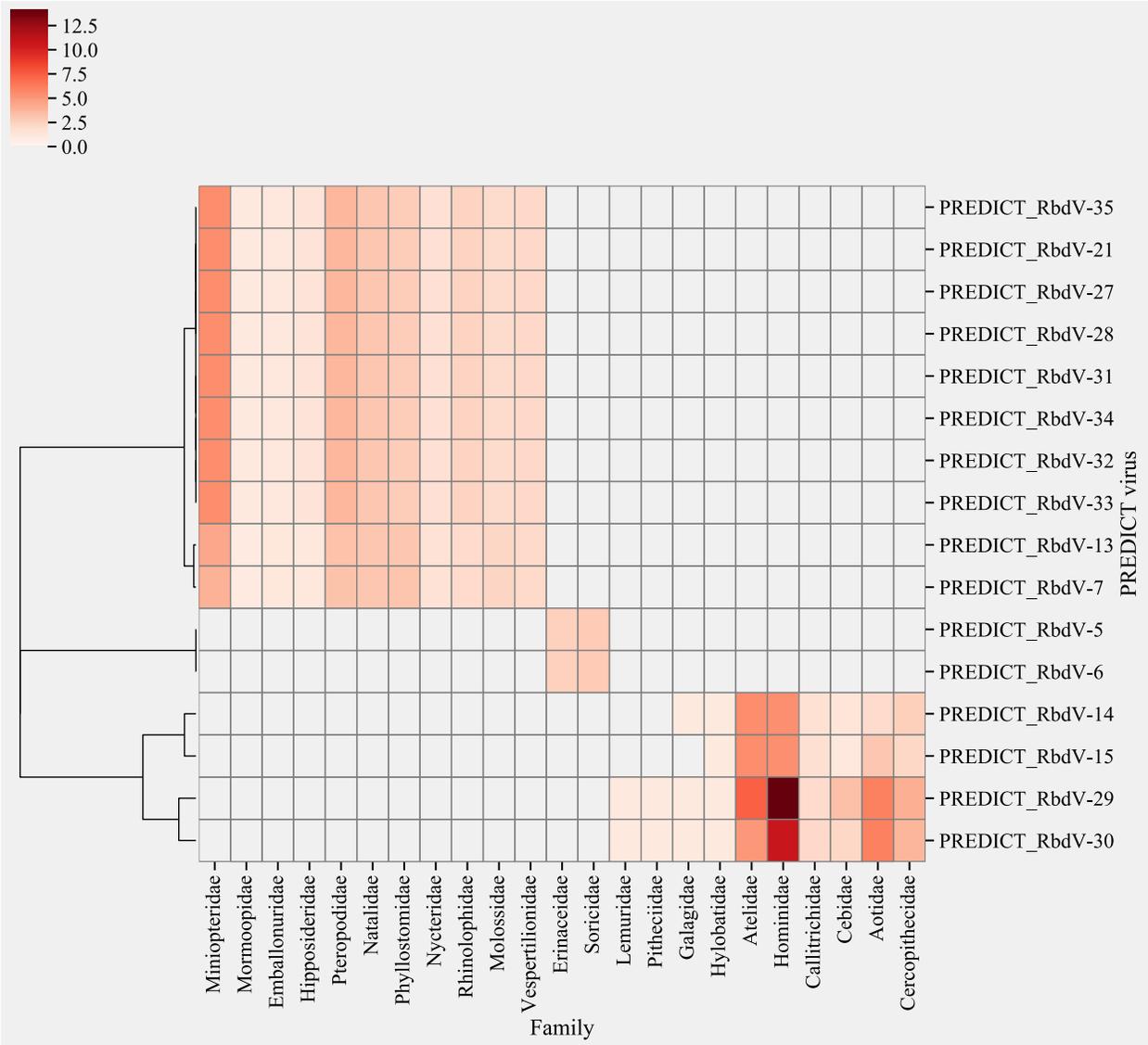
**Fig. S 28:** Surveillance targets for PREDICT picobirnaviruses based on predicted sharing of hosts with known viruses than their hosts. Red color represents the evidence towards species in the taxonomic family (frequency of links and probability). Clustering is based on Bray-curtis dissimilarity index.



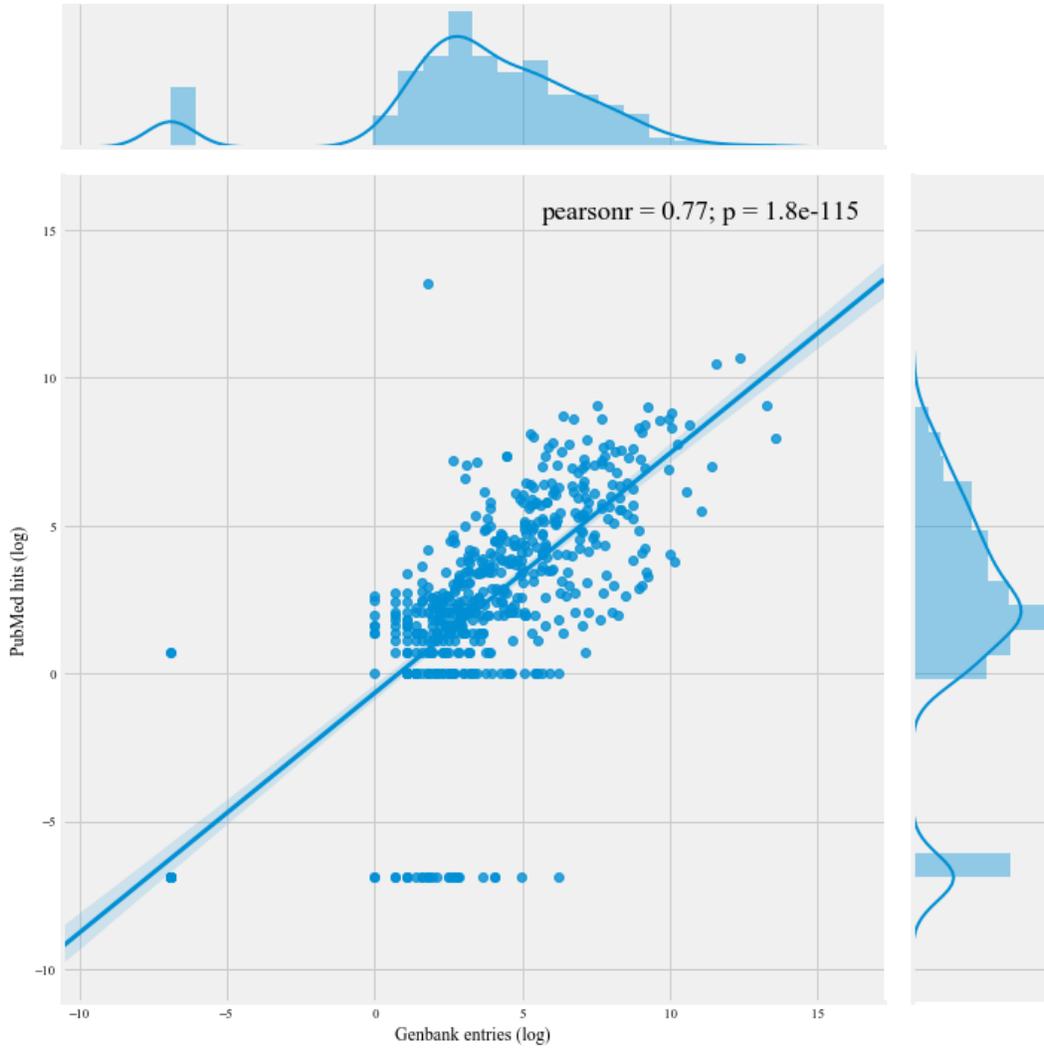
**Fig. S 29:** Surveillance targets for PREDICT picornaviruses based on predicted sharing of hosts with known viruses than their hosts. Red color represents the evidence towards species in the taxonomic family (frequency of links and probability). Clustering is based on Bray-curtis dissimilarity index.



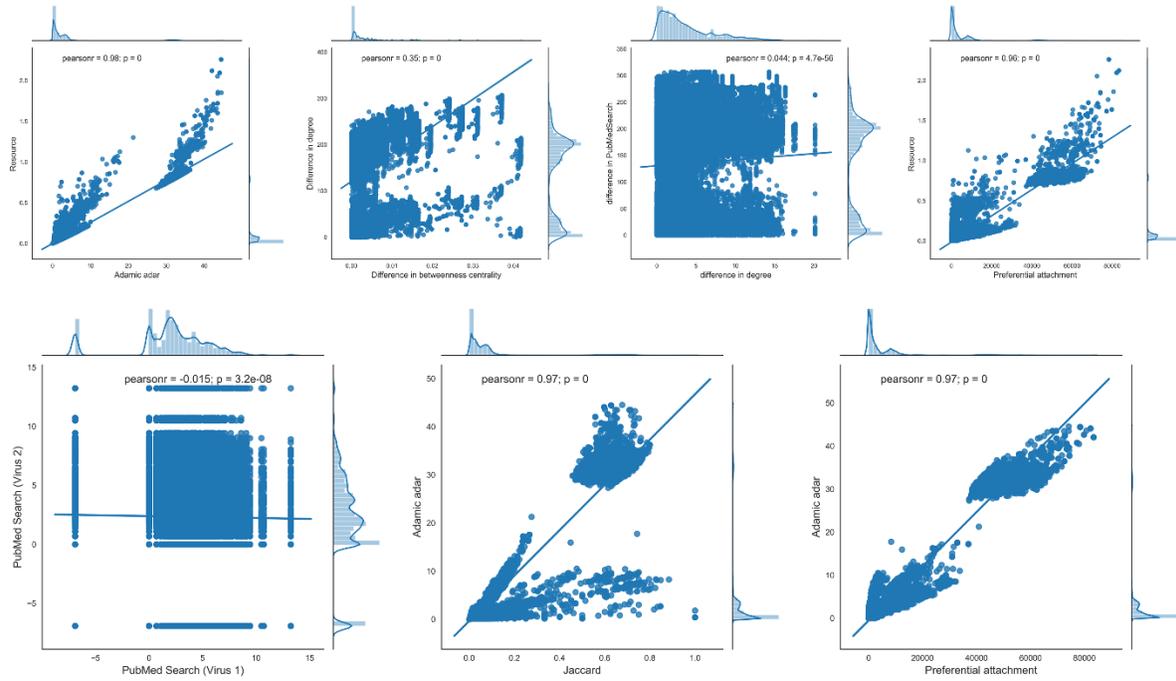
**Fig. S 30:** Surveillance targets for PREDICT polyomaviruses based on predicted sharing of hosts with known viruses than their hosts. Red color represents the evidence towards species in the taxonomic family (frequency of links and probability). Clustering is based on Bray-curtis dissimilarity index.



**Fig. S 31:** Surveillance targets for PREDICT rhabdoviruses based on predicted sharing of hosts with known viruses than their hosts. Red color represents the evidence towards species in the taxonomic family (frequency of links and probability). Clustering is based on Bray-curtis dissimilarity index.



**Fig. S 32:** Pearson Correlation between PubMed hits (PMH1) and GenBank Entries (PMH2) for individual viruses in the known network. Marginal plots show distribution of individual axes values.



**Fig. S 33:** Jointplots showing correlations of network topological features for all node pairs in observed network ( $G_c$ ).

**Number of predicted links of novel viruses with known viruses generated due to sharing of humans as host**

<i>Predictors</i>	<i>Incidence Rate Ratios</i>	<i>CI</i>	<i>p</i>
(Intercept)	41.37	16.16 – 105.89	<b>&lt;0.001</b>
Adenoviridae	<i>Reference</i>		
Arenaviridae	0.48	0.11 – 2.16	0.338
Astroviridae	0.84	0.68 – 1.04	0.113
Caliciviridae	0.79	0.17 – 3.74	0.764
Coronaviridae	0.84	0.66 – 1.07	0.167
Filoviridae	0.35	0.07 – 1.86	0.218
Flaviviridae	0.72	0.36 – 1.45	0.356
Herpesviridae	1.59	1.28 – 1.98	<b>&lt;0.001</b>
Papillomaviridae	1.64	0.49 – 5.48	0.425
Paramyxoviridae	1.12	0.91 – 1.36	0.281
Parvoviridae	0.70	0.14 – 3.51	0.664
Picobirnaviridae	0.53	0.40 – 0.69	<b>&lt;0.001</b>
Picornaviridae	0.46	0.27 – 0.81	<b>0.007</b>
Polyomaviridae	0.75	0.53 – 1.06	0.100
Reoviridae	0.85	0.31 – 2.28	0.741
Retroviridae	1.38	0.38 – 5.02	0.620
Rhabdoviridae	0.56	0.37 – 0.85	<b>0.007</b>
CARNIVORA	<i>Reference</i>		
CETARTIODACTYLA	0.39	0.08 – 2.00	0.258
CHIROPTERA	0.48	0.19 – 1.21	0.119
EULIPOTYPHLA	0.42	0.16 – 1.15	0.092
GALLIFORMES	1.19	0.34 – 4.25	0.784

PRIMATES	0.28	0.11 – 0.73	<b>0.009</b>
RODENTIA	0.52	0.20 – 1.34	0.174
PubMed	1.03	0.98 – 1.08	0.240

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Observations                      535

**Table S 1:** Negative binomial Generalized linear mixed model to understand association between virus family, taxonomic order of the host and PubMed Hists on the predicted number of links of novel viruses generated with known viruses formed due to sharing of humans as hosts  
**Number of human links ~ Virus Family + host Taxonomic Order + PubMed hits, family = negative binomial**

<b>Mean probability of virus links formed due to humans as hosts</b>			
<i>Predictors</i>	<i>Risk Ratios</i>	<i>CI</i>	<i>p</i>
(Intercept)	3.10	1.28 – 9.06	<b>0.020</b>
Adenoviridae	<i>Reference</i>		
Arenaviridae	0.61	0.28 – 1.32	0.198
Astroviridae	0.98	0.87 – 1.12	0.809
Caliciviridae	0.91	0.43 – 1.97	0.799
Coronaviridae	1.00	0.86 – 1.17	0.949
Filoviridae	1.36	0.59 – 3.51	0.492
Flaviviridae	0.98	0.66 – 1.48	0.923
Herpesviridae	1.07	0.92 – 1.25	0.393
Papillomaviridae	0.79	0.37 – 1.69	0.539
Paramyxoviridae	1.17	1.02 – 1.33	<b>0.026</b>
Parvoviridae	0.92	0.43 – 2.01	0.831
Picobirnaviridae	0.79	0.68 – 0.92	<b>0.002</b>
Picornaviridae	1.14	0.89 – 1.47	0.314
Polyomaviridae	1.03	0.84 – 1.27	0.786
Reoviridae	1.34	0.75 – 2.50	0.333
Retroviridae	1.39	0.63 – 3.32	0.435
Rhabdoviridae	0.69	0.56 – 0.85	<b>0.001</b>
CARNIVORA	<i>Reference</i>		
CETARTIODACTYLA	0.21	0.06 – 0.67	<b>0.011</b>
CHIROPTERA	0.58	0.20 – 1.40	0.268
EULIPOTYPHLA	0.53	0.18 – 1.33	0.211
GALLIFORMES	1.26	0.30 – 5.77	0.753
PRIMATES	0.47	0.16 – 1.13	0.121

RODENTIA	0.66	0.23 – 1.61	0.402
PubMed	1.02	0.98 – 1.05	0.346

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Observations                      535

**Table S 2:** Quasibinomial Generalized linear mixed model to understand association between virus family, taxonomic order of the host and PubMed Hists on the predicted mean probability of sharing a human as a link with another virus.

**probability of link being human ~ Virus Family + host Taxonomic Order + PubMed hits,  
family = quasibinomial**

Feature	Calculation/explanation	Variable type
Jaccard coefficient	$\frac{ \Gamma(virus_1) \cap \Gamma(virus_2) }{ \Gamma(virus_1) \cup \Gamma(virus_2) }$ where $\Gamma(virus)$ denotes set of neighbors of <i>virus</i>	Numeric
The difference in degree centrality	The absolute difference in the number of connections (edges) of viruses in the virus pair.	Numeric
The difference in betweenness centrality	Betweenness centrality of a node ( <i>virus</i> ) is the sum of the fraction of all-pairs shortest paths that pass through <i>virus</i> and is calculated as $c_B(virus) = \sum_{s,t \in viruses} \frac{\sigma(s,t virus)}{\sigma(s,t)}$ The difference was calculated as $ c_B(virus_1) - c_B(virus_2) $	Numeric
Same cluster	Modularity partition was estimated using Louvain clustering algorithm to check if both the viruses in the pair fall under the same modularity/cluster	Binary (1, 0)
Viral family virus 1	Taxonomical family of virus 1 in the pair	Categorical
Viral family virus 2	Taxonomical family of virus 2 in the pair	Categorical
Same viral families	A binary variable depicting if viruses in the pair belong to the same taxonomical family.	Binary (1, 0)
Difference in PubMedHits	The absolute difference in the number of sequences of both viruses in a pair detected in PubMed	Numeric
PubMedHits virus 1 (log)	Number of nucleotide sequences in PubMed for virus 1 in the pair	Numeric
PubMedHits virus 2 (log)	Number of nucleotide sequences in PubMed for virus 2 in the pair	Numeric

**Table S 3.** Network features, their definitions, and type of feature used in the training of GBM models.

Feature	Calculation/explanation	Variable type
Jaccard coefficient	$\frac{ \Gamma(virus_1) \cap \Gamma(virus_2) }{ \Gamma(virus_1) \cup \Gamma(virus_2) }$ where $\Gamma(virus)$ denotes set of neighbors of <i>virus</i>	Numeric
The difference in degree centrality	The absolute difference in the number of connections (edges) of viruses in the virus pair.	Numeric
The difference in betweenness centrality	Betweenness centrality of a node ( <i>virus</i> ) is the sum of the fraction of all-pairs shortest paths that pass through <i>virus</i> and is calculated as $c_B(virus) = \sum_{s,t \in viruses} \frac{\sigma(s,t virus)}{\sigma(s,t)}$ The difference was calculated as $ c_B(virus_1) - c_B(virus_2) $	Numeric
Same cluster	Modularity partition was estimated using Louvain clustering algorithm to check if both the viruses in the pair fall under the same modularity/cluster	Binary (1, 0)
Viral family virus 1	Taxonomical family of virus 1 in the pair	Categorical
Viral family virus 2	Taxonomical family of virus 2 in the pair	Categorical
Same viral families	A binary variable depicting if viruses in the pair belong to the same taxonomical family.	Binary (1, 0)
Difference in PubMedHits	The absolute difference in the number of sequences of both viruses in a pair detected in PubMed	Numeric
PubMedHits virus 1 (log)	Number of nucleotide sequences in PubMed for virus 1 in the pair	Numeric
PubMedHits virus 2 (log)	Number of nucleotide sequences in PubMed for virus 2 in the pair	Numeric

**Table S 4.** Network features, their definitions, and type of feature used in the training of GBM models.

<b>XGboost parameter</b>	<b>Parameter explanation</b>	<b>Parameter values tested</b>
Learning rate	Step size shrinkage used in the update to prevents overfitting.	0.05, 0.10, 0.20, 0.30
Max depth	Maximum depth of a tree.	3, 4, 8, 12, 15
Minimum child weight	The minimum sum of instance weight (hessian) needed in a child.	1, 3, 5, 7
Gamma	Minimum loss reduction that is required to make a further partition on a leaf node of the tree. The larger gamma is, the more conservative the algorithm will be.	0.0, 0.1, 0.3, 0.5
Colsample by tree	Parameters for subsampling of columns.	0.3, 0.5 , 0.7

**Table S 5:** Hyper-tuning of XGboost model parameters. The parameter name and values testes in the grid search to find the best performing parameters for the model.

<b>Country</b>	<b>Animal Class</b>	<b>No. of Animals</b>
<b>Bangladesh</b>	<b>Domestic</b>	956
	<b>Wildlife</b>	8120
<b>Bolivia</b>	<b>Wildlife</b>	717
<b>Brazil</b>	<b>Wildlife</b>	826
<b>Cambodia</b>	<b>Domestic</b>	894
	<b>Wildlife</b>	5084
<b>Cameroon</b>	<b>Wildlife</b>	7379
<b>China</b>	<b>Wildlife</b>	4912
<b>DR Congo</b>	<b>Domestic</b>	34
	<b>Wildlife</b>	4277
<b>Egypt</b>	<b>Wildlife</b>	1003
<b>Ethiopia</b>	<b>Wildlife</b>	836
<b>Gabon</b>	<b>Wildlife</b>	3127
<b>Ghana</b>	<b>Wildlife</b>	2784
<b>Guinea</b>	<b>Domestic</b>	95
	<b>Wildlife</b>	4411
<b>India</b>	<b>Wildlife</b>	19
<b>Indonesia</b>	<b>Wildlife</b>	2956
<b>Ivory Coast</b>	<b>Wildlife</b>	776
<b>Jordan</b>	<b>Wildlife</b>	1080
<b>Kenya</b>	<b>Domestic</b>	100
	<b>Wildlife</b>	811
<b>Lao PDR</b>	<b>Domestic</b>	479
	<b>Wildlife</b>	3068
<b>Liberia</b>	<b>Wildlife</b>	3795
<b>Malaysia, Peninsular</b>	<b>Domestic</b>	670
	<b>Wildlife</b>	1962
<b>Malaysia, Sabah</b>	<b>Wildlife</b>	2672
<b>Mexico</b>	<b>Wildlife</b>	1074
<b>Mongolia</b>	<b>Domestic</b>	2
	<b>Wildlife</b>	3236
<b>Myanmar</b>	<b>Wildlife</b>	1062
<b>Nepal</b>	<b>Wildlife</b>	3228
<b>Peru</b>	<b>Wildlife</b>	425
<b>Republic of Congo</b>	<b>Wildlife</b>	999
<b>Rwanda</b>	<b>Wildlife</b>	1341
<b>Senegal</b>	<b>Wildlife</b>	874
<b>Sierra Leone</b>	<b>Domestic</b>	267
	<b>Wildlife</b>	7587
<b>South Sudan</b>	<b>Wildlife</b>	271

<b>Tanzania</b>	<b>Domestic</b>	372
	<b>Wildlife</b>	4578
<b>Thailand</b>	<b>Domestic</b>	205
	<b>Wildlife</b>	3819
<b>Uganda</b>	<b>Domestic</b>	94
	<b>Wildlife</b>	1630
<b>Vietnam</b>	<b>Domestic</b>	488
	<b>Wildlife</b>	3984

**Table S 6:** Number of wild and domestic animals sampled for detection of viruses in 34 countries.

<b>Virus Family / Taxonomic unit</b>	<b>PCR tests: protocols used</b>	<b>Number of Tests</b>
<b>Adenoviridae</b>	<b>Casas, Hexon gene</b>	2875
	<b>Wellehan et al, Pol gene</b>	6632
	<b>other-Casas, Hexon gene</b>	88
	<b>other-Felix Dressler, Bonn Institute Protocol</b>	13
	<b>other-Heim et al., 2003</b>	262
	<b>other-Wellehan et al, Pol gene</b>	88
<b>Arenaviridae</b>	<b>Lonzano et al, S gene</b>	26390
	<b>Unpublished UCD</b>	2159
	<b>other-</b>	833
	<b>other-Drosten et al., 2007</b>	191
	<b>other-Eric Leroy, CIRMF Protocol</b>	12
	<b>other-UCD unpublished</b>	48
	<b>other-Vieth et al., 2007 (Modified)</b>	2527
	<b>other-Vieth et al., L-gene</b>	2558
<b>Astroviridae</b>	<b>Atkins et al, RdRp gene</b>	6255
	<b>Chu et al, RdRp gene</b>	10763
	<b>other</b>	130
	<b>other-Atkins et al, RdRp gene</b>	88
	<b>other-Chu et al, RdRp gene</b>	88
	<b>other-Felix Dressler, Bonn Institute Protocol</b>	6
	<b>other-Logan et al., 2007</b>	262
<b>Bunyavirales</b>	<b>Briese et al, L gene</b>	14390
	<b>Briese et al, M gene</b>	9122
	<b>Briese et al, S gene</b>	10605
	<b>other-</b>	1276
	<b>other-Briese et al, S, M and L segments</b>	88
	<b>other-UCD unpublished</b>	51
<b>Caliciviridae</b>	<b>Reid et al, Capsid</b>	458
	<b>other-Felix Dressler, Bonn Institute Protocol</b>	6
	<b>other-Hoehne et al., 2006</b>	262
	<b>other-Logan et al., 2007</b>	262
<b>Coronaviridae</b>	<b>Ge et al., RdRp</b>	153
	<b>Modified Watanabe et al, RdRp gene</b>	133872
	<b>Quan et al, RdRp gene</b>	134105
	<b>Quan   Watanabe</b>	60
	<b>Tong S, et al. EID 2009, RdRp gene</b>	120
	<b>other</b>	167
	<b>other-</b>	1630
	<b>other-CDC</b>	822
	<b>other-Dare et al., 2007</b>	3359
	<b>other-Felix Dressler, Bonn Institute Protocol</b>	10

	<b>other-de Souza Luna et al, 2007, RdRp gene</b>	1483
<b>Filoviridae</b>	<b>Goldstein</b>	27136
	<b>Jaaskelainen et al, L gene</b>	33208
	<b>Modified Zhai et al, L gene</b>	70
	<b>Modified Zhai et al, L gene, Filo A</b>	8142
	<b>Modified Zhai et al, L gene, Filo A   B   C   D</b>	7710
	<b>Modified Zhai et al, L gene, Filo B</b>	8115
	<b>Modified Zhai et al, L gene, Filo C</b>	7999
	<b>Modified Zhai et al, L gene, Filo D</b>	8003
	<b>Modified Zhai et al, L gene, Filo MOD</b>	16049
	<b>Towner et al, NP gene</b>	41929
	<b>Unpublished CII</b>	35711
	<b>Unpublished, UCD</b>	154833
	<b>other</b>	417
	<b>other-</b>	1615
	<b>other-Eric Leroy, CIRMF Protocol</b>	12
	<b>other-Grard et al., 2011</b>	1719
	<b>other-Grolla, PHAC</b>	3
	<b>other-New filo</b>	51
	<b>other-Public Health Agency Canada</b>	556
	<b>other-Towner et al., 2006</b>	1719
<b>Flaviviridae</b>	<b>Modified Moureau et al, NS5 gene</b>	28326
	<b>Moureau et al, NS5 gene</b>	86391
	<b>Sanchez-Seco et al, NS5 gene</b>	5045
	<b>Unpublished UCD, NS5 gene</b>	1295
	<b>other-</b>	1556
	<b>other-Eric Leroy, CIRMF Protocol</b>	12
	<b>other-Lanciotti et al 1992</b>	1
	<b>other-Lanciotti_ West Nile virus</b>	151
	<b>other-Linssen, 2000</b>	1
	<b>other-Makuwa, M., Souquire, S., Clifford, S. L., Mouinga-Ondeme, A., Bawe-Johnson, M., Wickings, E. J., Latour, S., Simon, F. &amp; Roques, P. (2005). Identification of hepatitis B virus genome in faecal sample from wild living chimpanzee (Pan troglodytes troglodytes) in Gabon. J Clin Virol 34, S83-S88.</b>	156
	<b>other-Peter Simmonds protocol</b>	555
	<b>other-Sharp/Simmonds</b>	1
	<b>Hantaviridae</b>	<b>Aitichou et al (2005)</b>
<b>Klempa et al, L-Segment</b>		5684
<b>Klempa et al, N gene</b>		7479
<b>Modified Aitichou et al, S gene</b>		16111

	<b>Modified Raboni et al, S gene</b>	8061
	<b>Modified Raboni et al, S segment</b>	1999
	<b>Modified Zhai et al, L gene, Filo D</b>	2
	<b>other</b>	17
	<b>other-</b>	1224
	<b>other-Araujo et al., 2011</b>	108
	<b>other-PCR-ICB, S gene</b>	68
	<b>other-qPCR-ICB, S gene</b>	74
<b>Hepadnairidae</b>	<b>other-Hepatitis A virus</b>	38
	<b>other-Hepatitis B virus</b>	38
<b>Herpesviridae</b>	<b>Chmielewicz et al, Term gene</b>	6232
	<b>Van DeVanter et al, Pol gene</b>	8290
	<b>other-</b>	773
	<b>other-Garner et al. 2009</b>	30
	<b>other-Herpes B</b>	828
	<b>other-Latimer et al. 2011</b>	90
<b>Orthomyxoviridae</b>	<b>Anthony et al, M gene</b>	115476
	<b>Liang et al, PB1 gene</b>	5278
	<b>Liang unpublished CII, PB1 gene</b>	110346
	<b>Unpublished CII, Influenza A, PB1 gene</b>	2458
	<b>other-</b>	2094
	<b>other-Gerloff 2016</b>	4890
	<b>other-Liang</b>	51
	<b>other-Liang et al, PB1 gene</b>	87
	<b>other-Liang unpublished CII, PB1 gene</b>	573
	<b>other-Protocol provided by Laboratory at National Institute of Infectious Diseases (NIID)</b>	4189
	<b>other-Unpublished CII, Influenza A, PB1 gene</b>	12
	<b>other-WHO-Avian Influenza A</b>	6
	<b>other-Wang, W., et al. 2009</b>	162
<b>Papillomaviridae</b>	<b>Forslund et al, L1 gene</b>	558
<b>Paramyxoviridae</b>	<b>Modified Feldman et al, L gene</b>	12582
	<b>Tong et al, PAR</b>	110940
	<b>Tong et al, Pol gene</b>	31542
	<b>Tong et al, RES</b>	5575
	<b>Wacharapluesadee et al, N gene</b>	5214
	<b>Wacharapluesadee et all, N gene</b>	587
	<b>other</b>	167
	<b>other-</b>	1371
	<b>other-Eric Leroy, CIRMF Protocol</b>	10
	<b>other-Gerloff 2016</b>	331

	<b>other-Nested conventional PCR targeting on the L-gene</b>	88
	<b>other-Protocol of HUA National Key Laboratory of Veterinary Biotechnology</b>	46
	<b>other-Tong, S et al, (2008)</b>	306
	<b>other-Unpublished methodology</b>	2324
	<b>other-adapted from Feldman et al., 2009</b>	822
<b>Parvoviridae</b>	<b>Kapoor et al, NS1 gene</b>	5001
	<b>other-Kapoor et al, NS1 gene</b>	88
	<b>other-Peter Simmonds protocol</b>	212
	<b>other-Sharp CP, LeBreton M, Kantola K, Nana A, Dikko Jle D, Djoko CF, Tamoufe U, Kiyang JA, Babila TG, Ngole EM, Pybus OG, Delwart E, Delaporte E, Peeters M, Soderlund-Venermo M, Hedman K, Wolfe ND, Simmonds P. (2010) Widespread infection with homologues of human parvoviruses B19, PARV4, and human bocavirus of chimpanzees and gorillas in the wild. Journal of Virology 84(19):10289-96.</b>	490
<b>Peribunyaviridae</b>	<b>Briese et al, L segment</b>	439
	<b>Unpublished UCD, L segment</b>	7275
<b>Phenuiviridae</b>	<b>Modified Sanchez-Seco et al, L gene</b>	1320
	<b>other-Drosten et al., 2002</b>	370
	<b>other-Modified Sanchez-Seco et al, L gene</b>	88
<b>Picornaviridae</b>	<b>Nix et al, VP3/VP1</b>	1401
	<b>Unpublished CII, 5 UTR</b>	4470
	<b>Unpublished CII, UTR gene</b>	2
	<b>Unpublished CII, VP4/2 gene</b>	1289
	<b>other-Breuer et al, VP2 gene</b>	3
	<b>other-Dierssen et al., 2007</b>	262
	<b>other-Encephalomyocarditis virus</b>	38
	<b>other-Felix Dressler, Bonn Institute Protocol</b>	25
	<b>other-Nix W.A. (2006), VP1 gene</b>	4
	<b>other-Nix W.A. 2006, VP1 gene</b>	10
	<b>other-Nix et al, VP3/VP1</b>	91
	<b>other-Nix, W.A. 2006</b>	6
	<b>other-Unpublished CII</b>	210
	<b>other-Unpublished CII, 5 UTR</b>	88
	<b>other-Unpublished CII, VP4/2 gene</b>	88
<b>Polyomaviridae</b>	<b>Johne, VP1 gene</b>	5345
	<b>other-Johne, VP1 gene</b>	88
<b>Poxviridae</b>	<b>Bracht et al, Pol and Topoisomerase genes</b>	370
	<b>Bracht et al, Topoisomerase gene</b>	1876

	<b>Bracht et al., Pol gene</b>	963
	<b>Inoshima et al, B2L gene</b>	3453
	<b>Nitsche et al, RdRp gene</b>	6660
	<b>Nollens et al, p42K gene</b>	95
	<b>other-Eric Leroy, CIRMF Protocol</b>	10
	<b>other-Inoshima et al, B2L gene</b>	88
	<b>other-Yu Li et al</b>	415
<b>Reoviridae</b>	<b>Palacios et al, VP1 gene</b>	1684
	<b>Unpublished CII, VP1 gene</b>	9126
	<b>other-</b>	750
	<b>other-Bourhy, H. et al</b>	519
	<b>other-Felix Dressler, Bonn Institute Protocol</b>	6
	<b>other-Hoehne et al., 2006</b>	262
	<b>other-Jothikumar, N., Kang, G., &amp; Hill, V. R. (2009)</b>	478
	<b>other-Palacios et al, VP1 gene</b>	88
	<b>other-Unpublished CII, VP1 gene</b>	88
	<b>other-Wellehan JF et al. , RdRp gene</b>	61
<b>Retroviridae</b>	<b>Courgnaud et al, Pol gene</b>	3221
	<b>Goldberg et al, LTR</b>	1409
	<b>Goldberg et al, Pol gene</b>	2562
	<b>other-</b>	390
	<b>other-Clewley et al, RdRp gene</b>	458
	<b>other-Courgnaud et al, Pol gene</b>	88
	<b>other-Duong et al</b>	84
	<b>other-Goldberg et al, Pol gene</b>	88
	<b>other-Vallejo et al, tax gene</b>	7
<b>Rhabdoviridae</b>	<b>Modified Vazquez-Moron et al, N gene</b>	1558
	<b>Unpublished CII, L pol gene</b>	21773
	<b>other-</b>	1084
	<b>other-Bourhy, H. et al</b>	1981
	<b>other-Grard, Plos Pathogens 2012, L gene</b>	1198
	<b>other-Institute Pasteur Paris, pol gene</b>	6869
	<b>other-Modified Vazquez-Moron et al, N gene</b>	88
	<b>other-Towner et al., 2006</b>	1641
	<b>other-Unpublished methodology</b>	419
<b>Togaviridae</b>	<b>Modified Sanchez-Seco et al, NSP4 gene</b>	7583
	<b>Sanchez-Seco et al, NS5 gene</b>	1
	<b>Sanchez-Seco et al, NSP4 gene</b>	2442
	<b>other-</b>	937
	<b>other-Eric Leroy, CIRMF Protocol</b>	12
	<b>other-Grywna et al., 2010</b>	3770
	<b>other-Lanciotti, 2000</b>	1

	<b>other-Linssen 2000_Eastern equine encephalitis virus</b>	151
	<b>other-Linssen 2000_Venezuelan equine encephalitis virus</b>	151
	<b>other-Linssen 2000_Western equine encephalitis virus</b>	151
	<b>other-PREDICT P-011</b>	64

**Table S 7:** Number of tests performed using various testing protocols of Conventional PCR, Conventional RT-PCR, Real-time PCR, and Next Generation Sequencing for detection of viruses in wildlife and domestic animals.

<i>Predictors</i>	<b>PubMed Search (Log)</b>		
	<i>Estimates</i>	<i>CI</i>	<i>p</i>
(Intercept)	-2.79	-4.05 – -1.53	<b>&lt;0.001</b>
Adenoviridae	<i>Reference</i>		
Arenaviridae	-0.18	-1.74 – 1.38	0.821
Arteriviridae	4.46	1.10 – 7.82	<b>0.009</b>
Asfarviridae	3.72	-1.86 – 9.30	0.191
Astroviridae	2.30	-0.67 – 5.26	0.129
Caliciviridae	0.82	-1.05 – 2.70	0.390
Circoviridae	2.40	-1.63 – 6.43	0.244
Coronaviridae	2.98	1.04 – 4.91	<b>0.003</b>
Filoviridae	3.80	1.78 – 5.82	<b>&lt;0.001</b>
Flaviviridae	2.43	1.05 – 3.81	<b>0.001</b>
Hantaviridae	-1.18	-2.84 – 0.48	0.163
Hepadnaviridae	4.10	1.13 – 7.07	<b>0.007</b>
Herpesviridae	1.08	-0.31 – 2.46	0.128
Nairoviridae	2.23	-0.48 – 4.94	0.106
Orthomyxoviridae	3.02	0.48 – 5.55	<b>0.020</b>
Papillomaviridae	1.15	-0.32 – 2.61	0.125

Paramyxoviridae	3.76	2.03 – 5.49	< <b>0.001</b>
Parvoviridae	3.49	1.68 – 5.29	< <b>0.001</b>
Peribunyaviridae	2.42	0.91 – 3.93	<b>0.002</b>
Phenuiviridae	3.88	1.50 – 6.25	<b>0.001</b>
Picobirnaviridae	1.24	-4.33 – 6.80	0.663
Picornaviridae	1.82	0.13 – 3.51	<b>0.035</b>
Pneumoviridae	3.35	0.37 – 6.33	<b>0.028</b>
Polyomaviridae	3.85	1.58 – 6.11	<b>0.001</b>
Poxviridae	3.89	2.28 – 5.50	< <b>0.001</b>
Reoviridae	1.83	0.15 – 3.51	<b>0.033</b>
Retroviridae	2.89	1.29 – 4.50	< <b>0.001</b>
Rhabdoviridae	2.73	1.17 – 4.29	<b>0.001</b>
Tobaniviridae	2.91	-0.45 – 6.26	0.090
Togaviridae	3.77	2.15 – 5.40	< <b>0.001</b>
GenBank Hits (Log)	0.72	0.61 – 0.82	< <b>0.001</b>
Observations	530		

**Table S 8:** Gaussian Generalized linear mixed model to understand association between virus family and PubMed Hits on the PubMed Search hits.

**PubMed Search (Log)~ Virus Family + PubMed hits, family = Gaussian**

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