

Supporting Information

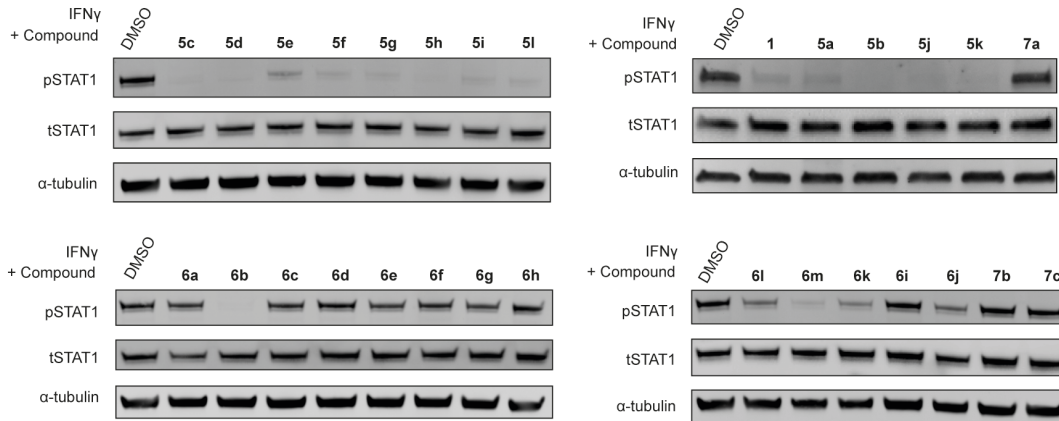
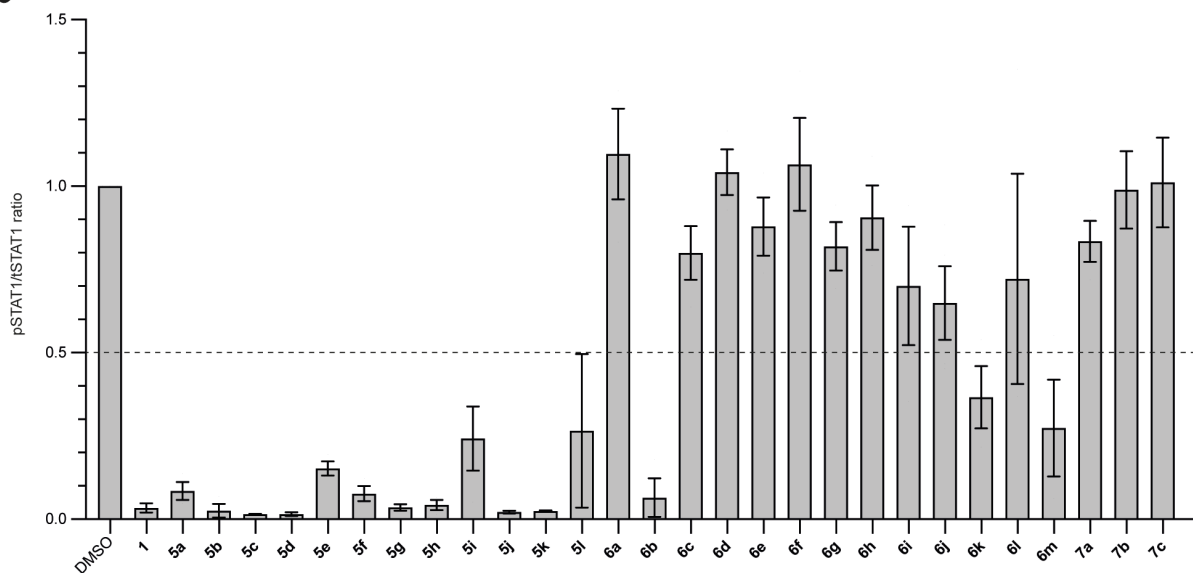
Small Molecule Inhibitors of Interferon-Induced JAK-STAT Signalling

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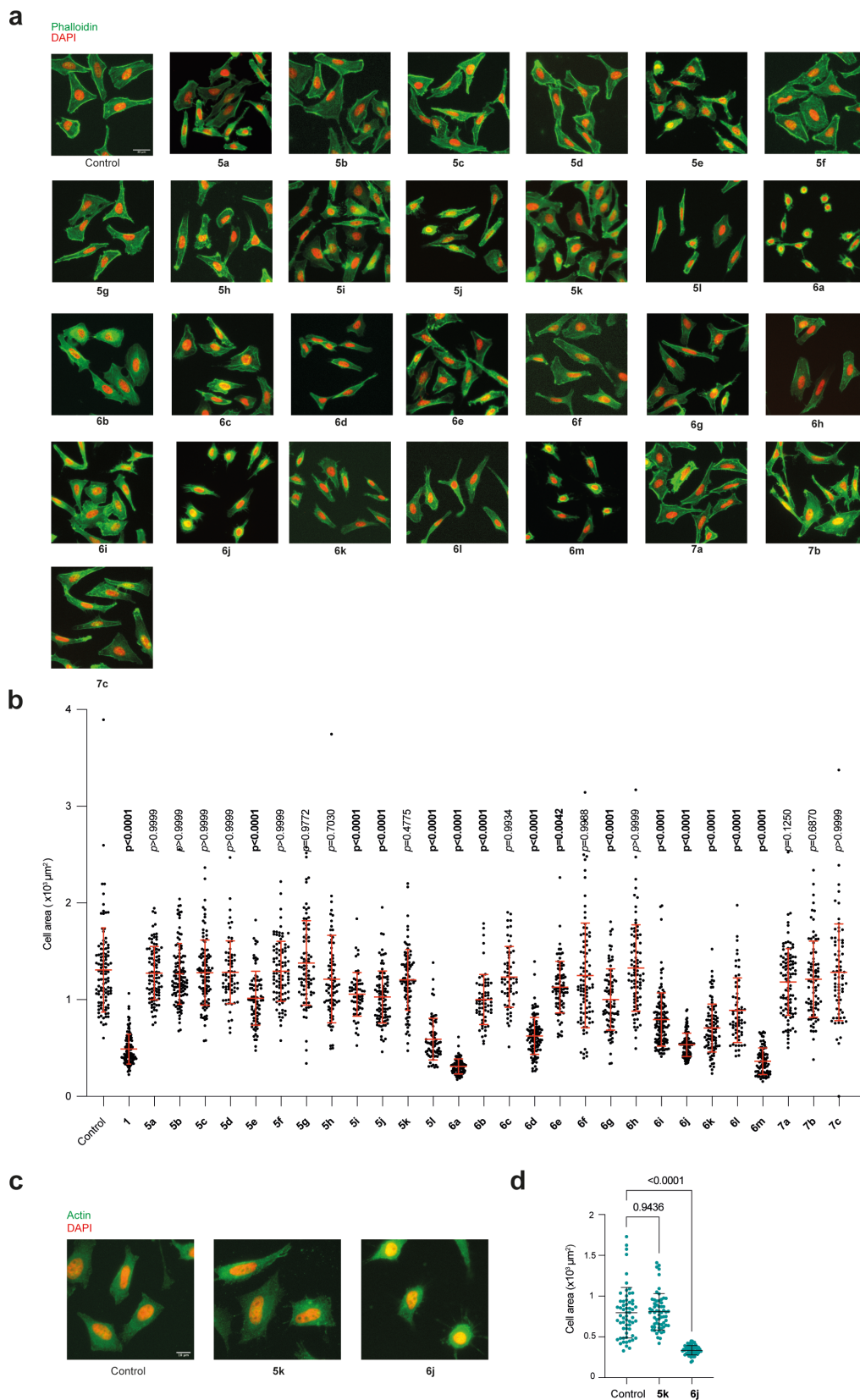
Supporting Information

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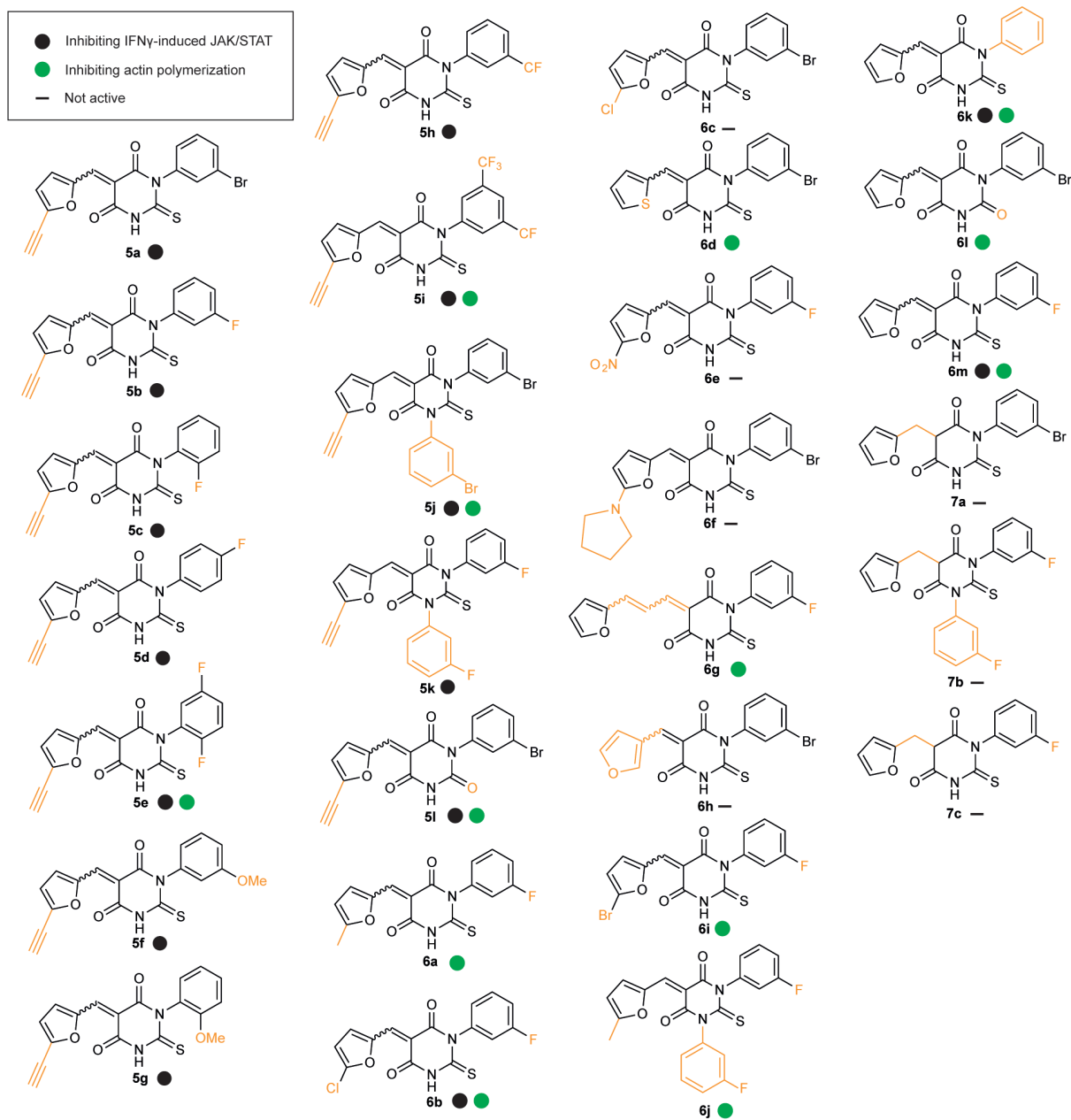
a**b**

Supplementary Figure 1 (S1). a) Anti-IFN γ activity screening performed using immunoblot analysis of pSTAT1 and tSTAT1 levels in HeLa cells treated with IFN γ (1000 U.ml⁻¹, 20 min), preincubated or not with compounds (40 μ M) for 20 min. Immunoblots on top right (screening of **1**, **5a**, **5b**, **5j**, **5k**, **7a**) are duplicates of Figure 2b. b) Quantification of pSTAT1/tSTAT1 of the immunoblot in **a**. $n = 3$ independent experiments.

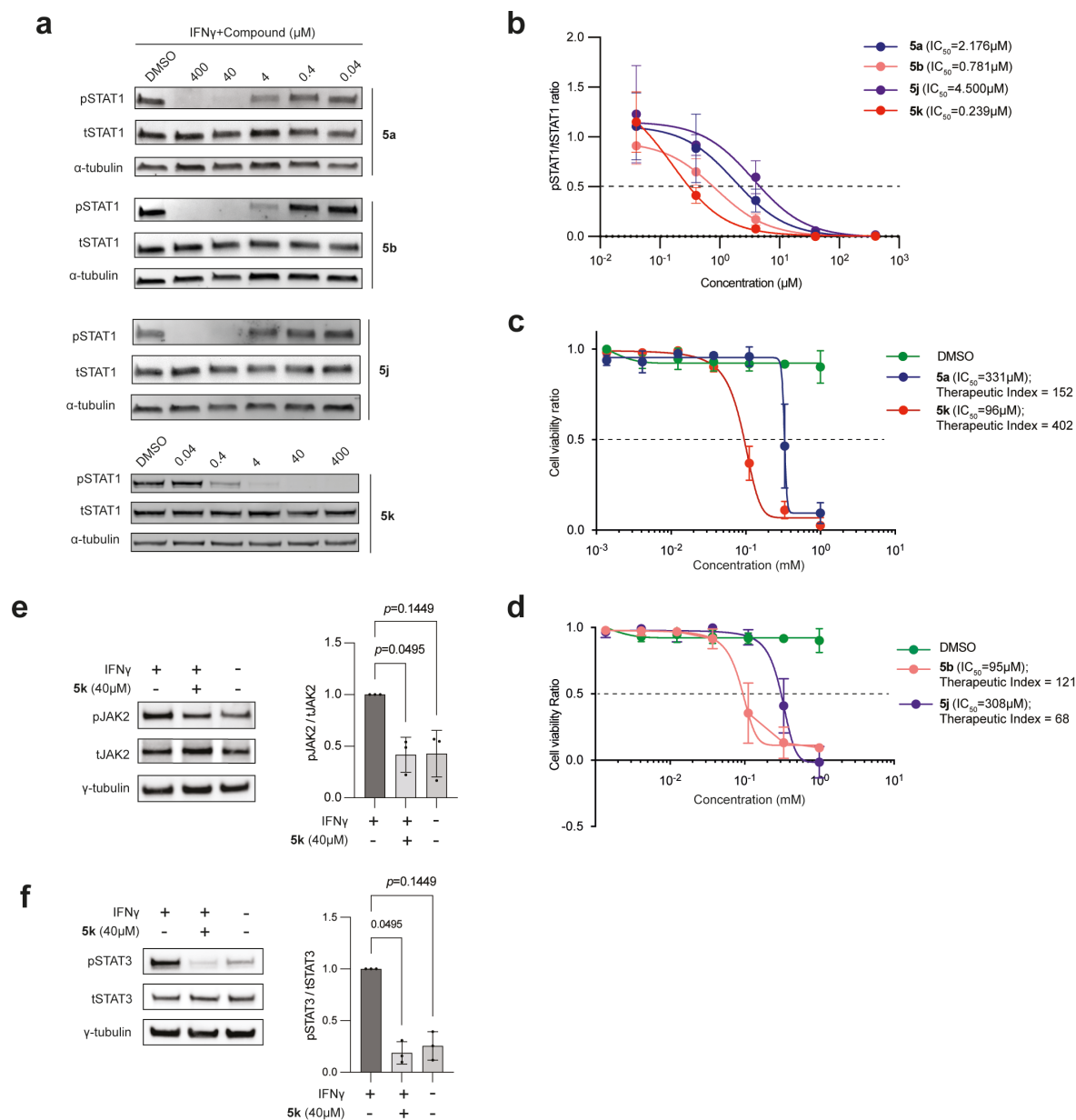


Supplementary Figure 2 (S2). a) Actin polymerization activity performed using fluorescence microscopy. Images of HeLa cells treated with small molecules ($40 \mu\text{M}$) for 20 min and stained for actin with phalloidin Alexa-488. Scale bar = $30 \mu\text{m}$. Images of Control, **5a**, **5b**, **5j**, **5k**, **7a** and **6j** are duplicates of Figure 2c. b) Quantification of cell area. Mean value \pm SD. Statistical analysis with one-way ANOVA. $n = 3$ independent experiments. c) Anti-actin polymerization activity screening (actin antibody) performed using fluorescence microscopy images of HeLa cells treated with compounds ($40 \mu\text{M}$) for 20

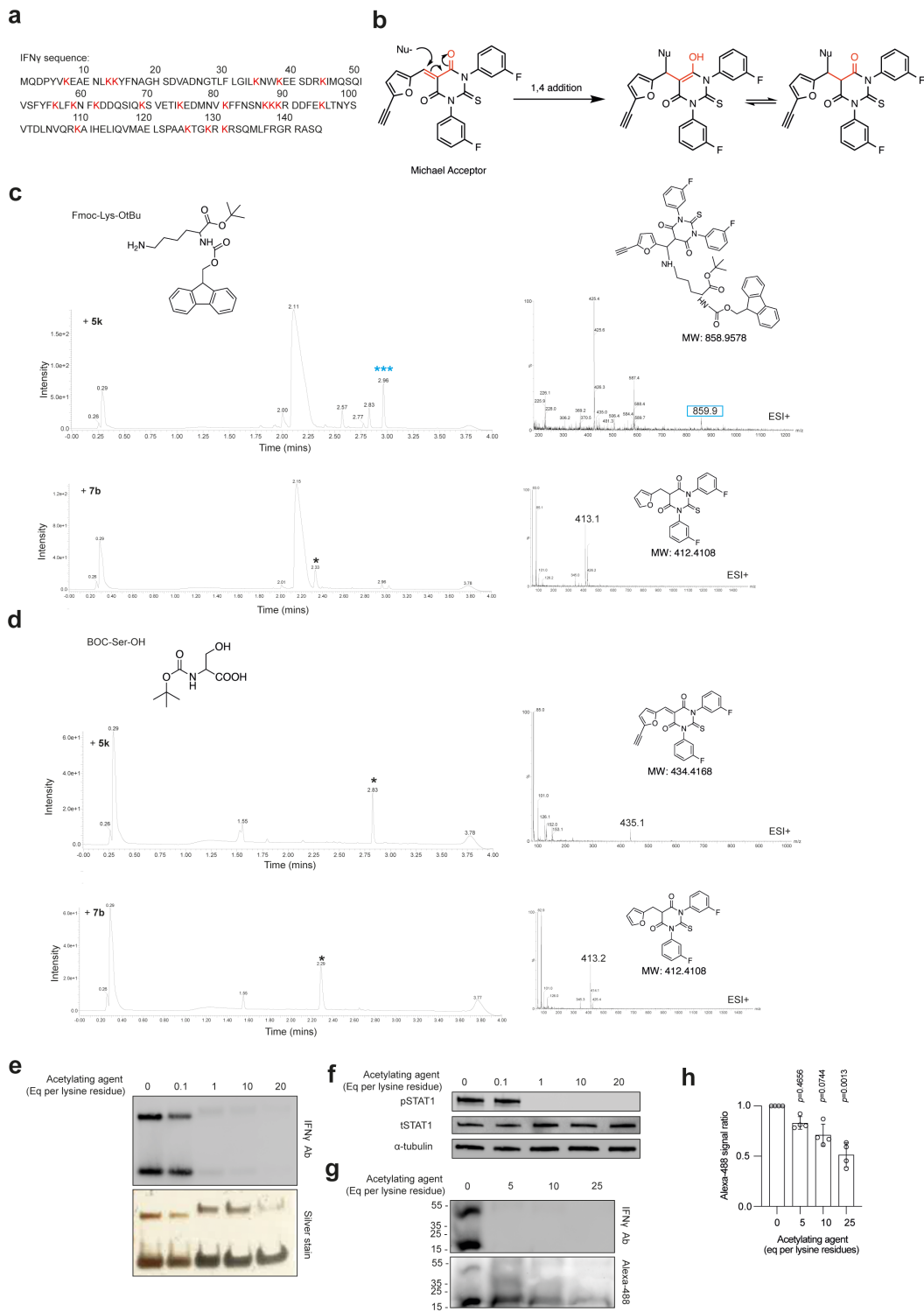
min. Scale bar = 10 μ m. d) Quantification of cell area from c. Mean value \pm SD. Statistical analysis with one-way ANOVA. $n = 3$ independent experiments.



Supplementary Figure 3 (S3). Structure of derivatives of **1** (SMIFH2) along with their activities as indicated.

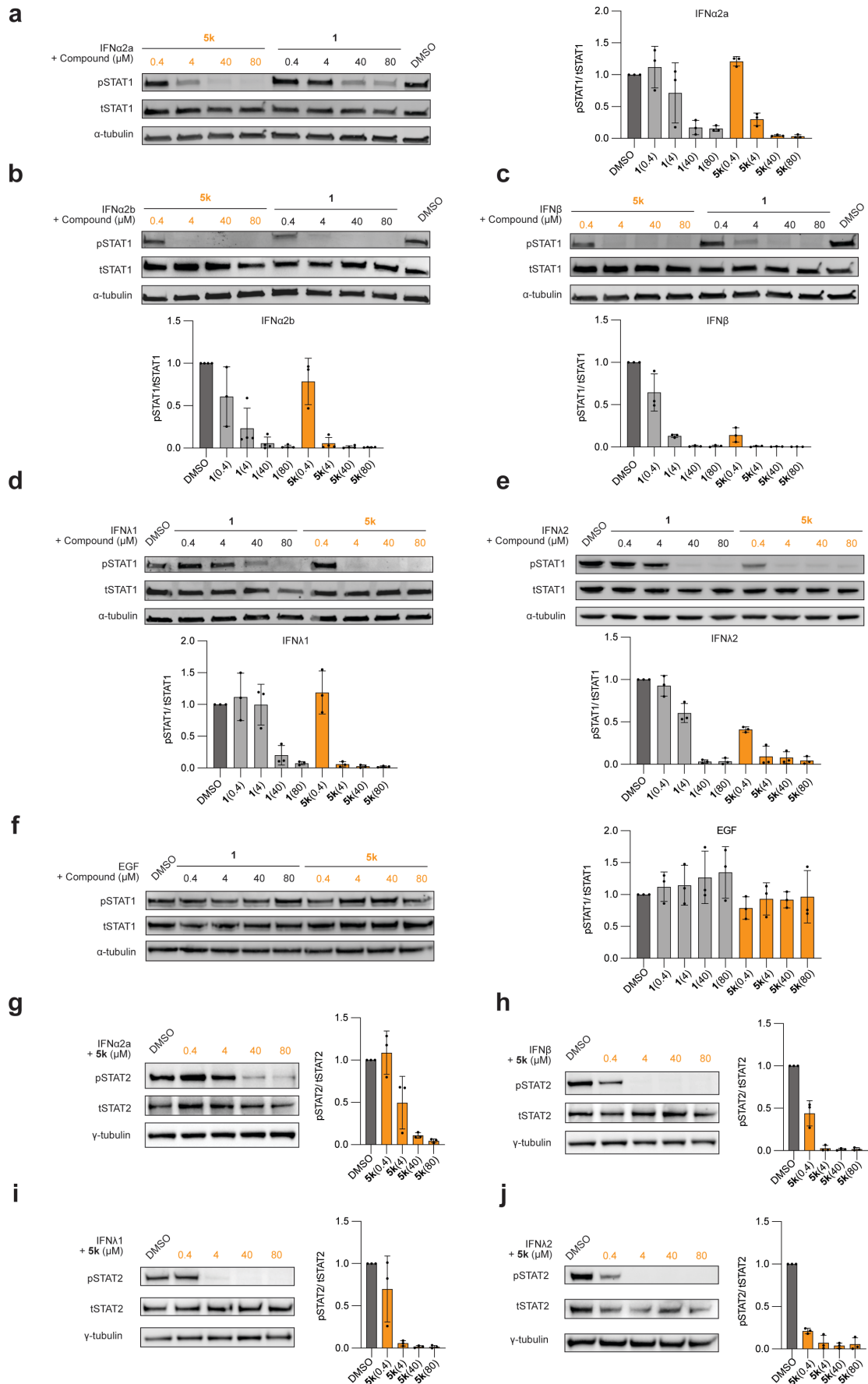


Supplementary Figure 4 (S4). a) Immunoblot analysis of pSTAT1 and tSTAT1 levels in HeLa cells treated for 20 min with IFN γ (1000 U.ml⁻¹), preincubated or not with different concentrations of indicated compounds for 20 min. Tubulin serves as a loading control. b) Quantification of pSTAT1/tSTAT1 of immunoblot in **a**. IC₅₀ towards IFN γ are determined by non-linear curve fit. $n = 3$ independent experiments. c) Dose-response viability curves of HeLa cells treated for 24 h with specified compounds. IC₅₀ are determined by non-linear curve fit. $n = 3$ independent experiments. Therapeutic Index = (IC₅₀ of Cell Viability/ IC₅₀ of pSTAT1 inhibition). d) Dose-response viability curves of HeLa cells treated for 24 h with specified compounds. IC₅₀ are determined by non-linear curve fit. $n = 3$ independent experiments. Therapeutic Index = (IC₅₀ of Cell Viability/ IC₅₀ of pSTAT1 inhibition). e) Immunoblot analysis of phosphorylation status of JAK2 (pJAK2) and total JAK2 (tJAK2) levels in HeLa cells treated for 20 min by IFN γ (1000 U.ml⁻¹), preincubated or not with **5k** (40 μ M) for 20 min as indicated. Tubulin serves as a loading control. The corresponding quantification as ratio of pJAK2/tJAK2. Mean value \pm SD. Statistical analysis with Kruskal-Wallis test with Dunn's post-test. $n = 3$ independent experiments. f) Immunoblot analysis of pSTAT3 and tSTAT3 levels in HeLa cells treated for 20 min by IFN γ (1000 U.ml⁻¹), preincubated or not with **5k** (40 μ M) for 20 min as indicated. Tubulin serves as a loading control. The corresponding quantification as ratio of pSTAT3/tSTAT3. Mean value \pm SD. Statistical analysis with Kruskal-Wallis test with Dunn's post-test. $n = 3$ independent experiments.



Supplementary Figure 5 (S5). a) IFN γ amino acid sequence. Lysine residues are highlighted in red. b) Reaction showing the nucleophilic attack on the Michael acceptor region (highlighted in red) of **5k** resulting in 1,4 Michael addition product. c) Chromatogram of UPLC-MS analysis of **5k** (up) and **7b** (down) incubated at 37°C in PBS for 12 h with Fmoc-Lysine-OtBu respectively and the corresponding ESI+ spectrum of the peak marked by asterisks (*). 1,4 Michael addition product is observed with **5k** and Fmoc-Lysine-OtBu indicated by ***. d) Chromatogram of UPLC-MS analysis of **5k** (up) and **7b** (down) incubated at 37°C in PBS for 24h with Boc-Serine-OH respectively and the corresponding ESI+ spectrum of the peak marked by asterisk (*). No 1,4 Michael addition product is observed with **5k**. e) Immunoblot showing the effect of acetylating agent on IFN γ . Top: IFN γ antibody. Bottom: silver staining. f) Immunoblot showing the effect of acetylated IFN γ on JAK-STAT signalling. g) Immunoblot analysis of IFN γ preincubated with analogue **5k** and acetylating agent as indicated. $n = 4$ independent

experiments. h) Alexa-488 signal ratio of immunoblot in **g**. Mean value \pm SD. Statistical analysis with Kruskal-Wallis test with Dunn's post-test.

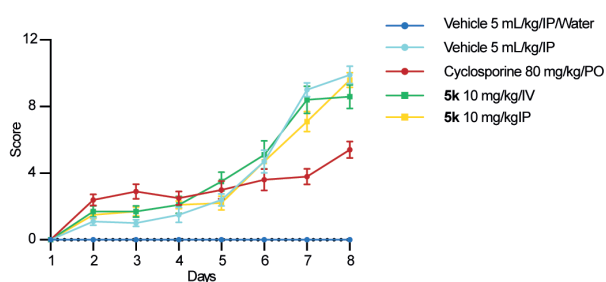


Supplementary Figure 6 (S6). Immunoblot analysis of pSTAT1 and tSTAT1 levels in HeLa cells treated for 20 min with a) IFN α 2a (1000 U.ml⁻¹), b) IFN α 2b (1000 U.ml⁻¹), c) IFN β (1000 U.ml⁻¹), d) IFN λ 1 (1000 U.ml⁻¹), e) IFN λ 2 (1000 U.ml⁻¹) and f) EGF (15.6 nM) preincubated with concentrations of **1** and **5k** for 20 min as indicated and corresponding pSTAT1/tSTAT1 ratio. $n = 3$ independent experiments. Immunoblot analysis of pSTAT2 and tSTAT2 levels in HeLa cells treated for 20 min with g) IFN α 2a (1000 U.ml⁻¹), h) IFN β (1000 U.ml⁻¹), i) IFN λ 1 (1000 U.ml⁻¹) and j) IFN λ 2 (1000 U.ml⁻¹) preincubated with concentrations of **5k** for 20 min as indicated and corresponding pSTAT2/tSTAT2. $n = 3$ independent experiments.

a

Route		IV	IP
Target Dose	(mg/kg)	2	10
$t_{1/2}$	(hr)	3.63	4.10
T_{max}	(hr)		2.00
C_{max}	(ng/mL)	3710	1442
C_0	(ng/mL)	4860	
AUC_{0-last}	(hr*ng/mL)	2072	8249
$AUC_{0-last}/Dose$	(hr*ng/mL)	1036	825
AUC_{0-inf}	(hr*ng/mL)	2085	8421
CL	mL/min/kg	16.0	
V_{ss}	L/kg	2.84	
relative F	%		79.6

b



Supplementary Figure 7 (S7). a) Pharmacokinetic profiling of compound in the C57BL/6N male mice following intravenous (IV) and intraperitoneal (IP) administration. b) Acute model of DSS-induced colitis in C57BL/6 mice. Disease clinical index (mean +/- SEM).

Material and methods:

A) Chemical synthesis

General information:

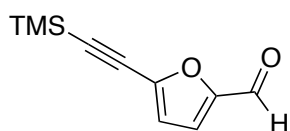
All solvents and chemicals were purchased from commercial sources and used without further purification. Solvents were dried under standard conditions. Reactions were monitored by thin layer chromatography (TLC) using precoated silica on aluminium sheets from Merck (60 F₂₅₄). TLC sheets were visualized with UV light. Products were purified on column chromatography with Silica gel 60 from Alfa Aesar (0.036-0.071 mm; 215-400 mesh), a Combiflash RF+ Teledyne Isco system fitted with pre-packed silica gel columns (Interchim, 4-300 g columns, 50 µm particle size), a preparative HPLC Quaternary Gradient 2545 equipped with a Photodiode Array detector (Waters) fitted with a reverse phase column (XBridge Prep C₁₈ 5 µm OBD 30×150 mm) in acetonitrile and water system.

NMR spectroscopy was performed on Bruker spectrometers. Spectra were run at 298 K unless stated otherwise. ¹H-NMR were recorded at 400 or 500 MHz, and chemical shifts δ are expressed in ppm using the residual non-deuterated solvent signal as internal standard and the coupling constants *J* are specified in Hz and E/Z to denote the signal from isomers. The following abbreviations are used: s, singlet; d, doublet; dd, doublet of doublets; t, triplet; td, triplet of doublets; q, quadruplet; m, multiplet; bs, broad signal. We only reported labile protons that could be clearly identified in the spectra, namely for compounds **2c**, **2f**, **7a**, **7b** and **7c**. ¹³C-NMR were recorded at 101 or 126 MHz, and chemical shifts δ ppm are expressed in ppm using deuterated solvent signal as internal standard and the coupling constants *J* with Fluorine are specified in Hz. ¹⁹F-NMR were recorded at 376 or 471 MHz, and chemical shifts δ are expressed in ppm. Molecular structures were characterized using a comprehensive dataset including ¹H- and ¹³C-NMR spectra (1D and 2D experiments including COSY, HMBC, and HSQC). For the final compounds which are in isomeric mixtures, the signals corresponding to either E or Z are separated by "/" and denoted by (E/Z). MestReNova V-10.0.1 was used to process the spectrum.

The purity of final compounds, determined to be >95% by UPLC-MS, and low-resolution mass spectra (LRMS) were recorded on a Waters Acquity H-class equipped with a Photodiode array detector and SQ Detector 2 fitted with a reverse phase column (Aquity UPLC® BEH C₁₈ 1.7 µm, 2.1x50 mm). Gradient used was H₂O/ACN-95/5 till 0.2 min, 95/5 to 0/100 till 2.5 min, 0/100 till 3 min, 0/100 to 95/5 till 3.10 min and 95/5 till 4 min. High resolution mass spectra (HRMS) were recorded on a Thermo Fisher Scientific Q-Exactive Plus equipped with a Robotic TriVersa NanoMate Advion.

Abbreviations used. ACN, acetonitrile; AcOH, acetic acid; aq., aqueous; DCM, dichloromethane; DMSO, dimethylsulfoxide; eq, equivalent(s); cHex, cyclohexane; EtOAc, ethyl acetate; Et₃N, triethylamine; ESI, electrospray ionization; HPLC, high pressure liquid chromatography; HRMS, high resolution mass spectroscopy; LRMS, low resolution mass spectroscopy; MeOH, methanol; min, minutes; h, hour; MS, mass spectrometry; NMR, nuclear magnetic resonance; r.t., room temperature; TMS, trimethylsilane; THF, tetrahydrofuran; TLC, thin-layer chromatography.

1. Synthesis of 5-((trimethylsilyl)ethynyl)furan-2-carbaldehyde **1a**



5-((trimethylsilyl)ethynyl)furan-2-carbaldehyde **1a** was synthesized according to published procedure.^[1] Trimethylsilylacetylene (835 μL , 1.05 eq) and triethylamine anhydrous (2 mL) were dissolved in THF (5 mL). The mixture was degassed three times and stirred under argon. 5-bromofuran-2-carbaldehyde (1.00 g, 5.71 mmol) and $\text{PdCl}_2(\text{PPh}_3)_2$ (80 mg, 2 mol%) were added. After 1 min of stirring, CuI (43.5 mg, 4 mol%) was added, then the mixture was stirred at r.t. for 16 h then concentrated under reduced pressure. The crude product was purified by flash chromatography on silica gel (cHex/EtOAc, 6/4) to give of designed product, pale yellow crystals. Percentage yield: 626.5 mg, 57%.

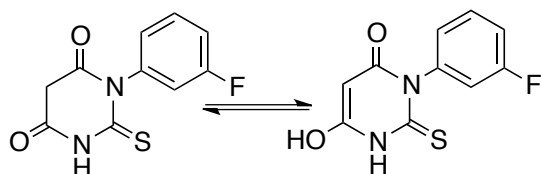
^1H NMR (400 MHz, CDCl_3) δ ppm: 9.61 (s, 1H), 7.19 (d, $J = 3.7$ Hz, 1H), 6.71 (d, $J = 3.7$ Hz, 1H), 0.27 (s, 9H). HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_{13}\text{O}_2\text{Si}^+$ ($\text{M}+\text{H}^+$): 193.0607; found: 193.0680

2. Synthesis of thiobarbituric acid derivatives

Analogues of thiobarbituric acids were synthesized according to published procedures^[2,3]. To a suspension of thiourea derivative (1 eq) in propanol (5 mL), diethyl malonate (2.5 eq) and sodium methoxide (25% wt in MeOH or powder, 2.5 eq) were added under Argon atmosphere. The reaction mixture was stirred under reflux (105°C) overnight. Completion of the reaction was checked by TLC analysis and UPLC-MS. The crude was then cooled to r.t., and quenched with acetic acid to reach about pH=6-7. The solvent was evaporated *in vacuo*, and the product was purified from the crude by flash chromatography on silica gel or using a CombiFlash system (DCM/MeOH, 8/2).

The Keto-Enol Tautomers are observed in the NMR spectra and their corresponding forms are stated in the description. In case of MeOD- d_4 as solvent, due to exchange with MeOD- d_4 , the protons involved in Keto-Enol tautomerization cannot be observed^[3], which is concordant with the literature^[3].

1-(3-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **2a**

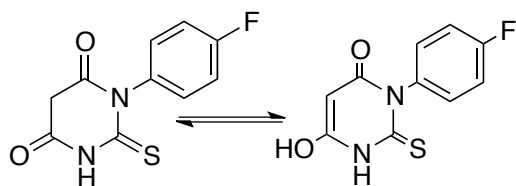


From 1-(3-fluorophenyl)thiourea (400 mg, 2.35 mmol), Diethyl malonate (896 μL), sodium methoxide (25% solution in MeOH) (1343 μL).

Light Yellow powder, Percentage Yield =89.6%

^1H NMR (400 MHz, $\text{DMSO}-d_6$) δ ppm: 10.66 (bs, 1H), 7.48 – 7.25 (m, 1H), 7.20 – 7.03 (m, 1H), 6.96 – 6.82 (m, 2H), 4.24 (s, 1H) *Enol Form*. ^{13}C NMR (101 MHz, $\text{DMSO}-d_6$) δ ppm: 176.5, 163.4, 162.3 (d, $J_{\text{C-F}} = 242.4$ Hz), 162.9, 143.4 (d, $J_{\text{C-F}} = 10.7$ Hz), 129.8 (d, $J_{\text{C-F}} = 8.7$ Hz), 126.6, 117.5 (d, $J_{\text{C-F}} = 22.1$ Hz), 114.1 (d, $J_{\text{C-F}} = 20.9$ Hz), 79.8. ^{19}F NMR (376 MHz, $\text{DMSO}-d_6$) δ ppm: -114.65. HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_8\text{FN}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}^+$): 239.0212; found: 239.0285

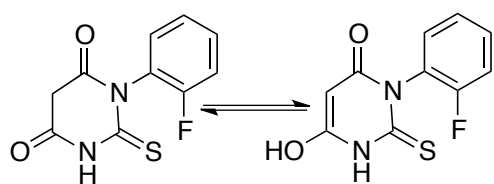
1-(4-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **2b**



From 1-(4-fluorophenyl)thiourea (250 mg, 1.47 mmoles), Diethyl malonate (558 μ L), sodium methoxide (25% solution in MeOH) (671 μ L). Yellow powder, Percentage Yield =248 mg, 71%.

^1H NMR (500 MHz, DMSO- d_6) δ ppm: 10.69 (bs, 1H), 7.16 (t, $J_{\text{C-F}} = 8.8$ Hz, 2H), 7.05 (dd, $J_{\text{C-F}} = 8.7$ Hz, $J = 5.0$ Hz, 2H), 4.27 (s, 1H, 82% *Enol*), 3.17 (d, $J = 4.5$ Hz, 1H, 17% *Keto*). ^{13}C NMR (126 MHz, DMSO- d_6) δ ppm: 176.8, 163.6, 163.0, 161.3 (d, $J_{\text{C-F}} = 242.1$ Hz), 137.9, 131.9 (d, $J_{\text{C-F}} = 8.7$ Hz, 2C), 115.3 (d, $J_{\text{C-F}} = 22.6$ Hz, 2C), 79.9, 49.0. HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_8\text{FN}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}^+$): 239.0212; found: 239.0285

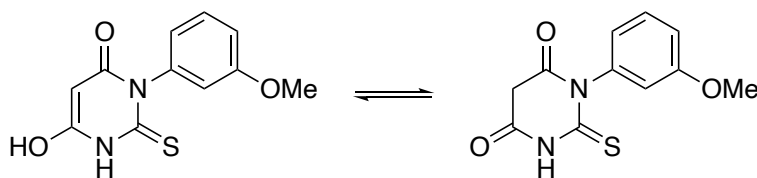
1-(2-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **2c**



From 1-(2-fluorophenyl)thiourea (250mg, 1.47 mmoles), Diethyl malonate (558 μ L), sodium methoxide (25% solution in MeOH) (671 μ L). Yellow powder, Percentage Yield=219 mg, 63%.

^1H NMR (500 MHz, MeOD- d_4) δ ppm: 7.35 – 7.23 (m, 1H), 7.19 – 7.01 (m, 3H), 3.25 (s, 1H, *Keto form*, 1H exchanged with MeOD- d_4). ^{13}C NMR (126 MHz, MeOD- d_4) δ ppm: 177.0, 165.4, 164.8, 158.2 (d, $J_{\text{C-F}} = 249.7$ Hz), 131.0, 129.6 (d, $J_{\text{C-F}} = 7.9$ Hz), 127.6 (d, $J_{\text{C-F}} = 13.4$ Hz), 123.9, 115.55 (d, $J_{\text{C-F}} = 20.0$ Hz), 48.5 (*Keto form*). ^{19}F NMR (376 MHz, MeOD- d_4) δ ppm: -123.64. HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_8\text{FN}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}^+$): 239.0212; found: 239.0285

1-(3-methoxyphenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **2d**

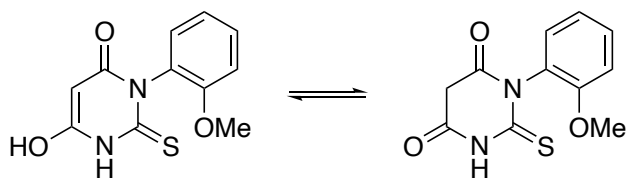


From 1-(3-methoxyphenyl)thiourea (250mg, 1.37 mmoles), Diethyl malonate (525 μ L), sodium methoxide (25% solution in MeOH) (630 μ L). Pale white powder,

Percentage Yield: 283 mg, 83%.

^1H NMR (400 MHz, DMSO- d_6) δ ppm: 10.61 (bs, 1H), 7.25 (t, $J = 8.0$ Hz, 2H), 6.84 (ddd, $J = 8.3, 2.5, 0.9$ Hz, 2H), 6.64 – 6.59 (m, 2H), 6.57 (t, $J = 2.2$ Hz, 2H), 4.26 (s, 1H, 50% *Enol*), 3.74 (s, 6H), 3.17 (s, 2H, 50% *Keto*). ^{13}C NMR (101 MHz, DMSO- d_6) δ ppm: 176.5, 163.6, 162.9, 159.7, 142.8, 129.1, 122.5, 115.9, 112.8, 79.9 (*Enol*), 55.6, 49.0 (*Keto*). HRMS(ESI+) m/z : calculated for $\text{C}_{11}\text{H}_{11}\text{N}_2\text{O}_3\text{S}^+$ ($\text{M}+\text{H}^+$): 251.0412; found: 251.0485

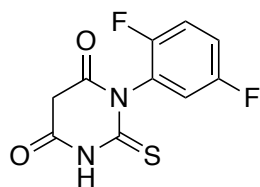
1-(2-methoxyphenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **2e**



From 1-(2-methoxyphenyl)thiourea (250 mg, 1.37 mmoles), Diethyl malonate (525 μ L), sodium methoxide (25% solution in MeOH) (630 μ L). Pale white powder, Percentage Yield: 311 mg, 91%.

^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ ppm: 10.62 (bs, 1H), 7.25 (ddd, $J=8.3, 6.9, 2.2$ Hz, 1H), 7.00 (d, $J=7.8$ Hz, 1H), 6.97 – 6.87 (m, 2H), 4.24 (s, 1H, 68% *Enol*), 3.69 (s, 3H), 3.17 (s, 1H, 32% *Keto*). ^{13}C NMR (101 MHz, $\text{DMSO-}d_6$) δ ppm: 176.7, 163.4, 162.9, 155.5, 131.1, 130.4, 128.8, 120.4, 112.3, 79.9 (*Enol*), 55.9, 49.0 (*Keto*). HRMS(ESI+) m/z : calculated for $\text{C}_{11}\text{H}_{11}\text{N}_2\text{O}_3\text{S}^+$ ($\text{M}+\text{H}$) $^+$: 251.0412; found: 251.0485

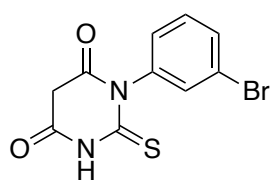
1-(2,5-difluorophenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **2f**



From 1-(2,5-difluorophenyl)thiourea (500 mg, 2.66 mmoles), Diethyl malonate (403 μL), sodium methoxide (25% solution in MeOH) (1518 μL). Light yellow powder, Percentage Yield: 572 mg, 84%.

^1H NMR (500 MHz, $\text{MeOD-}d_4$) δ ppm: 7.06 (2H, ddd, $J = 15.1, 8.2, 4.0$ Hz), 6.97 – 6.84 (1H, m), 3.25 (1H, s, *Keto form*, 1H exchanged with $\text{MeOD-}d_4$). ^{13}C NMR (126 MHz, $\text{MeOD-}d_4$) δ ppm: 176.9, 165.3, 165.0, 158.4 (d, $J_{\text{C-F}} = 241.1$ Hz), 154.8 (d, $J_{\text{C-F}} = 244.4$ Hz), 128.6 (dd, $J_{\text{C-F}} = 15.3, 11.6$ Hz), 117.9 (d, $J_{\text{C-F}} = 25.4$ Hz), 116.3 (dd, $J_{\text{C-F}} = 22.9, 9.4$ Hz), 115.9 (dd, $J_{\text{C-F}} = 24.1, 8.1$ Hz), 48.5. ^{19}F NMR (376 MHz, $\text{MeOD-}d_4$) δ ppm: -120.90 (d, $J_{\text{F-F}} = 16.0$ Hz), -128.75 (d, $J_{\text{F-F}} = 16.1$ Hz). HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_7\text{F}_2\text{N}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}$) $^+$: 257.0118; found: 257.0189

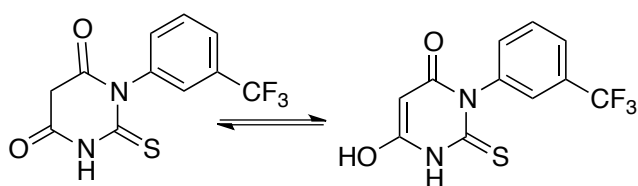
1-(3-bromophenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **2g**



From 1-(3-bromophenyl)thiourea (200 mg, 0.87 mmoles), Diethyl malonate (129 μL), sodium methoxide (25% solution in MeOH) (495 μL). Light pale yellow powder. Percentage Yield: 115 mg, 52%.

^1H NMR (400 MHz, CD_3CN) δ ppm: 10.73 (bs, 1H), 7.53 (m, 1H), 7.40 (t, $J = 1.9$ Hz, 1H), 7.35 (t, $J = 8.0$ Hz, 1H), 7.20 (m, 1H), 3.30 (s, 2H), 1.99 (s, 1H, Acetic Acid). ^{13}C NMR (101 MHz, CD_3CN) δ ppm 177.7, 174.0 (Acetic Acid), 165.3, 165.1, 142.9, 133.1, 131.0, 130.7, 129.3, 121.5, 49.5, 20.7 (Acetic Acid). HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_8\text{BrN}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}$) $^+$: 298.9412; found: 300.9464.

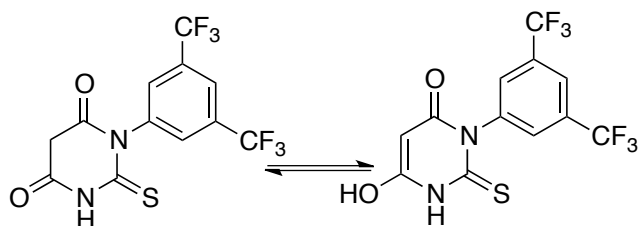
2-thioxo-1-(3-(trifluoromethyl)phenyl)dihydropyrimidine-4,6(1*H*,5*H*)-dione **2h**



From 1-(3-(trifluoromethyl)phenyl)thiourea (200 mg, 0.91 mmoles), Diethyl malonate (207 μL), sodium methoxide (25% solution in MeOH) (520 μL). Light yellow powder. Percentage Yield: 259 mg, 99%.

^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ ppm: 10.80 (bs, 1H), 7.69 – 7.55 (m, 2H), 7.46 – 7.32 (m, 2H), 4.27 (s, 1H). ^{13}C NMR (101 MHz, $\text{DMSO-}d_6$) δ ppm: 176.6, 163.4, 162.9, 142.5, 134.6, 129.8, 129.4 (q, $J_{\text{C-F}} = 31.9$ Hz), 126.98/ 126.94, 124.6 (q, $J_{\text{C-F}} = 272.0$ Hz), 124.1, 79.8. ^{19}F NMR (376 MHz, $\text{DMSO-}d_6$) δ ppm: -60.84. HRMS(ESI+) m/z : calculated for $\text{C}_{11}\text{H}_8\text{F}_3\text{N}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}$) $^+$: 289.0180; found: 289.0253.

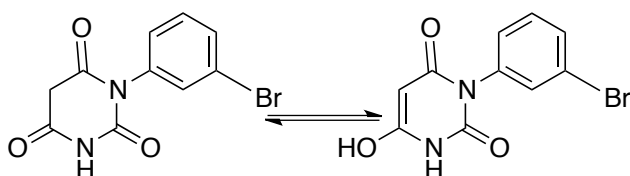
1-(3,5-bis(trifluoromethyl)phenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **2i**



From 1-(3,5-bis(trifluoromethyl)phenyl)thiourea (200 mg, 0.69 mmoles), Diethyl malonate (116 μ L), sodium methoxide (94 mg). Light yellow powder. Percentage Yield: 64 mg, 26%

^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ ppm: 11.01 (bs, 1H), 8.09 (s, 1H), 7.87 (d, $J = 1.5$ Hz, 2H), 4.35 (s, 1H). ^{13}C NMR (101 MHz, $\text{DMSO-}d_6$) δ ppm: 176.5, 163.1, 162.9, 143.8, 131.7(2C), 130.6 (d, $J_{\text{C-F}} = 33.0$ Hz, 2C), 123.7 (d, $J_{\text{C-F}} = 272.8$ Hz, 2C), 121.2, 79.8. ^{19}F NMR (376 MHz, $\text{DMSO-}d_6$) δ ppm: -61.12. HRMS(ESI+) m/z : calculated for $\text{C}_{12}\text{H}_7\text{F}_6\text{N}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}$) $^+$: 357.0054; found: 357.0127.

1-(3-bromophenyl)pyrimidine-2,4,6(1*H*,3*H*,5*H*)-trione **2j**

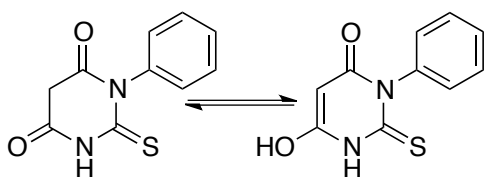


From 1-(3-bromophenyl)urea (200 mg, 0.93 mmoles), Diethyl malonate (353 μ L), sodium methoxide (25% solution in MeOH) (531 μ L). Light pale white powder; Yield: 169.7 mg, 65%. *Product not very pure, however*

engaged in next step.

^1H NMR (500 MHz, $\text{DMSO-}d_6$) δ ppm: 11.50 (s, 1H), 7.66 – 7.56 (m, 1H), 7.53 – 7.39 (m, 2H), 7.26 (d, $J = 7.8$ Hz, 1H), 4.13 (s, 1H). ^{13}C NMR (126 MHz, $\text{DMSO-}d_6$) δ ppm: 168.56 (2C), 167.5, 158.5, 151.9, 132.4, 131.1, 129.0, 121.4, 66.5. HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_8\text{BrN}_2\text{O}_3^+$ ($\text{M}+\text{H}$) $^+$: 282.9640; found: 282.9713.

1-phenyl-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **2k**



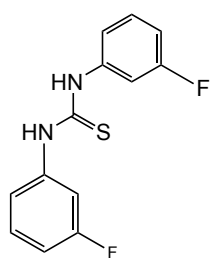
From 1-phenylthiourea (500 mg, 3.28 mmoles), Diethyl malonate (1250 μ L), sodium methoxide (25% solution in MeOH) (1500 μ L). Light pale white powder; Yield: 423 mg, 60%.

^1H NMR (400 MHz, $\text{DMSO-}d_6$) δ ppm: 10.50 (bs, 1H), 7.39 – 7.31 (m, 2H), 7.29 – 7.21 (m, 1H), 7.07 – 6.92 (m, 2H), 4.22 (s, 1H, *Enol form*). ^{13}C NMR (101 MHz, $\text{DMSO-}d_6$) δ ppm: 176.7, 163.5, 162.8, 141.9, 130.1 (2C), 128.5 (2C), 127.1, 79.8. HRMS(ESI+) m/z : calculated for $\text{C}_{10}\text{H}_9\text{N}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}$) $^+$: 221.0306; found: 221.0376.

3. Synthesis of Bis-phenyl-thiourea

The protocol was adapted from previously described procedure^[4]. Fluoro/Bromo-3-isothiocyanatobenzene and 3-Fluoro/Bromoaniline in 1:1 ratio were mixed mechanically using pestle and mortar for about 15-20 minutes until cream white colour paste is formed. The crude was washed with DCM and a white colour solid powder product is obtained.

1,3-bis(3-fluorophenyl)thiourea **3a**

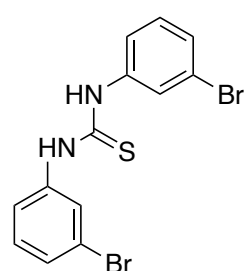


1-fluoro-3-isothiocyanatobenzene (9 g, 7.09 mL, 58.75 mmoles), 3-fluoroaniline (5.65 mL, 1 eq); Yield : 14.38 g, 74%.

^1H NMR (400 MHz, Acetone- d_6) δ ppm: 9.34 (s, 2H), 7.68 – 7.49 (m, 2H), 7.46 – 7.34 (m, 2H), 7.36 – 7.26 (m, 2H), 6.95 (tdd, $J = 8.2, 2.6, 1.0$ Hz, 2H). ^{13}C NMR (101 MHz, Acetone- d_6) δ ppm: 180.24, 162.54 (d, $J_{\text{C-F}} = 242.9$ Hz, 2C), 141.01 (d, $J_{\text{C-F}} = 10.6$ Hz, 2C), 130.07 (d, $J_{\text{C-F}} = 9.5$ Hz, 2C), 119.56 (d, $J_{\text{C-F}} = 2.9$ Hz, 2C), 112.40 – 109.72 (m, 4C).

^{19}F NMR (376 MHz, Acetone- d_6) δ ppm: -113.90. HRMS(ESI+) m/z : calculated for $\text{C}_{13}\text{H}_{11}\text{F}_2\text{N}_2\text{S}^+$ (M+H) $^+$: 265.0533; found: 265.0606

1,3-bis(3-bromophenyl)thiourea **3b**



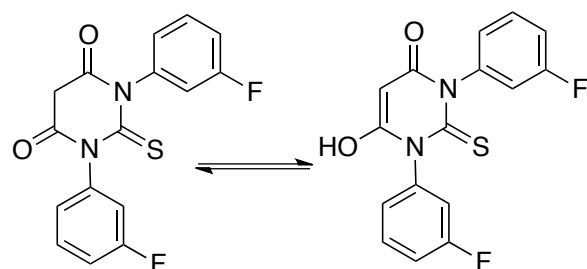
1-bromo-3-isothiocyanatobenzene (100 mg, 0.47 mmoles), 3-bromoaniline (48 μL , 1 eq). Yield: 110 mg, 61%.

^1H NMR (500 MHz, Acetone- d_6) δ ppm: 9.30 (s, 2H), 7.88 (t, $J = 2.0$ Hz, 2H), 7.58 – 7.48 (m, 2H), 7.43 – 7.18 (m, 4H). ^{13}C NMR (126 MHz), Acetone- d_6) δ ppm: 180.53, 140.80 (2C), 130.31 (2C), 127.92 (2C), 127.01 (2C), 123.04 (2C), 121.42 (2C). LRMS(ESI+) m/z : calculated for $\text{C}_{13}\text{H}_{10}\text{Br}_2\text{N}_2\text{S}^+$ (M+H) $^+$: 384.9; found: 384.9

4. Synthesis of Diphenyl-Thiobarbituric acid

1,3-bis(3-fluoro/bromophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione was synthesized as per previously described protocol^[3]. Briefly, to 1,3-bis(3-fluorophenyl/bromo)thiourea (1 eq) in anhydrous CHCl_3 , 1.2-1.3 eq of Malonic acid and 2 eq of Phosphoryl trichloride were added. The reaction mixture was put on reflux at 55°C for about 48 h. The crude was then washed with H_2O and recrystallized in ethanol to obtain crystals.

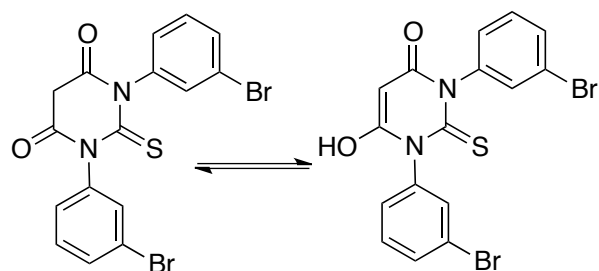
1,3-bis(3-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **4a**



3a (1 g, 3.79 mmoles), Malonic acid (473 mg, 1.2 eq) and Phosphoryl trichloride (691 μL , 2 eq). Light yellow tinge crystals; Yield= 755 mg, 60%. ^1H NMR (500 MHz, DMSO- d_6) δ ppm: 7.39 (q, $J = 7.7$ Hz, 2H), 7.12 (t, $J = 7.5$ Hz, 2H), 7.05 – 6.91 (m, 4H), 4.48 (s, 1H, *Enol*), 3.18 (s, 1H, *Keto*). ^{13}C NMR (126 MHz, DMSO- d_6) δ ppm: 178.7, 162.5

(d, $J_{\text{C-F}} = 242.4$ Hz, 2C), 162.5 (2C), 144.0 (2C), 123.0 (d, $J_{\text{C-F}} = 8.8$ Hz, 2C), 126.5 (2C), 117.4 (d, $J_{\text{C-F}} = 22.4$ Hz, 2C), 114.2 (d, $J_{\text{C-F}} = 20.7$ Hz, 2C), 80.0 (*Enol*), 49.07 (*Keto*). ^{19}F NMR (471 MHz, DMSO- d_6) δ ppm: -114.51. HRMS(ESI+) m/z : calculated for $\text{C}_{16}\text{H}_{11}\text{F}_2\text{N}_2\text{O}_2\text{S}^+$ (M+H) $^+$: 333.0431; found: 333.0504.

1,3-bis(3-bromophenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **4b**



3b (62 mg, 0.16 mmoles), Malonic acid (22 mg, 1.3 eq) and Phosphoryl trichloride (35 μ L, 2 eq). Yellow crystals; Yield= 51.5 mg, 70%.

^1H NMR (400 MHz, Acetone- d_6) δ ppm: 7.45 – 7.39 (m, 2H), 7.35 (dt, J = 8.9, 1.9 Hz, 2H), 7.29 (t, J = 7.9 Hz, 2H), 7.17 (t, J = 7.2 Hz, 2H), 4.61 (s, 1H). ^{13}C NMR (101 MHz, Acetone- d_6) δ ppm:

179.2, 162.8 (2C), 144.3 (2C), 133.0 (2C), 129.6 (2C), 129.4 (2C), 129.1 (2C), 120.6 (2C), 79.8. HRMS(ESI+) m/z : calculated for $\text{C}_{16}\text{H}_{11}\text{Br}_2\text{N}_2\text{O}_2\text{S}^+$ ($\text{M}+\text{H}$) $^+$: 452.8830; found: 454.8882.

5. Synthesis of analogues with alkyne group by Knoevenagel condensation

The following protocols were adapted from literature^{5,6}.

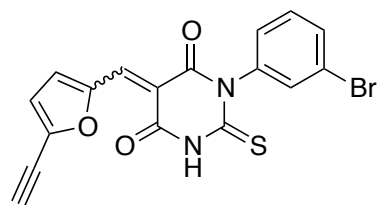
Step 1: Two methods were used:

Method A^[5] : The thiobarbituric acid derivatives (1 eq) and the 5-((trimethylsilyl)ethynyl)furan-2-carbaldehyde (1 eq) were dissolved in dry MeOH. To the reaction mixture was added catalytic amount of pyridine (1-2 drops) and the reaction was stirred at 60 $^\circ\text{C}$ for 2 hours. Then, the solvent and pyridine were removed under reduced pressure to obtain the crude product.

Method B^[6] : The thiobarbituric acid derivatives (1 eq) and the 5-((trimethylsilyl)ethynyl)furan-2-carbaldehyde (1 eq) were dissolved in distilled water and refluxed for 1 hour. Then, the water was removed under reduced pressure to obtain the crude product.

Step 2: The crude product from step 1 was dissolved in dry MeOH (2 ml) and K_2CO_3 (2 eq) was added. The reaction was stirred for 2 hours at r.t. and the solvent was removed under reduced pressure. The crude product was purified by preparative HPLC using acetonitrile and water and the solvents were removed under reduced pressure at 30-33 $^\circ\text{C}$ as freeze drying leads to degradation of the product. In some analogues, flash chromatography on silica gel using a CombiFlash system were used for the purification.

1-(3-bromophenyl)-5-((5-ethynylfuran-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **5a**



Using Method B of Step 1; **2g** (46 mg, 0.15 mmoles) and **1a** (41 mg, 1.02 eq) were dissolved in distilled water (2 mL), Brown powder, Yield= 4.9 mg, 7.9% (E/Z mixture; 50:50).

^1H NMR (500 MHz, THF- d_8) δ ppm: 12.04 (bs, 0.5H)/11.98 (bs, 0.5H) (E/Z), 8.74 (d, J = 3.9 Hz, 0.5H)/8.55 (d, J = 3.9 Hz, 0.5H) (E/Z), 8.25

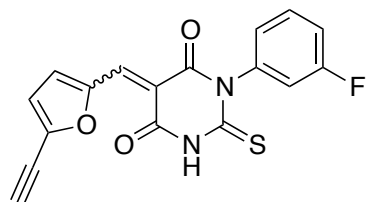
(s, 0.5H)/ 8.18 (s, 0.5H) (E/Z), 7.59 (t, J = 7.1 Hz, 1H), 7.48 (dt, J = 13.7, 2.0 Hz, 1H), 7.39 (m, 1H), 7.25 (m, 1H), 7.06 (d, J = 3.9 Hz, 0.5H)/7.01 (d, J = 3.9 Hz, 0.5H) (E/Z), 4.48(s, 0.5H)/4.47(s, 0.5H) (E/Z).

^{13}C NMR (126 MHz, THF- d_8) δ ppm: 179.37, 161.36/159.79/159.71/158.60 (E/Z, 2C), 151.45/151.30 (E/Z), 142.43, 140.70/140.45 (E/Z), 137.49/137.16 (E/Z), 132.39/132.25 (E/Z), 131.23, 130.07,

128.39/128.21 (E/Z), 127.59/127.54 (E/Z), 121.52/121.50 (E/Z), 120.13/120.11 (E/Z), 114.49/114.29 (E/Z), 87.54, 72.91.

HRMS(ESI+) *m/z*: calculated for C₁₇H₁₀BrN₂O₃S⁺ (M+H)⁺ : 402.2340; found: 402.9570

5-((5-ethynylfuran-2-yl)methylene)-1-(3-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **5b**



Using Method A of Step 1: **2a** (100 mg, 0.42 mmoles) and **1a** (81 mg, 1.01 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 25.5 mg, 17.8% (E/Z mixture; 50:50).

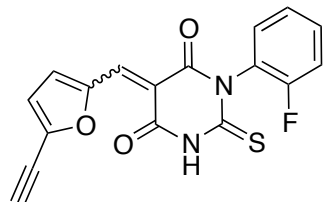
¹H NMR (500 MHz, THF-*d*₈) δ ppm : 12.04 (bs, 0.5H)/11.98 (bs, 0.5H) (E/Z), 8.74 (d, *J* = 3.9 Hz, 0.5H)/8.54 (d, *J* = 4.0 Hz, 0.5H) (E/Z), 8.25 (s, 0.5H)/8.19 (s, 0.5H) (E/Z), 7.50 – 7.44 (m, 1H), 7.22 – 7.16 (m, 1H), 7.12 – 7.00 (m, 3H), 4.48 (s, 0.5H)/4.47 (s, 0.5H) (E/Z).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 179.34/179.25 (E/Z), 162.87 (d, *J*_{C-F} = 245.1 Hz), 161.34/159.77 (E/Z), 159.73/158.62 (E/Z), 151.45/151.31 (E/Z), 142.41, 140.78 (d, *J*_{C-F} = 10.4 Hz)/ 140.53 (d, *J*_{C-F} = 10.3 Hz) (E/Z), 137.48/137.12 (E/Z), 129.82 (d, *J*_{C-F} = 8.7 Hz), 127.52, 125.32 (d, *J*_{C-F} = 21.1 Hz), 120.12, 116.73 (d, *J*_{C-F} = 23.7 Hz)/ 116.58 (d, *J*_{C-F} = 23.4 Hz) (E/Z), 115.13/114.96 (E/Z), 114.43 (d, *J* = 21.8 Hz), 87.54, 72.92.

¹⁹F NMR (471 MHz, THF-*d*₈) δ ppm: -114.11.

HRMS(ESI+) *m/z*: calculated for C₁₇H₁₀FN₂O₃S⁺ (M+H)⁺ : 341.0318; found: 341.0391.

5-((5-ethynylfuran-2-yl)methylene)-1-(2-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **5c**



Using Method A of Step 1: **2c** (50 mg, 0.21 mmoles) and **1a** (41 mg, 1.02 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 5.4 mg, 7.6% (E/Z mixture; 50:50).

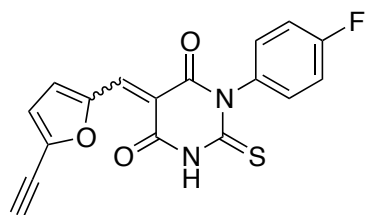
¹H NMR (500 MHz, THF-*d*₈) δ ppm : 12.13 (bs, 0.5H)/12.06 (bs, 0.5H) (E/Z), 8.76 (d, *J* = 3.9 Hz, 0.5H)/8.55 (d, *J* = 3.9 Hz, 0.5H) (E/Z), 8.28 (s, 0.5H)/8.20 (s, 0.5H) (E/Z), 7.46 (t, *J* = 6.9 Hz, 1H), 7.37 – 7.30 (m, 1H), 7.29 – 7.22 (m, 2H), 7.04 (dd, *J* = 26.9 (E/Z), 3.9 Hz, 1H), 4.48 (s, 1H).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 178.73, 160.89/159.27 (E/Z), 159.6, 158.23 (d, *J*_{C-F} = 233.0 Hz), 151.46/151.31 (E/Z), 142.55, 137.90/137.60 (E/Z), 131.06 (d, *J*_{C-F} = 20.7 Hz), 130.37 (d, *J*_{C-F} = 6.7 Hz), 127.75, 124.21, 120.15, 115.79 (d, *J*_{C-F} = 19.8 Hz), 114.46, 114.01/113.80 (E/Z), 87.59, 72.91.

¹⁹F NMR (471 MHz, THF-*d*₈) δ ppm: -122.65/-122.77 (E/Z).

HRMS(ESI+) *m/z*: calculated for C₁₇H₁₀FN₂O₃S⁺ (M+H)⁺ : 341.0318; found: 341.0391.

5-((5-ethynylfuran-2-yl)methylene)-1-(4-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **5d**



Using Method A of Step 1: **2b** (50 mg, 0.21 mmoles) and **1a** (41 mg, 1.01 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 6.7 mg, 9.3% (E/Z mixture; 50:50).

¹H NMR (500 MHz, THF-*d*₈) δ ppm : 12.02 (bs, 0.5H)/11.96 (bs, 0.5H) (E/Z), 8.73 (d, *J* = 3.9 Hz, 0.5H)/8.54 (d, *J* = 3.9 Hz, 0.5H) (E/Z), 8.24

(s, 0.5H)/8.18 (s, 0.5H) (E/Z), 7.29 – 7.24 (m, 2H), 7.22 – 7.18 (m, 2H), 7.06 (d, $J = 3.9$ Hz, 0.5H)/7.01 (d, $J = 3.9$ Hz, 0.5H) (E/Z), 4.47 (s, 1H).

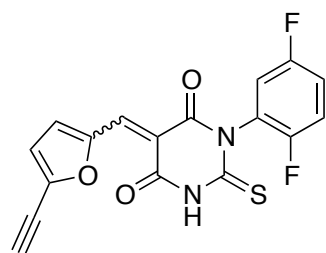
^{13}C NMR (126 MHz, THF- d_8) δ ppm: 179.70/179.60 (E/Z), 162.37 (d, $J_{\text{C-F}} = 245.7$ Hz), 161.52/158.61 (E/Z), 159.96/159.71 (E/Z), 151.47/151.33 (E/Z), 142.35, 137.47/137.05 (E/Z), 135.47/135.19 (E/Z), 131.10 (d, $J_{\text{C-F}} = 8.9$ Hz), 130.93 (d, $J_{\text{C-F}} = 8.7$ Hz), 127.42, 120.09, 115.39 (d, $J_{\text{C-F}} = 23.1$ Hz, 2C), 114.61/114.45 (E/Z), 87.49, 72.92.

^{19}F NMR (471 MHz, THF- d_8) δ ppm: -115.05/-115.06 (E/Z).

HRMS(ESI+) m/z : calculated for $\text{C}_{17}\text{H}_{10}\text{FN}_2\text{O}_3\text{S}^+$ (M+H) $^+$: 341.0318; found: 341.0391.

1-(2,5-difluorophenyl)-5-((5-ethynylfuran-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione

5e



Using Method A of Step 1: **2f** (100 mg, 0.39 mmoles) and **1a** (75 mg, 1.01 eq) was dissolved in anhydrous MeOH (3 mL). Purified by Flash chromatography in (cHex/EtOAc). Brown powder, Yield: 14.3 mg, 10.2%. Traces of grease in sample due to cHex (E/Z mixture; 50:50).

^1H NMR (500 MHz, THF- d_8) δ ppm: 12.19 (bs, 0.5H)/12.13 (bs, 0.5H) (E/Z), 8.76 (d, $J = 3.9$ Hz, 0.5H)/8.56 (d, $J = 3.9$ Hz, 0.5H) (E/Z), 8.29 (s, 0.5H)/8.21 (s, 0.5H) (E/Z), 7.33 – 7.23 (m, 2H), 7.22 – 7.14 (m, 1H), 7.08

(d, $J = 3.9$ Hz, 0.5H)/7.03 (d, $J = 3.9$ Hz, 0.5H)(E/Z), 4.49 (s, 1H).

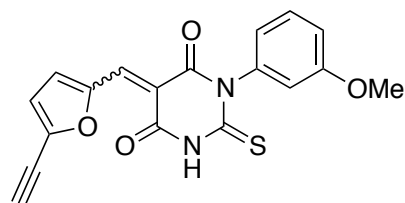
^{13}C NMR (126 MHz, THF- d_8) δ ppm: 178.58/ 178.45 (E/Z), 160.83/159.64 (E/Z), 158.41 (d, $J_{\text{C-F}} = 242.6$ Hz), 159.17/158.54 (E/Z), 154.85 (d, $J = 248.1$ Hz)/ 154.72 (d, $J = 245.7$ Hz)(E/Z), 151.41/151.25 (E/Z), 142.76/ 142.71 (E/Z), 138.05/137.83 (E/Z), 128.03/127.96 (E/Z), 127.57 – 127.00 (m), 120.22, 118.02 (d, $J = 17.8$ Hz)/ 117.82 (d, $J = 17.7$ Hz)(E/Z), 117.09-116.60 (m, 2C), 113.76/113.54 (E/Z), 87.76/87.73 (E/Z), 72.88.

^{19}F NMR (471 MHz, THF- d_8) δ ppm: -119.29 (d, $J = 2.9$ Hz)/-119.32 (d, $J = 2.7$ Hz) (E/Z), -127.66 (d, $J_{\text{F-F}} = 16.2$ Hz)/-127.80 (d, $J_{\text{F-F}} = 16.2$ Hz) (E/Z).

HRMS(ESI+) m/z : calculated for $\text{C}_{17}\text{H}_9\text{F}_2\text{N}_2\text{O}_3\text{S}^+$ (M+H) $^+$: 358.0224; found: 359.0296.

5-((5-ethynylfuran-2-yl)methylene)-1-(3-methoxyphenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione

5f



Using Method A of Step 1: **2d** (43 mg, 0.17 mmoles) and **1a** (33 mg, 1 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 5.4 mg, 8.9% (E/Z mixture; 50:50).

^1H NMR (500 MHz, THF- d_8) δ ppm: 11.96 (bs, 0.5H)/11.90 (bs, 0.5H) (E/Z), 8.73 (d, $J = 3.9$ Hz, 0.5H)/8.55 (d, $J = 3.9$ Hz, 0.5H)

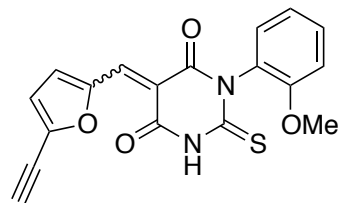
(E/Z), 8.24 (s, 0.5H)/8.17 (s, 0.5H) (E/Z), 7.33 (td, $J = 8.0, 5.3$ Hz, 1H), 7.05 (d, $J = 3.9$ Hz, 0.5H)/7.00 (d, $J = 4.0$ Hz, 0.5H) (E/Z), 6.99 – 6.94 (m, 1H), 6.85 – 6.78 (m, 2H), 4.46 (s, 1H), 3.80 (s, 3H).

^{13}C NMR (126 MHz, THF- d_8) δ ppm: 179.31, 161.24/158.62 (E/Z), 160.38, 159.72, 151.51/ 151.37 (E/Z), 142.22, 140.35/140.06 (E/Z), 137.33/136.90 (E/Z), 129.02, 127.31/127.24(E/Z), 121.25/ 121.09 (E/Z), 120.04, 114.85/114.73 (E/Z), 114.56, 113.77, 87.38, 72.95, 54.53.

HRMS(ESI+) m/z : calculated for $C_{18}H_{13}N_2O_4S^+$ ($M+H$) $^+$: 353.0518; found: 353.0591

5-((5-ethynylfuran-2-yl)methylene)-1-(2-methoxyphenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione

5g



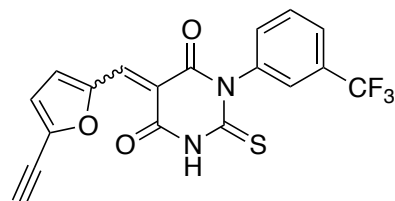
Using Method A of Step 1: **2e** (30 mg, 0.12 mmoles) and **1a** (23 mg, 1 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 5.9 mg, 13.9% (E/Z mixture; 50:50).

1H NMR (500 MHz, THF- d_6) δ ppm: 11.97 (bs, 0.5H)/11.90 (bs, 0.5H) (E/Z), 8.74 (d, $J = 4.0$ Hz, 0.5H)/8.54 (d, $J = 3.9$ Hz, 0.5H) (E/Z), 8.25 (s, 0.5H)/8.17 (s, 0.5H) (E/Z), 7.38 (q, $J = 7.1, 6.5$ Hz, 1H), 7.20 – 7.13 (m, 1H), 7.12 – 7.04 (m, 1H), 7.03 – 6.96 (m, 2H), 4.46 (s, 1H), 3.78 (s, 3H).

^{13}C NMR (126 MHz, THF- d_6) δ ppm: 179.08 /179.00 (E/Z), 160.73/159.71 (E/Z), 159.22/158.62 (E/Z), 155.27/155.12 (E/Z), 151.54/151.40 (E/Z), 142.29/142.25 (E/Z), 137.69/137.27 (E/Z), 130.13/129.99 (E/Z), 129.61, 127.88/127.58 (E/Z), 127.42/127.33 (E/Z), 120.13, 120.05/120.01 (E/Z), 114.32/114.15 (E/Z), 111.67, 87.38, 72.95, 54.97.

HRMS(ESI+) m/z : calculated for $C_{18}H_{13}N_2O_4S^+$ ($M+H$) $^+$: 353.0518; found: 353.0591.

5-((5-ethynylfuran-2-yl)methylene)-2-thioxo-1-(3-(trifluoromethyl)phenyl)dihydropyrimidine-4,6(1*H*,5*H*)-dione **5h**



Using Method A of Step 1: **2h** (30 mg, 0.11 mmoles) and **1a** (23 mg, 1 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 5.9 mg, 13.9% (E/Z mixture; 50:50).

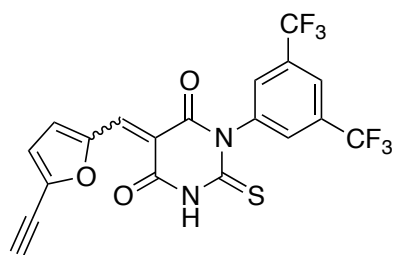
1H NMR (500 MHz Acetone- d_6) δ ppm: 11.80 (bs, 0.5H)/11.75 (bs, 0.5H) (E/Z), 8.72 (d, $J = 3.9$ Hz, 0.5H)/8.50 (d, $J = 4.0$ Hz, 0.5H) (E/Z), 8.21 (s, 0.5H)/8.15 (s, 0.5H) (E/Z), 7.88 – 7.77 (m, 3H), 7.72 (ddt, $J = 13.4, 8.0, 1.6$ Hz, 1H), 7.21 (dd, $J = 4.0, 0.8$ Hz, 0.5H)/7.15 (dd, $J = 4.0, 0.8$ Hz, 0.5H) (E/Z), 4.67 (s, 0.5H)/4.67 (s, 0.5H) (E/Z).

^{13}C NMR (126 MHz, Acetone- d_6) δ ppm: 179.63/179.52 (E/Z), 161.95/158.69 (E/Z), 159.98/160.16 (E/Z), 151.27/151.13 (E/Z), 142.35, 140.35/140.11 (E/Z), 137.90/137.50 (E/Z), 133.66/133.49 (E/Z), 131.02 – 130.53 (m), 130.21/130.16 (E/Z), 128.11/128.02 (E/Z), 126.50/126.36 (E/Z), 125.34, 124.12 (d, $J_{C-F} = 271.5$ Hz), 120.76, 114.67/114.55 (E/Z), 88.21, 72.85.

^{19}F NMR (471 MHz, Acetone- d_6) δ ppm: -63.02/ -63.04 (E/Z).

HRMS(ESI+) m/z : calculated for $C_{18}H_{10}F_3N_2O_3S^+$ ($M+H$) $^+$: 391.0286; found: 391.0359.

1-(3,5-bis(trifluoromethyl)phenyl)-5-((5-ethynylfuran-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **5i**



Using Method A of Step 1: **2i** (41 mg, 0.12 mmol) and **1a** (22 mg, 1 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 5.2 mg, 9.8% (E/Z mixture; 50:50).

¹H NMR (500 MHz, THF-*d*₈) δ ppm: 12.24 (bs, 0.5H)/12.18 (bs, 0.5H) (E/Z), 8.77 (d, *J* = 4.0 Hz, 0.5H)/8.55 (d, *J* = 3.9 Hz, 0.5H) (E/Z), 8.31 (s, 0.5H)/8.22 (s, 0.5H) (E/Z), 8.15 (d, *J* = 7.1 Hz, 1H),

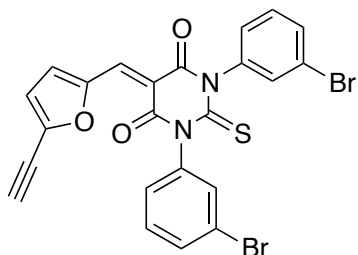
8.00 (s, 1H), 7.97 (s, 1H), 7.10 (d, *J* = 3.9 Hz, 0.5H)/7.04 (d, *J* = 3.9 Hz, 0.5H) (E/Z), 4.52 (s, 0.5H)/4.51 (s, 0.5H)(E/Z).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 179.36/179.23 (E/Z), 161.57/158.57 (E/Z), 159.90/159.69 (E/Z), 151.33/151.17 (E/Z), 142.86, 140.97, 137.91/137.73 (E/Z), 132.02 (d, *J*_{C-F} = 34.2 Hz, 2C), 130.65/130.49 (E/Z), 128.19, 123.26 (d, *J*_{C-F} = 272.4 Hz, 2C), 122.28, 120.32/120.28 (E/Z), 113.97/113.75 (E/Z), 87.88, 72.84.

¹⁹F NMR (471 MHz, THF-*d*₈) δ ppm: -63.57, -63.59.

HRMS(ESI+) *m/z*: calculated for C₁₉H₉F₆N₂O₃S⁺ (M+H)⁺: 459.0160; found: 459.0233.

1,3-bis(3-bromophenyl)-5-((5-ethynylfuran-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **5j**



Using Method B of Step 1: **4b** (30 mg, 0.07 mmol) and **1a** (12 mg, 1 eq) was dissolved in distilled water (2 mL). Brown powder, Yield: 4.8 mg, 13.06%.

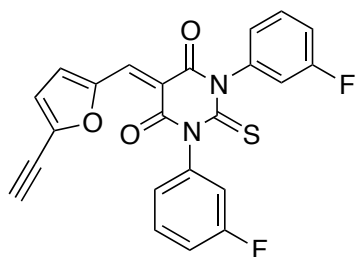
¹H NMR (500 MHz, THF-*d*₈) δ ppm: 8.62 (d, *J* = 4.0 Hz, 1H), 8.31 (s, 1H), 7.65 – 7.56 (m, 2H), 7.50 (dt, *J* = 13.4, 2.0 Hz, 2H), 7.45 – 7.38 (m,

2H), 7.32 – 7.22 (m, 2H), 7.05 (d, *J* = 3.9 Hz, 1H), 4.50 (s, 1H).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 180.2, 160.5, 159.0, 151.4, 142.7, 141.5, 141.2, 138.4, 132.2, 132.1, 131.2 (2C), 130.3 (2C), 128.2 (2C), 128.0, 121.7 (2C), 120.3, 114.2, 87.9, 72.9.

HRMS(ESI+) *m/z*: calculated for C₂₃H₁₃Br₂N₂O₃S⁺ (M+H)⁺: 557.2280; found: 556.8988.

5-((5-ethynylfuran-2-yl)methylene)-1,3-bis(3-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **5k**



Using Method B of Step 1: **4a** (296 mg, 0.89 mmol) and **1a** (171 mg, 1 eq) was dissolved in distilled water (5 mL). Orange-brown powder, Yield: 54.3 mg, 14.1%.

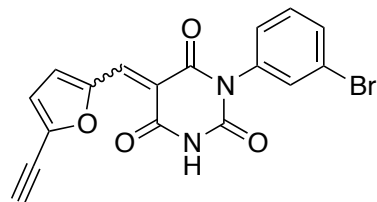
¹H NMR (500 MHz, THF-*d*₈) δ ppm: 8.61 (d, *J* = 4.0 Hz, 1H), 8.31 (s, 1H), 7.54 – 7.41 (m, 2H), 7.23 – 7.15 (m, 2H), 7.15 – 7.06 (m, 4H), 7.05 (d, *J* = 3.9 Hz, 1H), 4.50 (s, 1H).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 180.1, 163.0 (d, *J*_{C-F} = 245.3 Hz, 2C), 160.5, 159.0, 151.4, 142.8, 141.6 (d, *J*_{C-F} = 10.6 Hz), 141.3 (d, *J*_{C-F} = 10.4 Hz), 138.3, 130.0 (d, *J*_{C-F} = 8.8 Hz, 2C), 128.0, 125.1 (d, *J*_{C-F} = 21.9 Hz, 2C), 120.3, 116.6 (d, *J*_{C-F} = 18.8 Hz), 116.4 (d, *J*_{C-F} = 19.0 Hz), 115.0 (d, *J*_{C-F} = 21.1 Hz, 2C), 114.4, 87.8, 72.9.

¹⁹F NMR (471 MHz, THF-*d*₈) δ ppm: -113.91, -113.94.

HRMS(ESI+) m/z : calculated for $C_{23}H_{13}F_2N_2O_3S^+$ (M+H)⁺ : 435.0537; found: 435.0609.

1-(3-bromophenyl)-5-((5-ethynylfuran-2-yl)methylene)pyrimidine-2,4,6(1*H*,3*H*,5*H*)-trione **5l**



Using Method A of Step 1: **4j** (67.8 mg, 0.24 mmol) and **1a** (46.5 mg, 1 eq) was dissolved in anhydrous MeOH (2 mL). Brown powder, Yield: 8.5 mg, 9.2% (E/Z mixture; 50:50).

¹H NMR (500 MHz, Acetone-*d*₆) δ ppm: 10.55 (bs, 0.5H)/10.48 (bs, 0.5H) (E/Z), 8.63 (d, *J* = 3.9 Hz, 0.5H)/8.46 (d, *J* = 3.8 Hz, 0.5H) (E/Z),

8.20 (s, 0.5H)/ 8.16 (s, 0.5H)(E/Z), 7.76 – 7.63 (m, 2H), 7.54 – 7.42 (m, 2H), 7.14 (dd, *J* = 4.0, 0.8 Hz, 0.5H)/7.09 (dd, *J* = 4.0, 0.8 Hz, 0.5H)(E/Z), 4.52 (s, 0.5H)/4.52 (s, 0.5H)(E/Z).

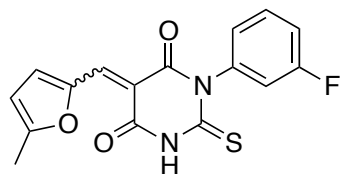
¹³C NMR (126 MHz, Acetone-*d*₆) δ ppm: 161.68, 161.41, 160.81, 151.11/151.00 (E/Z), 149.72/149.63 (E/Z), 141.77, 137.32/137.00 (E/Z), 132.41/132.25 (E/Z), 131.46, 130.51, 128.57/128.40 (E/Z), 127.10/126.96 (E/Z), 121.31/121.25 (E/Z), 120.29, 114.56/114.47 (E/Z), 87.43, 72.84.

HRMS(ESI+) m/z : calculated for $C_{17}H_{10}BrN_2O_4^+$ (M+H)⁺ : 384.9746; found: 384.9818.

6. Synthesis of analogues by Knoevenagel condensation in acidic condition.^[7]

1 equivalent of thiourea derivatives was dissolved in 2 mL of acetic acid, the solution became yellow in most cases. Furan derivative (1 eq) was added, and the solution mixture was stirred at rt for 15 minutes to 1 hour. Completion of the reaction was followed by TLC in 2 solvents systems (Hex/EA and DCM/MeOH to make sure the starting material was converted) and by UPLC-MS. When the reaction was completed, the solvent was evaporated *in vacuo*, and the crude solid was mixed with 10 mL of a mixture of cHex/EtOAc (4:6) and sonicated. The resultant solution was filtered with the same cHex/EtOAc mixture as before. The liquid was then evaporated to obtain a pure product. Further purifications are carried out for impure product and purification methods are written against the analogues.

1-(3-fluorophenyl)-5-((5-methylfuran-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **6a**



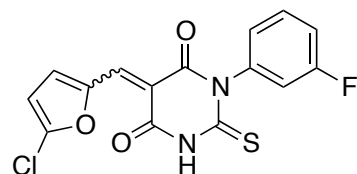
50 mg (0.21 mmol) of **2a** and 5-methylfuran-2-carbaldehyde (23.5 mg, 1 eq) were dissolved in 2 mL of acetic acid. Reaction time = 15 minutes. Yellow-orange powder, Yield: 9.2 mg, 13.2% (E/Z mixture; 50:50).

¹H NMR (500 MHz, Acetone-*d*₆) δ ppm: 11.58 (bs, 0.5H)/11.52 (bs, 0.5H) (E/Z), 8.77 (d, *J* = 3.8 Hz, 0.5H)/8.57 (d, *J* = 3.8 Hz, 0.5H) (E/Z), 8.25 (s, 0.5H)/ 8.18 (s, 0.5H)(E/Z), 7.67 – 7.48 (m, 1H), 7.39 – 7.12 (m, 3H), 6.69 (dt, *J* = 3.8, 0.8 Hz, 0.5H)/6.64 (dt, *J* = 3.8, 0.8 Hz, 0.5H) (E/Z), 2.53 (s, 3H).

¹³C NMR (126 MHz, Acetone-*d*₆) δ ppm: 179.42/179.33 (E/Z), 164.44/ 166.40 (E/Z), 163.74/163.69 (E/Z), 162.74 (d, *J*_{C-F} = 243.9 Hz), 161.80/160.22 (E/Z), 160.40/158.92 (E/Z), 150.35/150.20 (E/Z), 141.08 (d, *J*_{C-F} = 10.5 Hz), 138.91/138.46 (E/Z), 130.33 (d, *J*_{C-F} = 8.9 Hz), 125.78/125.62 (E/Z), 116.78 (d, *J*_{C-F} = 23.5 Hz), 115.26 (d, *J*_{C-F} = 21.1 Hz), 113.59, 110.83/110.73 (E/Z), 13.69.

HRMS(ESI+) m/z : calculated for $C_{16}H_{12}FN_2O_3S^+$ (M+H)⁺ : 331.0474; found: 331.0547

5-((5-chlorofuran-2-yl)methylene)-1-(3-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **6b**



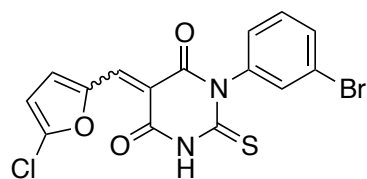
50 mg (0.21 mmoles) of **2a** and 5-methylfuran-2-carbaldehyde (27.5 mg, 1eq) were dissolved in 2 mL of acetic acid. Reaction time = 25 minutes. Yellow-orange powder, Yield: 8.2 mg, 11.1% (E/Z mixture; 50:50).

¹H NMR (Acetone-*d*₆, 500 MHz) δ ppm: 11.71 (bs, 0.5H) / 11.66 (bs, 0.5H)(E/Z), 8.75 (d, *J* = 3.9 Hz, 0.5H)/8.56 (d, *J* = 4.0 Hz, 0.5H) (E/Z), 8.18 (s, 0.5H)/8.12 (s, 0.5H) (E/Z), 7.64 – 7.53 (m, 1H), 7.37 – 7.16 (m, 3H), 6.94 (dd, *J* = 3.9, 0.7 Hz, 0.5H)/6.88 (dd, *J* = 4.0, 0.7 Hz, 0.5H) (E/Z).

¹³C NMR (126 MHz, Acetone-*d*₆) δ ppm: 179.46/179.38 (E/Z), 162.72 (d, *J*_{C-F} = 246.2 Hz), 161.75/159.95 (E/Z), 160.10/158.77 (E/Z), 151.05/150.93 (E/Z), 145.22, 141.00 (d, *J*_{C-F} = 10.0 Hz) / 140.73 (d, *J*_{C-F} = 10.4 Hz) (E/Z), 137.72/137.29 (E/Z), 130.45 (d, *J*_{C-F} = 5.8 Hz) / 130.38 (d, *J*_{C-F} = 5.6 Hz) (E/Z), 130.23/130.14 (E/Z), 125.70/125.54 (E/Z), 116.80 (d, *J*_{C-F} = 19.8 Hz) / 116.61 (d, *J*_{C-F} = 19.9 Hz) (E/Z), 115.40 (d, *J*_{C-F} = 21.0 Hz), 113.55/113.45 (E/Z), 113.00.

HRMS(ESI+) *m/z*: calculated for C₁₅H₉ClFN₂O₃S⁺ (M+H)⁺ : 350.9928; found: 351.0001.

5-((5-chlorofuran-2-yl)methylene)-1-(3-bromophenyl)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **6c**



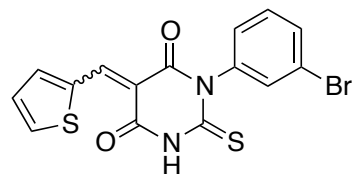
60 mg (0.20 mmoles) of **2g** and 5-chlorofuran-2-carbaldehyde (20 mg, 1 eq) were dissolved in 2 mL of acetic acid. Reaction time = 45 minutes. Yellow powder, Yield: 52.1 mg, 63.1% (E/Z mixture; 50:50).

¹H NMR (500 MHz, THF-*d*₈) δ ppm: 12.04 (bs, 0.5H) / 11.98 (bs, 0.5H) (E/Z), 8.79 (d, *J* = 3.9 Hz, 0.5H) / 8.60 (d, *J* = 3.9 Hz, 0.5H) (E/Z), 8.24 (s, 0.5H) / 8.17 (s, 0.5H) (E/Z), 7.59 (t, *J* = 6.2, 1H), 7.48 (d, *J* = 14.0 Hz, 1H), 7.44 – 7.35 (m, 1H), 7.29 – 7.19 (m, 1H), 6.81 (d, *J* = 3.9 Hz, 0.5H) / 6.76 (d, *J* = 3.9 Hz, 0.5H) (E/Z).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 179.37/179.27 (E/Z), 161.34/159.90 (E/Z), 159.69/158.72 (E/Z), 151.30/151.18 (E/Z), 145.03/144.98 (E/Z), 140.69/140.43 (E/Z), 137.35/137.01 (E/Z), 132.39/ 132.25 (E/Z), 131.24, 130.08, 129.75/129.68 (E/Z), 128.39/128.21 (E/Z), 121.53, 113.51/113.33 (E/Z), 112.48.

HRMS(ESI+) *m/z*: calculated for C₁₅H₉BrClN₂O₃S⁺ (M+H)⁺ : 412.6540; found: 412.9180.

1-(3-bromophenyl)-5-(thiophen-2-ylmethylene)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **6d**



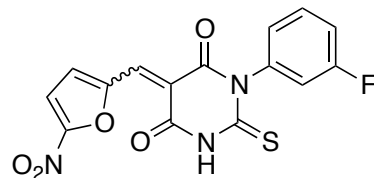
150 mg (0.50 mmoles) of **2g** and thiophene-2-carbaldehyde (49 μL, 1.05 eq) were dissolved in 1.5 mL of acetic acid. Reaction time = 40 minutes. Yellow powder, Yield: 32 mg, 16.2% (E/Z mixture; 50:50).

¹H NMR (500 MHz, THF-*d*₈) δ ppm: 11.93 (bs, 1H), 8.79 (s, 0.5H) / 8.71 (s, 0.5H) (E/Z), 8.25 – 8.16 (m, 1H), 8.15 – 8.08 (m, 1H), 7.63 – 7.56 (m, 1H), 7.49 (dt, *J* = 17.8, 2.0 Hz, 1H), 7.39 (q, *J* = 8.1 Hz, 1H), 7.34 (q, *J* = 4.8 Hz, 1H), 7.29 – 7.19 (m, 1H).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 179.52/179.44 (E/Z), 161.78, 160.70, 147.78/147.44 (E/Z), 145.37, 142.34, 140.87/140.37 (E/Z), 137.48/137.23 (E/Z), 132.49/132.33 (E/Z), 131.20/131.14 (E/Z),

130.05/130.01 (E/Z), 128.47/128.28 (E/Z), 128.13, 121.52 /121.46 (E/Z), 112.14/111.98 (E/Z). HRMS(ESI+) *m/z*: calculated for C₁₅H₁₀BrN₂O₂S₂⁺ (M+H)⁺ : 392.9289; found: 392.9362.

1-(3-fluorophenyl)-5-((5-nitrofuran-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **6e**



50mg (0.21 mmoles) of **2a** and 5-nitrofuran-2-carbaldehyde (23 mg, 1eq) were dissolved in 2 mL of acetic acid. Reaction time = 25 minutes. Purified by flash chromatography on silica using EtOAc-cHex. Yellow powder, Yield: 4 mg, 5.2% (E/Z mixture; 50:50).

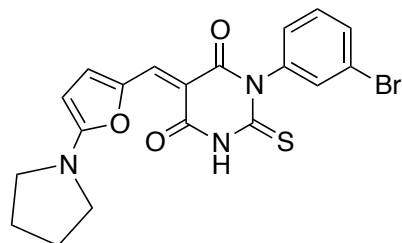
¹H NMR (500 MHz, THF-*d*₈) δ ppm: 12.23 (bs, 0.5H)/12.17 (bs, 0.5H) (E/Z), 8.70 (d, *J* = 4.1 Hz, 0.5H)/8.52 (d, *J* = 4.1 Hz, 0.5H) (E/Z), 8.27 (s, 0.5H)/8.21 (s, 0.5H) (E/Z), 7.67 (d, *J* = 4.1 Hz, 0.5H)/7.64 (d, *J* = 4.1 Hz, 0.5H)(E/Z), 7.56 – 7.40 (m, 1H), 7.31 – 7.17 (m, 1H), 7.14 – 7.00 (m, 2H).

¹³C NMR (126 MHz, THF-*d*₈) δ ppm: 179.32/179.23 (E/Z), 162.87 (d, *J*_{C-F} = 242.8 Hz), 160.69/159.63 (E/Z), 159.03/158.38 (E/Z), 154.03, 150.89/150.76 (E/Z), 140.34 (d, *J*_{C-F} = 21.9 Hz), 136.41 (d, *J*_{C-F} = 41.0 Hz), 130.02/129.95 (E/Z), 126.30, 125.21 (d, *J*_{C-F} = 20.6 Hz), 119.78/119.58 (E/Z), 116.63 (d, *J*_{C-F} = 17.3 Hz)/116.44 (d, *J*_{C-F} = 16.8 Hz) (E/Z), 115.28 (d, *J*_{C-F} = 21.1 Hz), 112.74.

¹⁹F NMR (471 MHz, THF-*d*₈) δ ppm: -113.85/ -113.88 (E/Z).

HRMS(ESI+) *m/z*: calculated for C₁₅H₉FN₃O₅S⁺ (M+H)⁺ : 362.0169; found: 362.0241

1-(3-bromophenyl)-5-((5-(pyrrolidin-1-yl)furan-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **6f**



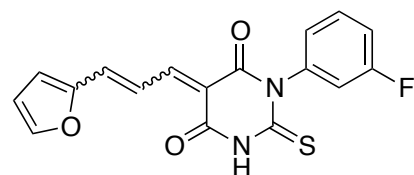
50 mg (0.17 mmoles) of **2g** and 5-(pyrrolidin-1-yl)furan-2-carbaldehyde (28 mg, 1.05 eq) were dissolved in 2 mL of acetic acid. Reaction time = 40 minutes. Purified by flash chromatography on silica using EtOAc-cHex. Yellow-Orange powder, Yield: 20.2 mg, 27.1%.

¹H NMR (500 MHz, CD₂Cl₂) δ ppm: 9.20 (s, 1H), 7.90 (s, 1H), 7.60 (d, *J* = 8.0 Hz, 1H), 7.48 – 7.37 (m, 2H), 7.24 (d, *J* = 7.9 Hz, 1H), 6.03 (d, *J* = 4.9 Hz, 1H), 3.70 (m, 4H), 2.14 (t, *J* = 4.1 Hz, 4H).

¹³C NMR (126 MHz, CD₂Cl₂) δ ppm: 177.5, 165.5 (3C), 147.4, 143.0, 141.1, 132.3, 131.4, 130.3, 128.2 (2C), 121.9, 102.0, 96.5, (49.68, 49.51, 49.34, 49.17, 49.00, 48.83, 48.66) (2C), 25.2 (2C).

HRMS(ESI+) *m/z*: calculated for C₁₉H₁₇BrN₃O₃S⁺ (M+H)⁺ : 446.0096; found: 446.0169.

(1-(3-fluorophenyl)-5-(3-(furan-2-yl)allylidene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **6g**



50 mg (0.21 mmoles) of **2a** and 3-(furan-2-yl)acrylaldehyde (26 mg, 1 eq) were dissolved in 2 mL of acetic acid. Reaction time = 45 min; Orange Red powder, Yield: 11.3 mg, 15.72% (E/Z mixture; 50:50).

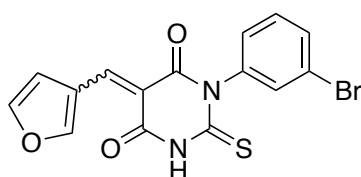
¹H NMR (500 MHz, THF-*d*₈) δ ppm: 11.81 (bs, 0.5H)/11.79 (bs, 0.5H) (E/Z), 8.52 (dd, *J* = 15.2, 12.3 Hz, 0.5H)/8.38 (dd, *J* = 15.1, 12.4 Hz, 0.5H) (E/Z), 8.17 (d, *J* = 12.4 Hz, 0.5H)/8.12 (d, *J* = 12.4 Hz, 0.5H)

(E/Z), 7.83 (d, $J = 1.8$ Hz, 0.5H)/ 7.78 (d, $J = 1.8$ Hz, 0.5H) (E/Z), 7.52 – 7.38 (m, 2H), 7.20 – 7.13 (m, 1H), 7.10 – 7.02 (m, 2H), 6.99 (t, $J = 3.3$ Hz, 1H), 6.67 (dd, $J = 3.5, 1.8$ Hz, 0.5H)/6.64 (dd, $J = 3.6, 1.8$ Hz, 0.5H) (E/Z).

^{13}C NMR (126 MHz, THF- d_8) δ ppm: 179.53, 162.81 (d, $J_{\text{C-F}} = 245.8$ Hz), 161.34/159.43 (E/Z), 160.50/159.81 (E/Z), 155.39/154.81 (E/Z), 152.68, 147.26/147.21 (E/Z), 1140.90 (d, $J_{\text{C-F}} = 10.4$ Hz)/ 140.68 (d, $J_{\text{C-F}} = 10.5$ Hz) (E/Z), 138.48/138.41 (E/Z), 129.66 (d, $J_{\text{C-F}} = 8.8$ Hz), 125.45/125.34 (E/Z), 123.26/ 123.06 (E/Z), 117.96/117.83 (E/Z), 116.78 (d, $J = 23.8$ Hz)/116.67 (d, $J_{\text{C-F}} = 23.4$ Hz)(E/Z), 115.46/115.39 (E/Z), 114.85 (d, $J_{\text{C-F}} = 21.2$ Hz), 113.30.

HRMS(ESI+) m/z : calculated for $\text{C}_{17}\text{H}_{12}\text{FN}_2\text{O}_3\text{S}^+$ (M+H) $^+$: 343.0474; found: 343.0546.

1-(3-bromophenyl)-5-(furan-3-ylmethylene)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **6h**

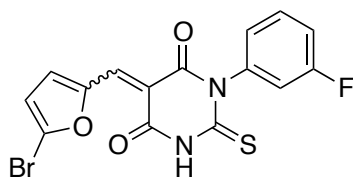


70 mg (0.23 mmoles) of **2g** and furan-3-carbaldehyde (19.5 μL , 1.05 eq) were dissolved in 1.5 mL of acetic acid. Reaction time = 45 minutes. Yellow powder, Yield: 42.8 mg, 48.4% (E/Z mixture; 50:50). ^1H NMR (500 MHz, THF- d_8) δ ppm: 11.95 (bs, 0.5H)/11.92 (bs, 0.5H) (E/Z), 8.94 (s, 0.5H)/8.86 (s, 0.5H) (E/Z), 8.44 (s, 0.5H)/8.37 (s, 0.5H)

(E/Z), 7.72 (s, 0.5H)/7.69 (s, 0.5H) (E/Z), 7.63 – 7.56 (m, 1H), 7.52 – 7.32 (m, 3H), 7.29 – 7.19 (m, 1H). ^{13}C NMR (126 MHz, THF- d_8) δ ppm: 179.57/179.48 (E/Z), 161.83/158.87 (E/Z), 160.14/160.06 (E/Z), 155.04, 146.31/146.06 (E/Z), 144.55, 140.82/140.56 (E/Z), 132.43/132.30 (E/Z), 131.18, 130.05, 128.42/128.25 (E/Z), 122.17/121.95 (E/Z), 121.52/121.48 (E/Z), 116.23/116.04 (E/Z), 113.74/113.68 (E/Z).

HRMS(ESI+) m/z : calculated for $\text{C}_{15}\text{H}_{10}\text{BrN}_2\text{O}_3\text{S}^+$ (M+H) $^+$: 376.9517; found: 376.9590.

5-((5-bromofuran-2-yl)methylene)-1-(3-fluorophenyl)-2-thioxodihydropyrimidine-4,6(1H,5H)-dione **6i**



50 mg (0.21 mmoles) of **2a** and 5-bromofuran-2-carbaldehyde (37 mg, 1 eq) were dissolved in 2 mL of acetic acid. Reaction time = 25 minutes. Yellow powder, Yield: 8.9 mg; 10.7% (E/Z mixture; 50:50). ^1H NMR (500 MHz, THF- d_8) δ ppm: 12.02 (bs, 0.5H)/11.96 (bs, 0.5H) (E/Z), 8.72 (d, $J = 3.9$ Hz, 0.5H)/8.53 (d, $J = 3.9$ Hz, 0.5H)

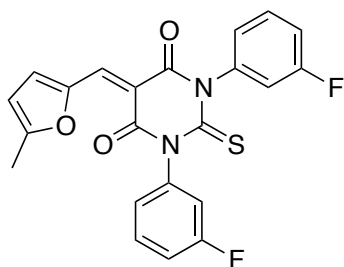
(E/Z), 8.27 (s, 0.5H)/8.20 (s, 0.5H) (E/Z), 7.55 – 7.39 (m, 1H), 7.27 – 7.14 (m, 1H), 7.14 – 7.02 (m, 2H), 6.92 (d, $J = 3.9$ Hz, 0.5H)/6.87 (d, $J = 3.9$ Hz, 0.5H) (E/Z).

^{13}C NMR (126 MHz, THF- d_8) δ ppm: 179.35, 162.82 (d, $J_{\text{C-F}} = 256.0$ Hz), 161.31/158.73 (E/Z), 159.87/159.70 (E/Z), 153.60/153.47 (E/Z), 140.85/140.58 (E/Z), 137.25/136.89 (E/Z), 132.80/132.85, 129.79 (d, $J_{\text{C-F}} = 8.8$ Hz), 129.48/129.46 (E/Z), 125.32 (d, $J_{\text{C-F}} = 19.2$ Hz), 117.39, 116.72 (d, $J_{\text{C-F}} = 23$ Hz)/116.58 (d, $J_{\text{C-F}} = 23.4$ Hz) (E/Z), 115.01 (d, $J_{\text{C-F}} = 20.6$ Hz), 113.31/ 113.15 (E/Z).

^{19}F NMR (471 MHz, THF- d_8) δ ppm: -114.16/ -114.16 (E/Z).

HRMS(ESI+) m/z : calculated for $\text{C}_{15}\text{H}_9\text{BrFN}_2\text{O}_3\text{S}^+$ (M+H) $^+$: 396.2024; found: 396.9475.

1,3-bis(3-fluorophenyl)-5-((5-methylfuran-2-yl)methylene)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione
6j



100 mg (0.30 mmoles) of **4a** and 5-methylfuran-2-carbaldehyde (29.6 μ L, 1 eq) were dissolved in 2 mL of acetic acid. Reaction time = 30 minutes. Purified by flash chromatography on silica using EtOAc-cHex. Yellow-orange powder, Yield: 28 mg, 21.8%.

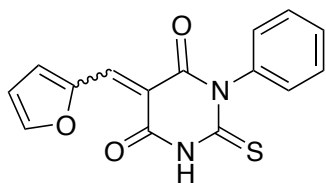
^1H NMR (500 MHz, THF- d_8) δ ppm: 8.64 (d, $J=3.8$ Hz, 1H), 8.33 (s, 1H), 7.53 – 7.32 (m, 2H), 7.28 – 6.95 (m, 6H), 6.52 (d, $J=3.7$ Hz, 1H), 2.46 (s, 3H).

^{13}C NMR (126 MHz, THF- d_8) δ ppm: 180.2, 164.2, 163.0 (d, $J_{\text{C-F}} = 244.8$ Hz, 2C), 160.9, 159.2, 150.6, 141.8 (d, $J_{\text{C-F}} = 10.3$ Hz), 141.5 (d, $J_{\text{C-F}} = 10.6$ Hz), 139.4, 131.3, 129.9 (d, $J_{\text{C-F}} = 8.9$ Hz, 2C), 125.3 (d, $J_{\text{C-F}} = 23.0$ Hz, 2C), 116.7 (d, $J_{\text{C-F}} = 19.7$ Hz), 116.5 (d, $J_{\text{C-F}} = 20.3$ Hz), 114.8 (d, $J_{\text{C-F}} = 21.0$ Hz, 2C), 113.2, 110.8, 13.4.

^{19}F NMR (471 MHz, THF- d_8) δ ppm: -114.09, -114.11.

HRMS(ESI+) m/z : calculated for $\text{C}_{22}\text{H}_{15}\text{F}_2\text{N}_2\text{O}_3\text{S}^+$ ($\text{M}+\text{H}^+$): 425.0693; found: 425.0766.

5-(furan-2-ylmethylene)-1-phenyl-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **6k**



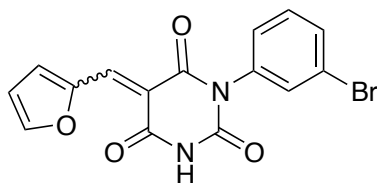
100 mg (0.45 mmoles) of **2k** and furan-2-carbaldehyde (39.6 μ L, 1 eq) were dissolved in 2 mL of acetic acid. Reaction time = 45 minutes. Brown powder, Yield: 25 mg, 18% (E/Z mixture; 50:50).

^1H NMR (500 MHz, THF- d_8) δ ppm: 11.93 (s, 1H)/11.87 (s, 1H) (E/Z), 8.79 (d, $J = 3.8$ Hz, 1H)/8.59 (d, $J = 3.8$ Hz, 1H) (E/Z), 8.36 (s, 1H)/8.28 (s, 1H) (E/Z), 8.11 (d, $J = 1.6$ Hz, 1H)/8.09 (d, $J = 1.6$ Hz, 1H) (E/Z), 7.50 – 7.35 (m, 6H), 7.30 – 7.18 (m, 4H), 6.86 (d, $J = 3.0$ Hz, 1H)/6.80 (d, $J = 3.1$ Hz, 1H) (E/Z).

^{13}C NMR (126 MHz, THF- d_8) δ ppm : 179.65/179.57 (E/Z), 161.62/158.76 (E/Z), 159.96, 151.45/151.30 (E/Z), 151.15/151.07 (E/Z), 139.58/139.31(E/Z), 138.91/138.48(E/Z), 129.18, 129.01, 128.51, 128.48, 127.89, 127.52/127.45 (E/Z), 114.96, 113.32/ 113.15 (E/Z).

HRMS(ESI+) m/z : calculated for $\text{C}_{15}\text{H}_{11}\text{N}_2\text{O}_3\text{S}^+$ ($\text{M}+\text{H}^+$): 299.0412; found :299.0485

1-(3-bromophenyl)-5-(furan-2-ylmethylene)pyrimidine-2,4,6(1*H*,3*H*,5*H*)-trione **6l**



100 mg (0.35 mmoles) of **2j** and furan-2-carbaldehyde (31 μ L, 1.05 eq) were dissolved in 2 mL of acetic acid. Reaction time = 15 minutes. Brown powder, Yield: 18 mg, 14% (E/Z mixture; 50:50).

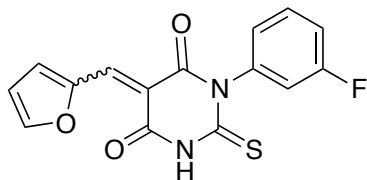
^1H NMR (500 MHz, DMSO- d_6) δ ppm : 11.77 (s, 0.5H)/11.69 (s, 0.5H) (E/Z), 8.55 (d, $J = 3.8$ Hz, 0.5H)/8.37 (d, $J = 3.8$ Hz, 0.5H) (E/Z), 8.31 (s, 0.5H)/8.29 (s, 0.5H) (E/Z), 8.16 (s, 0.5H)/8.09 (s, 0.5H) (E/Z), 7.72 – 7.56 (m, 2H), 7.52 – 7.43 (m, 1H), 7.42 – 7.32 (m, 1H), 6.96 (s, 0.5H)/6.89 (s, 0.5H) (E/Z).

^{13}C NMR (126 MHz, DMSO- d_6) δ ppm: 162.92, 162.34, 161.36/161.26 (E/Z), 151.64/ 151.60 (E/Z), 150.22/150.10/149.99 (E/Z), 137.57/137.39 (E/Z), 137.00/136.72 (E/Z), 132.13/131.98 (E/Z), 131.29,

130.69/130.64 (E/Z), 128.62/128.46 (E/Z), 127.11/126.93 (E/Z), 121.01/120.94 (E/Z), 115.41/115.38 (E/Z), 112.97/ 112.84 (E/Z).

HRMS(ESI+) m/z : calculated for $C_{15}H_{10}BrN_2O_4^+$ (M+H) $^+$: 360.9746; found :360.9818

1-(3-fluorophenyl)-5-(furan-2-ylmethylene)-2-thioxodihydropyrimidine-4,6(1*H*,5*H*)-dione **6m**



30 mg (0.13 mmoles) of **2g** and furan-2-carbaldehyde (12 μ L, 1.1 eq) were dissolved in 1.5 mL of acetic acid. Reaction time = 45 minutes. Yellow powder, Yield: 10 mg, 25% (E/Z mixture; 50:50).

1H NMR (500 MHz, THF- d_8) δ ppm: 11.99 (bs, 0.5H)/11.93 (bs, 0.5H) (E/Z), 8.79 (dt, J = 3.8, 0.5 Hz, 0.5H)/8.60 (dt, J = 3.9, 0.5 Hz, 0.5H)

(E/Z), 8.36 (s, 0.5H)/ 8.29 (s, 0.5H) (E/Z), 8.12 (dd, J = 1.7, 0.6 Hz, 0.5H)/8.11 (dd, J = 1.7, 0.6 Hz, 0.5H) (E/Z), 7.55 – 7.41 (m, 1H), 7.24 – 7.14 (m, 1H), 7.13 – 7.03 (m, 2H), 6.87 (ddd, J = 3.8, 1.7, 0.8 Hz, 0.5H)/6.81 (ddd, J = 3.9, 1.7, 0.9 Hz, 0.5H)(E/Z).

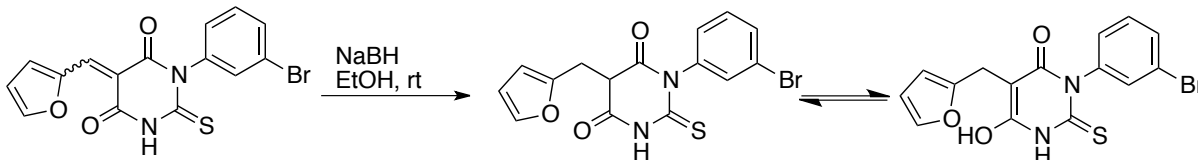
^{13}C NMR (126 MHz, THF- d_8) δ ppm : 180.44/180.35 (E/Z), 163.90 (d, J_{C-F} = 245.3 Hz)/ 163.86 (d, J_{C-F} = 245.3 Hz)(E/Z), 162.58/160.97 (E/Z) , 160.90/159.77 (E/Z), 152.43/152.28 (E/Z), 152.39/152.32 (E/Z), 141.83 (d, J_{C-F} = 30.6 Hz)/ 141.74 (d, J_{C-F} = 30.8 Hz) (E/Z), 140.05/139.70 (E/Z), 130.81 (d, J_{C-F} =8.4 Hz), 128.74, 126.46/126.29 (E/Z), 117.81 (d, J = 19.6 Hz)/117.62 (d, J = 19.6 Hz)(E/Z), 116.09, 115.92, 114.04 (d, J = 22.7 Hz).

^{19}F NMR (471 MHz, THF- d_8) δ ppm: -114.19/-114.20 (E/Z).

HRMS(ESI+) m/z : calculated for $C_{15}H_{10}FN_2O_3S^+$ (M+H) $^+$: 317.0318; found: 317.0393.

7. Reduction of some analogues^[7] :

3-(3-bromophenyl)-5-(furan-2-ylmethyl)-6-hydroxy-2-thioxo-2,3-dihydropyrimidin-4(1*H*)-one **7a**



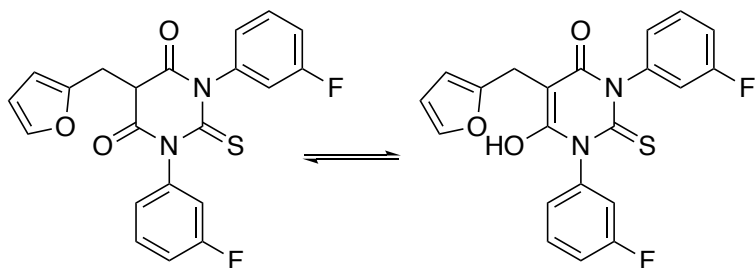
To SMIFH2 (23.1 mg, 0.06 mmoles) in EtOH (2mL) at 0°C, NaBH₄ (9.3 mg, 4 eq) was added, and the reaction was brought to r.t. gradually for 1.5 hours and quenched it with acetic acid. The solvents were evaporated under reduced pressure and the prep-HPLC in water/ACN was performed for purification. Colourless product was formed. Yield: 6 mg, 26.1%.

1H NMR (500 MHz, MeOD- d_4) δ ppm: 7.55 (dd, J = 8.1, 1.8 Hz, 1H), 7.43 (t, J = 1.9 Hz, 1H), 7.38 (t, J = 8.0 Hz, 1H), 7.27 – 7.21 (m, 2H), 6.21 (dd, J = 3.1, 1.9 Hz, 1H), 5.99 (d, J = 3.1 Hz, 1H), 3.75 (s, 2H), 3.37 (s, 1H, exchanged with Tautomer).

^{13}C NMR (126 MHz, MeOD- d_4) δ ppm: 175.5, 164.7 (2C), 155.6, 141.9, 139.9, 132.3, 130.6, 129.9, 128.2, 121.4, 109.7, 104.1, 90.8, 21.9.

HRMS(ESI+) m/z : calculated for $C_{15}H_{12}BrN_2O_3S^+$ (M+H) $^+$: 380.2284; found: 380.9726

1,3-bis(3-fluorophenyl)-5-(furan-2-ylmethyl)-6-hydroxy-2-thioxo-2,3-dihydropyrimidin-4(1*H*)-one **7b**



4a (100 mg, 0.12 mmoles) and furan-2-carbaldehyde (24.6 μ L, 1 eq) were solubilized in 2 mL Acetic Acid and stirred for 30 minutes at rt. The crude mixture was dried under vacuum and purified by flash

chromatography on silica using cHex-EtOAc. The product obtained (50 mg, 1 eq) was dissolved in EtOH (2mL) at 0°C, then NaBH₄(19 mg, 4 eq) was added, and the reaction was brought to r.t. gradually for 1.5 hours and then quenched with acetic acid. The solvents were evaporated under reduced pressure and the flash chromatography on silica gel, DCM MeOH (8/2). Colourless product was formed. Yield: 13.9 mg; 28%.

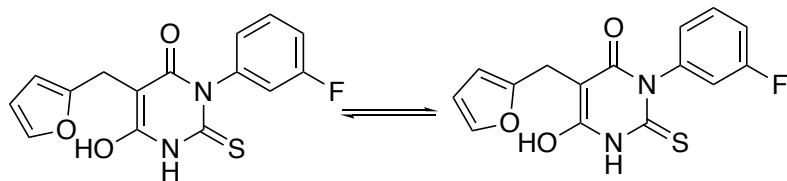
¹H NMR (500 MHz, MeOD-*d*₄) δ ppm: 7.46 – 7.38 (m, 2H), 7.28 (dd, *J* = 1.8, 0.9 Hz, 1H), 7.12 – 6.96 (m, 6H), 6.24 (dd, *J* = 3.1, 1.9 Hz, 1H), 6.01 (dd, *J* = 3.1, 1.0 Hz, 1H), 3.71 (s, 2H).

¹³C NMR (126 MHz, MeOD-*d*₄) δ ppm: 177.6, 163.4 (2C), 163.0 (d, *J*_{C-F} = 244.3 Hz, 2C), 155.8, 143.0 (d, *J*_{C-F} = 10.3 Hz, 2C), 139.8, 129.5 (d, *J*_{C-F} = 8.8 Hz, 2C), 125.2, 116.6 (d, *J*_{C-F} = 23.0 Hz, 2C), 113.9 (d, *J*_{C-F} = 21.2 Hz, 2C), 109.7, 104.1, 90.4, 22.4.

¹⁹F NMR (376 MHz, MeOD-*d*₄) δ ppm: -115.66, -115.68.

HRMS(ESI+) *m/z*: calculated for C₂₁H₁₅F₂N₂O₃S⁺ (M+H)⁺ : 413.0693; found: 413.0763.

3-(3-fluorophenyl)-5-(furan-2-ylmethyl)-6-hydroxy-2-thioxo-2,3-dihydropyrimidin-4(1H)-one **7c**



To **6m** (30.1 mg, 0.09 mmoles) in EtOH (2mL) at 0°C, NaBH₄ (11 mg, 3 eq) was added, and the reaction was brought to r.t. gradually for 1.5 hours and then

quenched with acetic acid. The solvents were evaporated under reduced pressure and the flash chromatography on silica, DCM/MeOH (8/2). Colourless product was formed. Yield: 9.2 mg; 30.5%.

¹H NMR (500 MHz, MeOD-*d*₄) δ ppm: 7.52 – 7.39 (m, 1H), 7.27 (s, 1H), 7.11 (td, *J* = 8.6, 2.5 Hz, 1H), 7.01 (d, *J* = 7.9 Hz, 1H), 6.96 (dt, *J* = 9.6, 2.3 Hz, 1H), 6.23 (t, *J* = 2.5 Hz, 1H), 5.96 (d, *J* = 3.1 Hz, 1H), 3.66 (s, 2H).

¹³C NMR (126 MHz, MeOD-*d*₄) δ ppm: 175.3, 163.2 (2C), 162.9 (d, *J*_{C-F} = 260.5 Hz), 155.7, 142.1 (d, *J*_{C-F} = 10.2 Hz), 139.8, 129.5 (d, *J*_{C-F} = 9.2 Hz), 125.2, 116.6 (d, *J*_{C-F} = 22.9 Hz), 114.2 (d, *J*_{C-F} = 21.3 Hz), 109.6, 103.9, 90.0, 21.7.

¹⁹F NMR (376 MHz, MeOD-*d*₄) δ ppm: -115.60.

HRMS(ESI+) *m/z*: calculated for C₁₅H₁₂FN₂O₃S⁺ (M+H)⁺ : 319.044; found: 319.0547.

9. Nucleophilic amino acids and **5k/7b** reaction:

Nucleophilic amino acids: Fmoc-Lysine-OtBu.HCl and Boc-Serine-OH 20 mM stock solution in MeOH and 1.75 mM stock solution in DMSO of **5k**, **7b** were prepared. In eppendorf tubes, 200 μ L of DPBS (1x) (Gibco), 50 μ L of **5k/7b** and 50 μ L of an amino acid were added and left at 37°C with sand bath for 12 hours. The samples were analysed by UPLC-MS after 2 and 12 hours.

B) CPMG filter NMR experiments

Compounds (1.75 mM stock solution) were made in deuterated DMSO and stored at -20°C. NMR samples were prepared in 558 μL D_2O , 12 μL (9 μM) of Compound **5k** and **6b** and 18.2 μL of DPBS (1x) or 18.2 μL (35 μM) of IFN γ (300 μM in PBS). Both samples were preincubated at 37°C for 20 minutes before running the NMR. All the ^{19}F NMR experiments were recorded at 37°C with a Bruker 500 spectrometer operating at a ^{19}F Larmor frequency of 470.5 MHz. The protocol was adapted from literature^[8]. The R2 filter experiments were recorded with the Carr-Purcell-Meibom-Gill scheme with a time interval of 40 ms between the 180° pulses and a length of 0.16 s. The spectra were acquired with proton decoupling using the Waltz-16 composite pulse sequence with a 90° pulse of 80 μs . The data were collected with a spectral width from 75 to 135 ppm and 384 scans recorded for each spectrum.

C) Biological experiments

Reagents: DMEM Glutamax (Gibco, Life Technologies), Alexa-488 and Alexa-568 labelled-Phalloidin (ThermoFisher Scientific), DAPI (Sigma-Aldrich), FluoromountG (ThermoFisher Scientific), Bovine Serum Albumin (BSA) (Euromedex), SuperSignal™ West Pico PLUS Chemiluminescent Substrate (ThermoFisher Scientific), Mini-PROTEAN® TGX™ Precast Gels (BioRad), Cell Titer-Blue® reagent (Promega), Click-iT™ EdU Cell Proliferation Kit for Imaging, Alexa Fluor™ 488 dye (ThermoFisher Scientific), Recombinant Human IFN γ (Biolegend), DPBS (1x) (Gibco, Life Technologies), bis(sulfosuccinimidyl)suberate (BS3) (ThermoFisher Scientific), Pierce™ Streptavidin Magnetic Beads (ThermoFisher Scientific), Pierce™ Silver Stain for Mass Spectrometry (ThermoFisher Scientific), Recombinant Human EGF (Miltenyi Biotec), Recombinant Human IFN α 2a (Miltenyi Biotec), Recombinant Human IFN α 2b (ThermoFisher Scientific), Recombinant Human IFN β -1a (Miltenyi Biotec), Recombinant Human IFN λ 1 (BioLegend), Recombinant Human IFN λ 2 (BioLegend).

Cell culture: HeLa cells were grown at 37°C under 5% CO_2 in DMEM high glucose Glutamax (Gibco, Life Technologies) complemented with 10% FBS (v/v) (Gibco, Life Technologies) and supplemented with 5 mM pyruvate (v/v) (Gibco, Life Technologies) and 1% penicillin- streptomycin (v/v) (Gibco, Life Technologies).

In vivo studies: Animal work was conducted at Selvita Ltd according to 2010/63/EU and National legislation regulating the use of laboratory animals in scientific research and for other purposes (Official Gazette 47/11). An institutional Committee on Animal Research Ethics (CARE-Zg) oversaw that animal-related procedures were not compromising the animal welfare. The accreditation number of the project is: KLASA:UP/1-322-01/19-01/82, URBROJ: 525-10/1241-20-4.

Primary antibodies (Western blot): phospho-STAT1 Tyr701 (BD Transduction Laboratories, 612132, 1:1000), STAT1 (Cell Signaling, 9172, 1:1000), alpha-tubulin (Sigma, clone B512, T5168, 1:5000); IFN γ (Abcam, EPR1108, ab133566, 1:1000); phospho-JAK2 Tyr1007/1008 (Cell Signaling, 3776, 1:1000); JAK2 (Cell Signaling, 3230, 1:1000); phospho-STAT2 Tyr689 (Merk, 07-224, 1:1000), STAT2 (Abcam, ab32367, 1:1000); phospho-STAT3 Tyr705 (Cell Signaling, 9145, 1:1000); STAT3 (Cell Signaling, 9132, 1:1000); Primary antibodies (Immunofluorescence): beta actin (Abcam, 8226, 1:200); Secondary

antibodies: anti-mouse-Alexa-488 (Invitrogen, A21202), anti-mouse-HRP (Jackson ImmunoResearch, 715-035-151) and anti-rabbit-HRP (Jackson ImmunoResearch, 715-035-152) were used at 1:5000 for western blot and anti-mouse-Alexa-488 (Invitrogen, A21202) was used at 1:200 for immunofluorescence.

JAK-STAT signalling activation assay

20 mM stock solution of compounds in DMSO were prepared and stored at -20°C.

JAK-STAT stimulation on HeLa cells were done using previously described protocol.^[9] For phosphorylation status of STAT1 (pSTAT1) in different cytokines: 1000 U.ml⁻¹ of IFN γ , IFN α 2a, IFN α 2b, IFN β , IFN λ 1, IFN λ 2 or 15.6 nM EGF in DMEM containing 0.2% BSA with different concentrations (0.4, 4, 40, 80 μ M) of compound **1** (SMIFH2) and **5k**, and DMSO (as control) were incubated for 20 min at 37°C and added to HeLa cells for 20 min at 37°C.

In case of screening compounds, 40 μ M compounds were preincubated with 1000 U.ml⁻¹ of IFN γ in DMEM containing 0.2% BSA for 20 min at 37°C before adding it to HeLa cells. 10-fold 7 or 5 serial dilutions from 400 μ M of compounds were made in DMEM containing 1000 U.ml⁻¹ of IFN γ and 0.2% BSA, and incubated for 20 min before stimulating JAK-STAT for finding IC₅₀ towards IFN γ . Similarly, for cross-linked IFN γ , HeLa cells were treated with the preincubated mixture at 37°C for 20 min for JAK-STAT signalling.

For comparing pSTAT1 status in different conditions of **5k** treatment, JAK-STAT stimulation on HeLa cells were made by adding mixture containing 1000 U.ml⁻¹ IFN γ in DMEM with 0.2% BSA and 40 μ M compound preincubated for 20 min at 37°C; similar mixture without preincubation; 40 μ M compound in DMEM for 20 min at 37°C, then followed by addition of 1000 U.ml⁻¹ IFN γ to the media and stimulated the JAK-STAT signalling for 20 min further.

Immunoblotting

Cells were lysed in sample buffer (62.5 mM Tris/HCl, pH 6.0, 2% v/v SDS, 10% glycerol v/v, 40 mM dithiothreitol, and 0.03% w/v phenol red). Samples were analysed by SDS-PAGE on 4-15% Mini-PROTEAN® TGX™ Precast Gels or on 4-15% Mini-PROTEAN® TGX™ Stain Free Gel (Bio-Rad) and immuno-blotted with the indicated primary antibodies and horseradish peroxidase or Alexa-488 conjugated secondary antibodies. Chemiluminescence signal was revealed using Pierce ECL Western Blotting Substrate, SuperSignal West Dura Extended Duration Substrate or SuperSignal West Femto Substrate (Thermo Scientific Life Technologies). Acquisition and quantification were performed with the ChemiDoc MP Imaging System (Bio-Rad). Phosphorylated protein over total ratio was determined on the same blot.

***In cellulo* actin nucleation assay**

HeLa cells grown on coverslips were treated with DMEM containing 0.2% BSA and the compounds (40 μ M) for 20 min at 37°C, washed with cold PBS (two times) and then fixed with 4% paraformaldehyde for 30 min at room temperature, quenched in 50 mM NH₄Cl for 10 min and permeabilised with 0.05% saponin in 0.2% BSA in PBS for 20 min. Cells were incubated with 165 nM phalloidin for 1 h at room

temperature. DAPI containing fluoromount-G was used to mount coverslips onto glass slide. Cell areas were measured with ImageJ software (NIH).

For anti-actin immunofluorescence assay, after cell permeabilization, primary antibody (1 μ L in 200 μ L of 0.05% saponin in PBS) was added for 1 h. It was washed with PBS for 5 min for 3 times and secondary antibody (1 μ L in 200 μ L of 0.05% saponin in PBS) was added for 1 h. Washed with PBS for 5 min for 3 times and DAPI containing fluoromount-G was used to mount coverslips onto glass slide for further analysis.

Cell viability assay

Cell viability assay was carried out by plating 10,000 cells/well in 96-well plates. 3-Fold 8 serial dilutions of the compounds from 3 mM were made in DMEM. HeLa cells were treated for 24 h with the compounds of different concentrations made. In case of preincubation with IFN γ , 6000 U.mL⁻¹ of IFN γ per concentration of compounds were used and incubated for 20 min. According to manufacturer's protocol, Cell Titer-Blue® reagent was added after 24 h treatment and cells were incubated for 3 h before recording fluorescence intensities (λ_{ex} . 560/20 nm; λ_{em} . 590/10 nm) using a Perkin Elmer Wallac 1420 Victor2 Microplate Reader.

Acetylation of IFN γ

Acetylation of Lysine and N-terminal amino groups were done according to previously described protocol.^[10] Briefly, 1 μ g of IFN γ was dissolved in 20 μ L of NH₄HCO₃ (50 mM). Acetic anhydride (5 or 10 or 25 eq per lysine residues (14 Lysine)) were added along with 20 eq of **5k** and incubated at 37°C for 30 min before adding the click reagents for further immunoblot analysis. Quantification was done using ImageJ software (NIH).

Click reaction for Native Gel

The protocol was adapted from previously described protocol on protein electrophoresis and in-gel fluorescence scanning.^[11] 6 μ L of IFN γ from stock solution (71 μ M final) were added on 120 μ L of NH₄HCO₃ (50 mM) and divided into 6 parts (20 μ L each). Parts 2 and 7 (control) were treated with 20 eq of **5k** (0.6 μ L of 2 mM stock in DMSO), Part 3 and 7 with 20 eq of **7b** (0.6 μ L of 2 mM stock in DMSO), Part 4 was co-treated with 20 eq of **5k** and **7b** (2 mM stock in DMSO), acetic anhydride (10 eq per lysine residues, 0.8 μ L) was added to part 5 and 6, and incubated at 37°C for 30 min. Then, all parts were added with 30 μ L of click reaction cocktail (prepared using the manufacturer's protocol of Click-iT™ EdU Cell Proliferation Kit for Imaging, Alexa Fluor™ 488 dye; 430 μ L of 1x Click-iT reaction buffer with 20 μ L of CuSO₄ solution, 1.2 μ L of Alexa-488-azide, 50 μ L of reaction buffer additive (sodium ascorbate)) and incubated for 45 min. Corresponding amount of Sample Solubilizing Buffer (1%(w/v) SDS, 100 mM Tris-HCl (pH 9.5)) to reach up to 100 μ L were added and 50 μ L were loaded on native gel (Mini-PROTEAN® TGX™ Precast Gel) using running buffer 1x (250 mM Tris, 1.92 M glycine, pH 9.2) to perform western blot analysis. Proteins were transferred on nitrocellulose membrane and scanned for Alexa-488 signal using ChemiDoc MP Imaging System (Bio-Rad). Then, further incubation with primary, then secondary antibodies was done for IFN γ immunoblotting.

Cross-linking of IFN γ

The cross-linking reactions were performed as previously described in literature.^[12] Briefly, 1 μ g of IFN γ in 10 μ L of PBS was mixed with 2 μ L of 10 mM solution of BS3 and the mixture was kept on ice for 1 h. The reaction was stopped by addition of 10 μ L of 1 M Tris-HCl, pH 6.8. The reaction product was analysed by SDS-PAGE and western blot analysis. To test the activity of the cross-linked IFN γ , 1.1 μ L of the reaction mixture was added to 1 mL DMEM (1000 U.ml⁻¹ final) containing 0.2% BSA, incubated at 37°C with or without **5k** (40 μ M final) and activation of JAK-STAT signalling was determined in HeLa cells stimulated for 20 min by lysate immunoblotting as mentioned above.

Pharmacokinetic profiling in the C57BL/6N mice

Each group of mice (C57BL/6 male, n = 9, Charles River Germany) were given with **5k** via intravenous (IV) at dose of 2 mg/kg and intraperitoneal (IP) administration at dose of 10mg/kg in the formulation of DMSO/cremaphor/PBS (10/10/80; v/v/v). Blood samples were analysed by LC-MS/MS (Shimadzu LC30AD/Shimadzu SIL30ACMP/Shimadzu CTO30A) at the interval of 0.05, 0.25, 0.5, 1, 2, 4, 8, 24 h.

Acute model of DSS-induced colitis in C57BL/6 mice

Mice (C57BL/6 male, n = 50, Charles River Germany) were grouped into 5. Groups 2-5 received 3% Dextran Sulfate Sodium (DSS) solution in drinking water for 7 full days and group 1 received water only. Animals in group 3 were administered with Cyclosporine A (pharmacological control) at 80 mg/kg per os (PO), once per day (QD). Animals in group 4 were administered intravenously (IV)/QD with 10 mg/kg of **5k**, while animals in group 5 received 10 mg/kg of **5k** given via intraperitoneal (IP) route once per day (QD). Animals in groups 1 and 2 were administered IP/QD with 5 mL/kg of DMSO/cremophor/PBS (10/10/80 v/v/v) vehicle solution. The efficacy of the therapeutic treatments was assessed daily through clinical monitoring consisting of body weight changes, stool consistency, colorectal bleeding condition. Colorectal bleeding score was defined with 0 for no presence of blood in stool or anal region, 2 for visible blood in stool pellets and 4 for gross bleeding and/or blood around anus region. Disease activity index (DAI) was calculated as combined score of a) weight loss b) stool consistency and c) colorectal bleeding daily with maximum score of 12. Mice were sacrificed by ketamine/xylazine overdose at day 8.

Statistical analysis

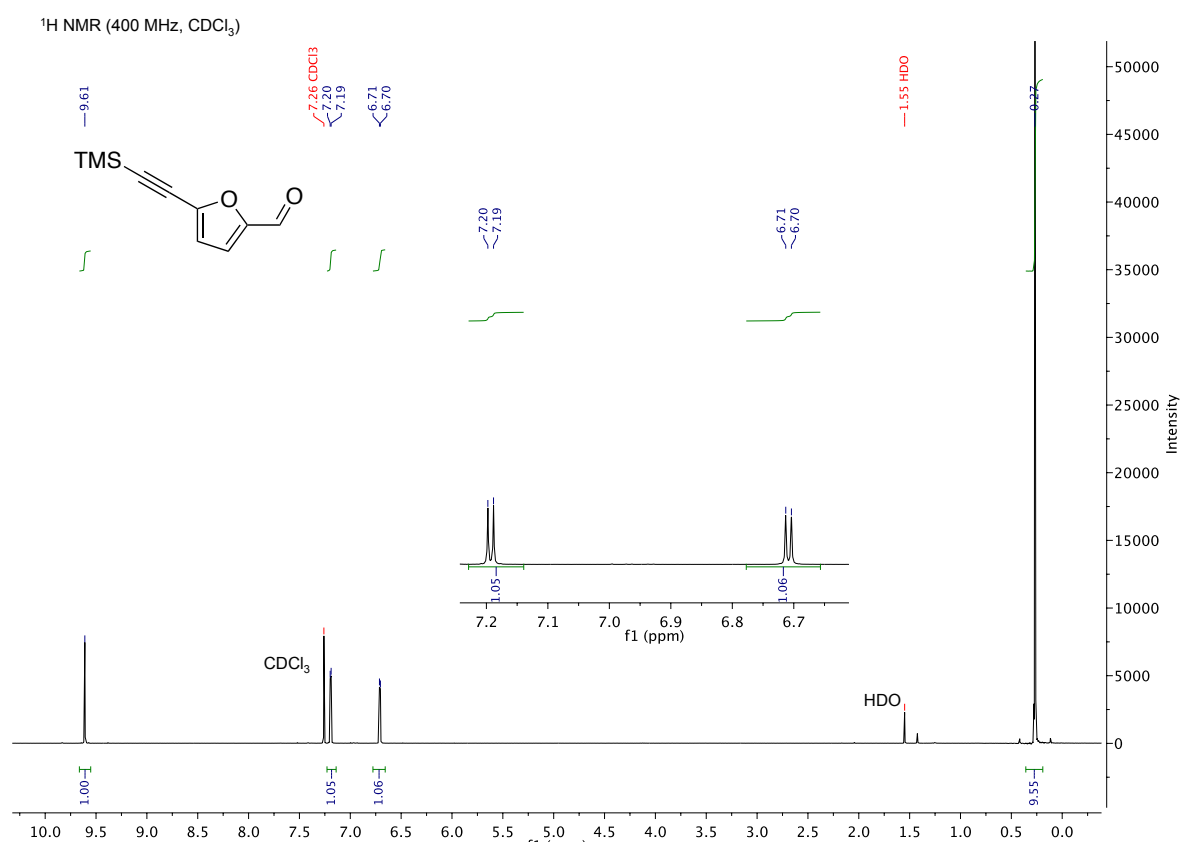
All analyses were performed using GraphPad Prism version 7.0 and 9.1.1, (GraphPad Software, La Jolla, CA, USA). One-way ANOVA, Two-way ANOVA with Dunnett's multiple comparison test and Kruskal-Wallis test with Dunn's post-test were performed. Significance of mean comparison is marked on the graphs by asterisks. Error bars denote SEM or SD.

References:

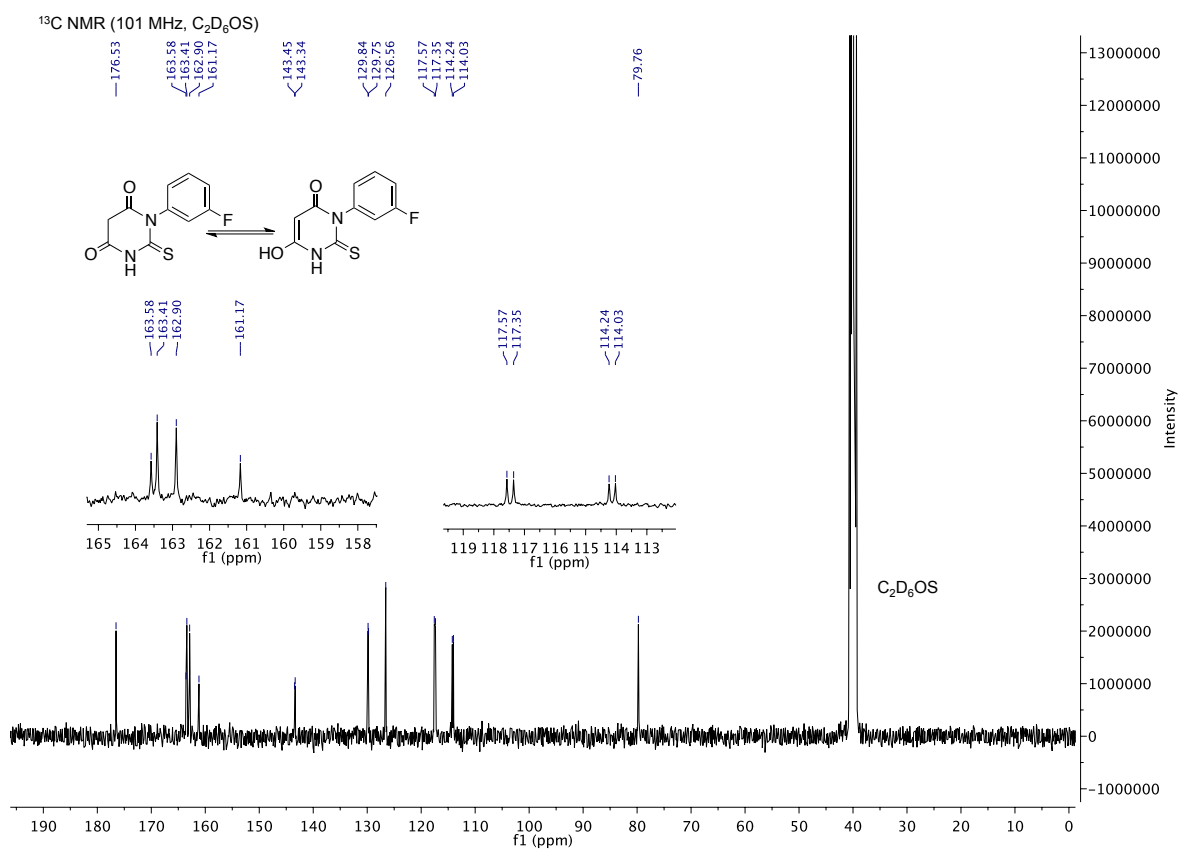
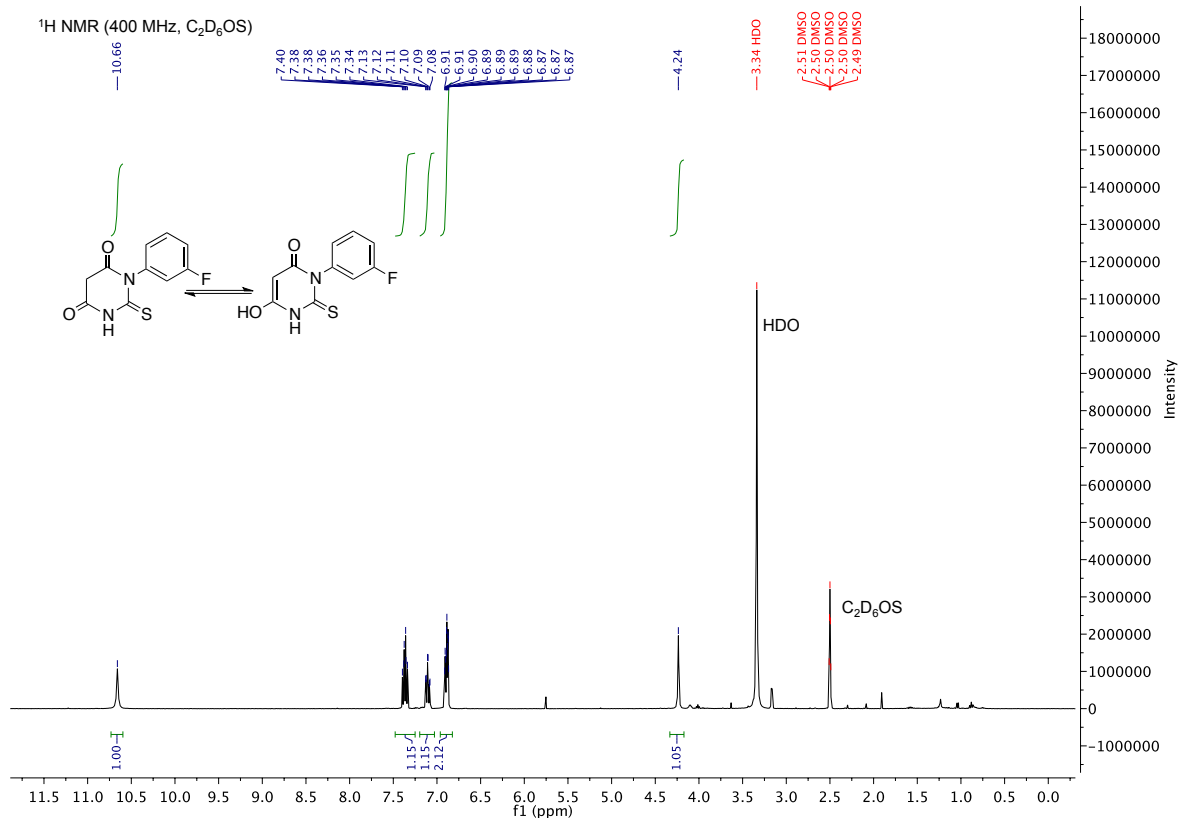
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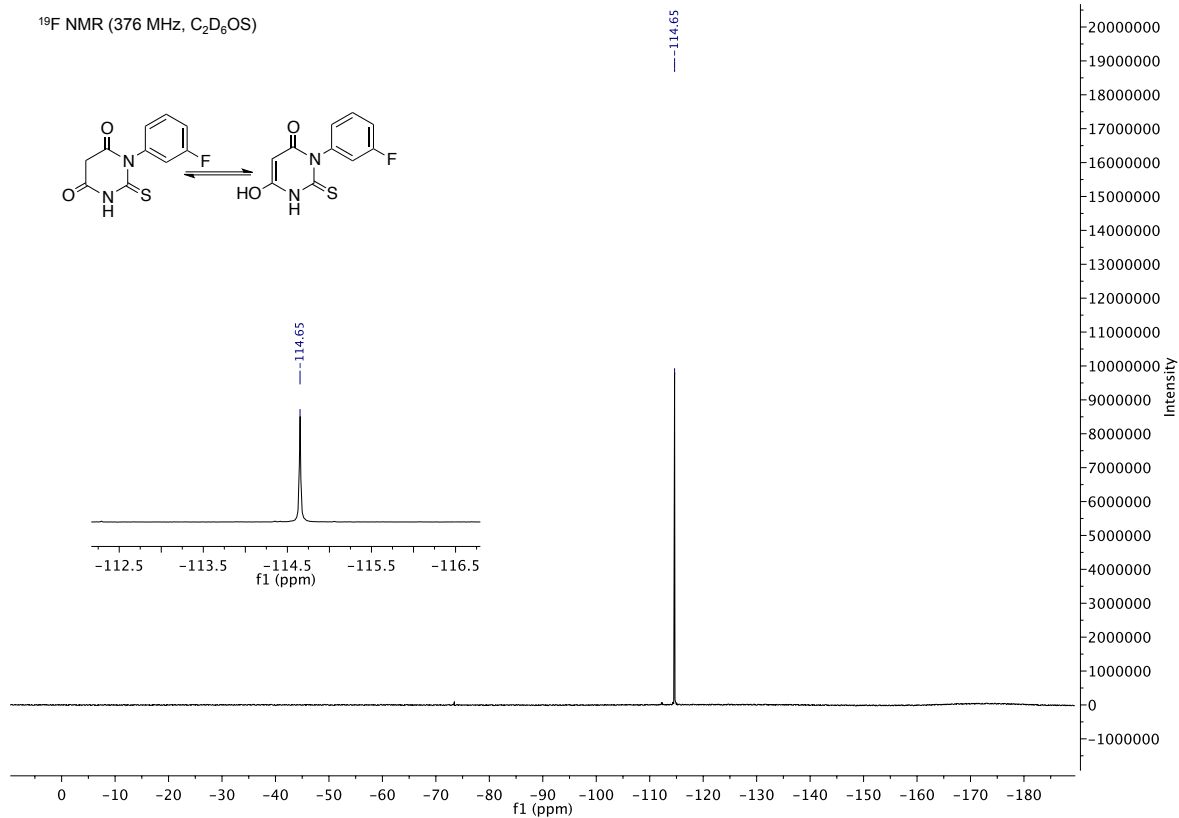
NMR spectra of all compounds (^1H , ^{13}C , ^{19}F spectra)

Compound 1a

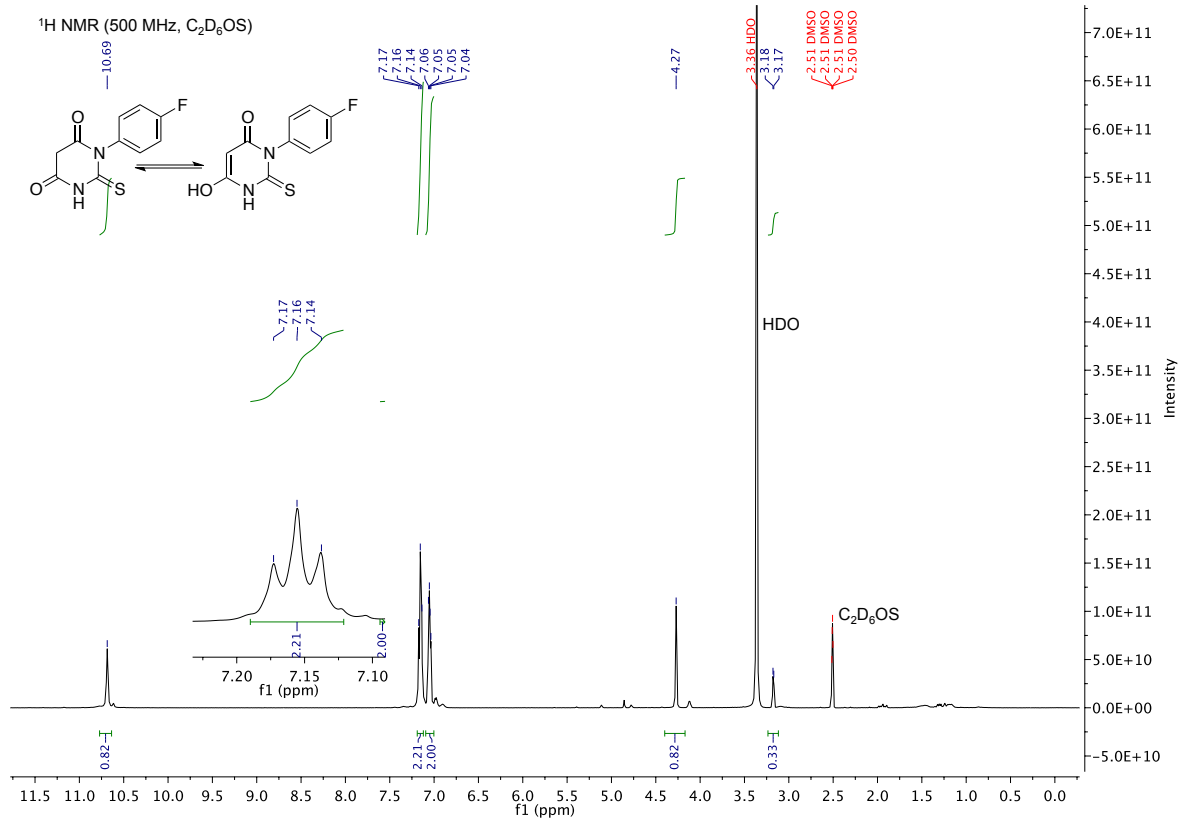


Compound 2a

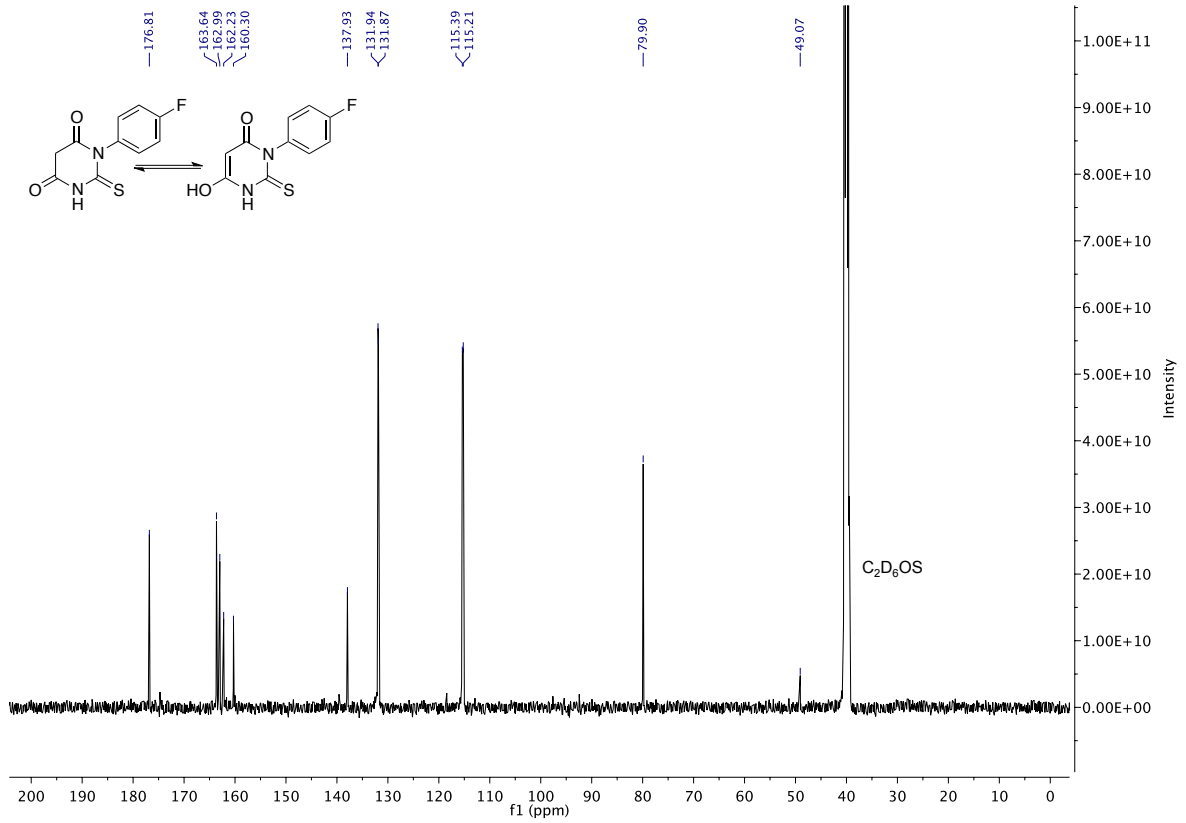




Compound 2b

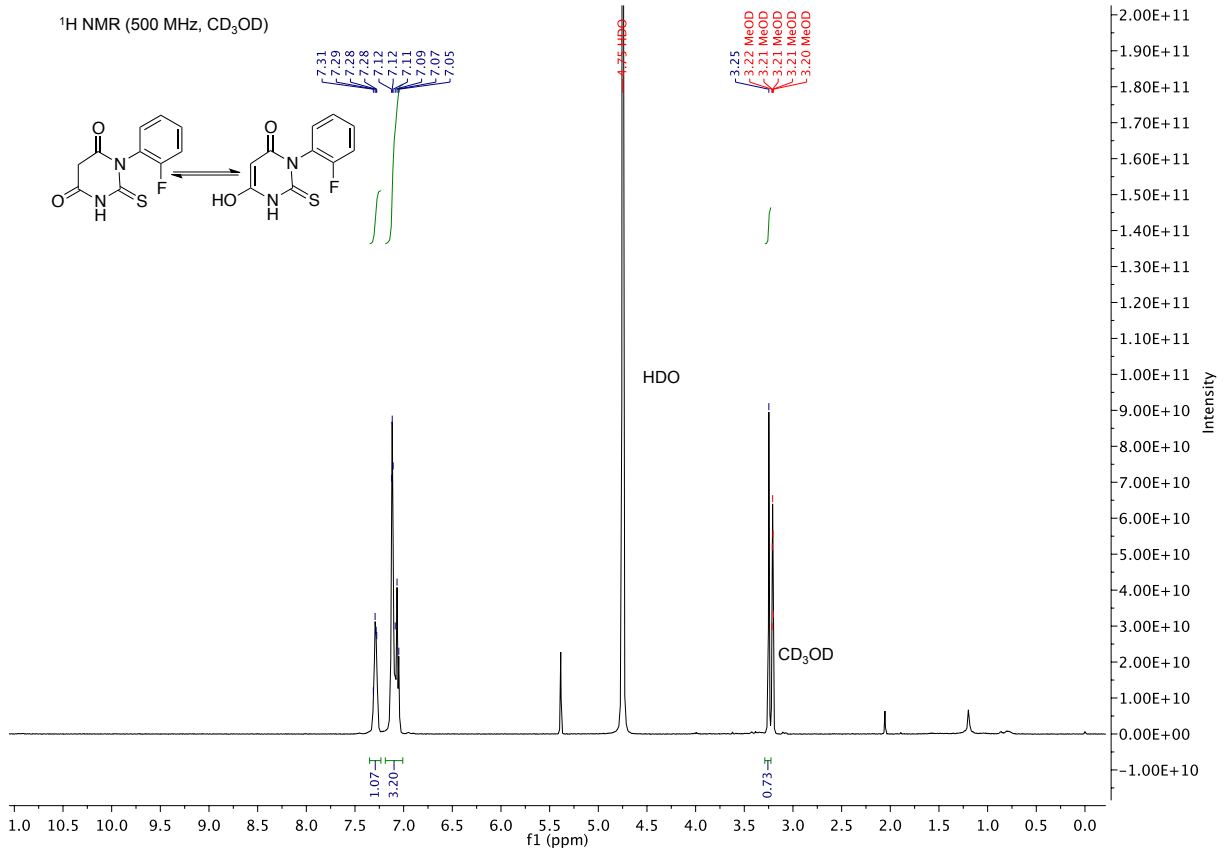


¹³C NMR (126 MHz, C₂D₆OS)

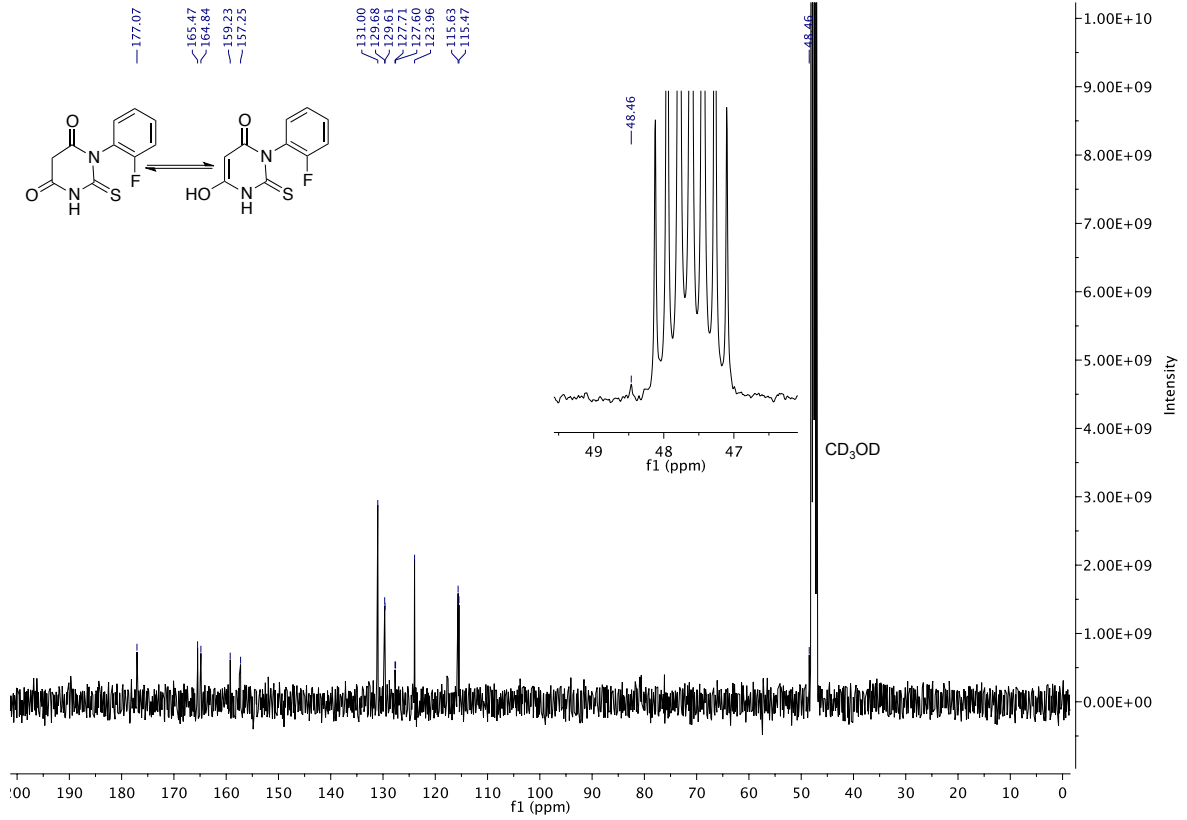


Compound 2c

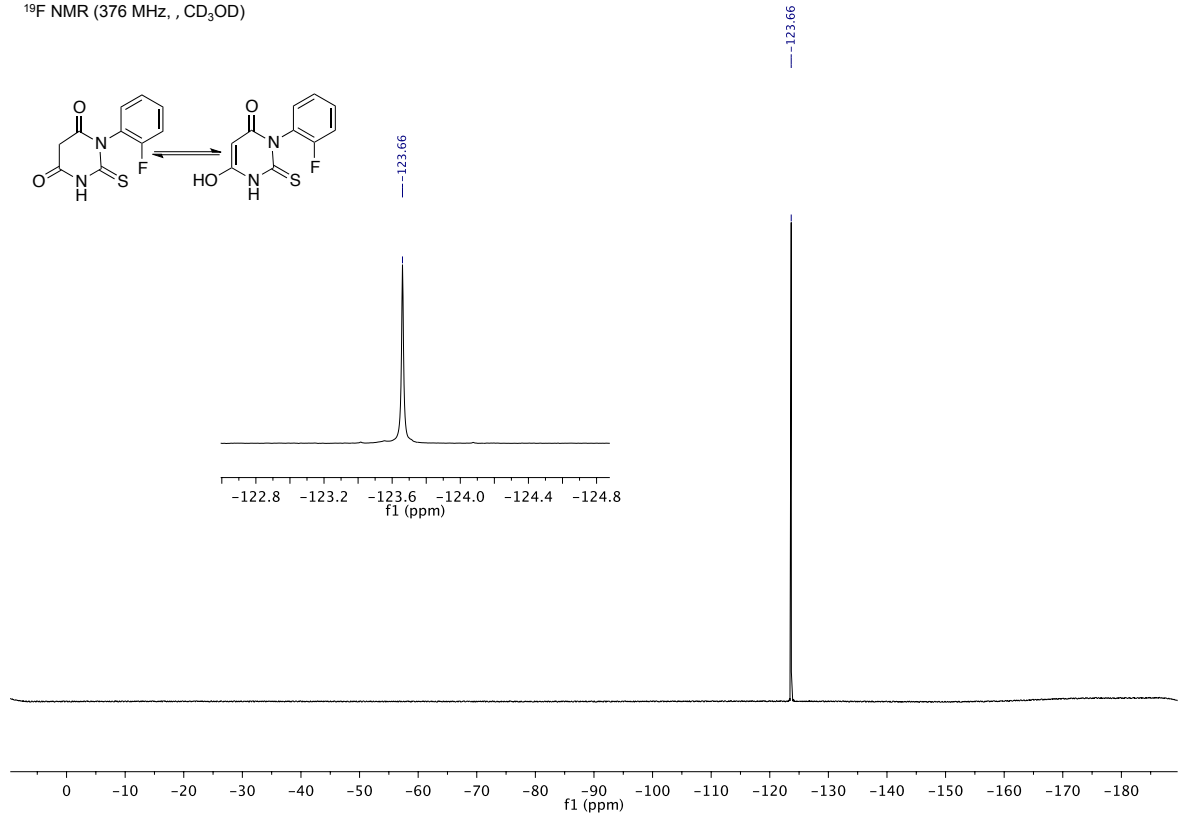
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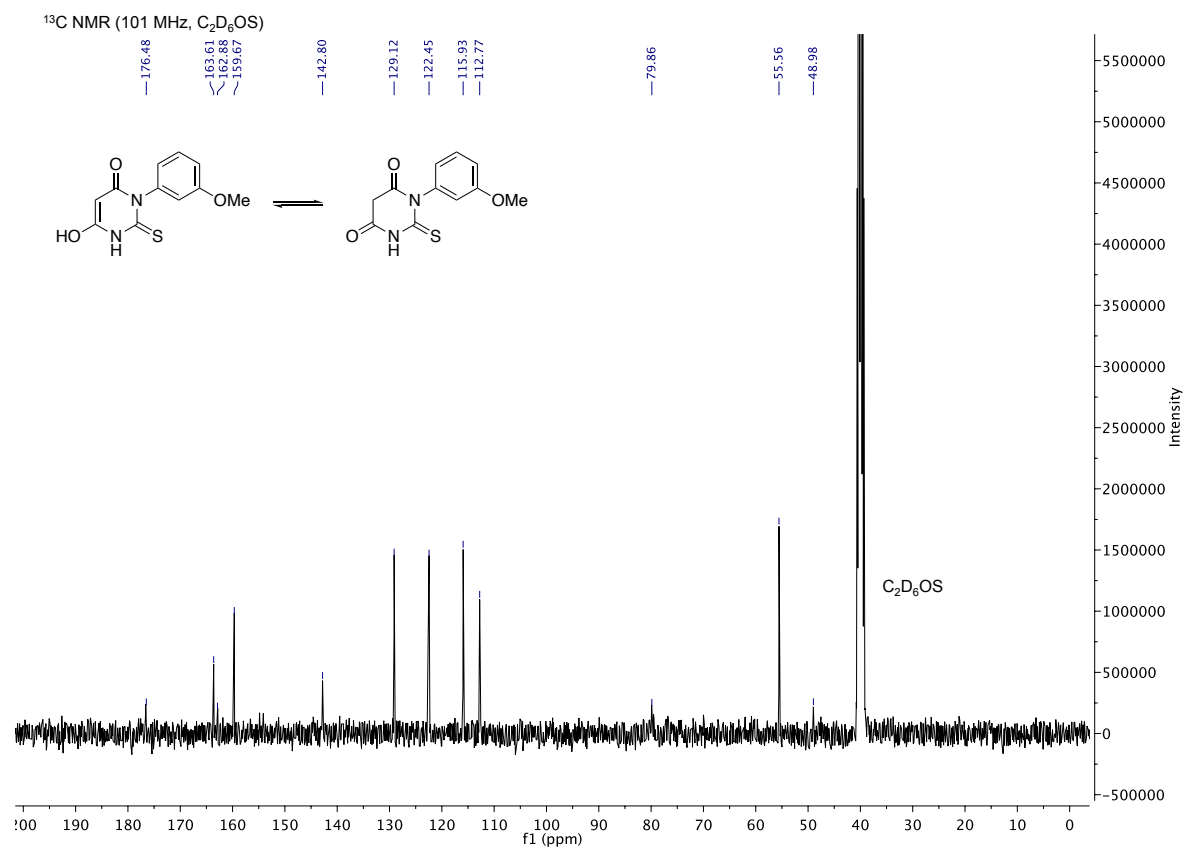
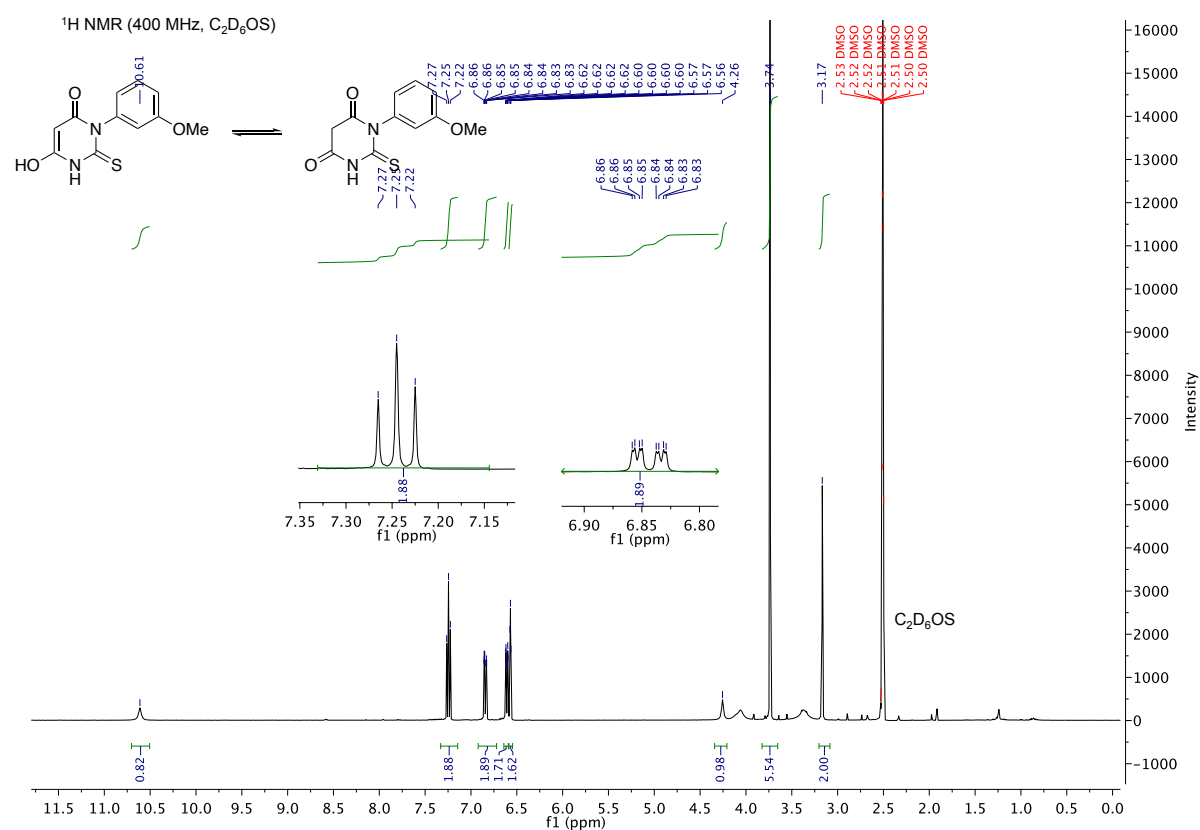
¹³C NMR (126 MHz, CD₃OD)



¹⁹F NMR (376 MHz, CD₃OD)

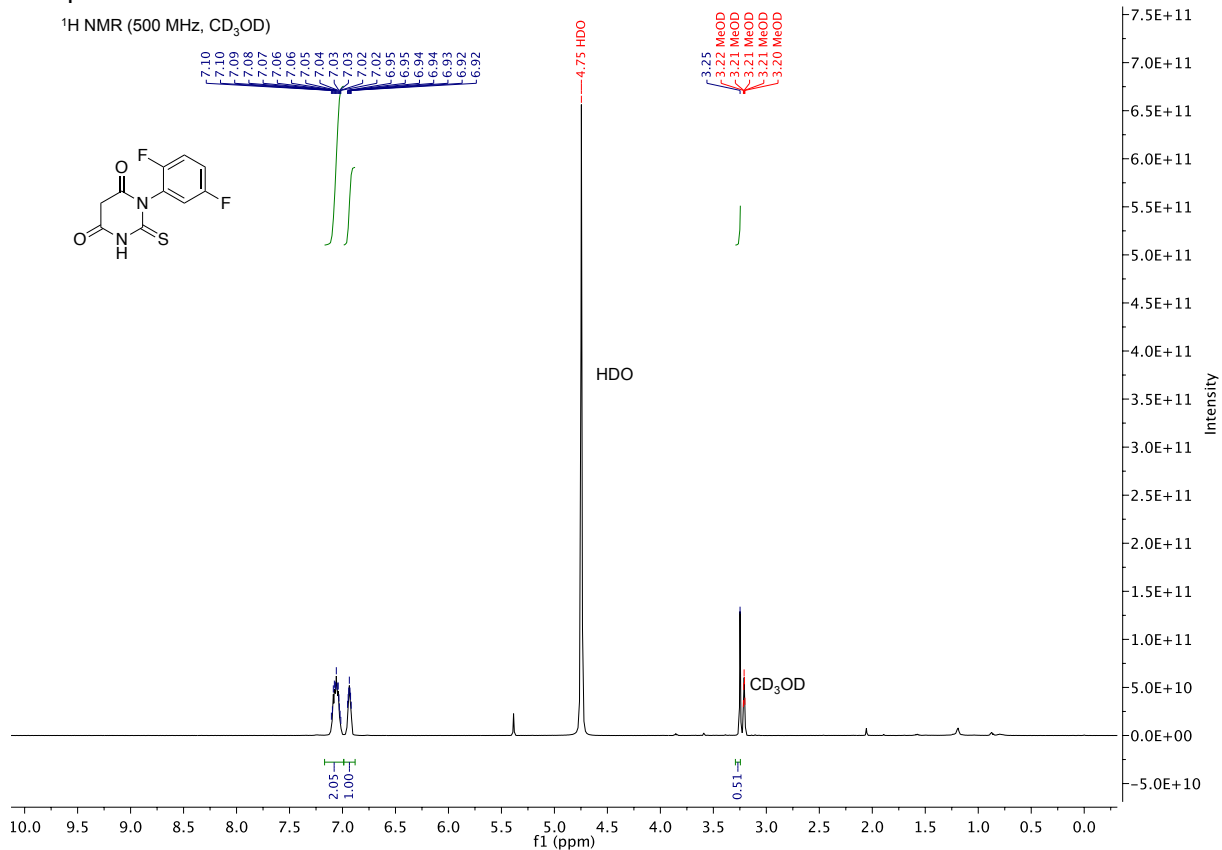


Compound 2d

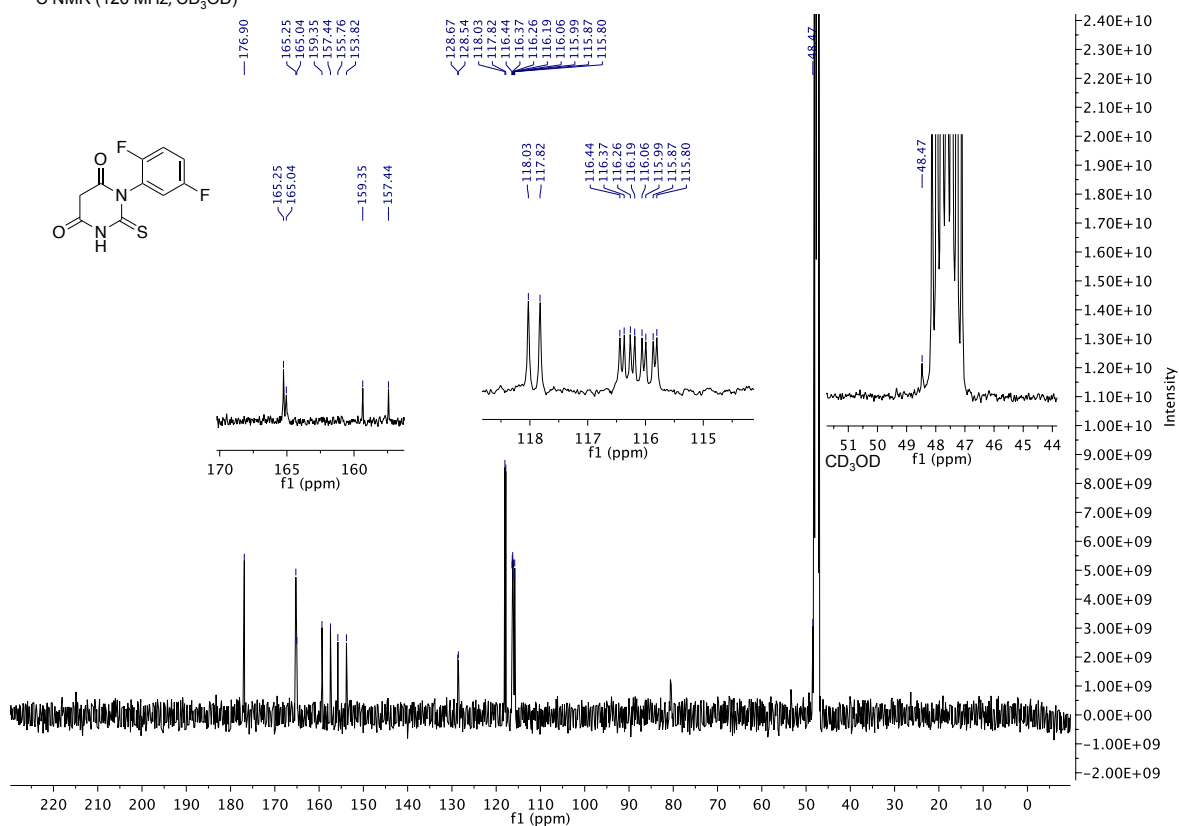


Compound 2f

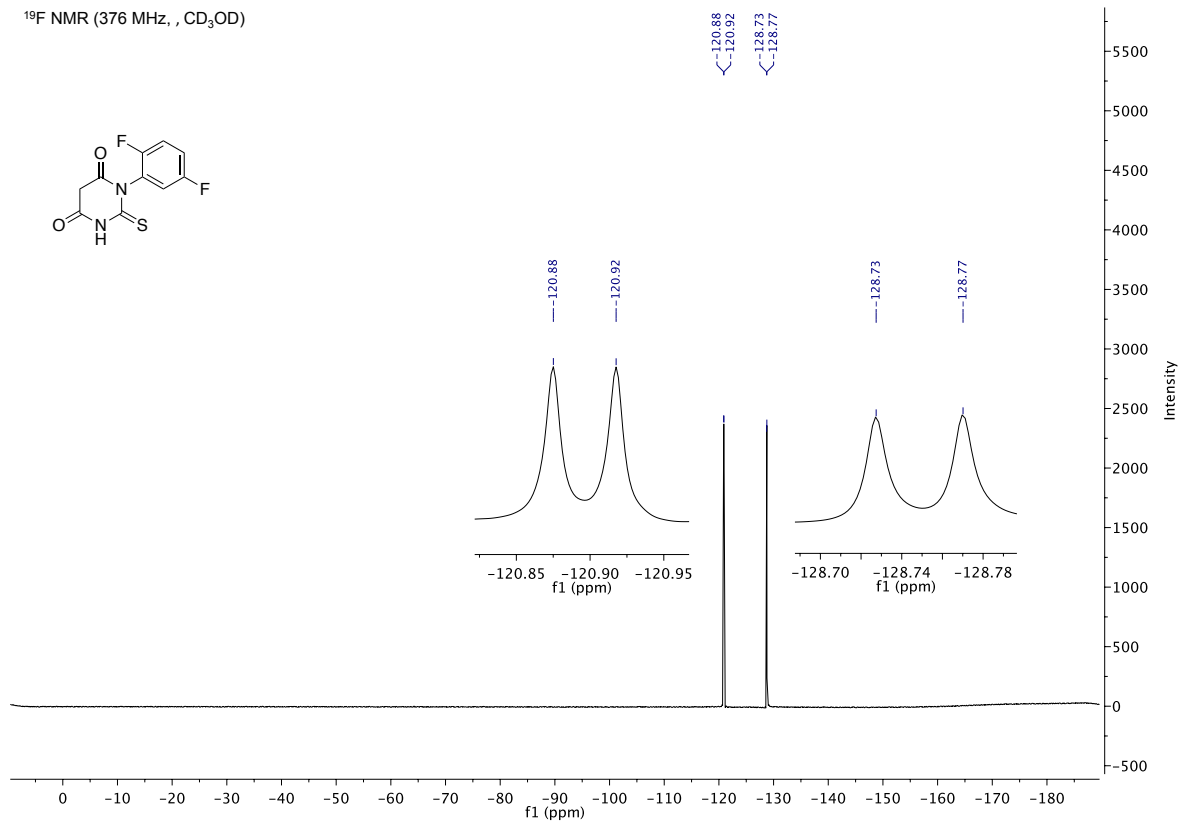
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¹³C NMR (126 MHz, CD₃OD)

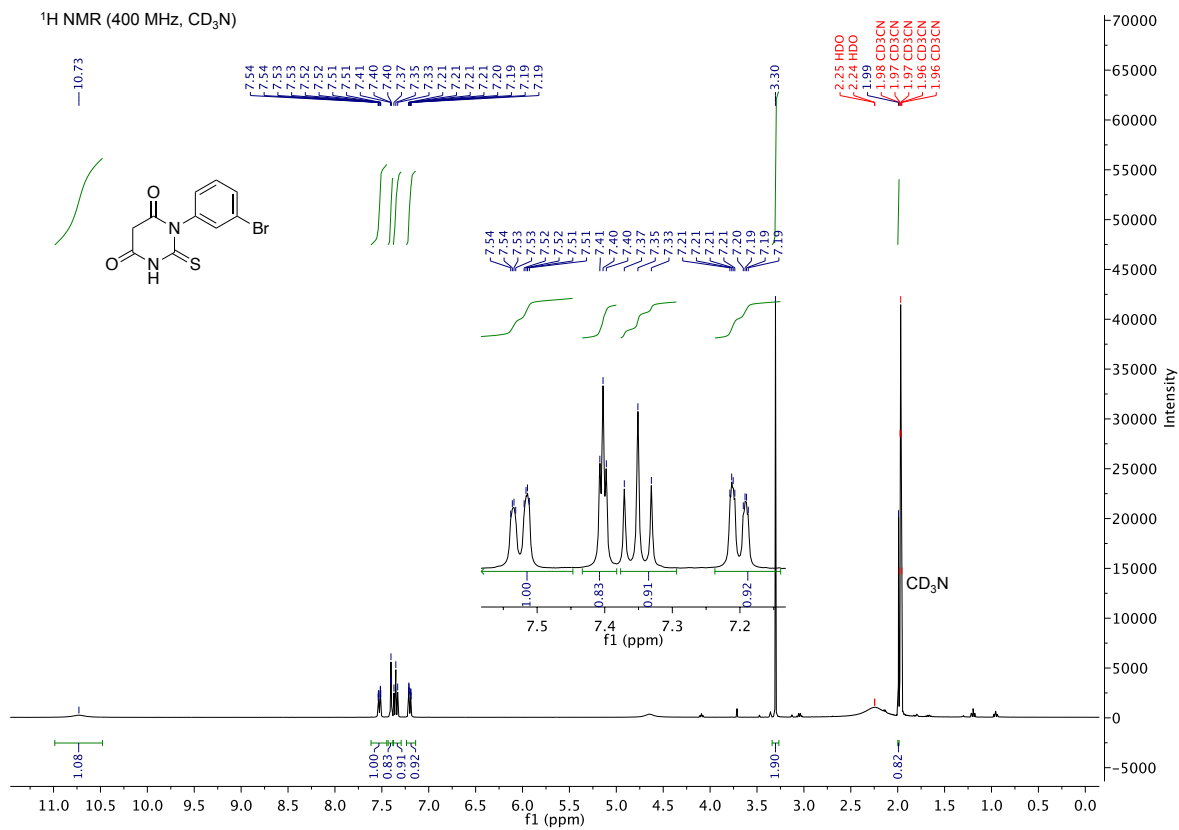


¹⁹F NMR (376 MHz, CD₃OD)

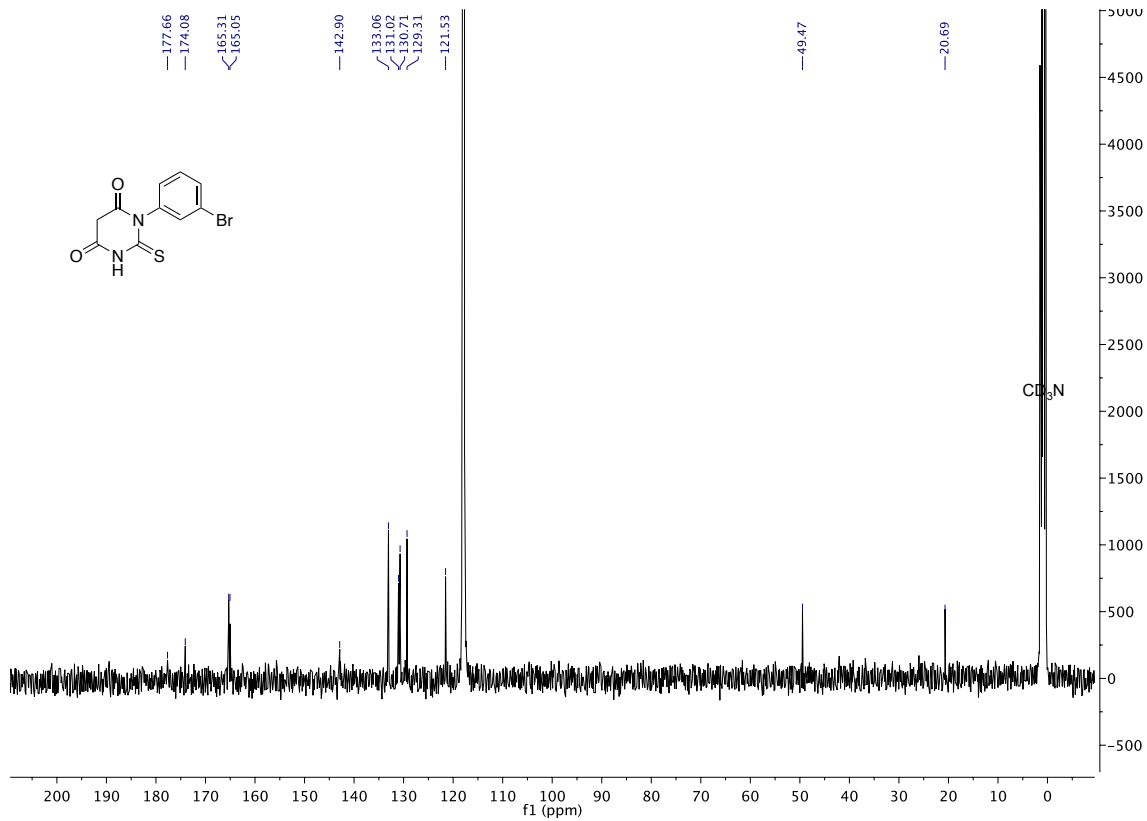


Compound 2g

¹H NMR (400 MHz, CD₃N)

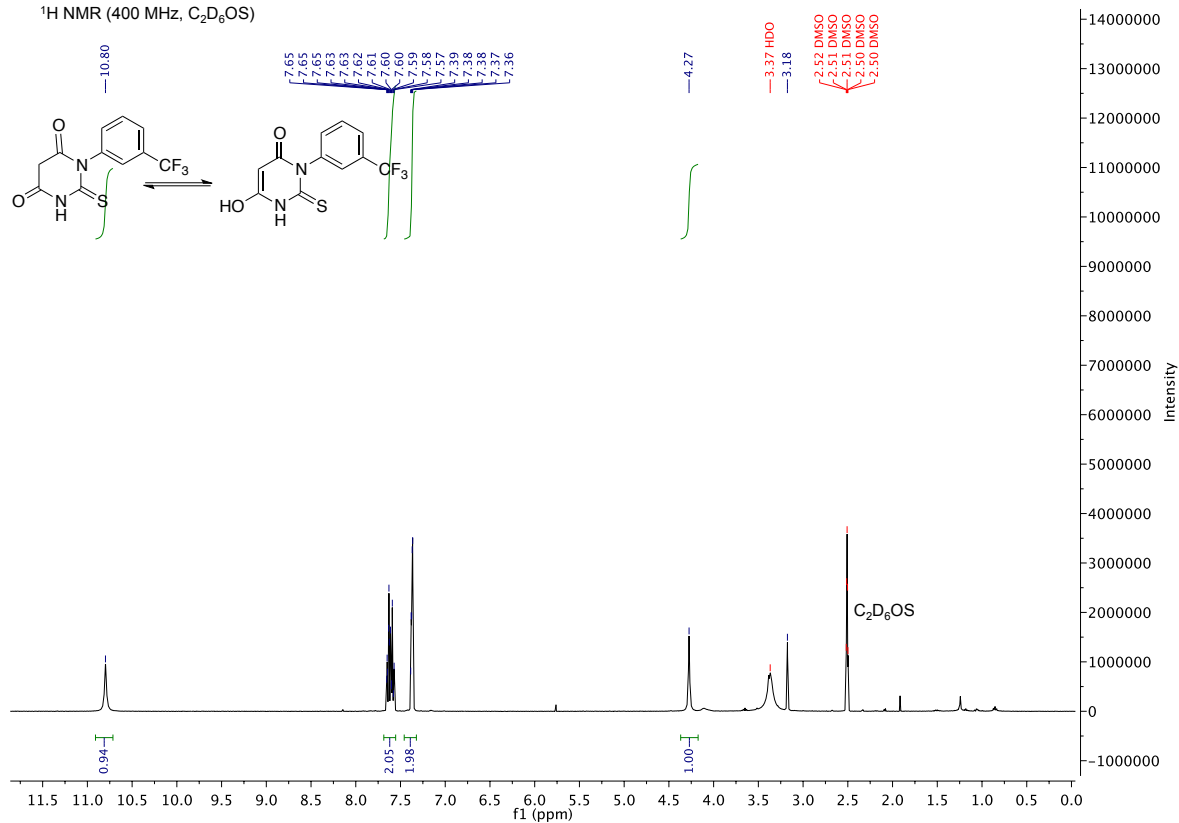


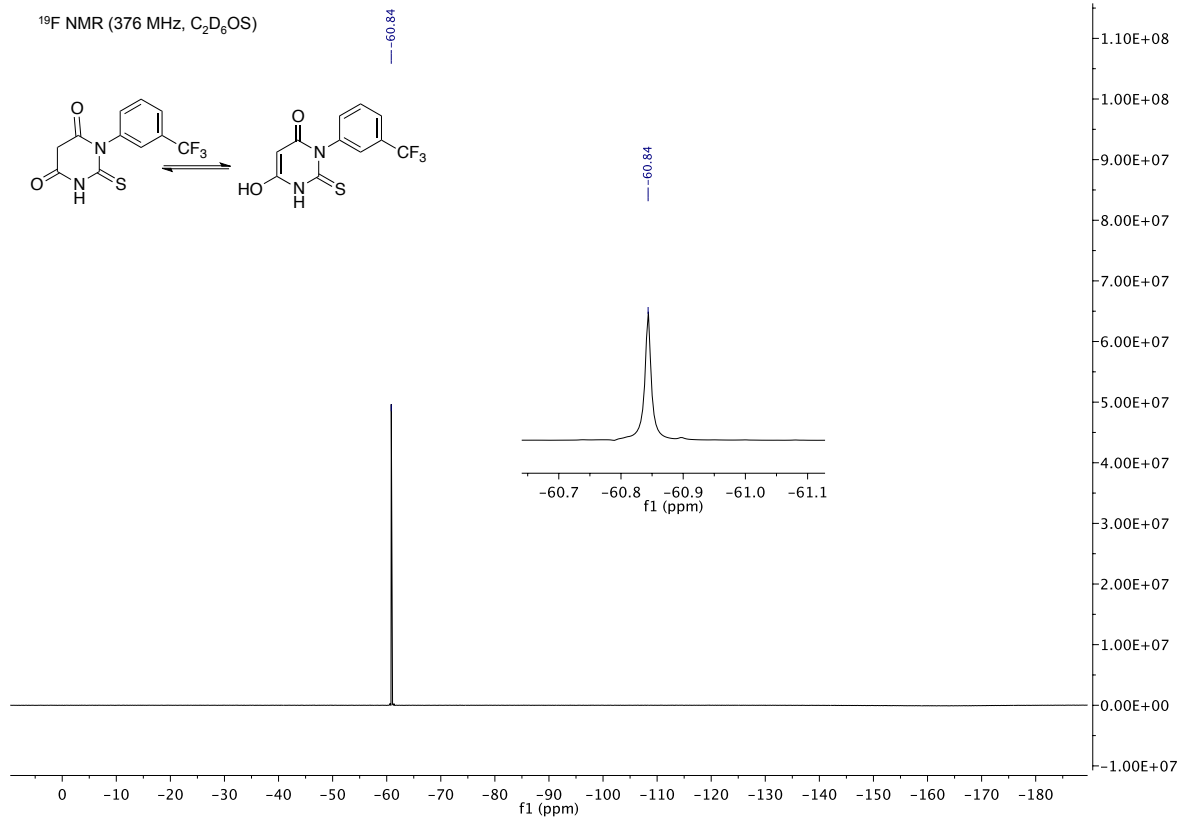
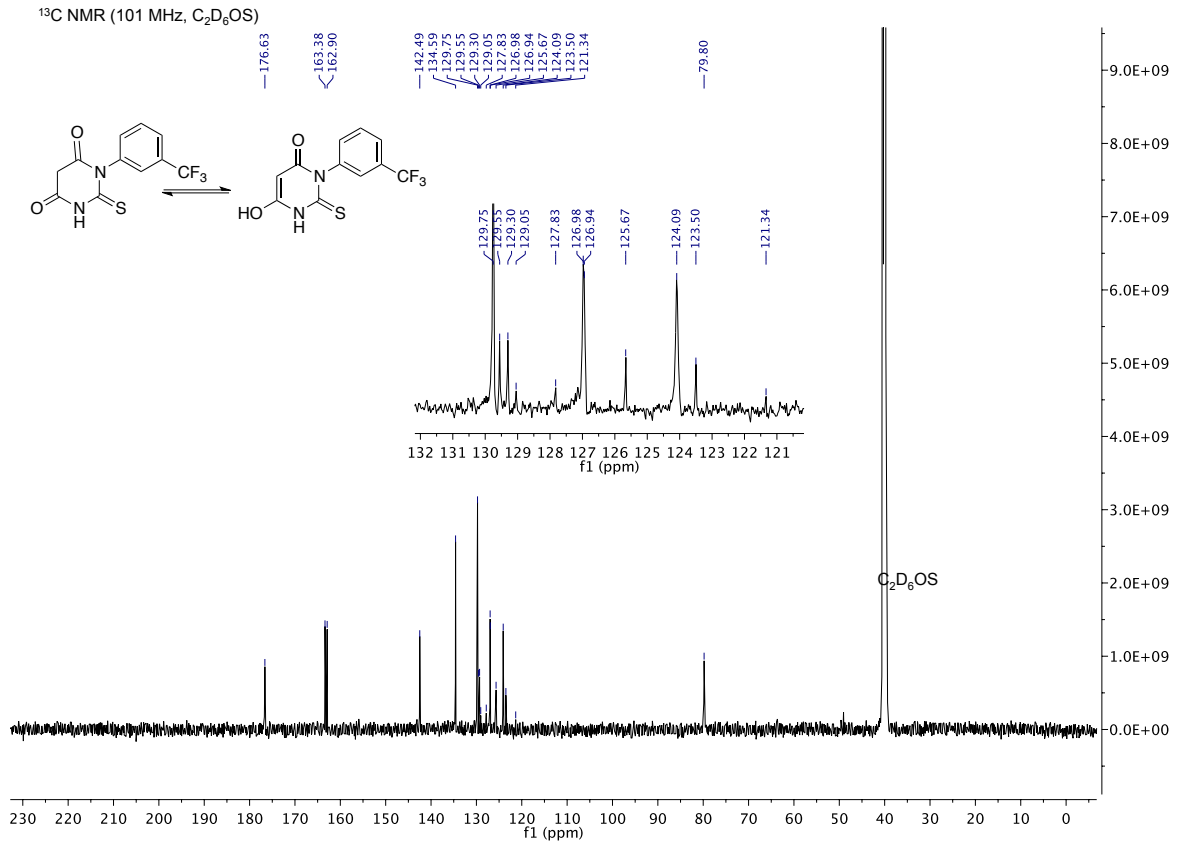
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Compound 2h

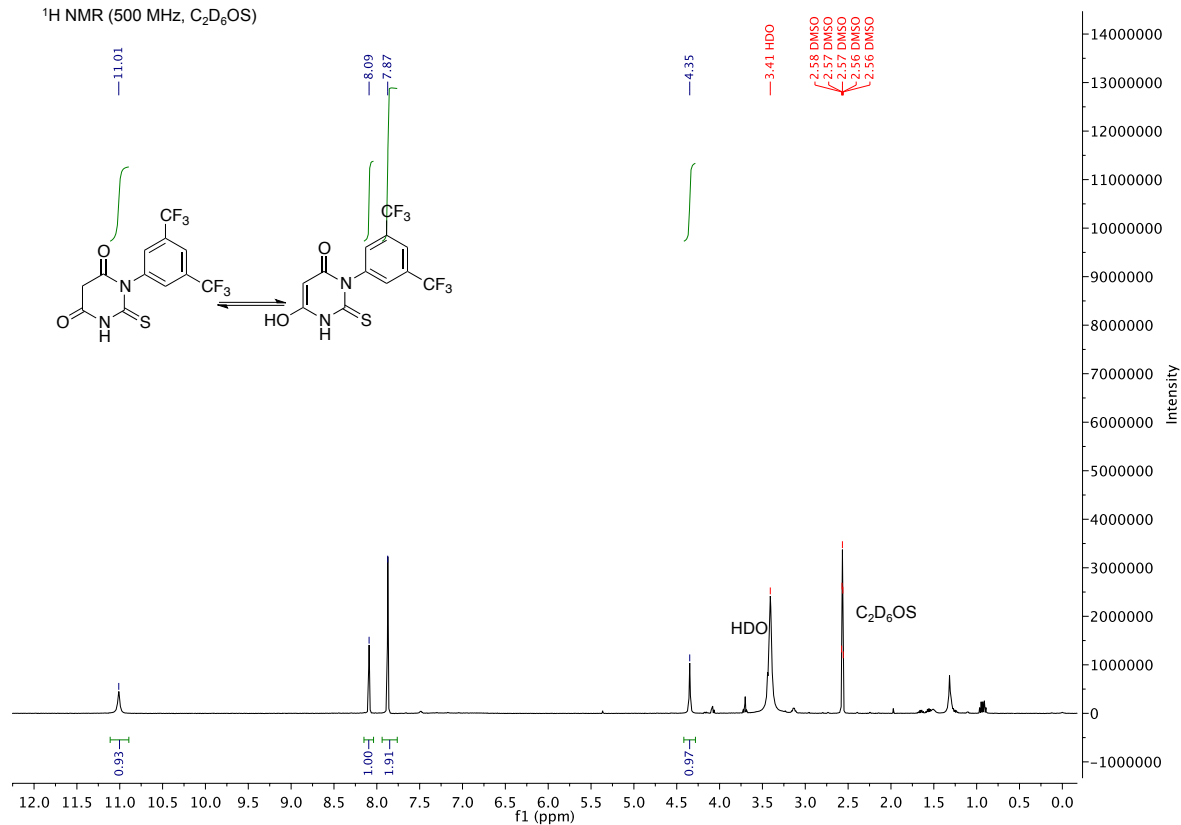
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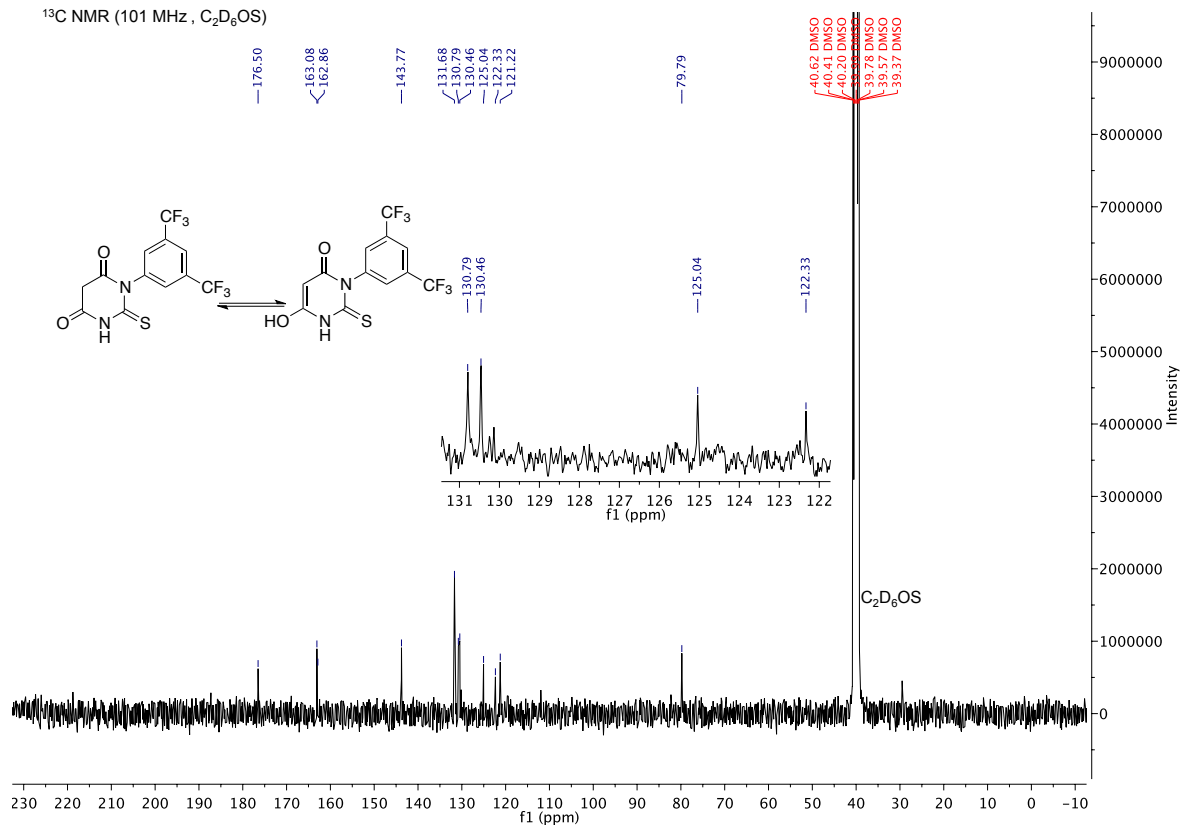


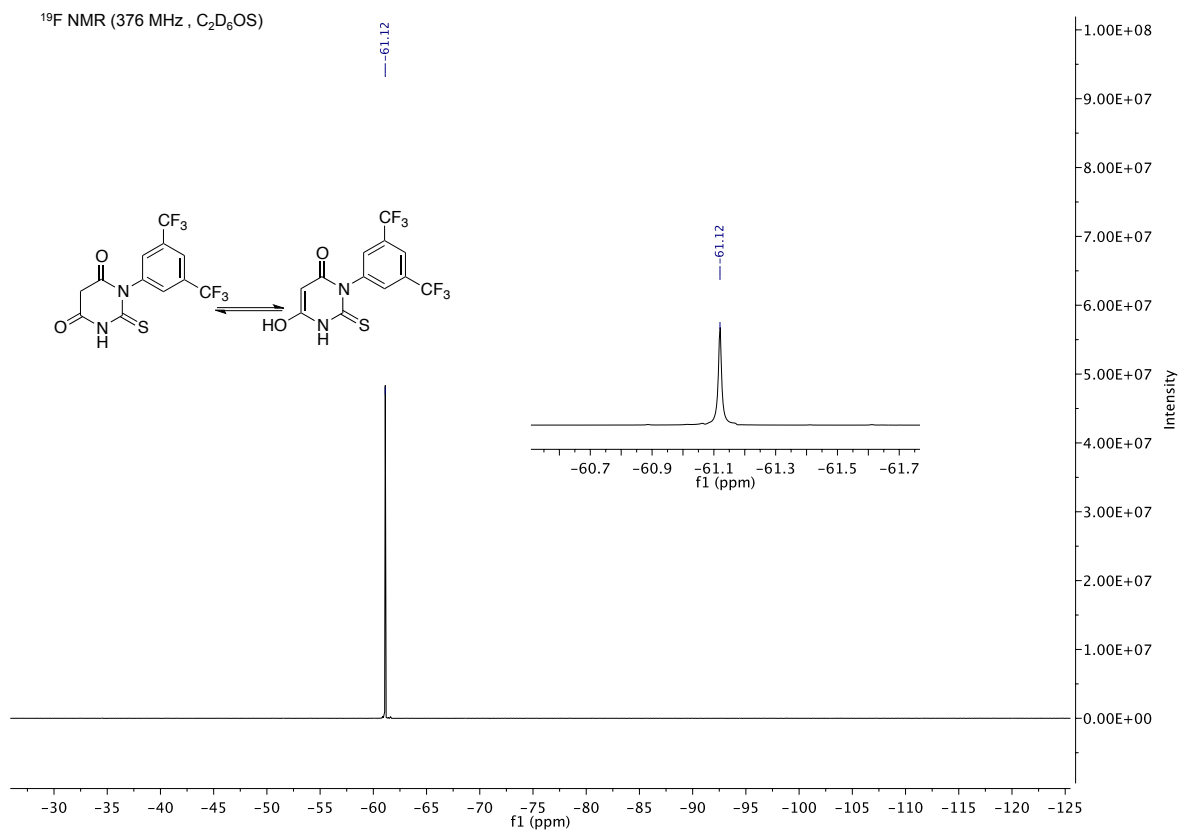
Compound 2i

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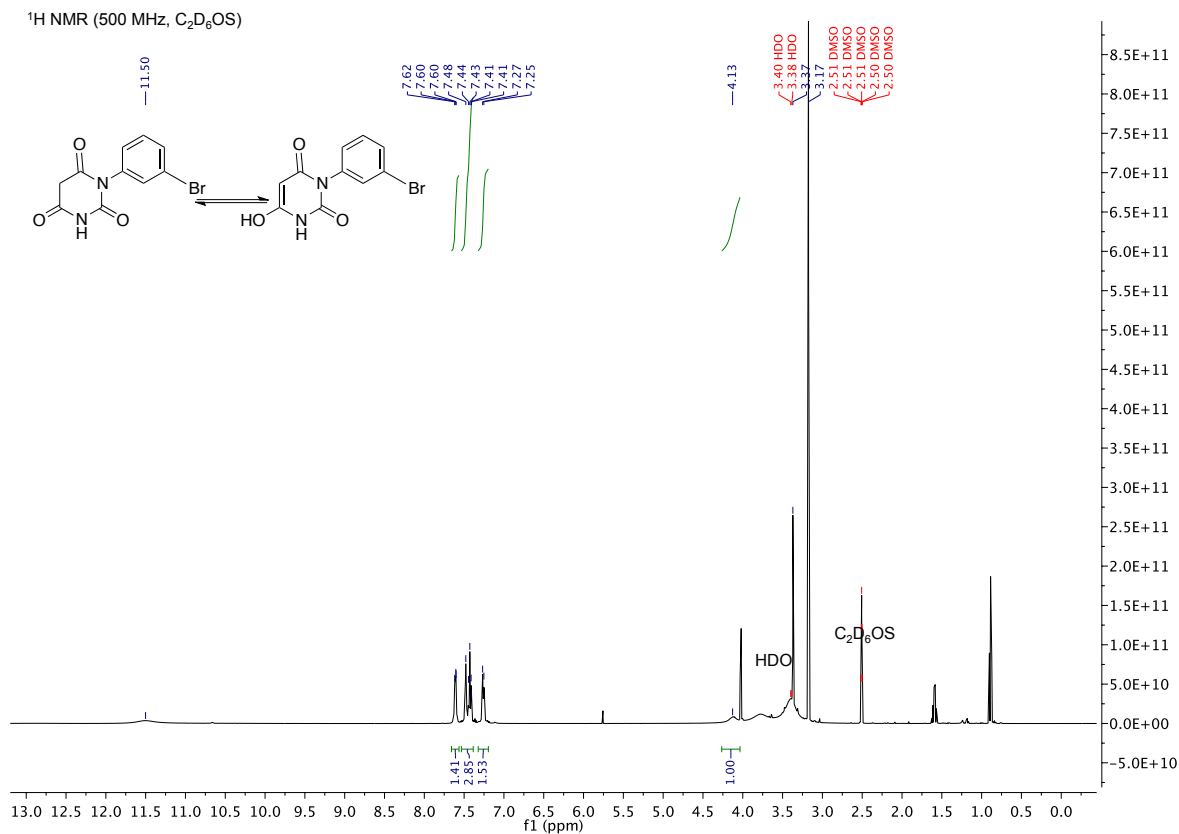


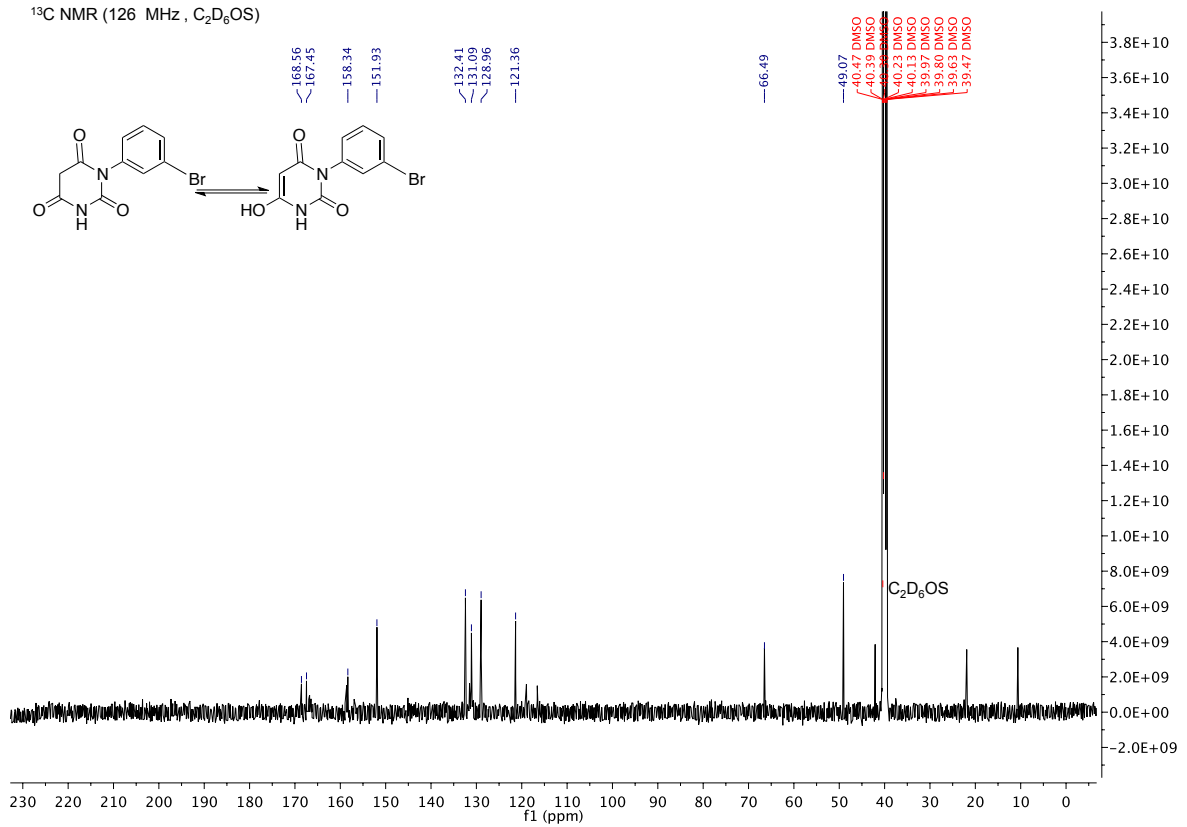
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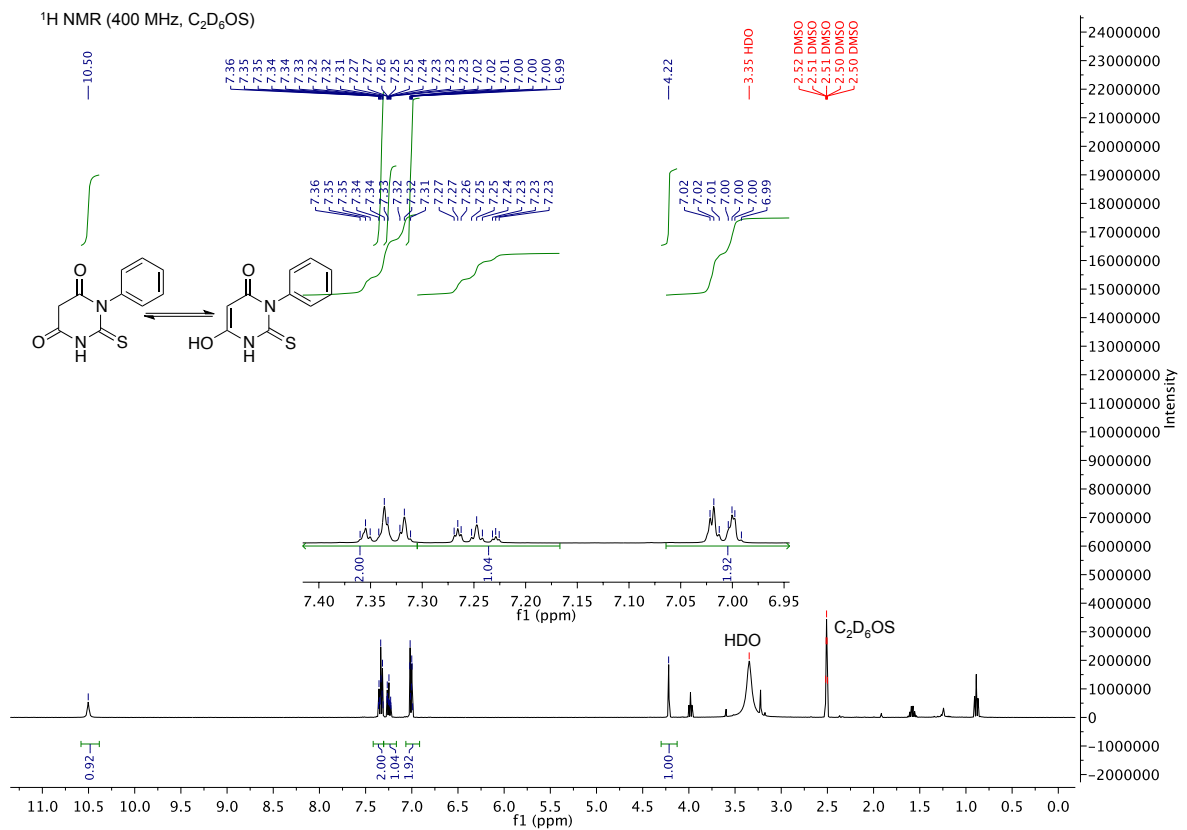


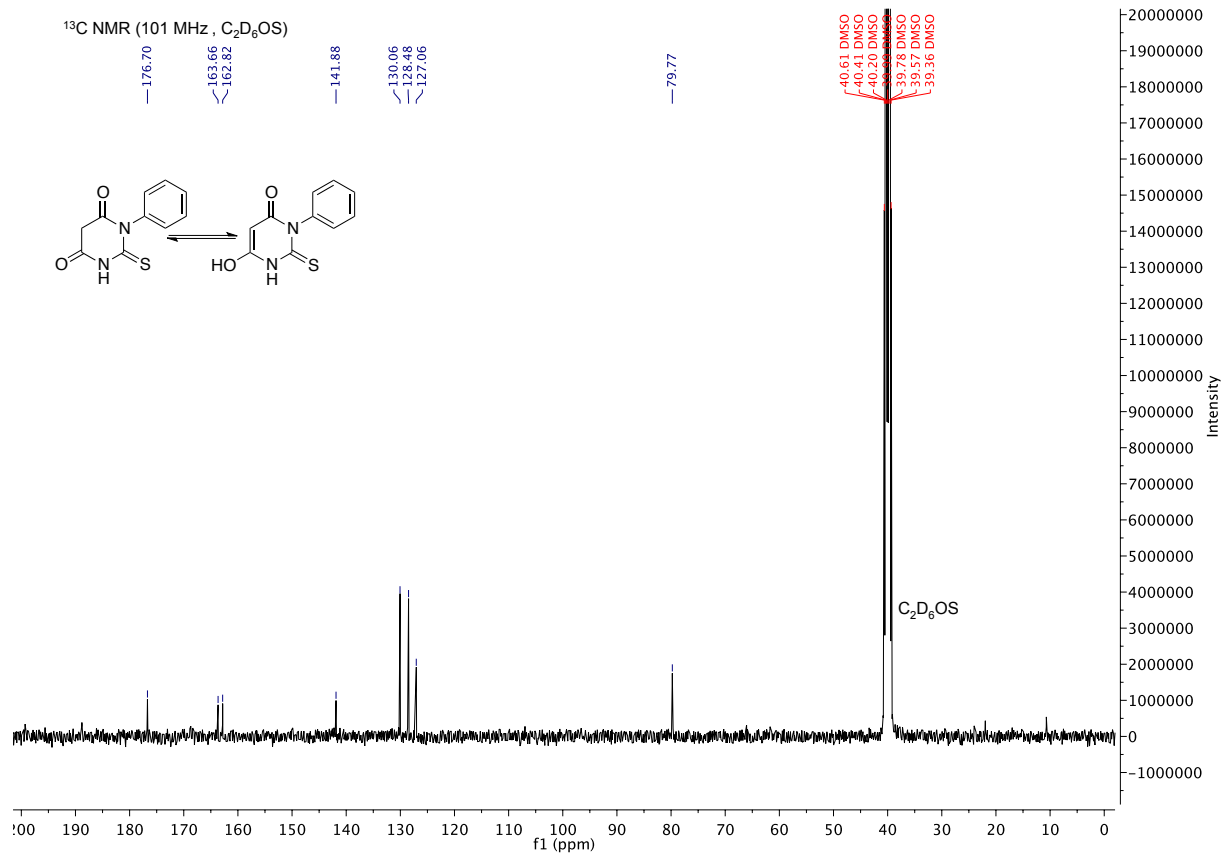
Compound 2j (Not pure)



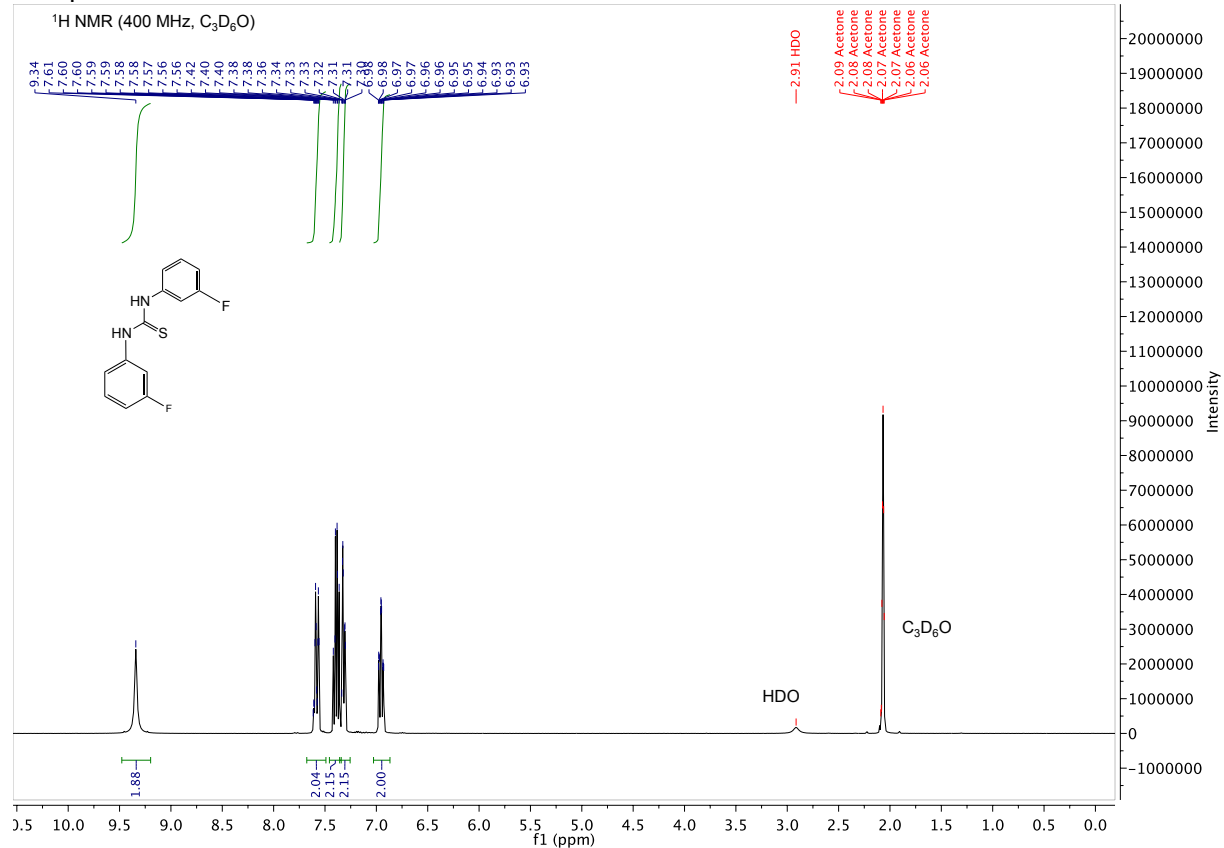


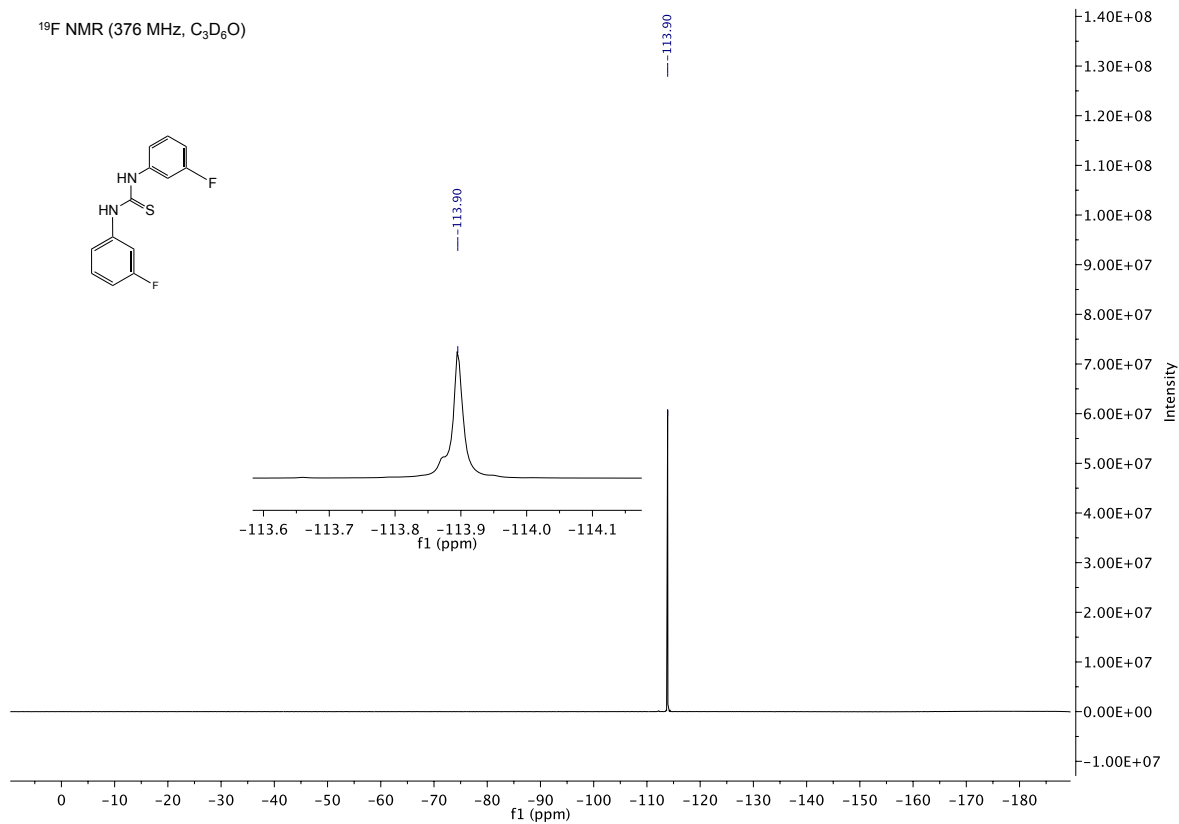
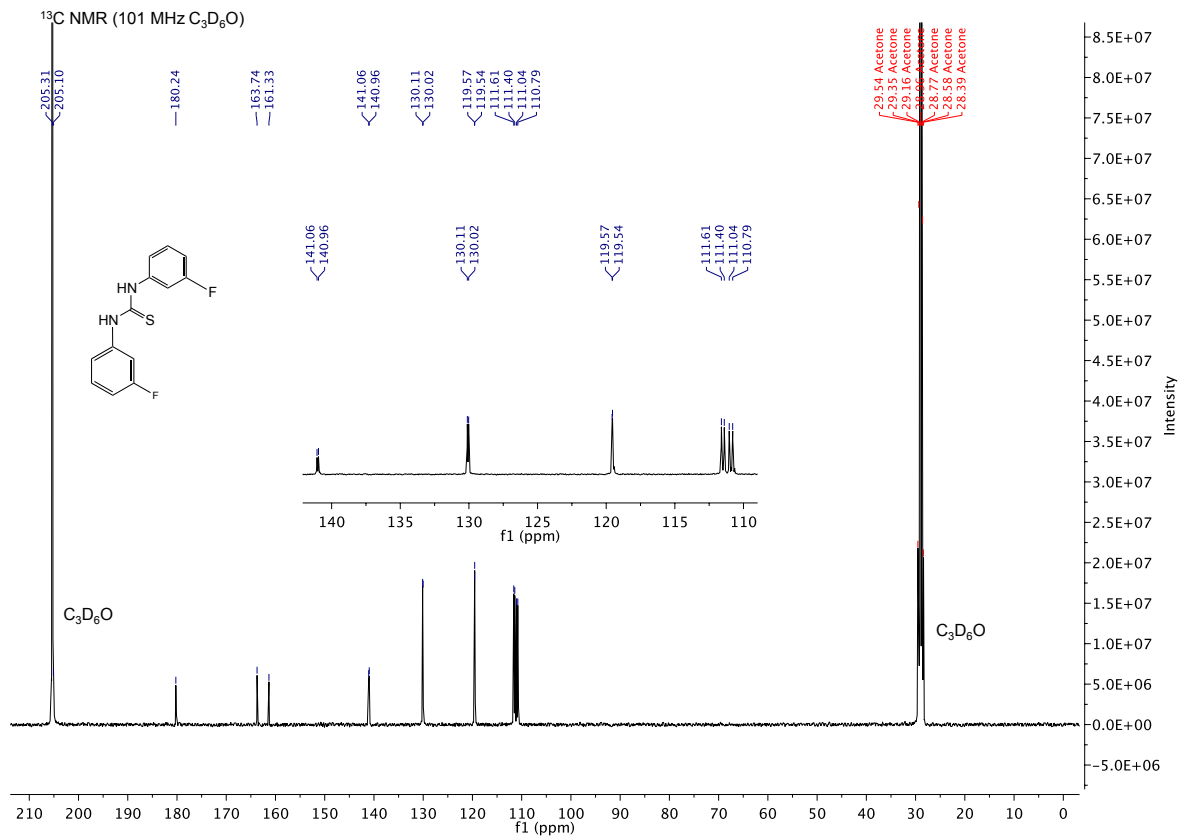
Compound 2k



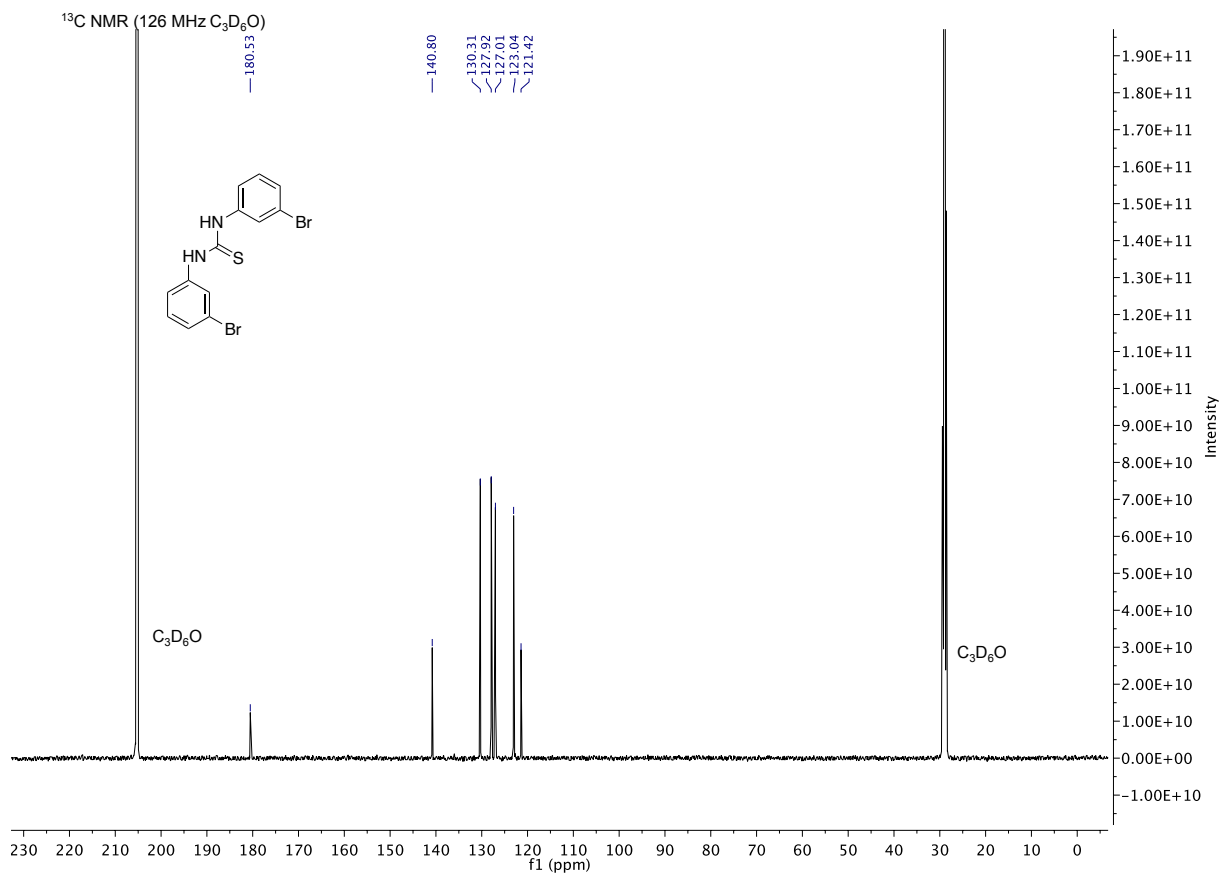
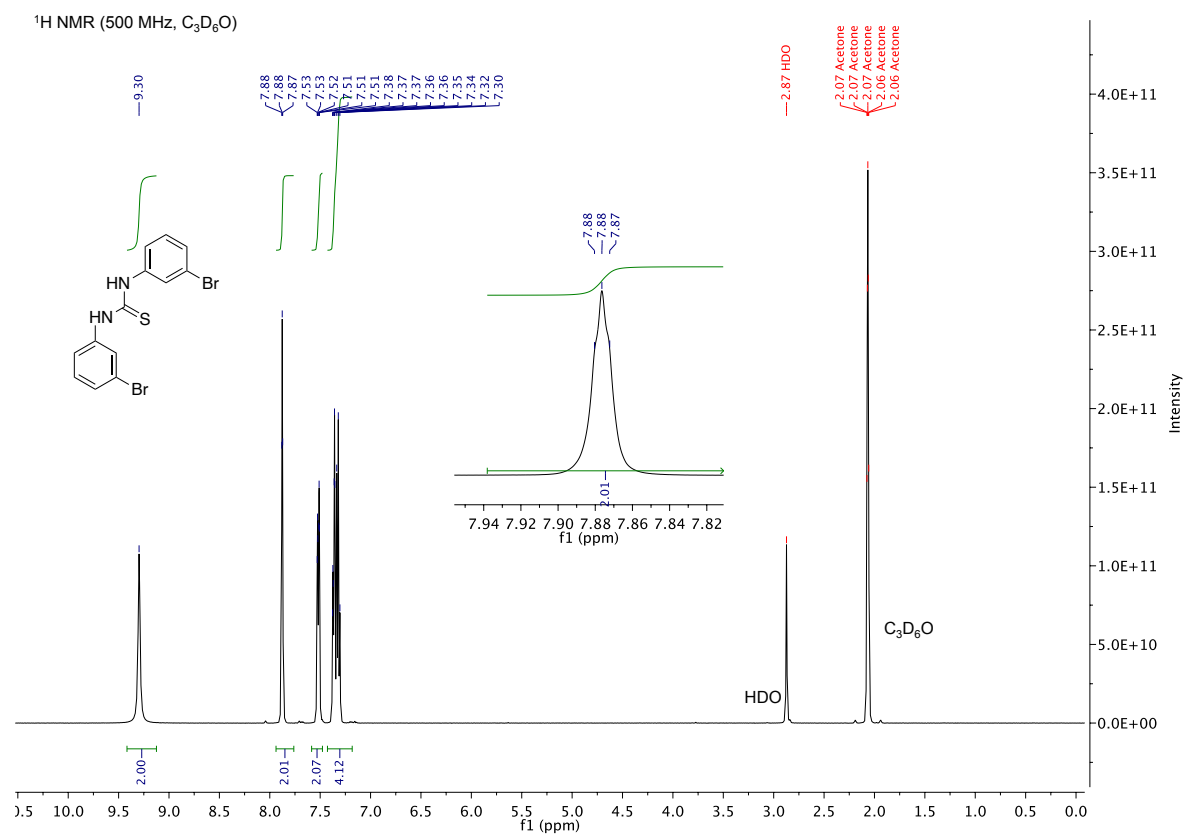


Compound 3a



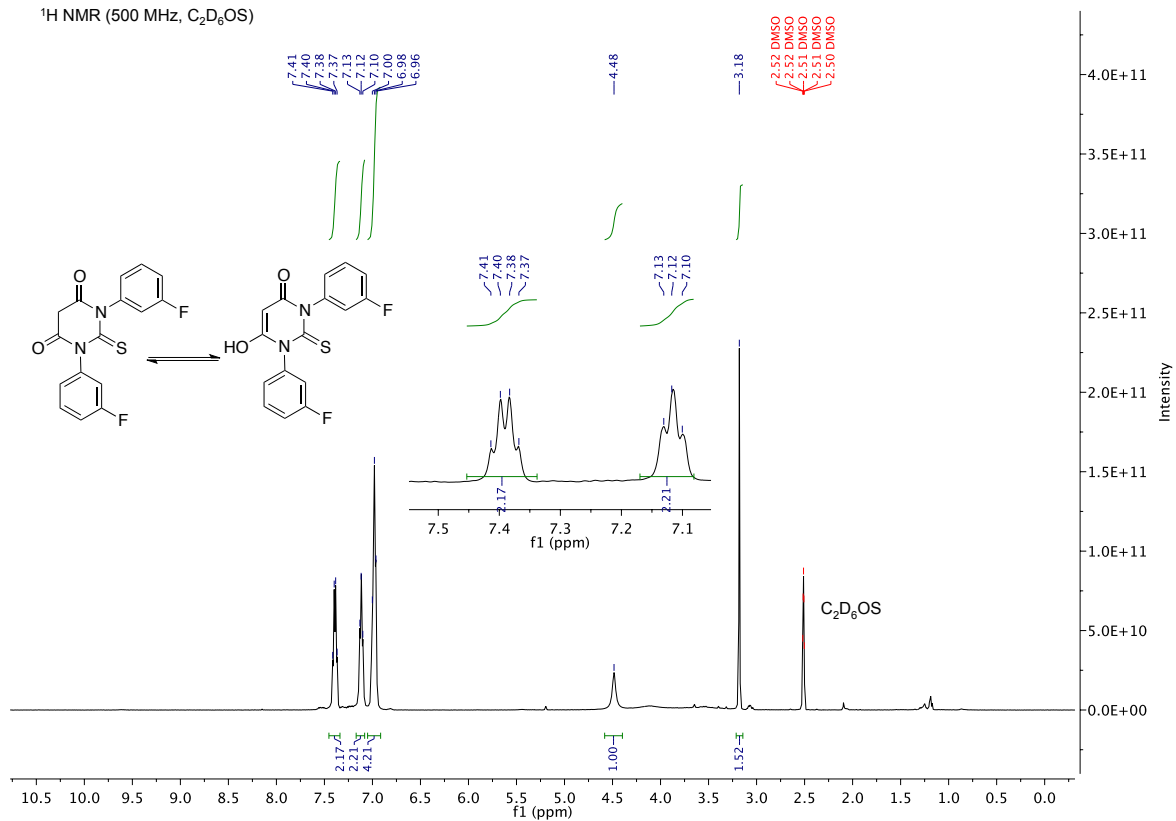


Compound 3b

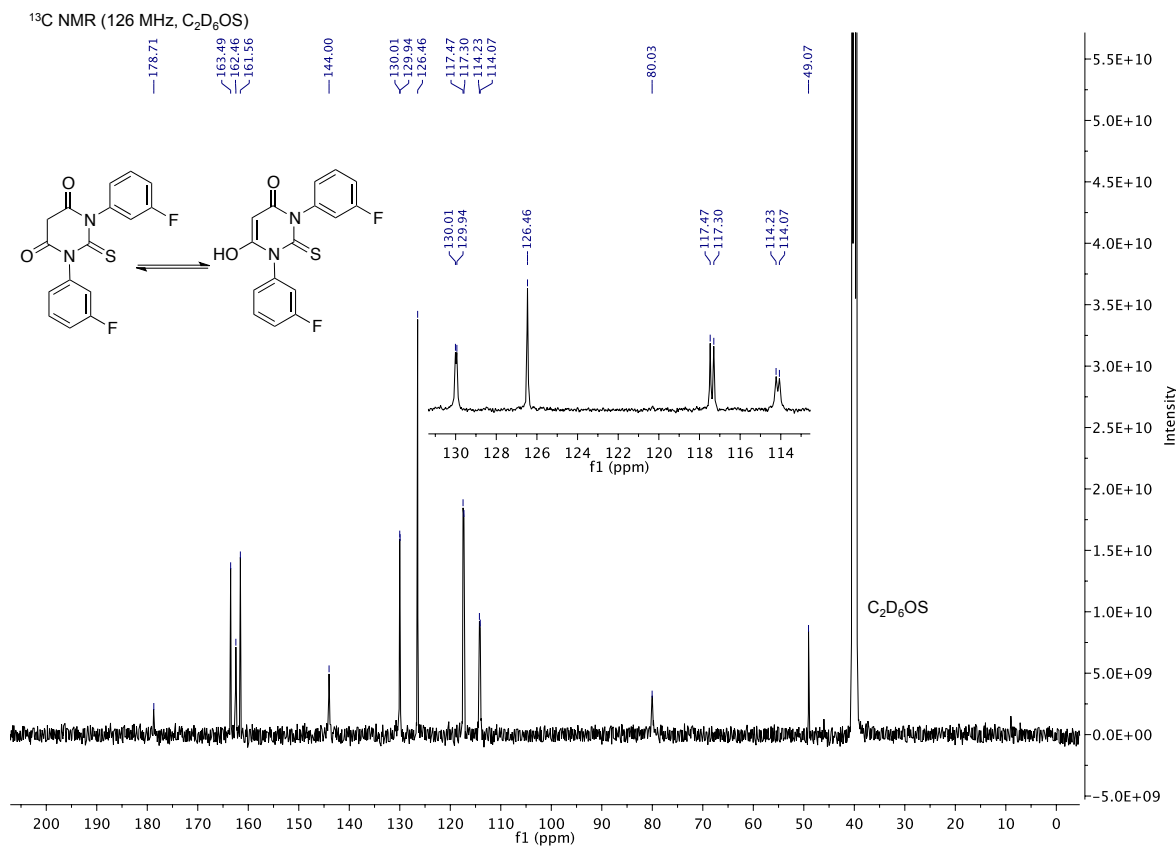


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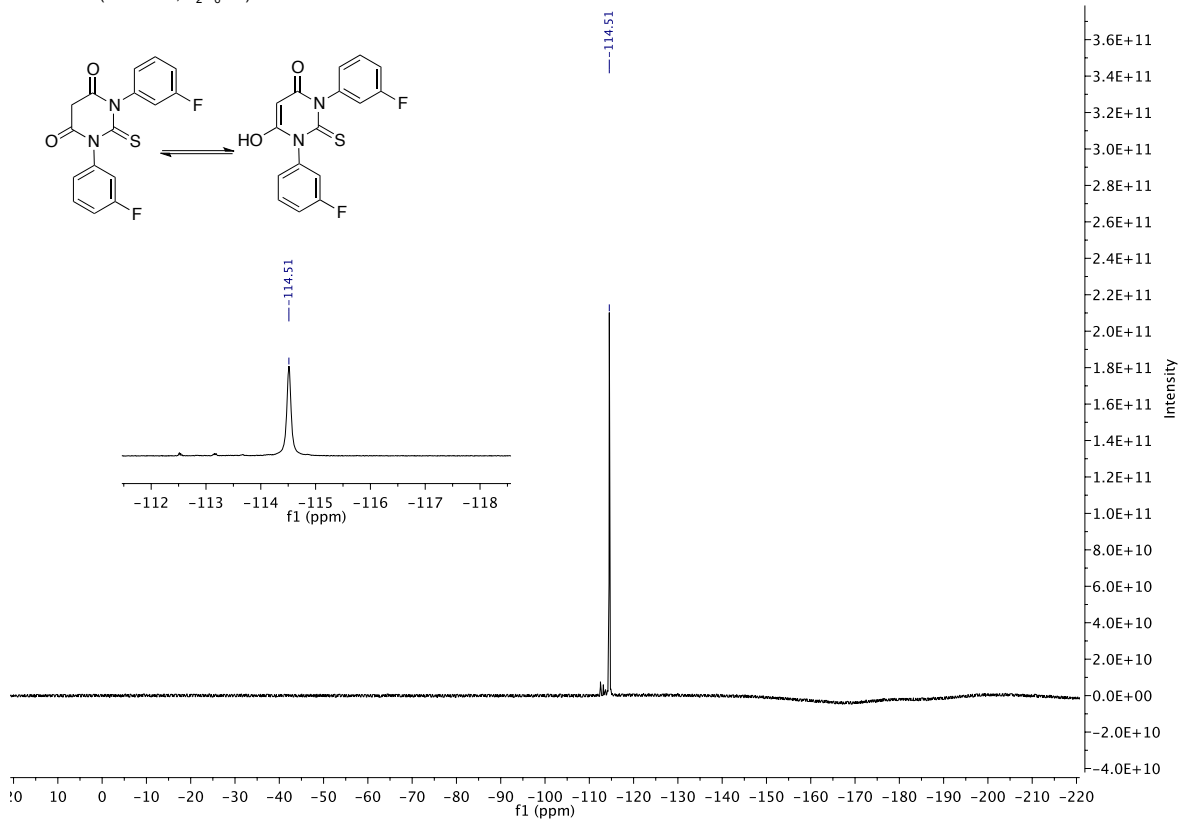
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¹³C NMR (126 MHz, C₂D₆OS)

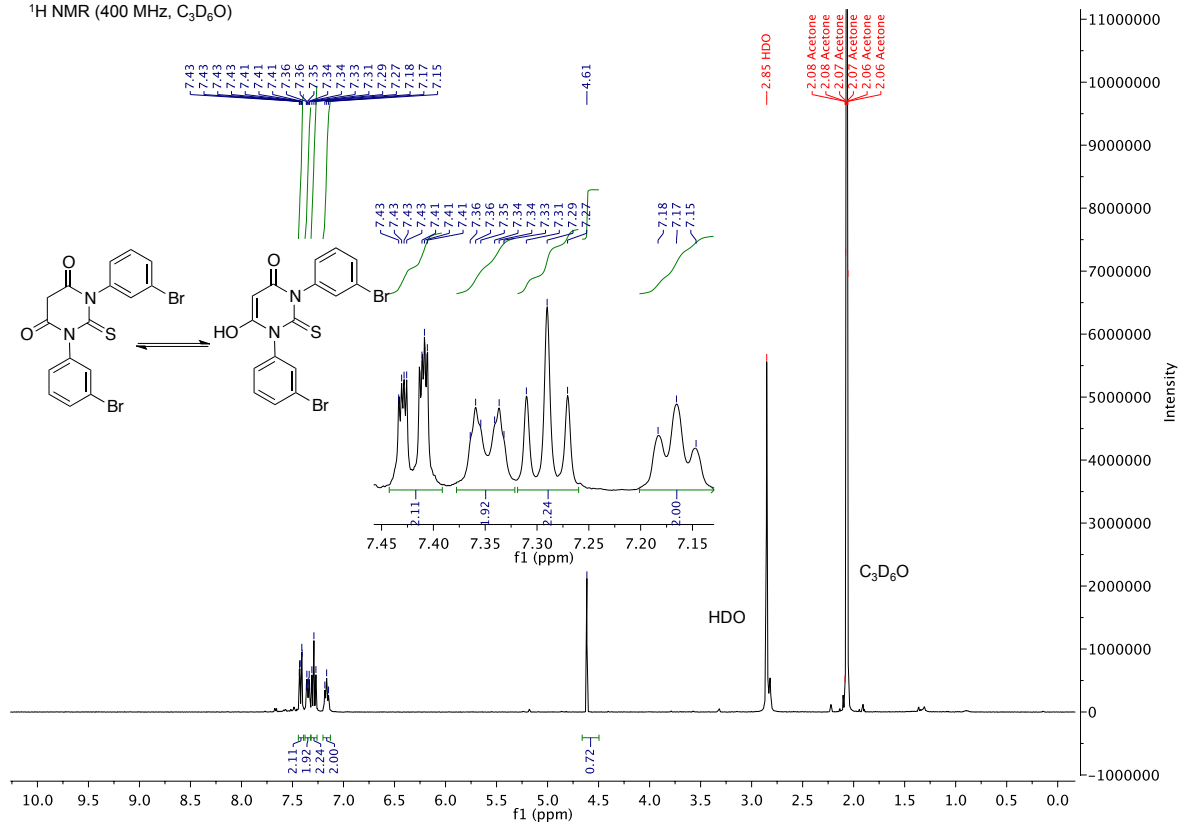


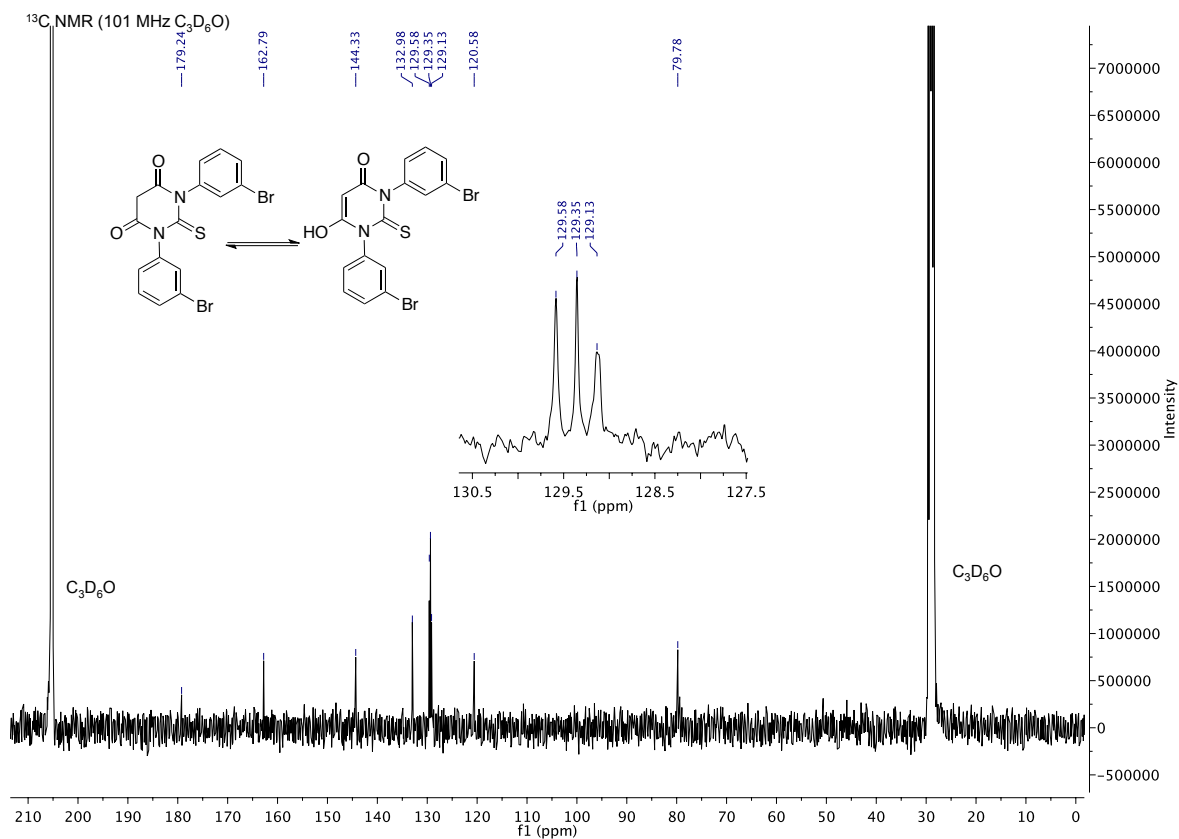
^{19}F NMR (471 MHz, $\text{C}_2\text{D}_6\text{OS}$)



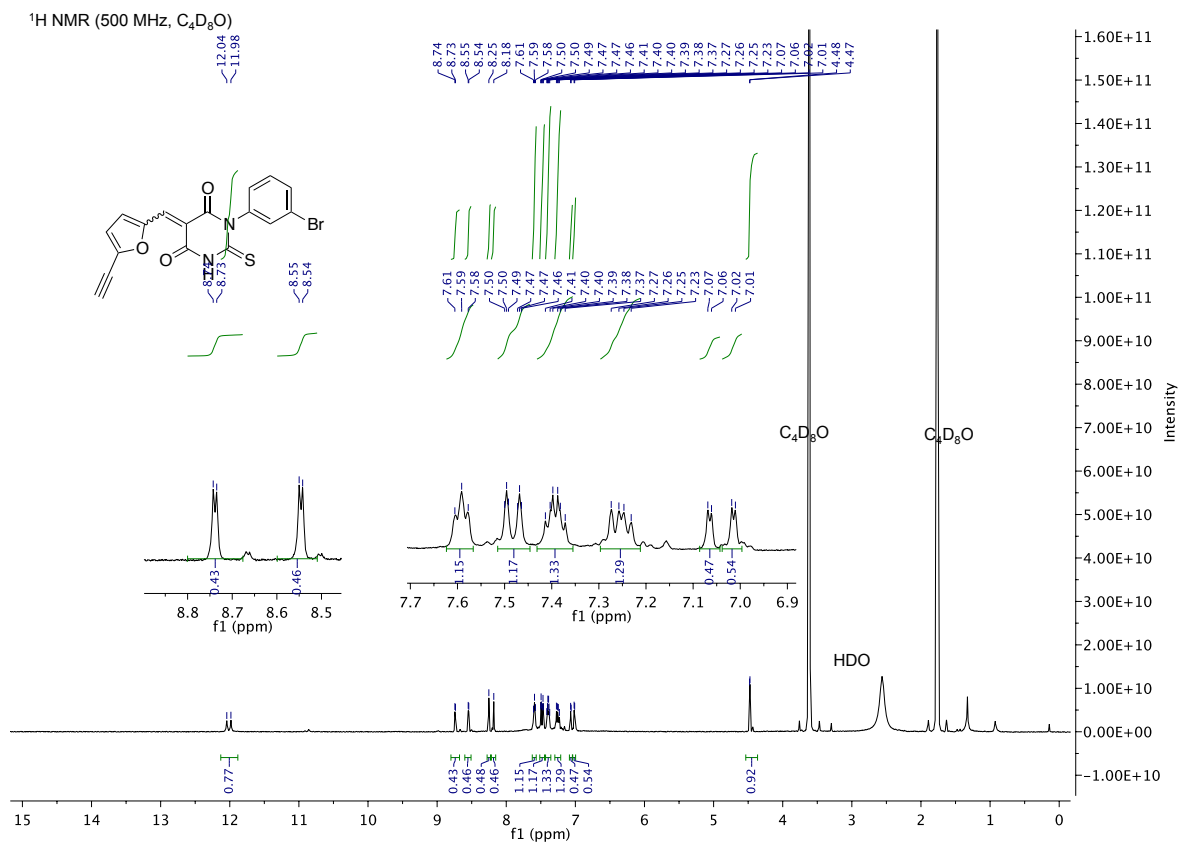
Compound 4b

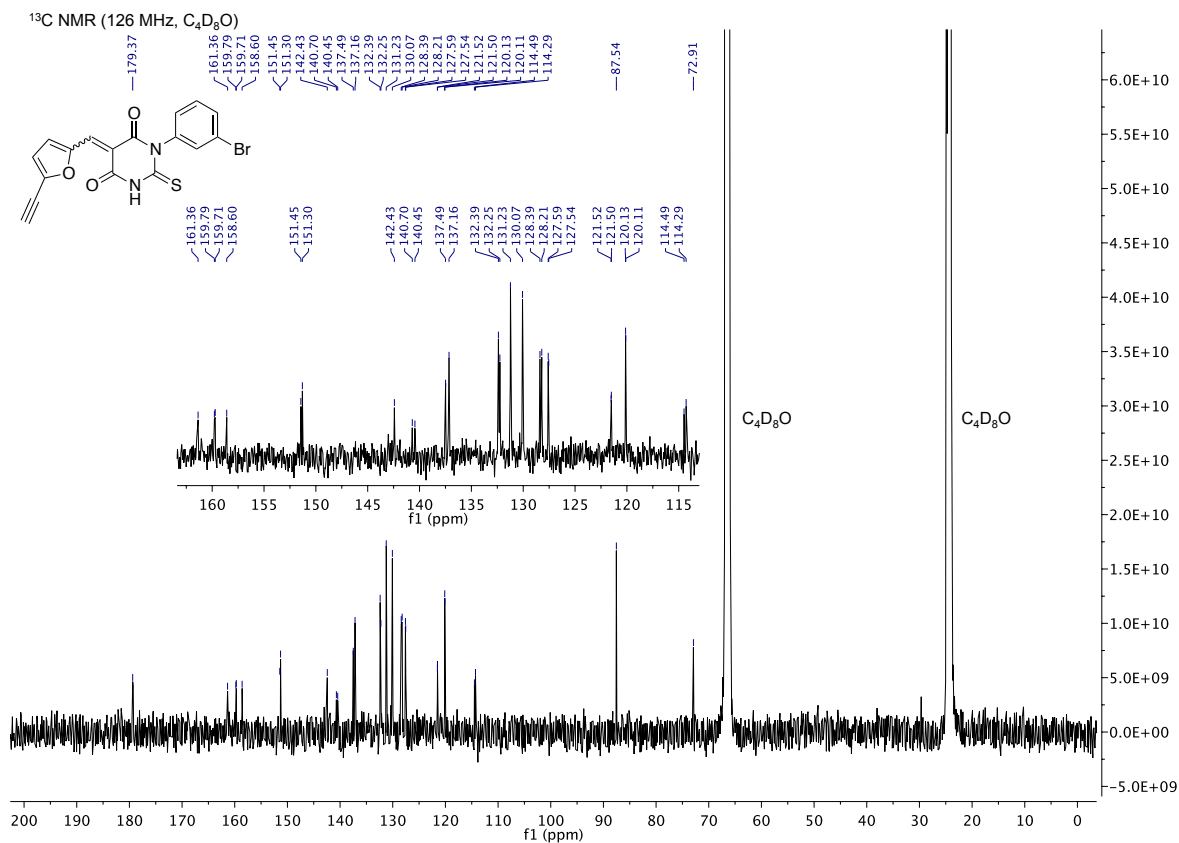
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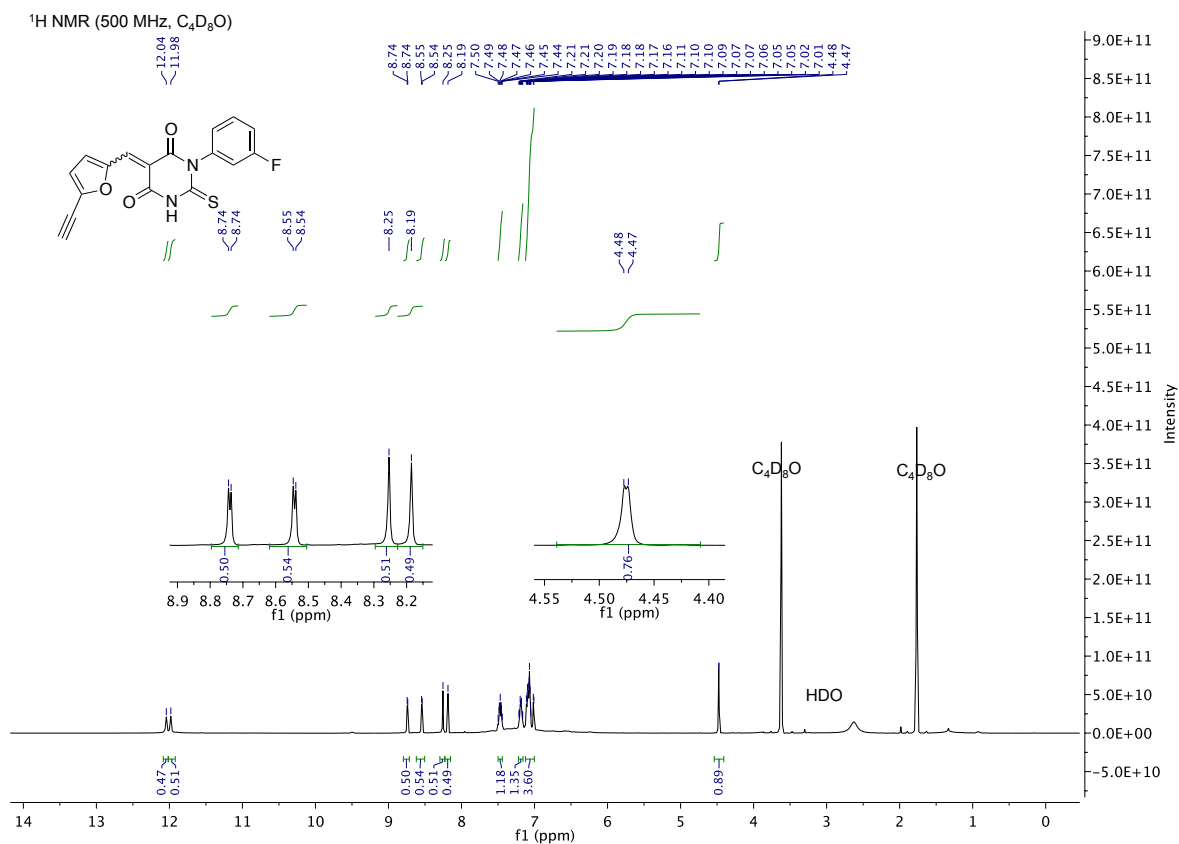


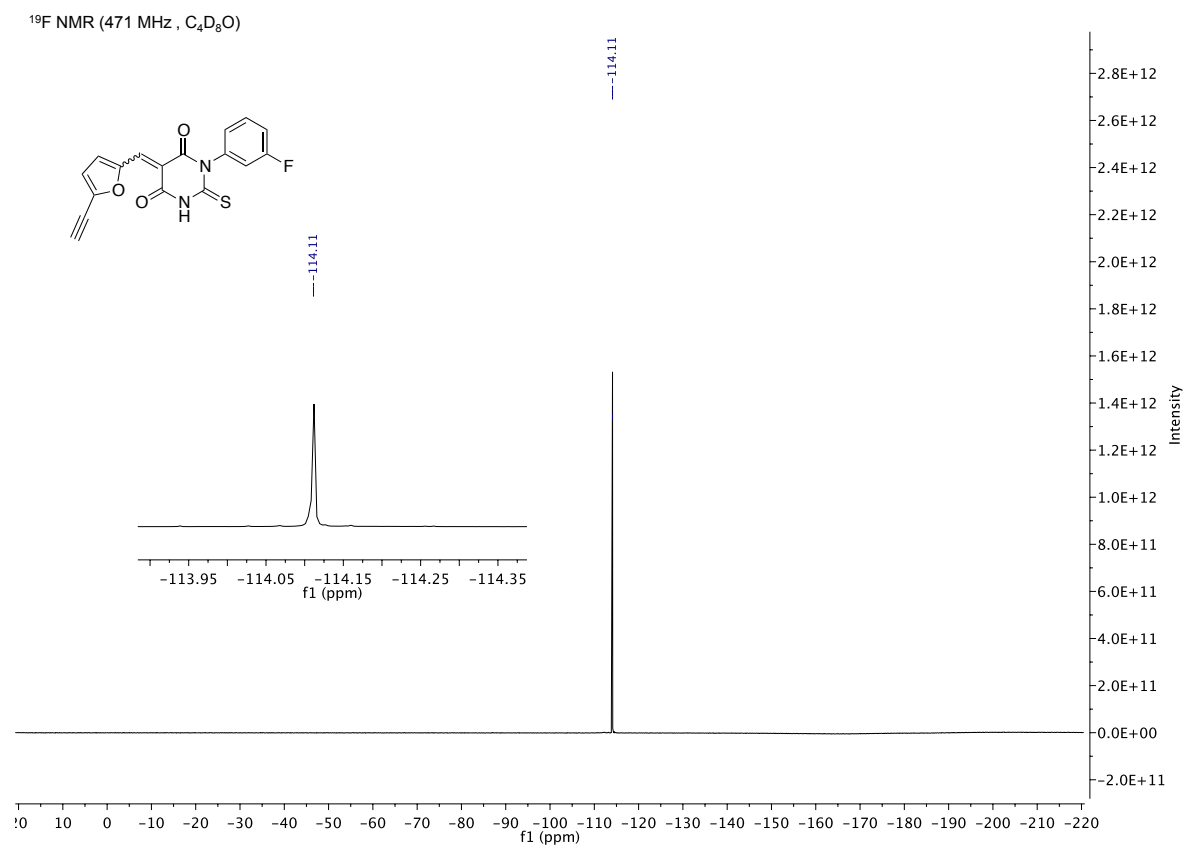
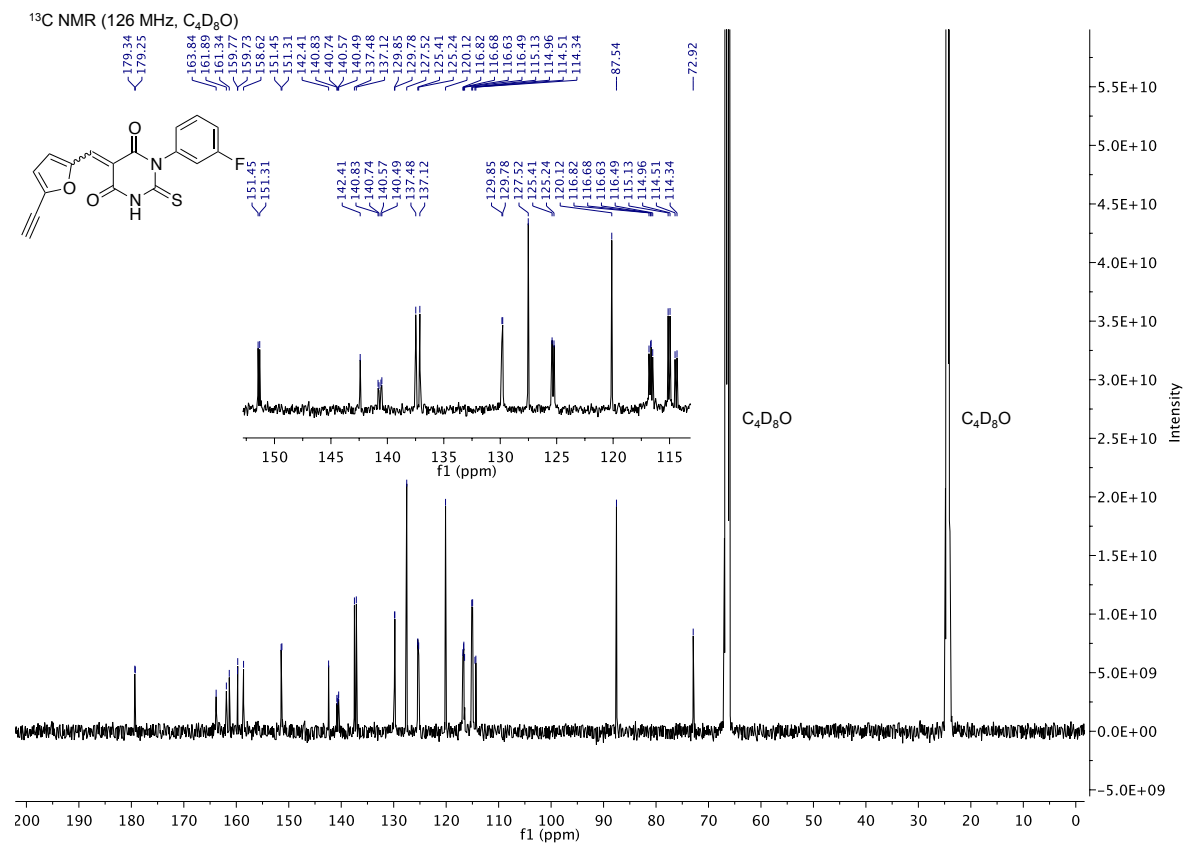
Compound 5a



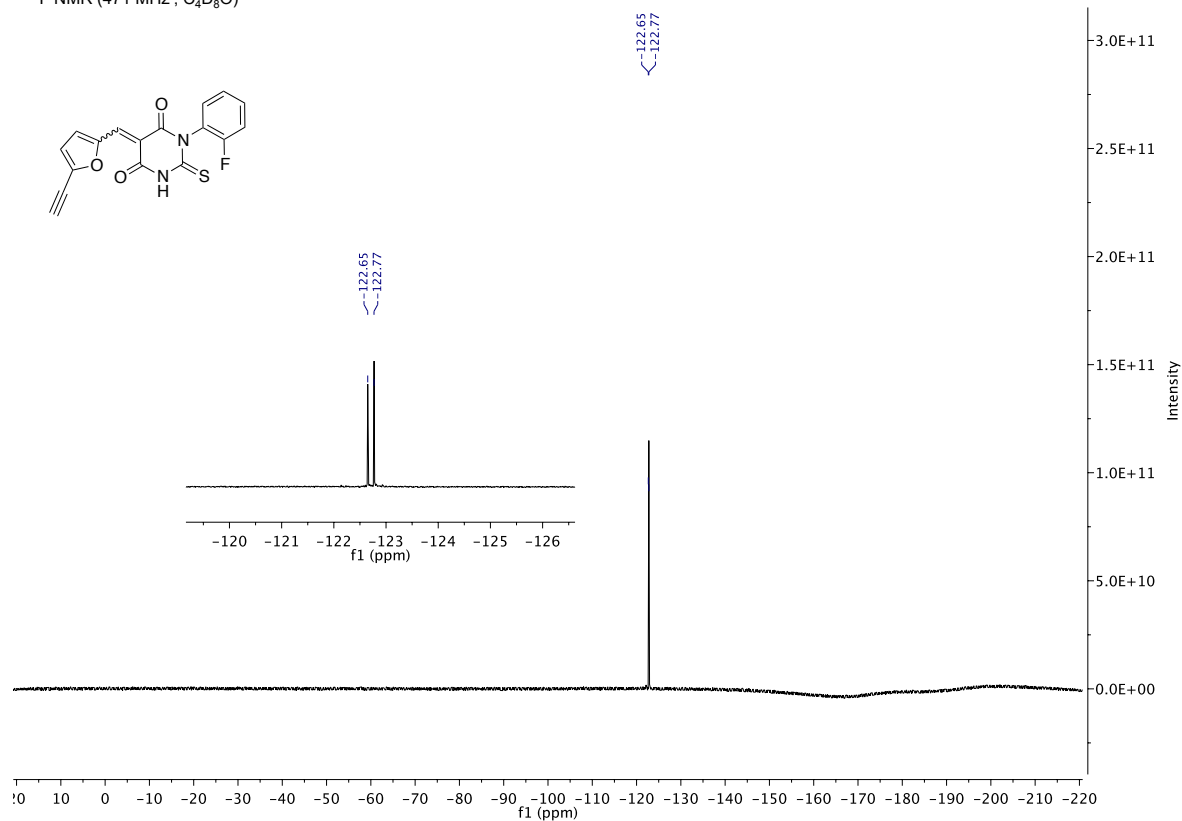


Compound 5b



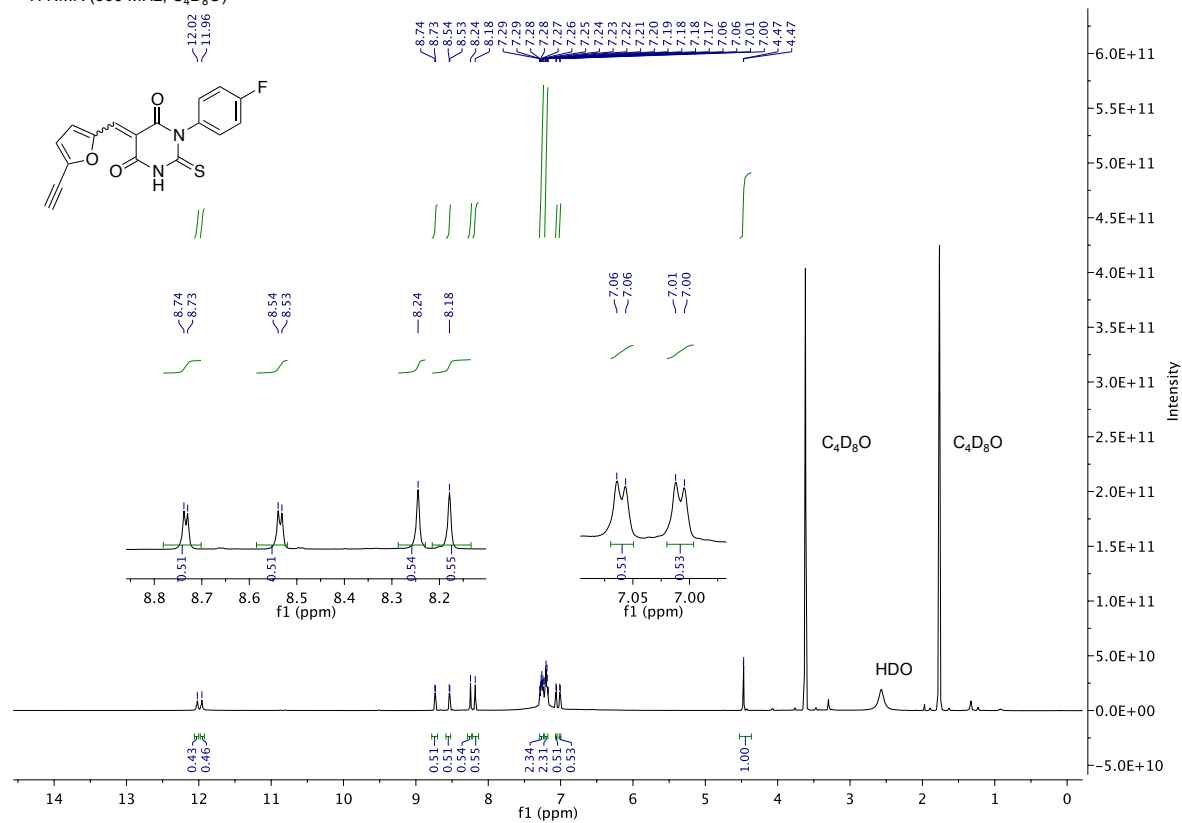


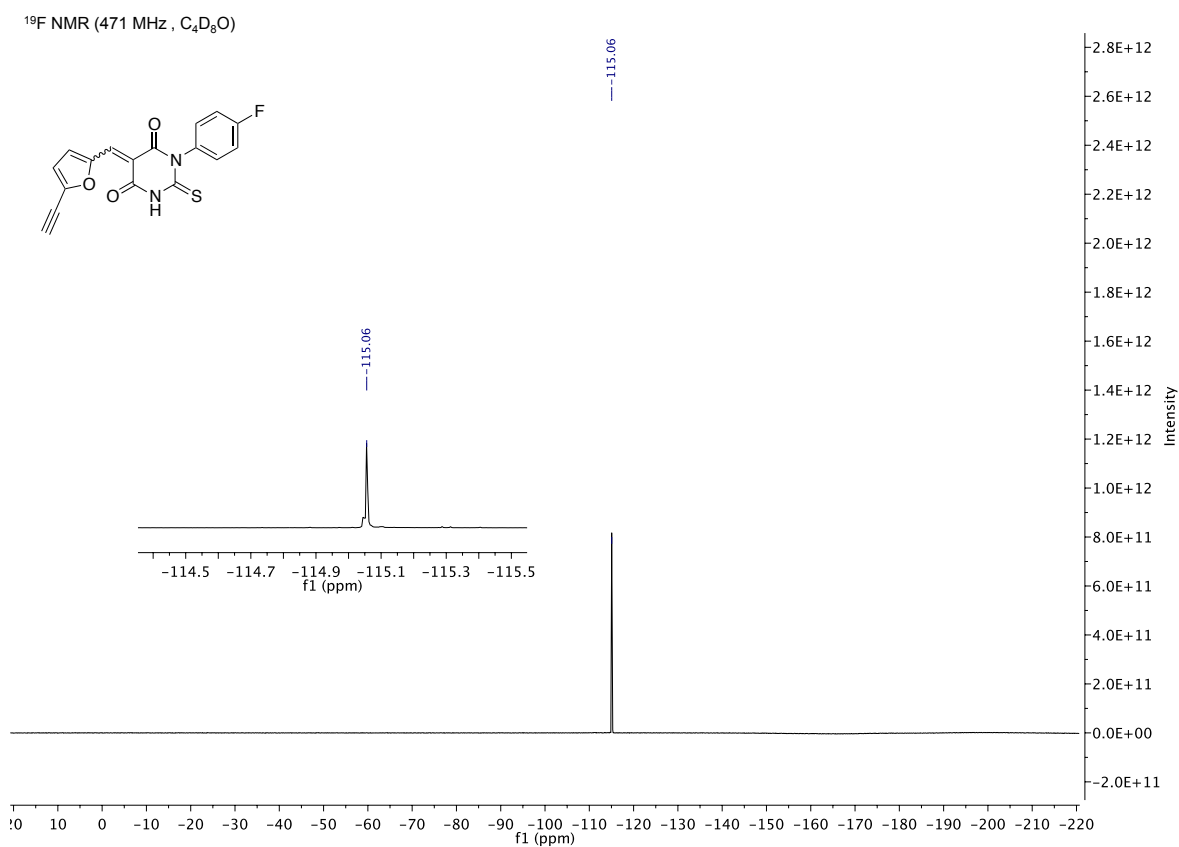
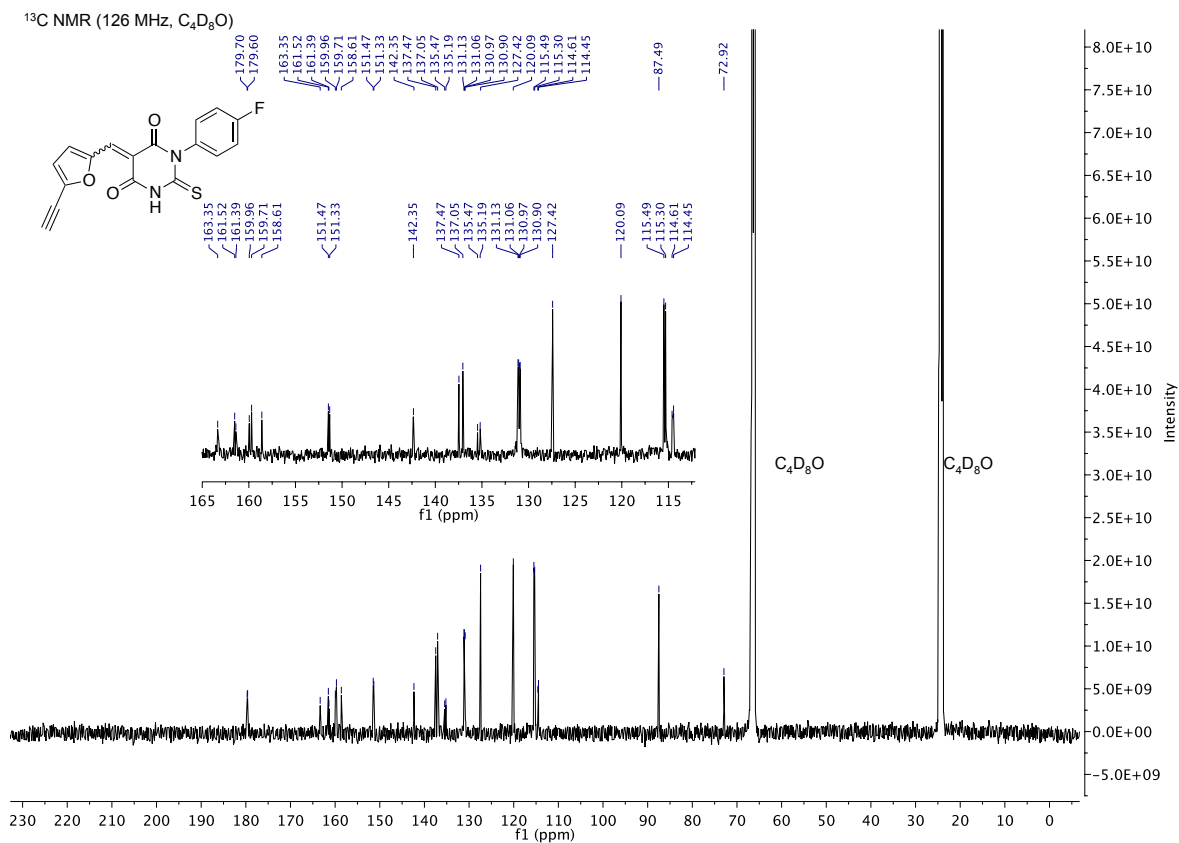
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Compound 5d

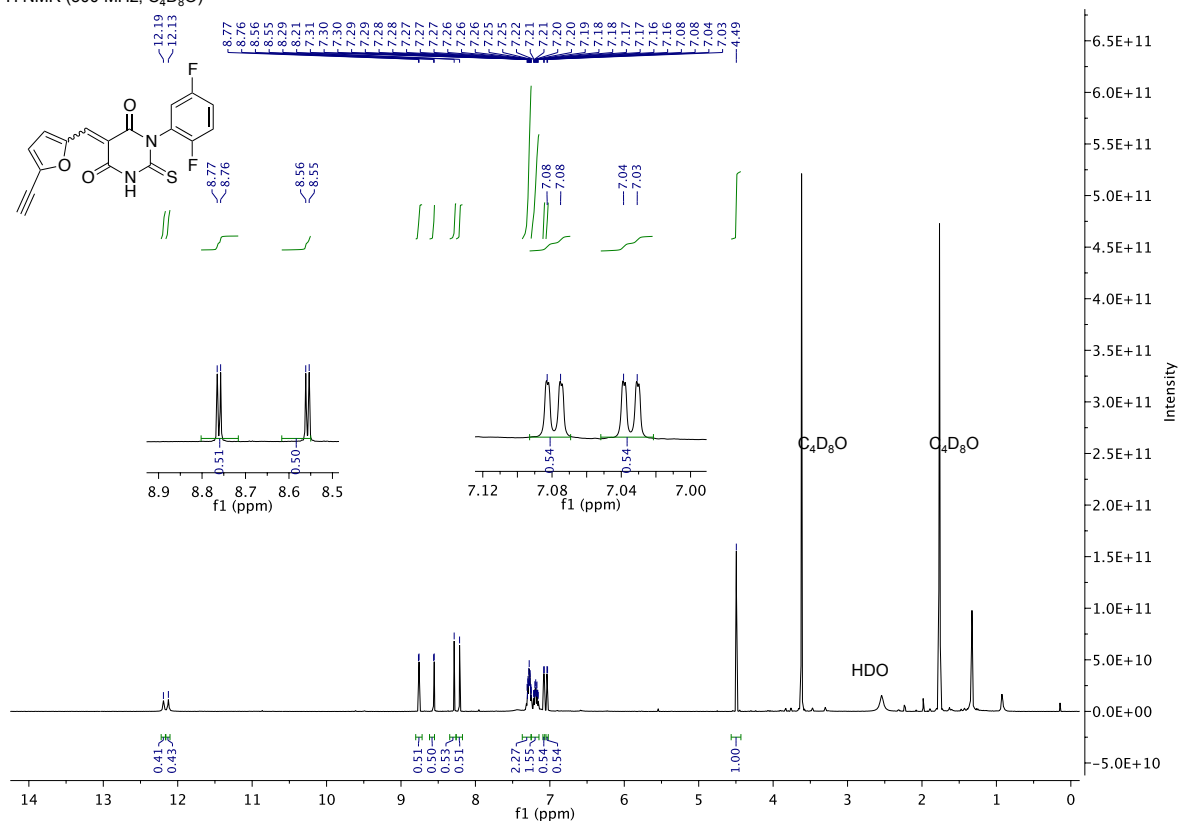
^1H NMR (500 MHz, $\text{C}_4\text{D}_8\text{O}$)



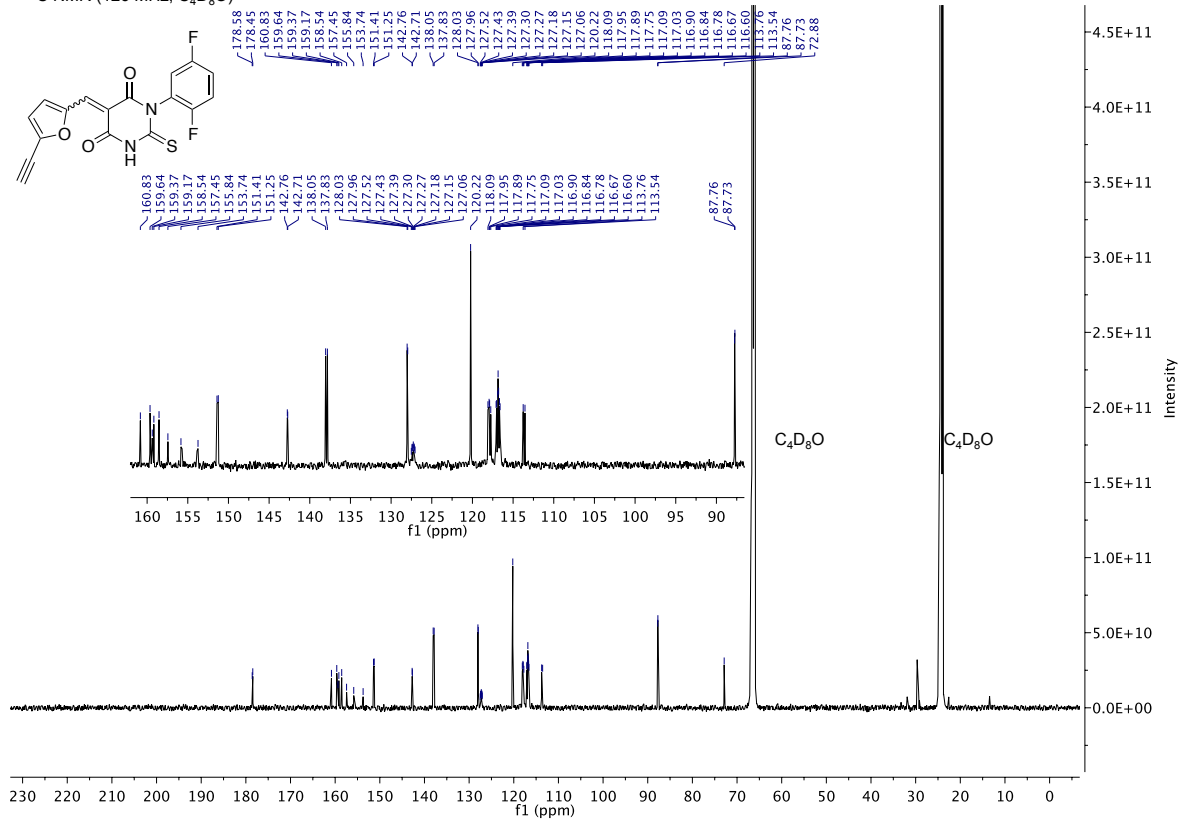


Compound 5e

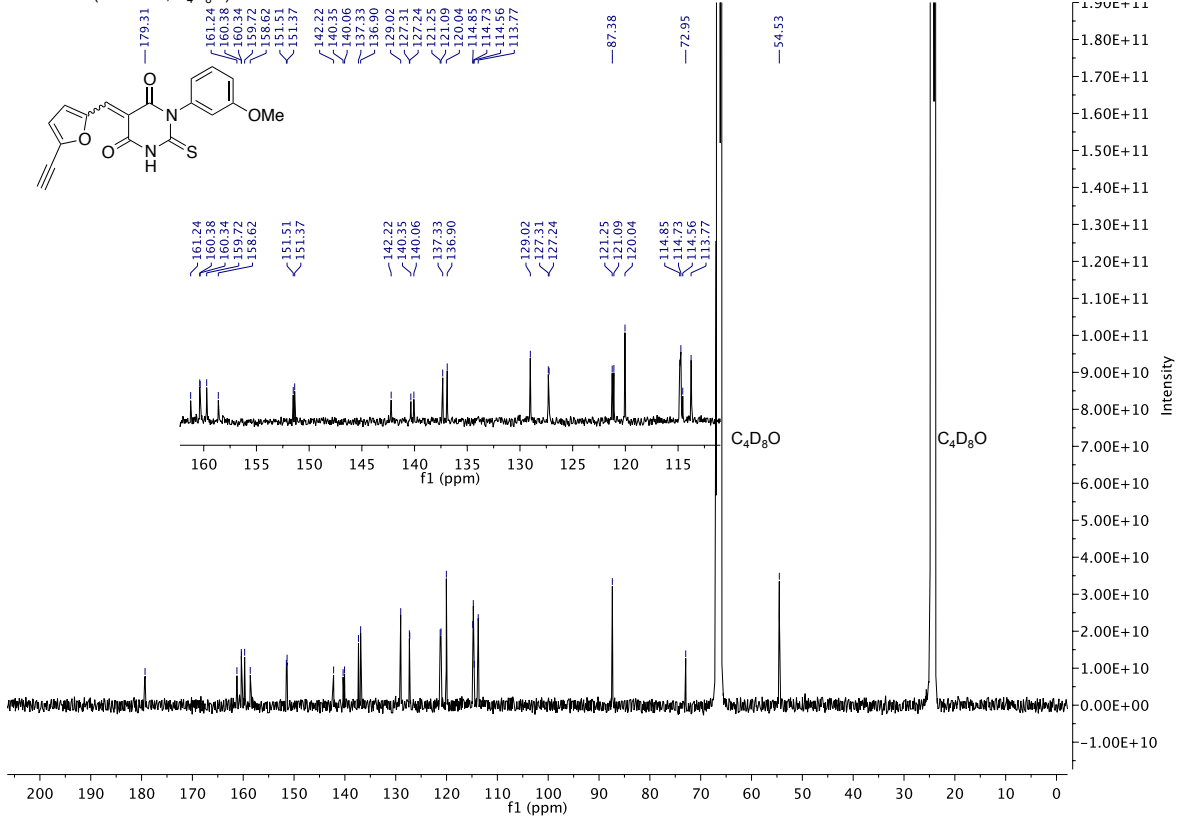
¹H NMR (500 MHz, C₄D₈O)



¹³C NMR (126 MHz, C₄D₈O)

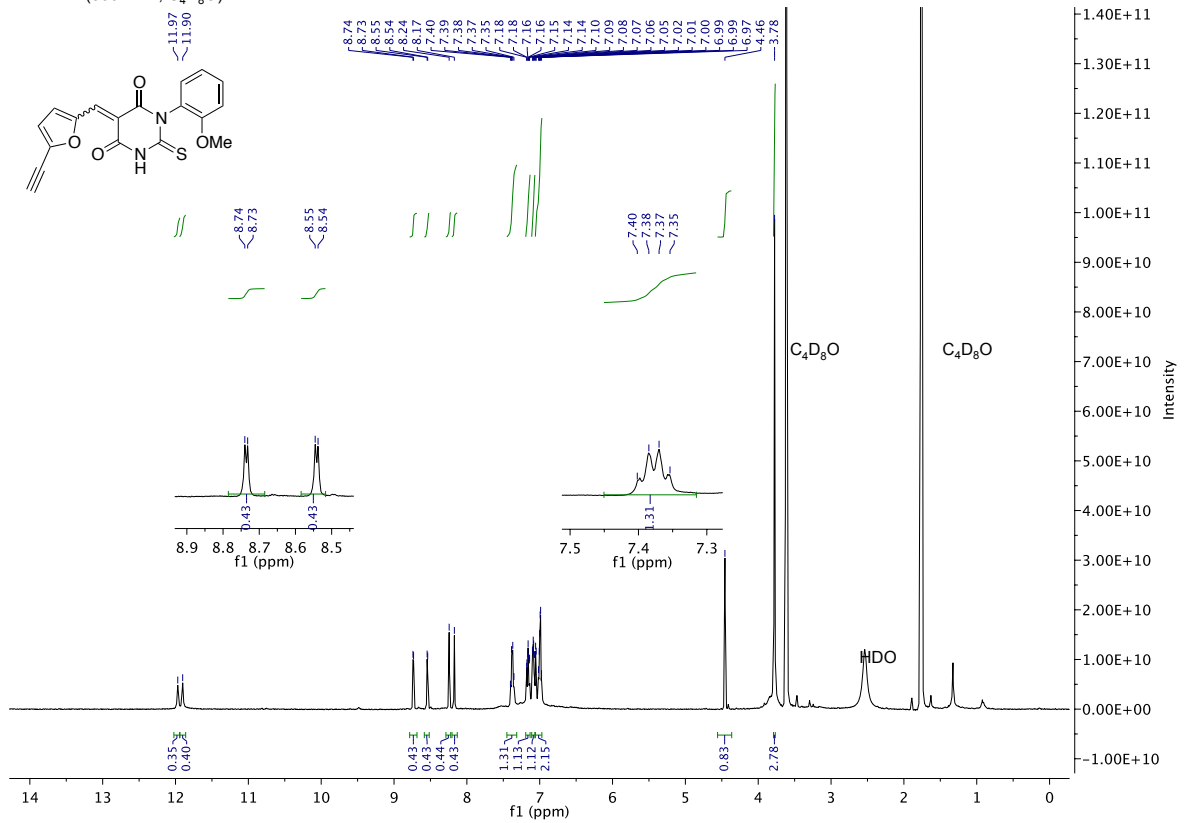


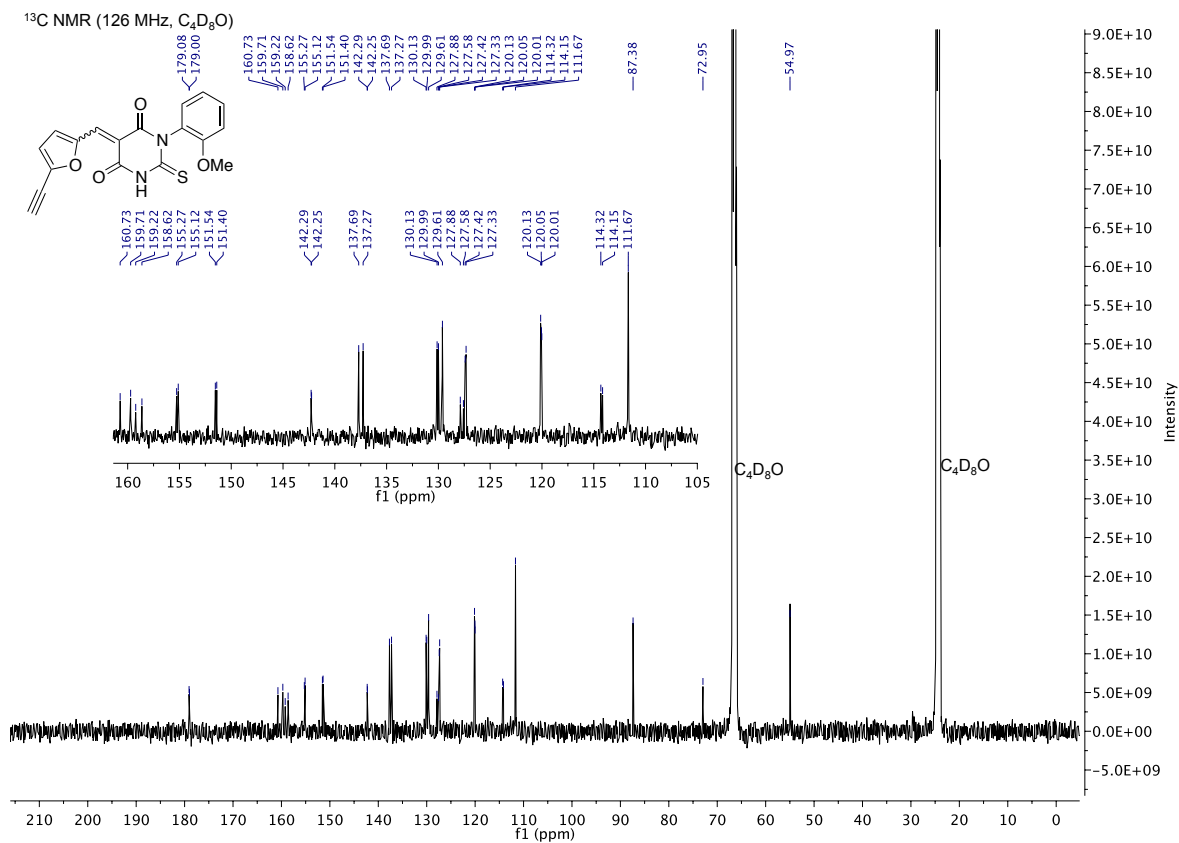
¹³C NMR (126 MHz, C₄D₈O)



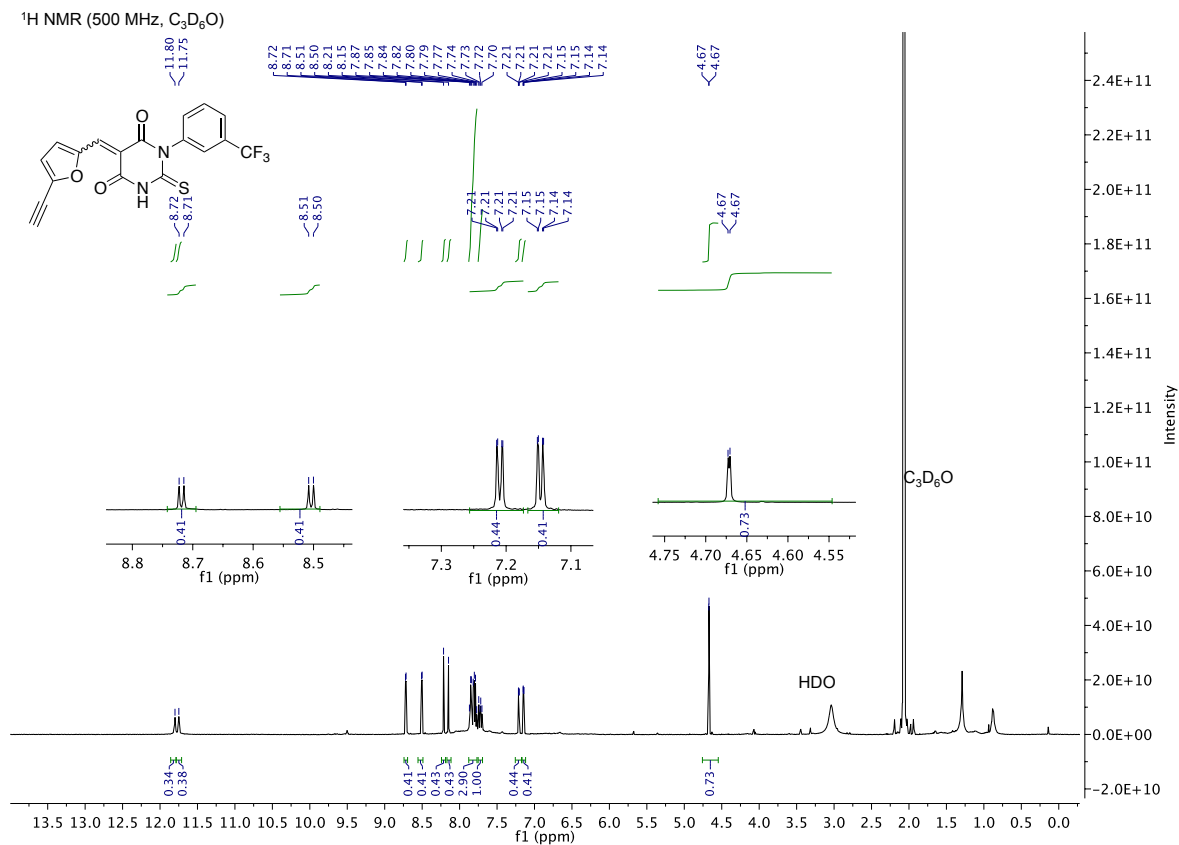
Compound 5g

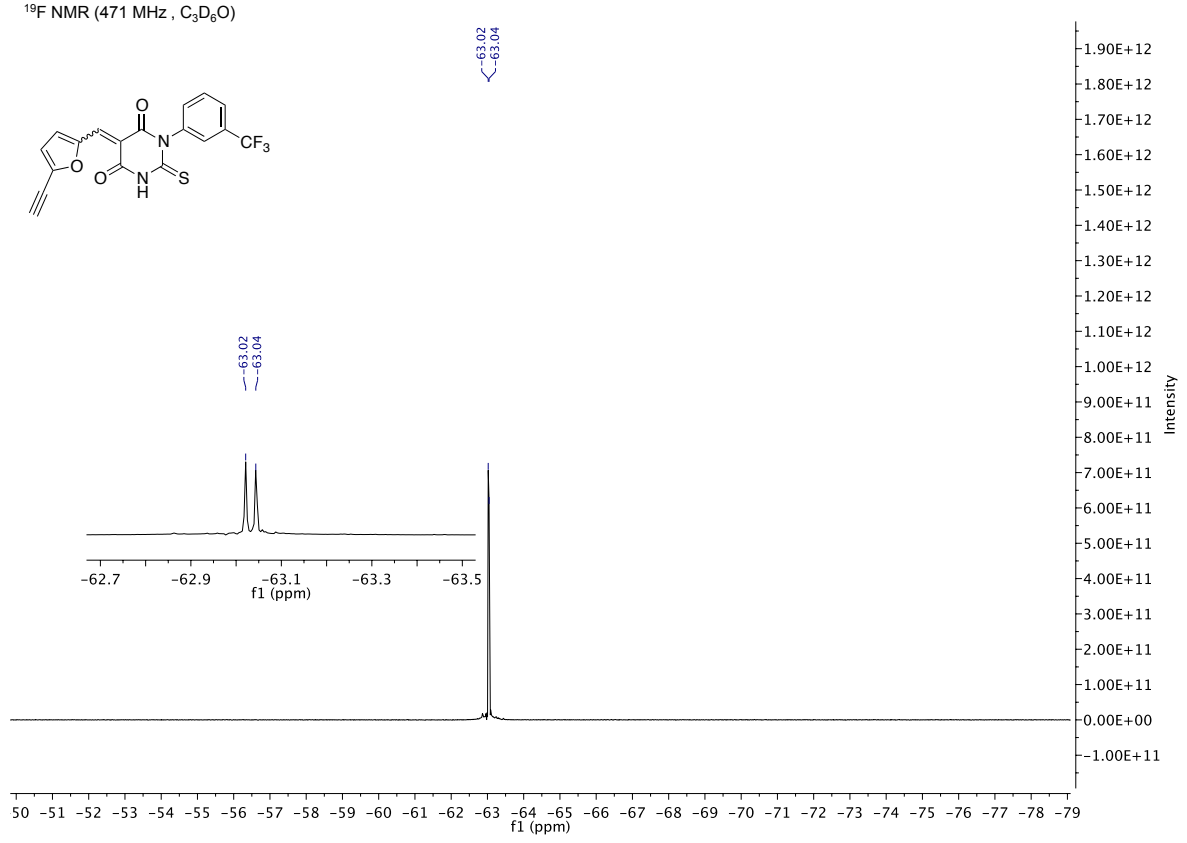
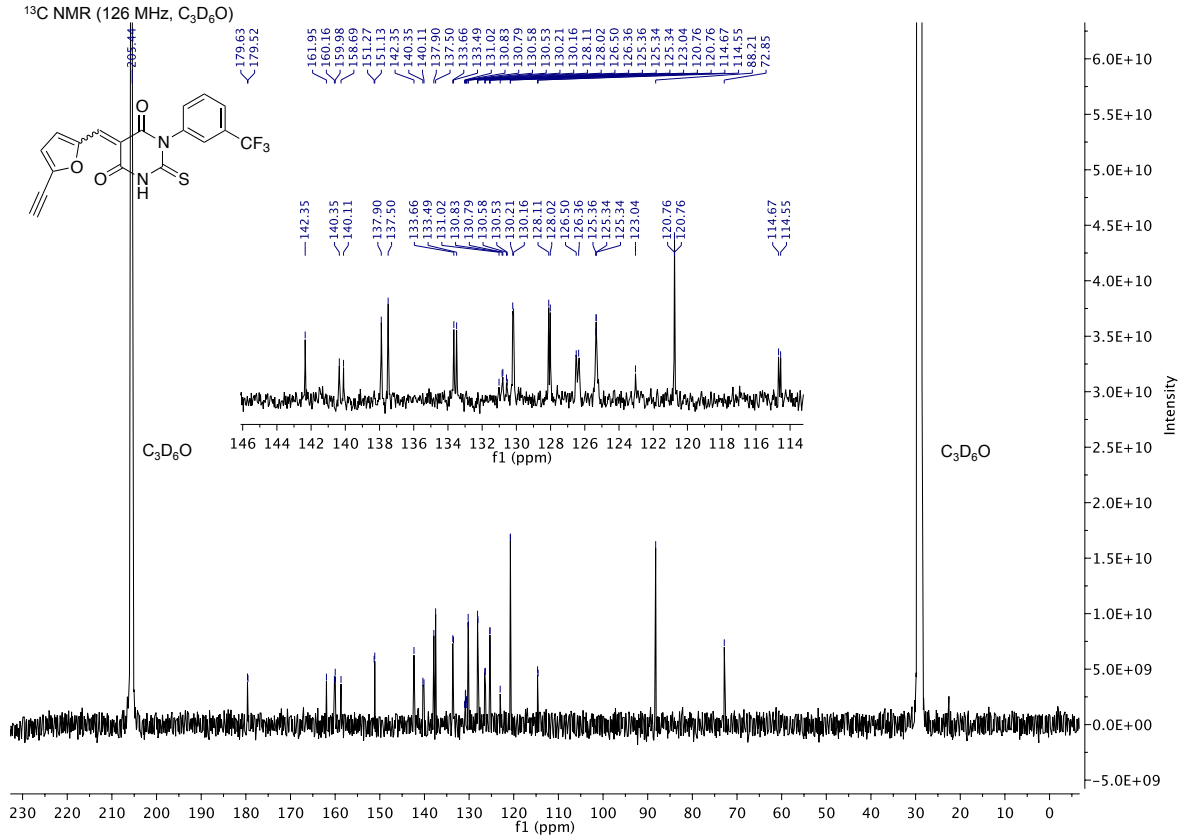
¹H NMR (500 MHz, C₄D₈O)





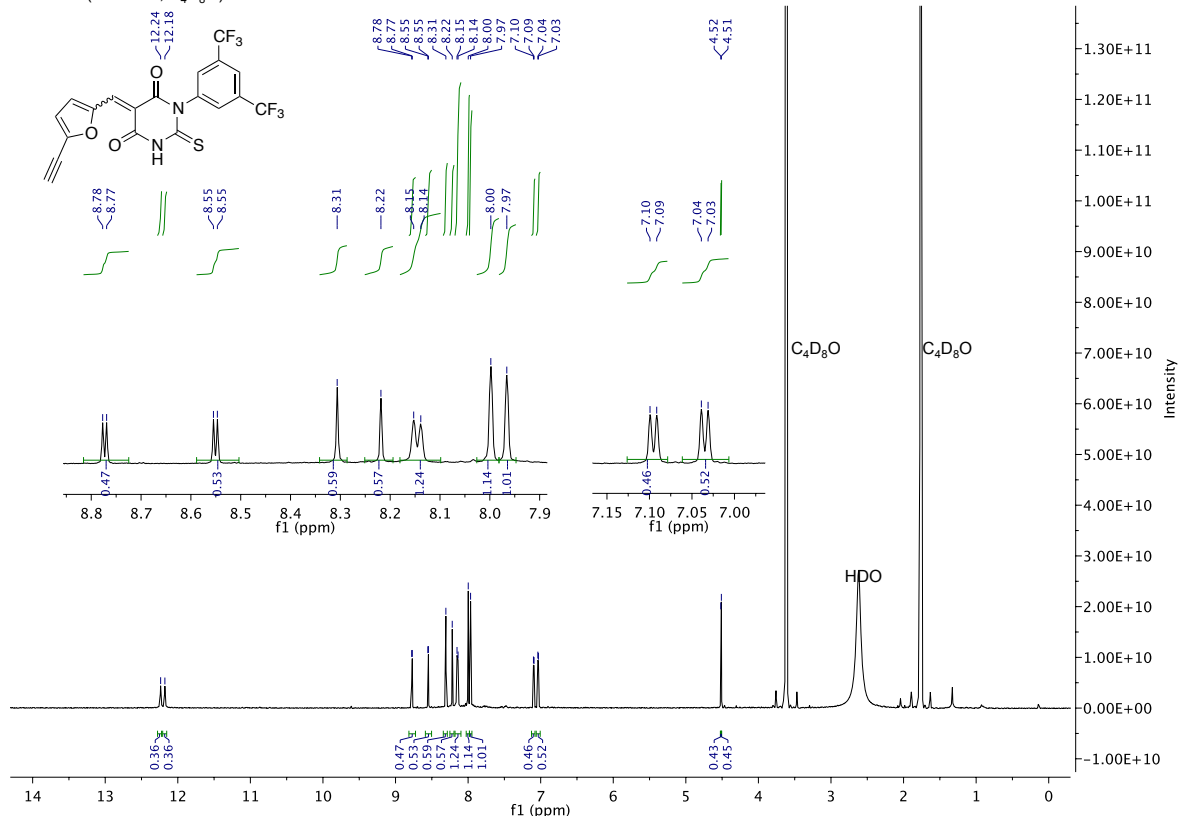
Compound 5h



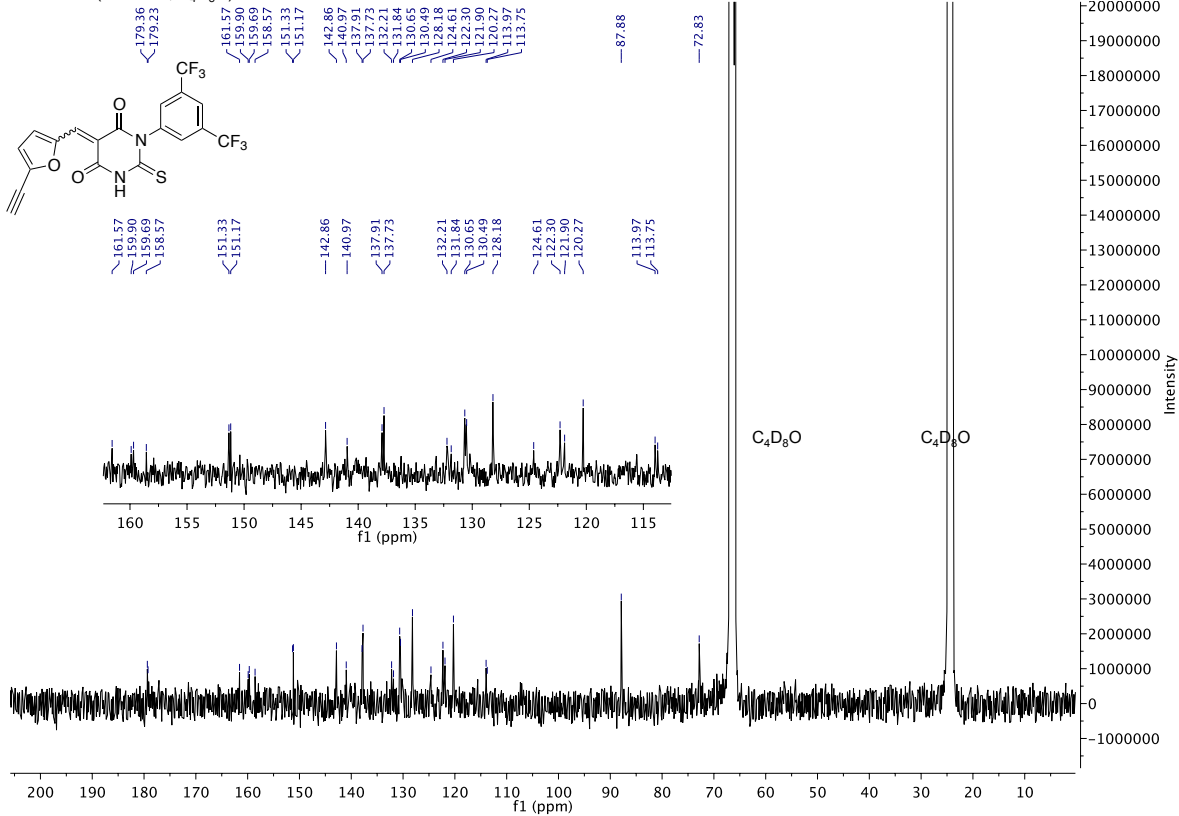


Compound 5i

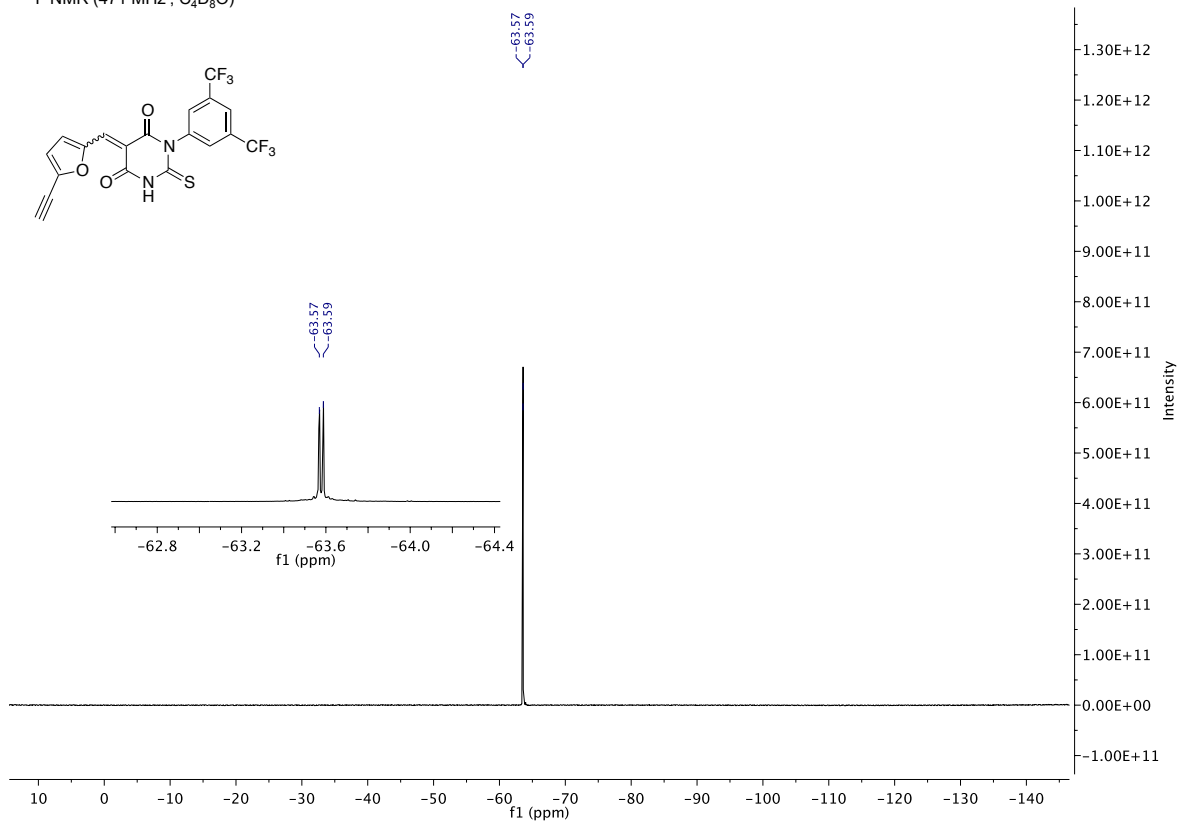
¹H NMR (500 MHz, C₄D₈O)



¹³C NMR (126 MHz, C₄D₈O)

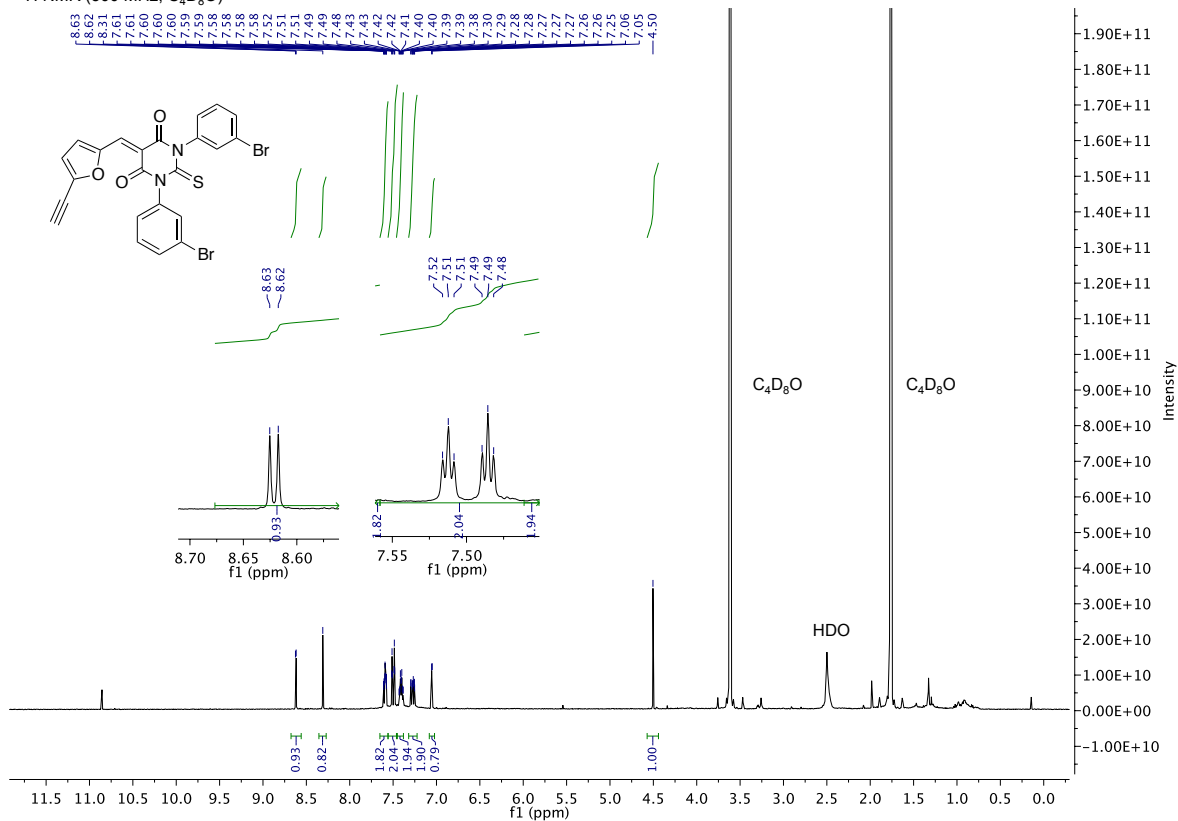


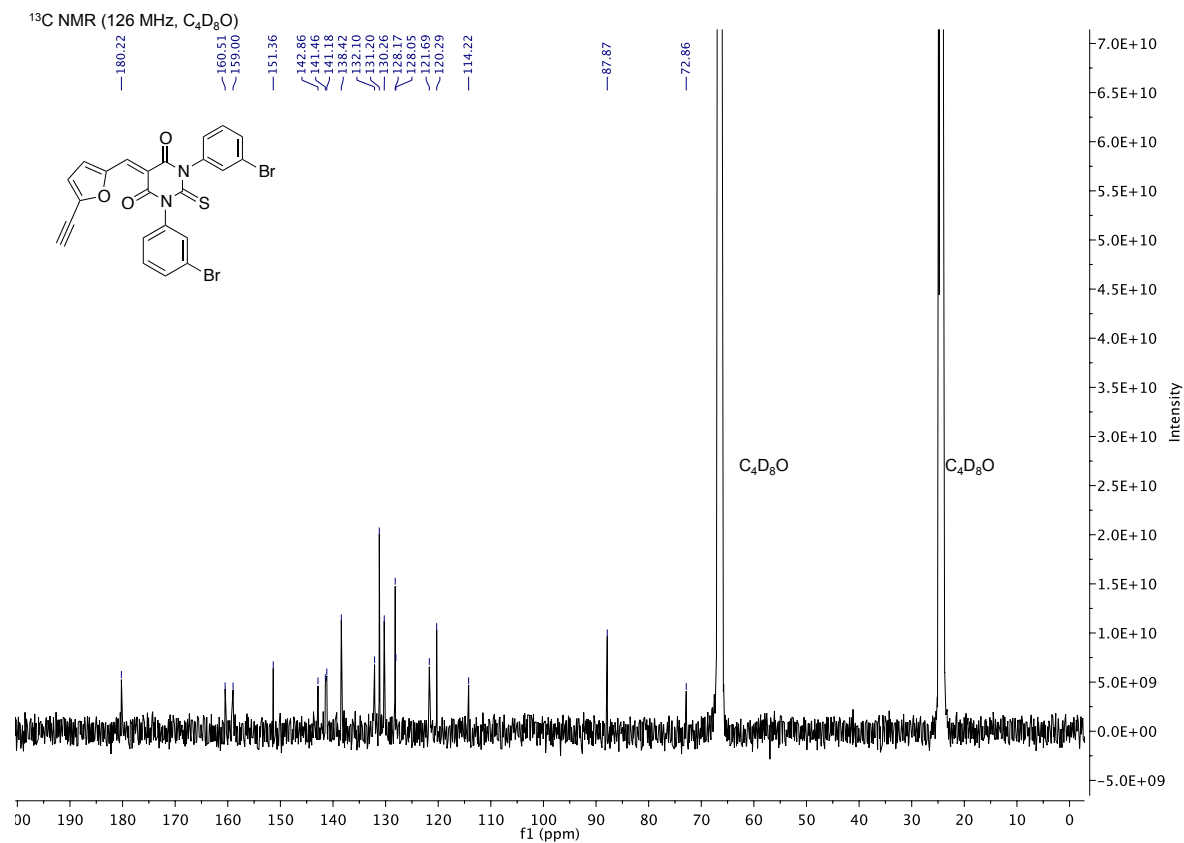
¹⁹F NMR (471 MHz, C₄D₈O)



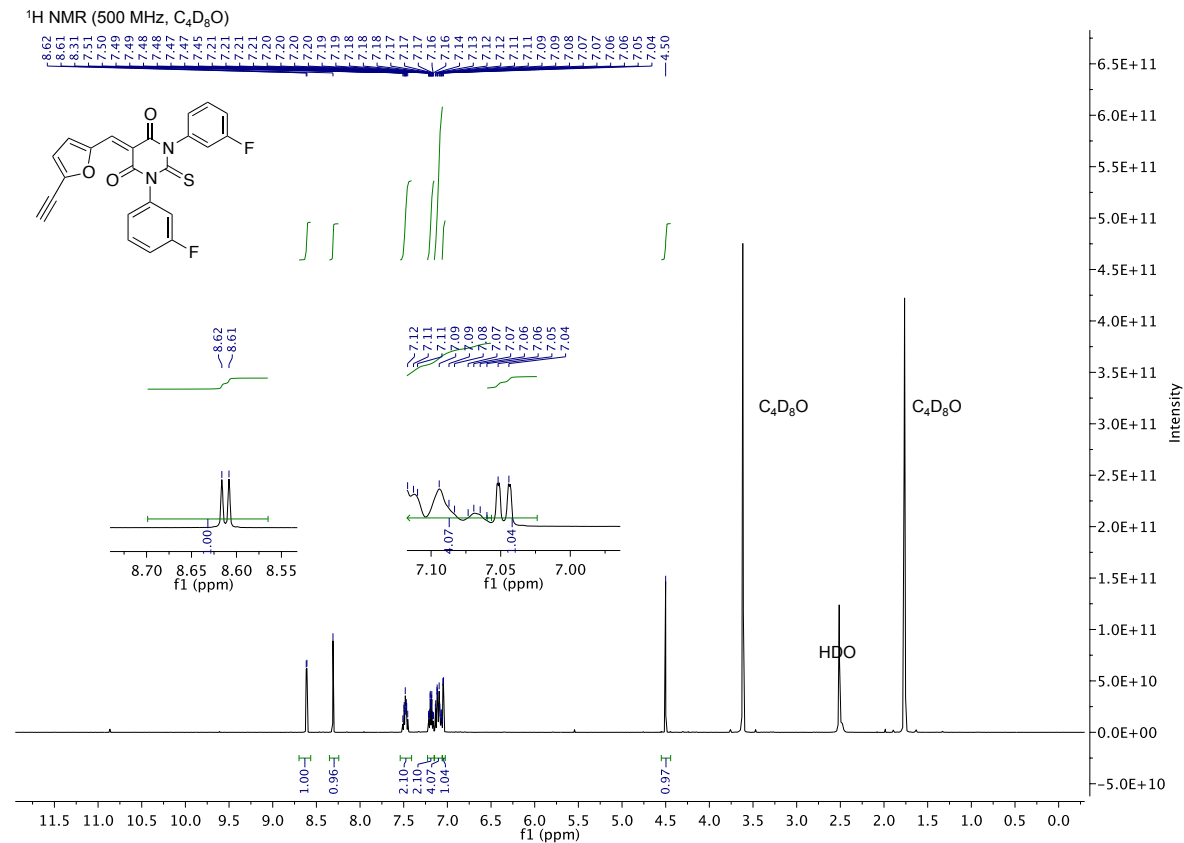
Compound 51

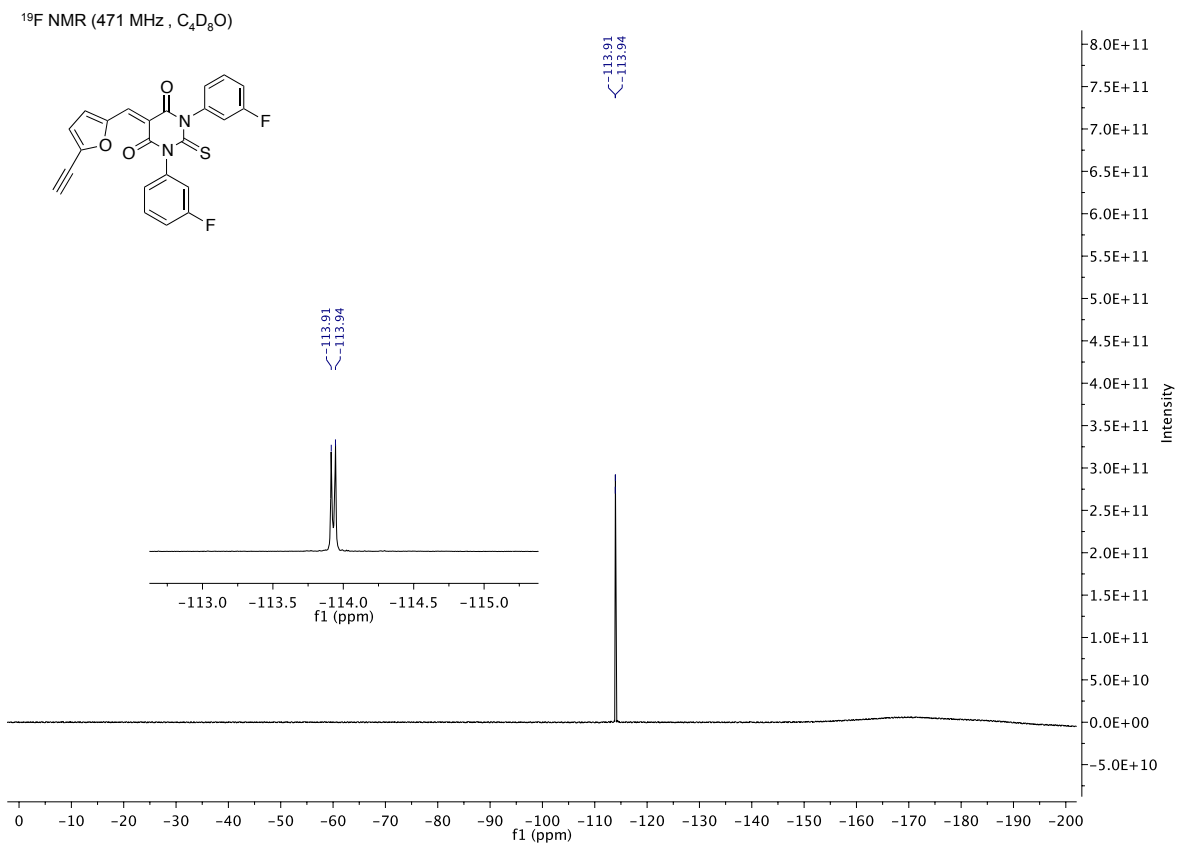
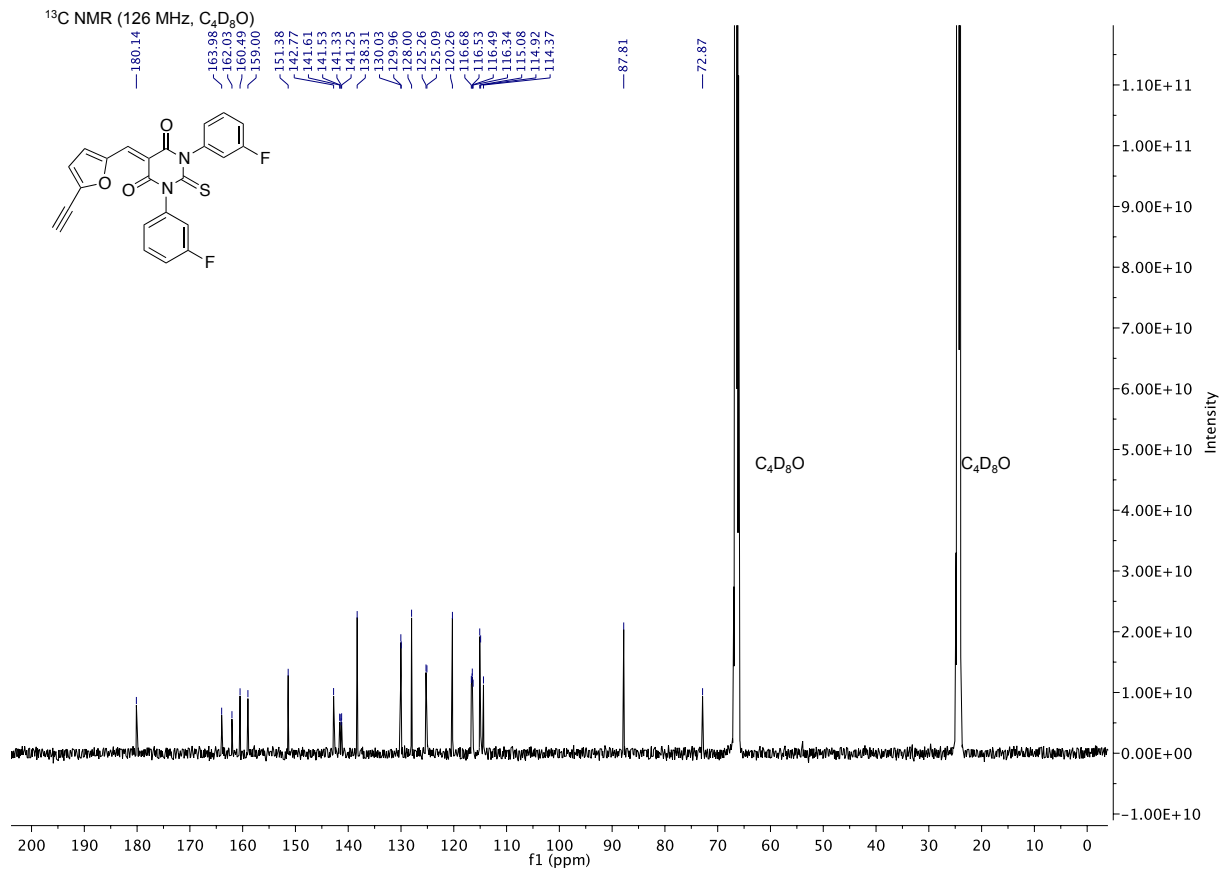
¹H NMR (500 MHz, C₄D₈O)





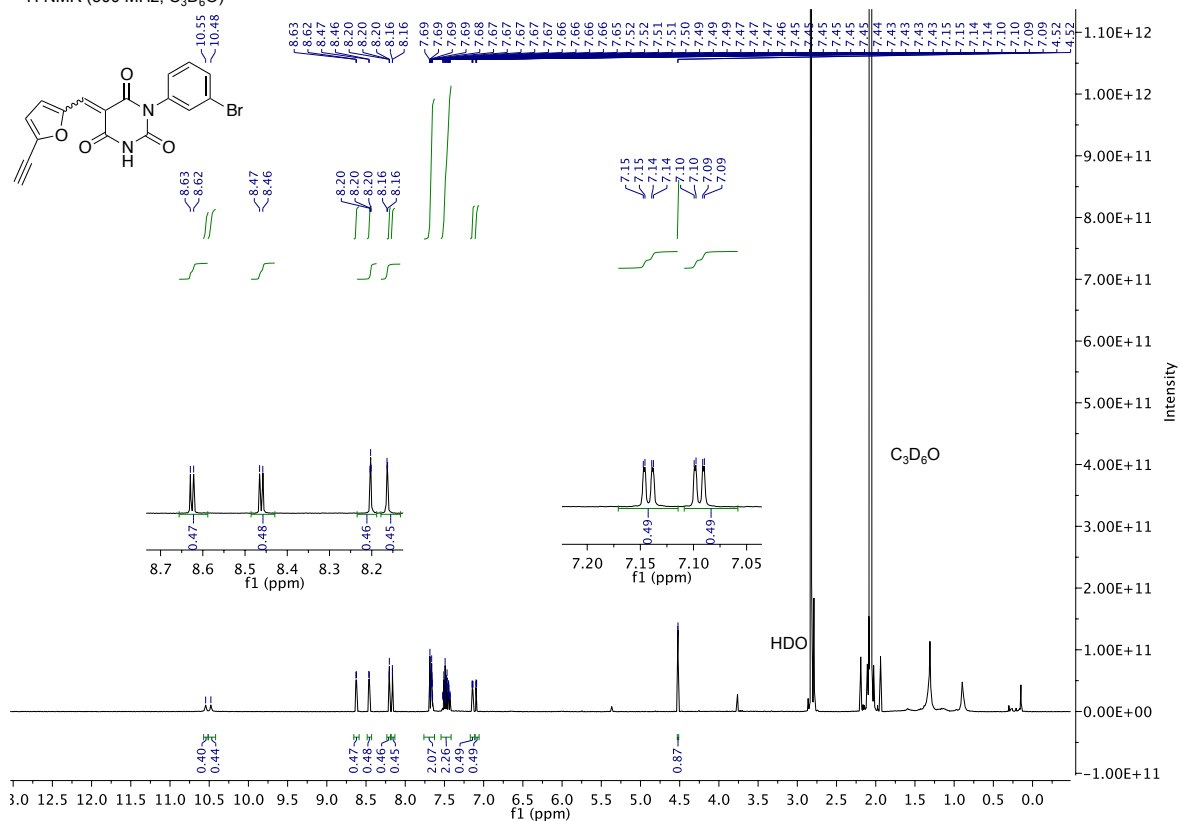
Compound 5k



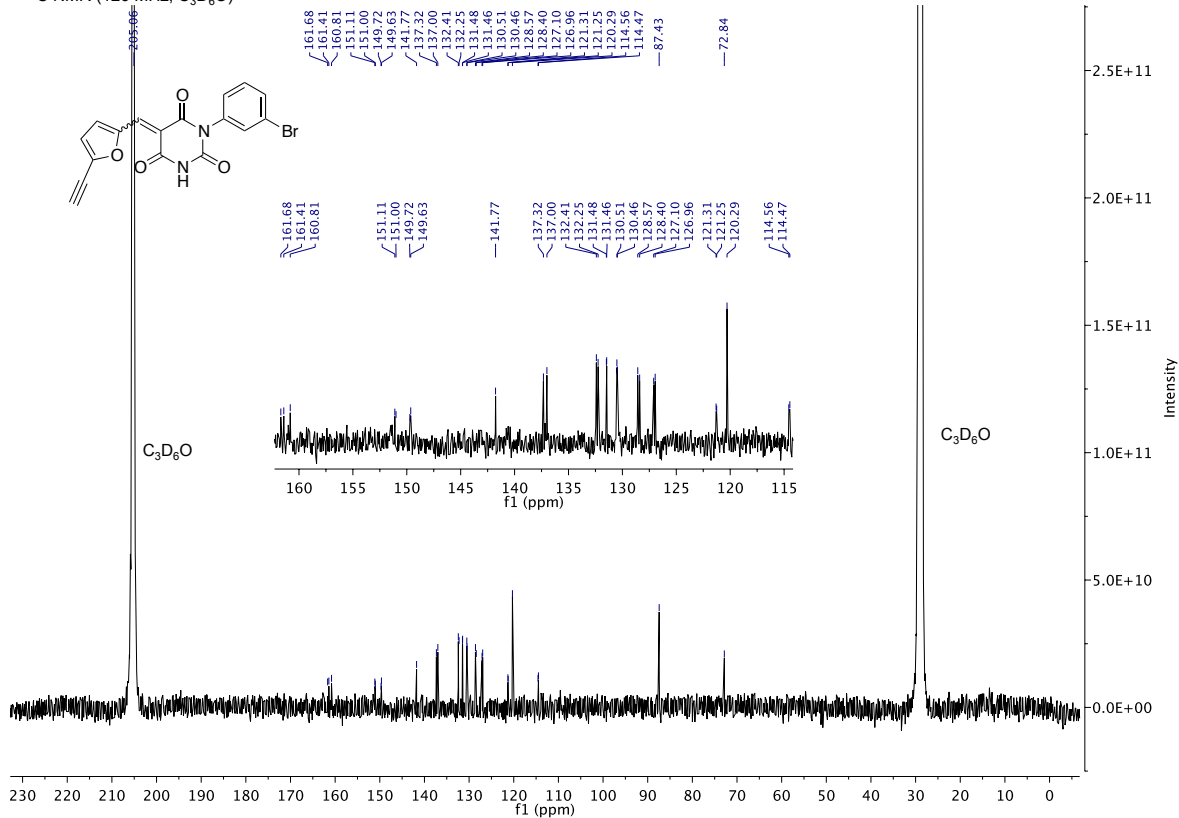


Compound 51

¹H NMR (500 MHz, C₃D₆O)

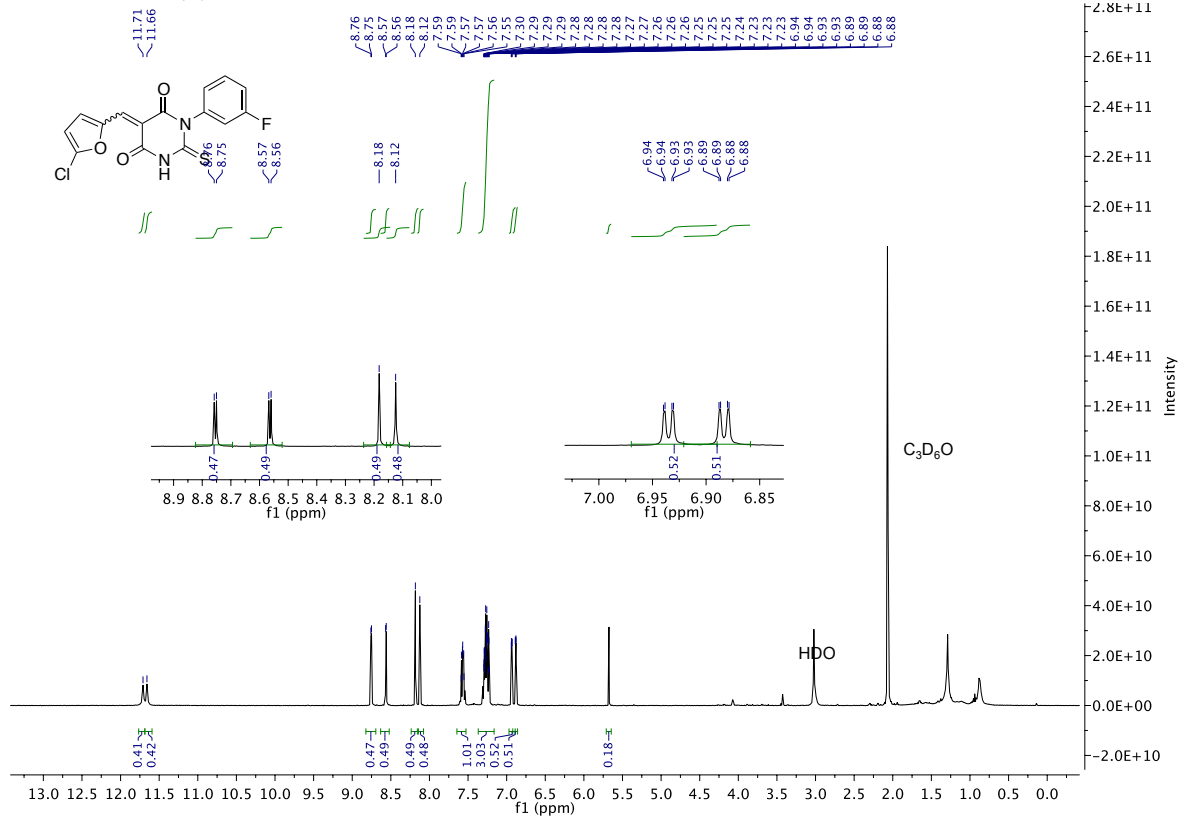


¹³C NMR (126 MHz, C₃D₆O)

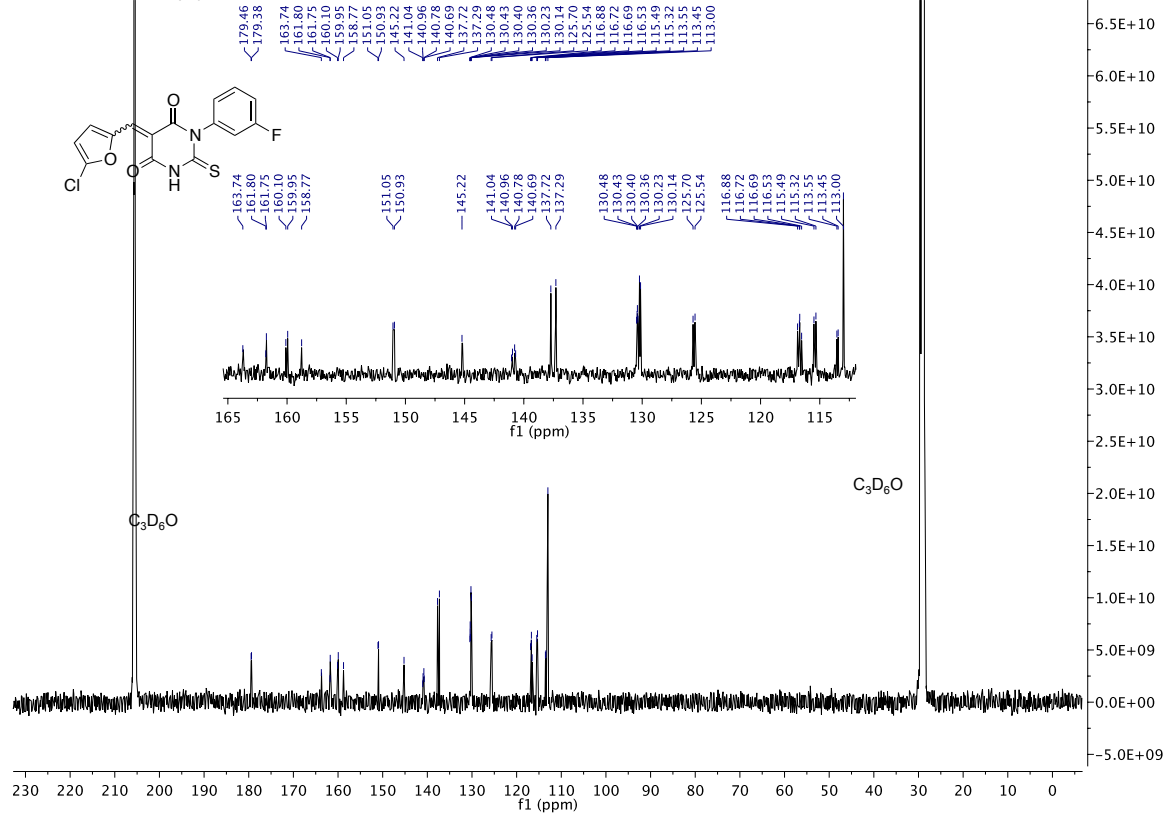


Compound 6b

¹H NMR (500 MHz, C₃D₆O)

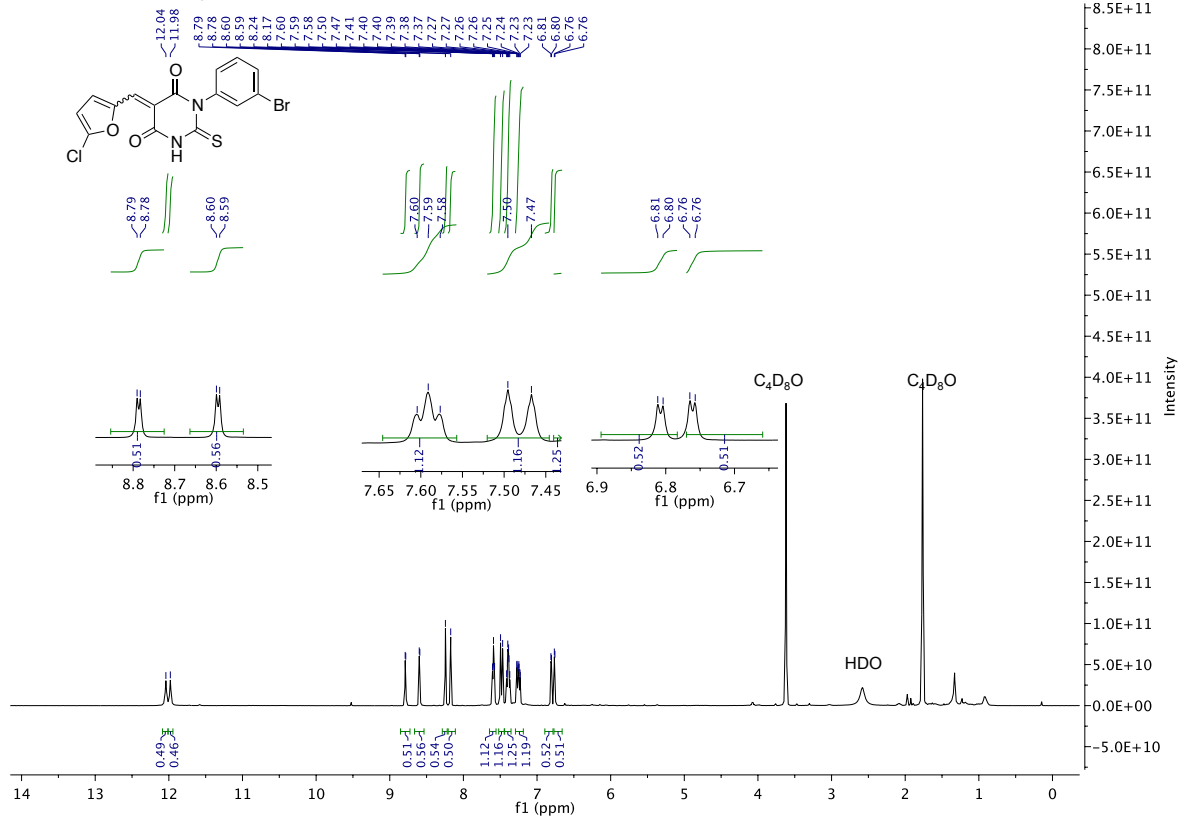


¹³C NMR (126 MHz, C₃D₆O)

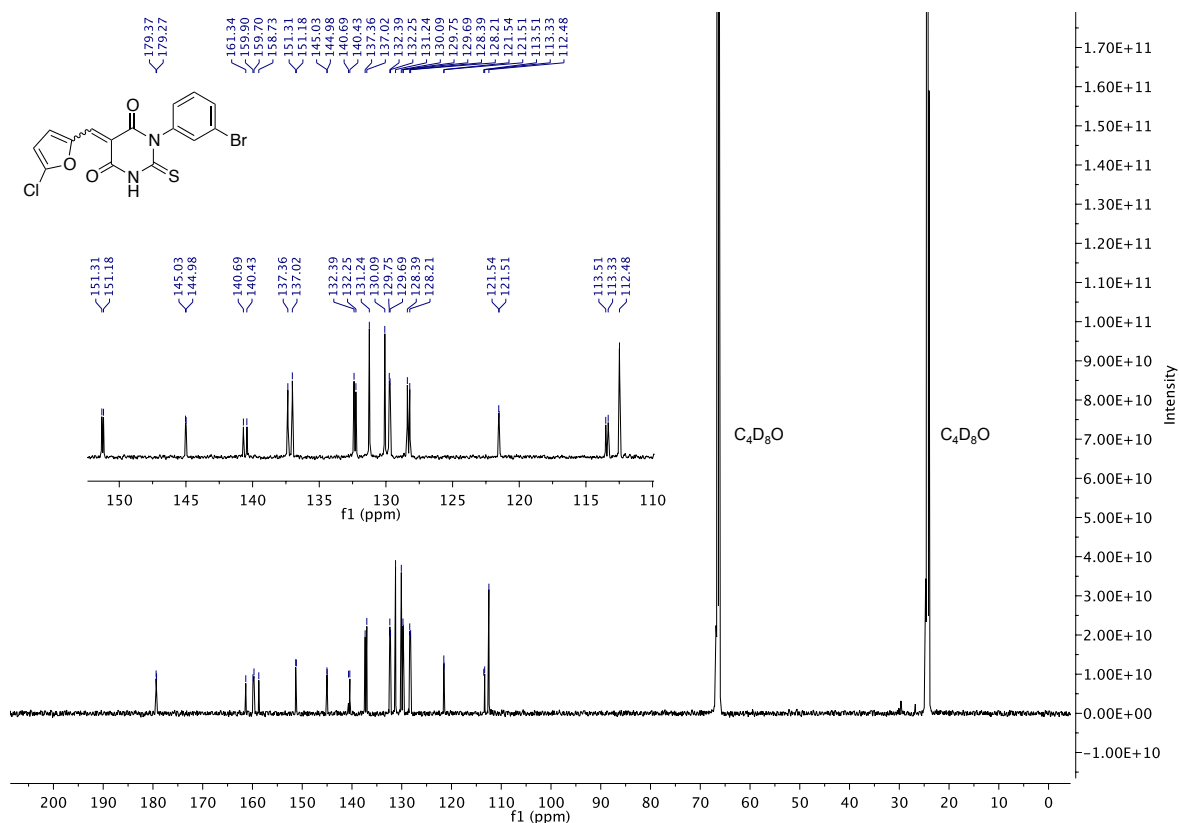


Compound 6c

¹H NMR (500 MHz, C₄D₈O)

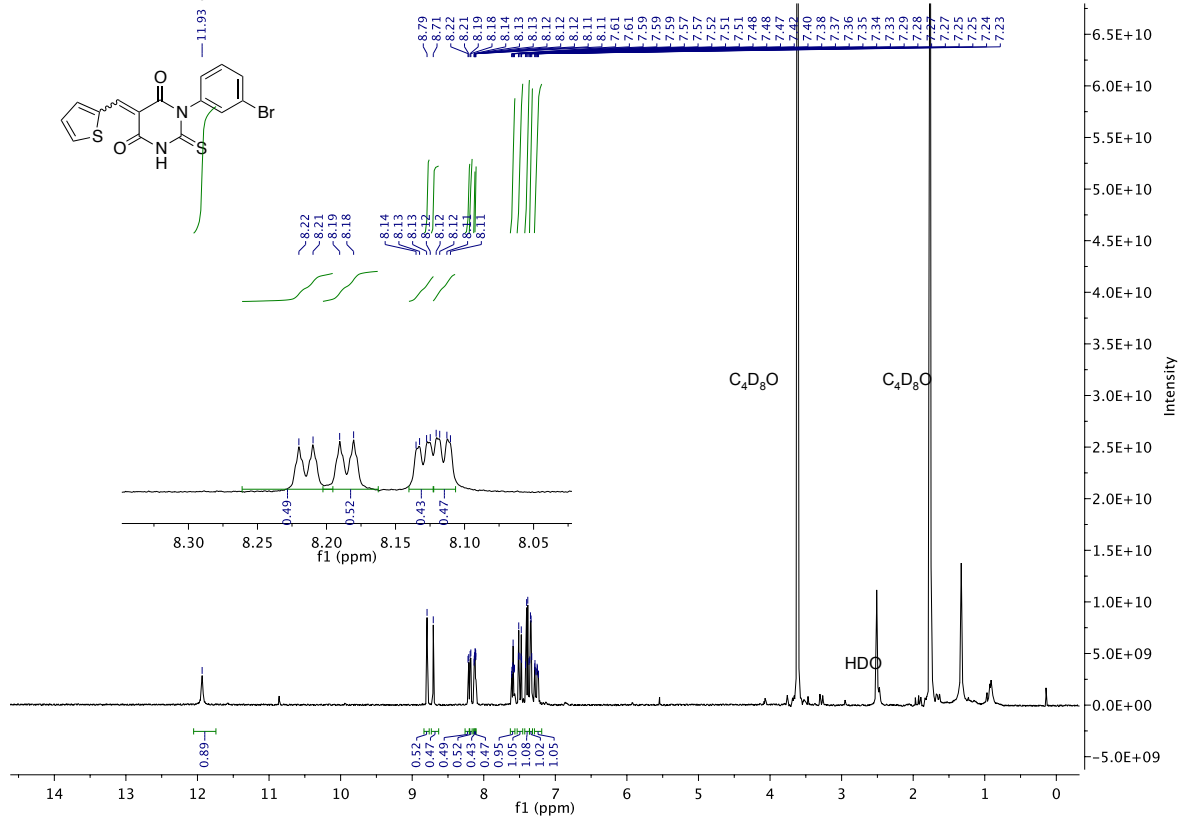


¹³C NMR (126 MHz, C₄D₈O)

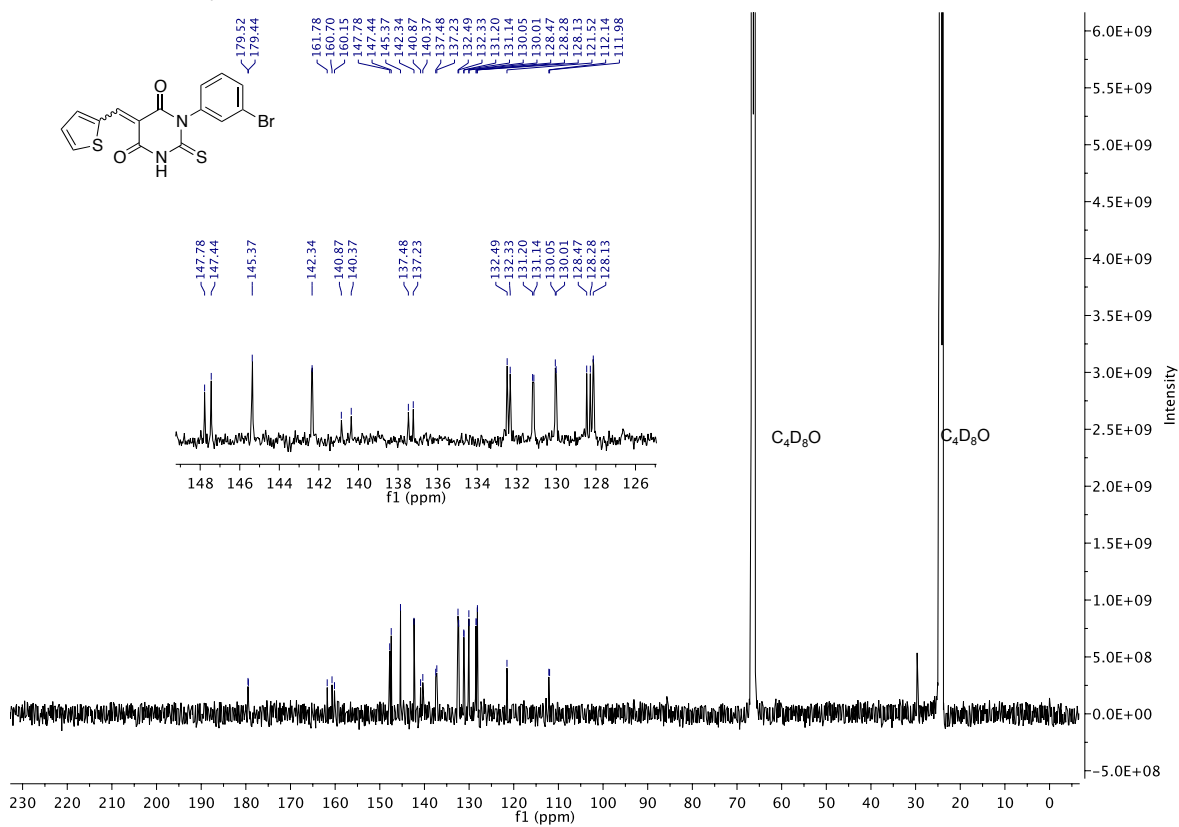


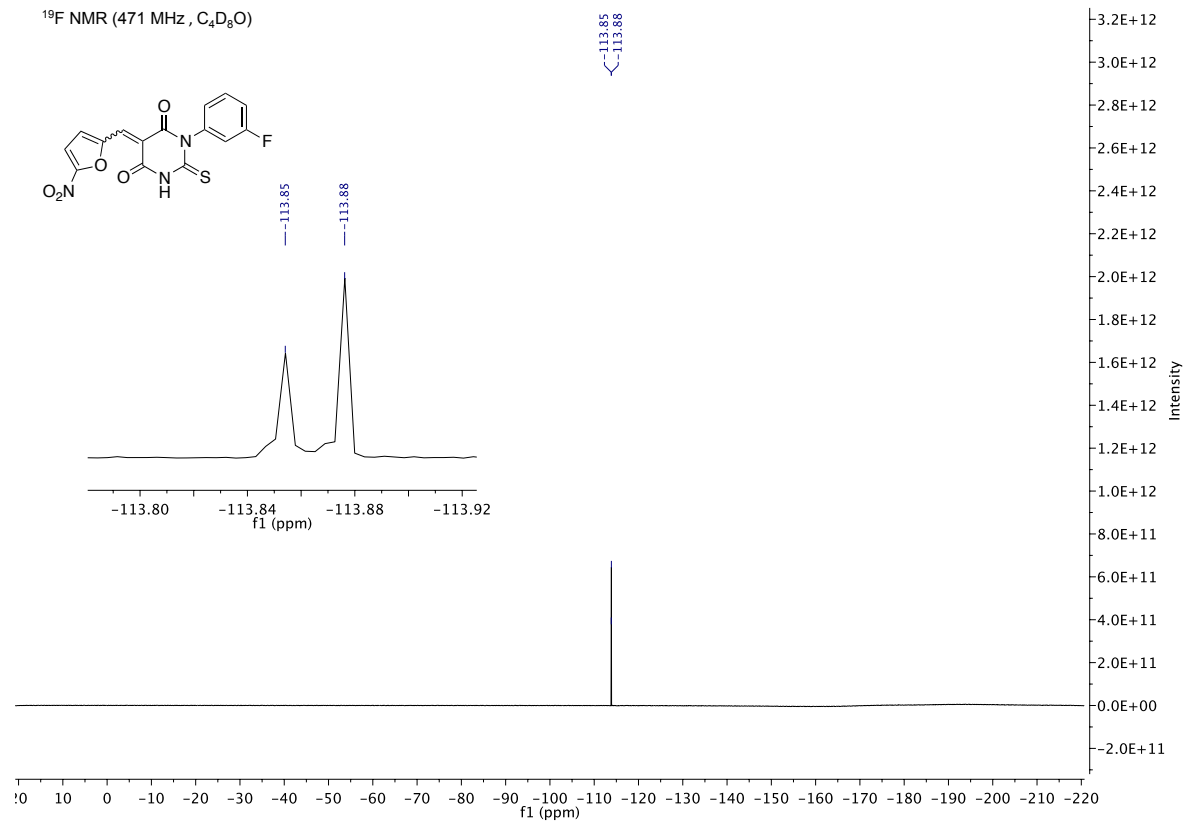
Compound 6d

¹H NMR (500 MHz, C₄D₈O)

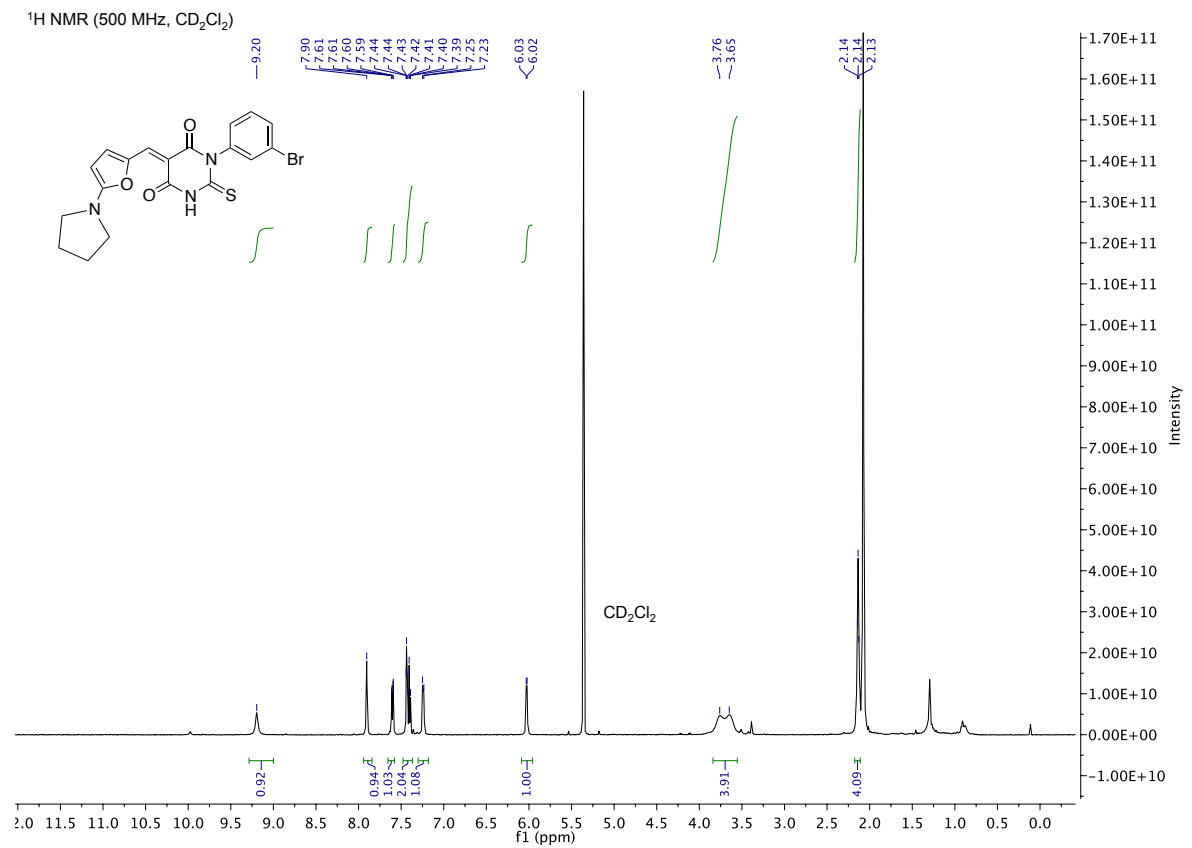


¹³C NMR (126 MHz, C₄D₈O)

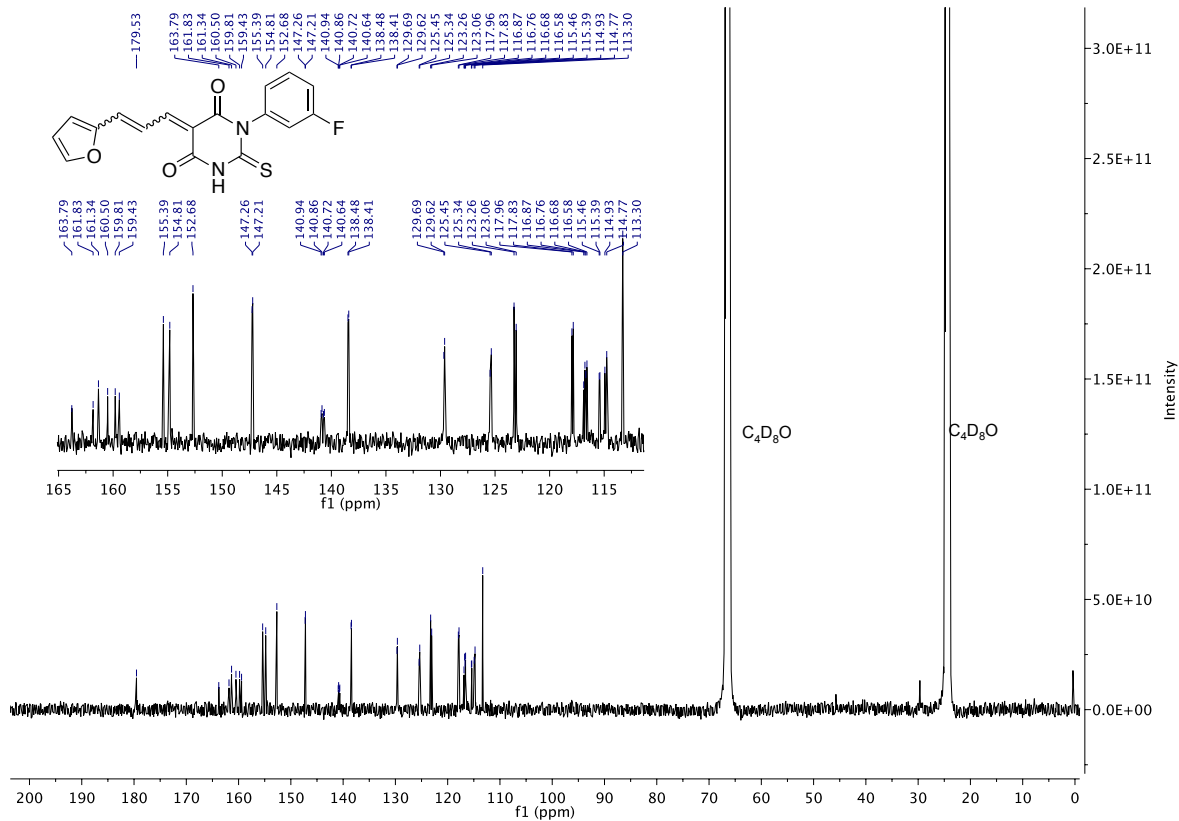




Compound 6f

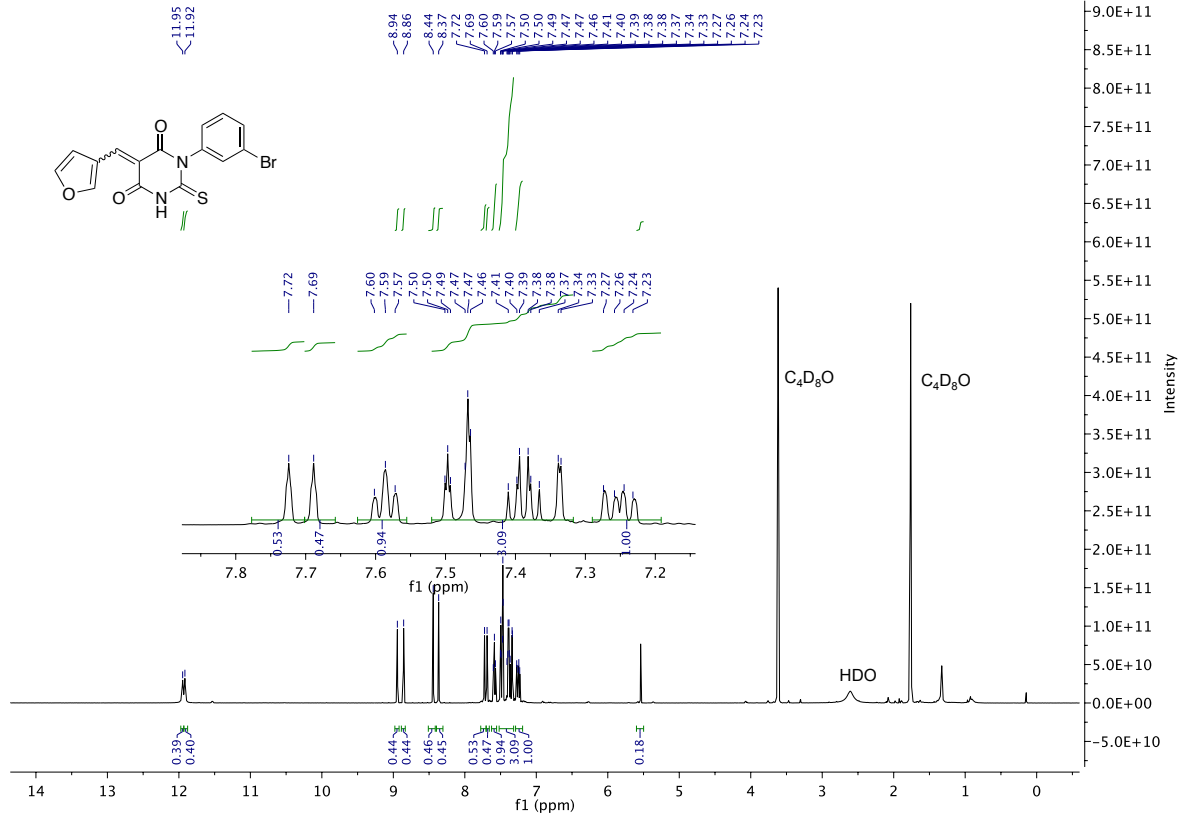


¹³C NMR (126 MHz, C₄D₈O)

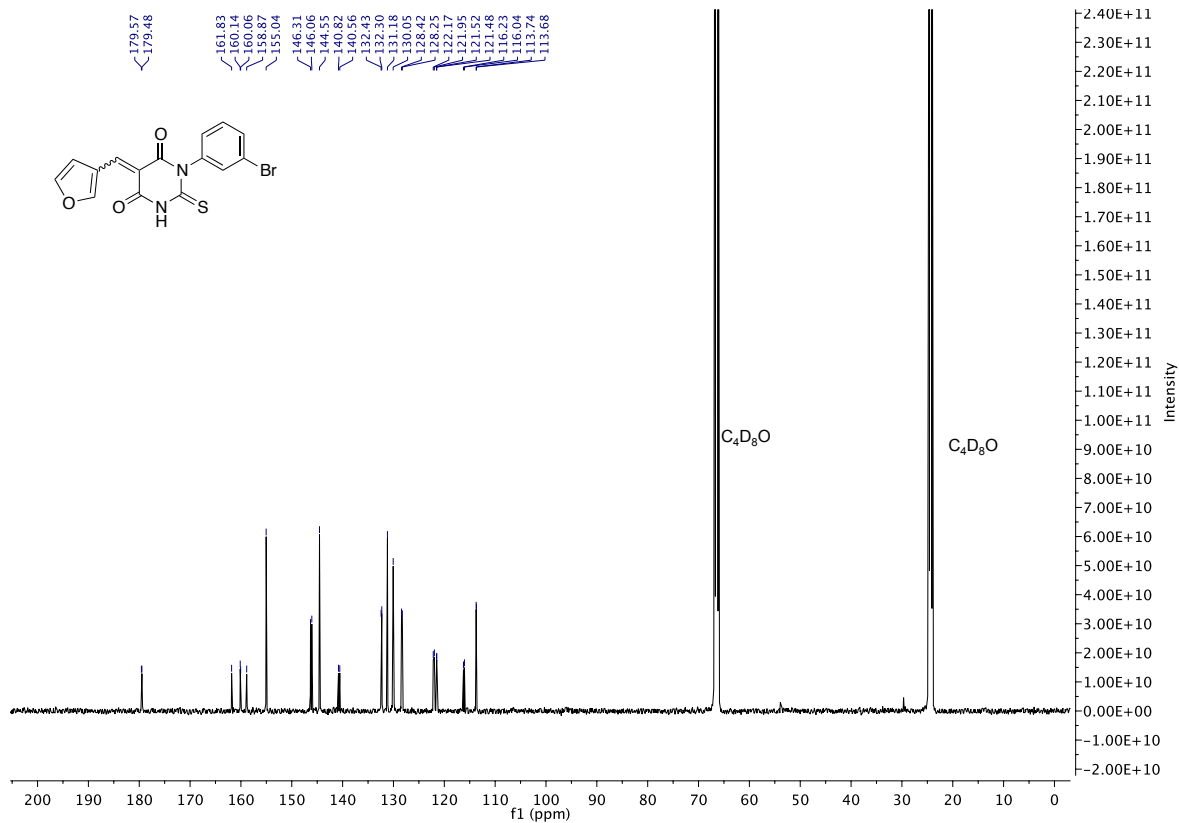


Compound 6h

¹H NMR (500 MHz, C₄D₈O)

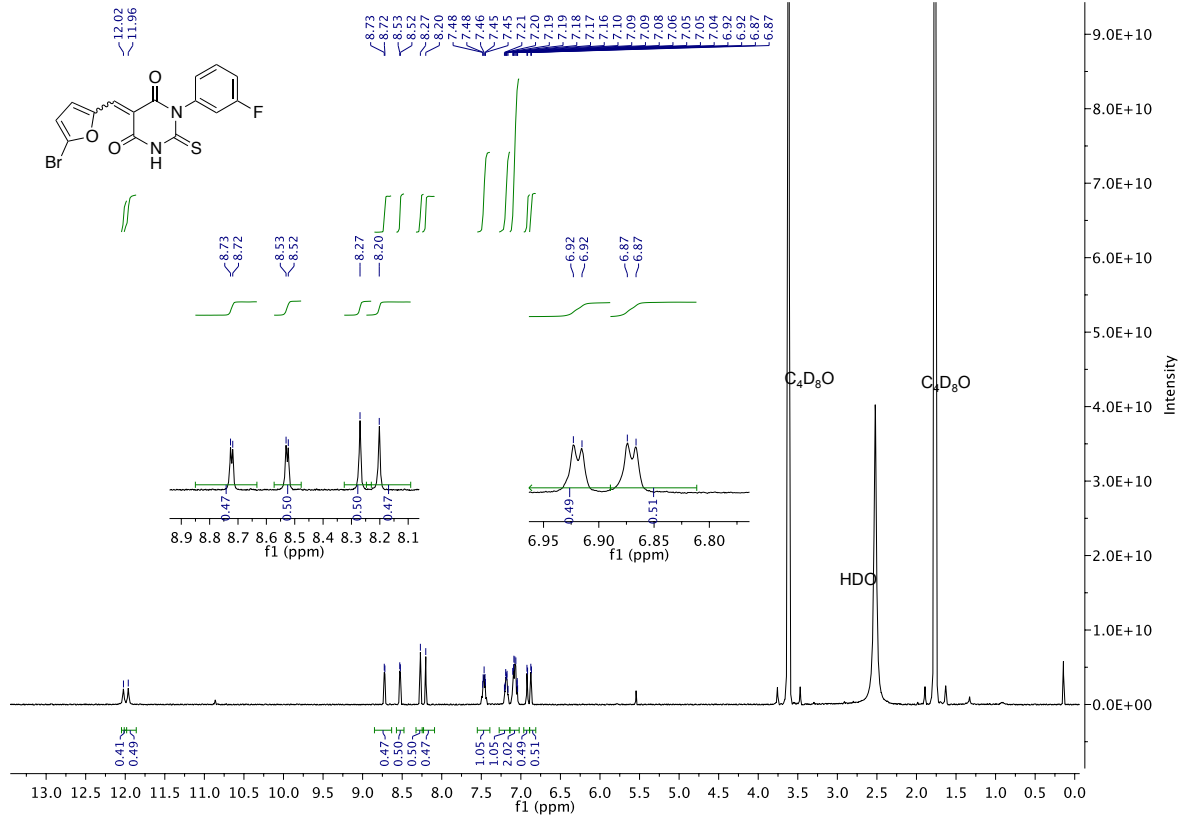


¹³C NMR (126 MHz, C₄D₈O)

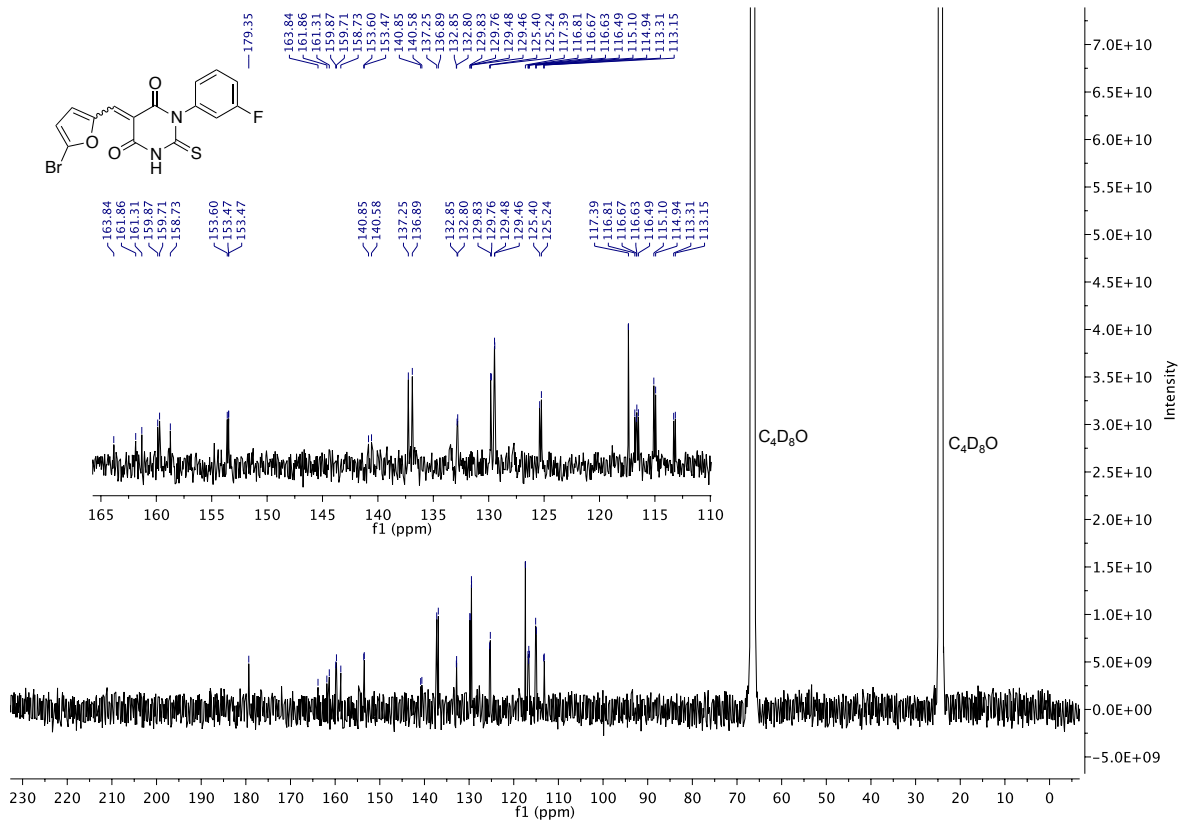


Compound 6i

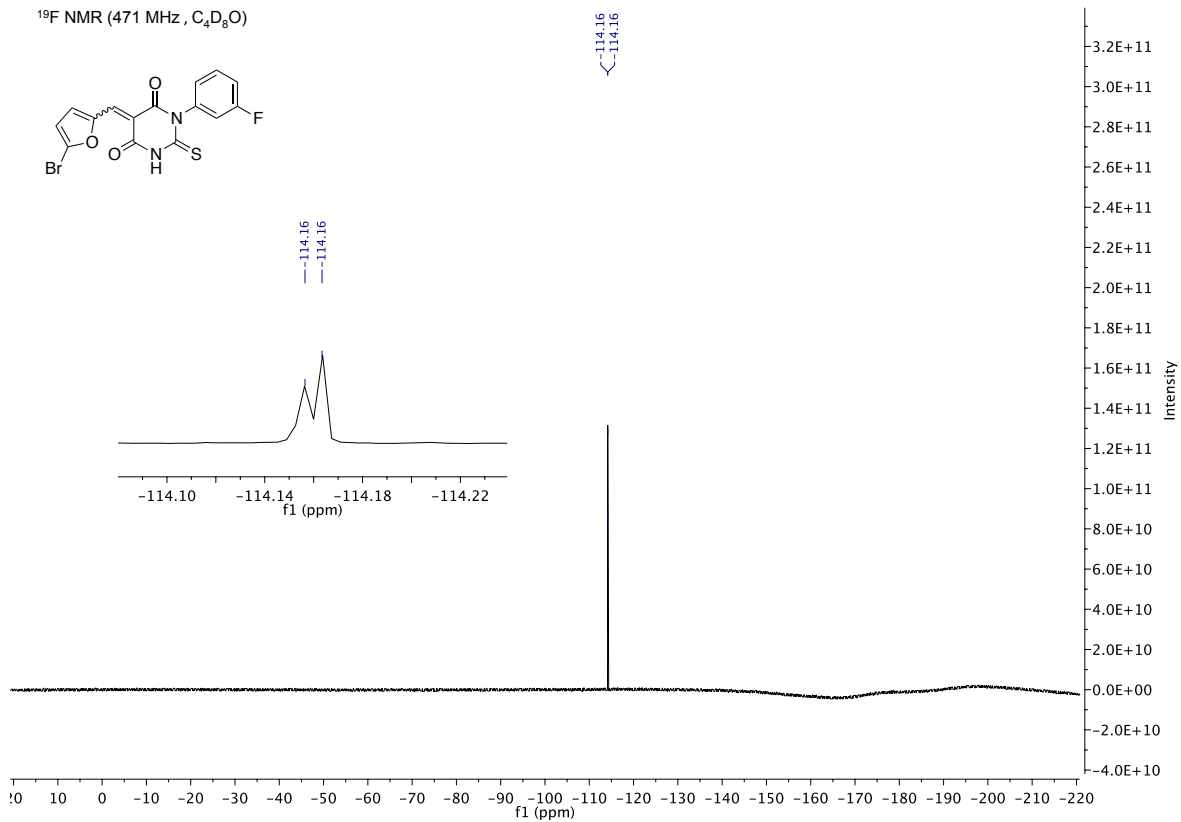
¹H NMR (500 MHz, C₄D₈O)



¹³C NMR (126 MHz, C₄D₈O)

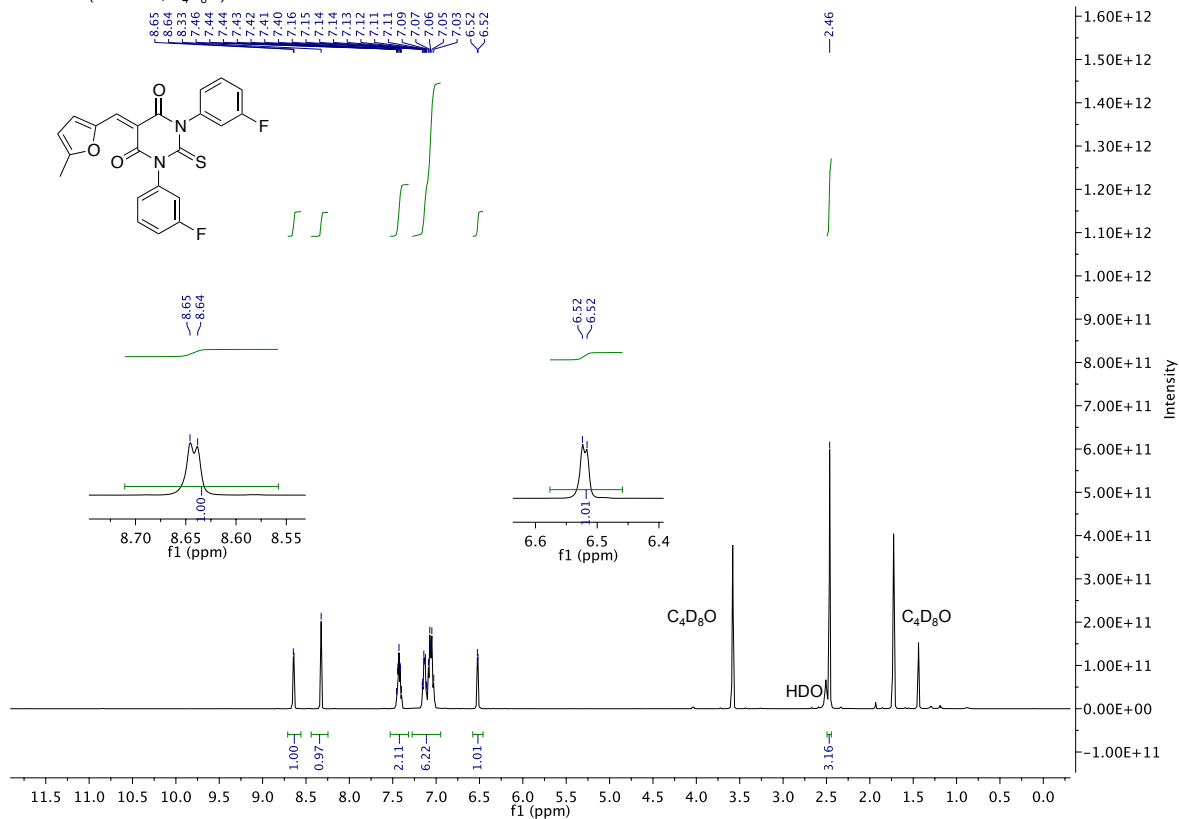


¹⁹F NMR (471 MHz, C₄D₈O)

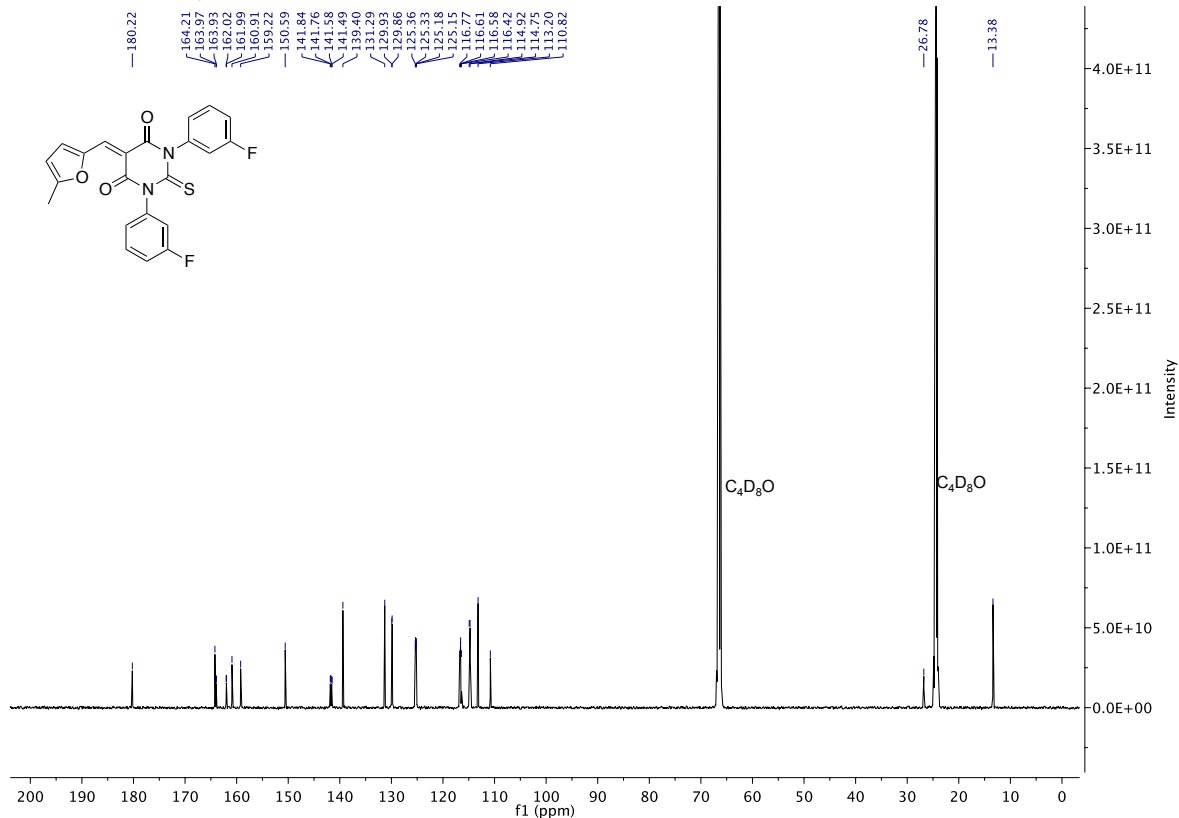


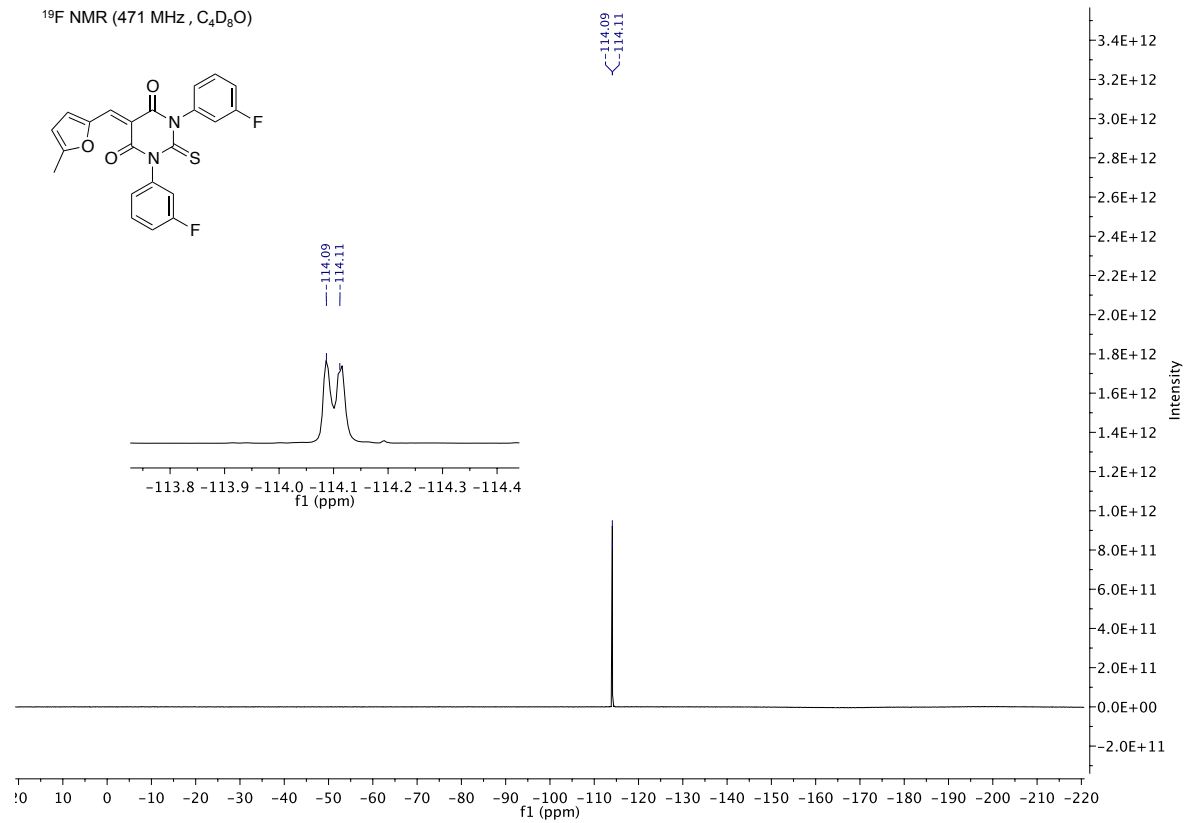
Compound 6j

¹H NMR (500 MHz, C₄D₈O)

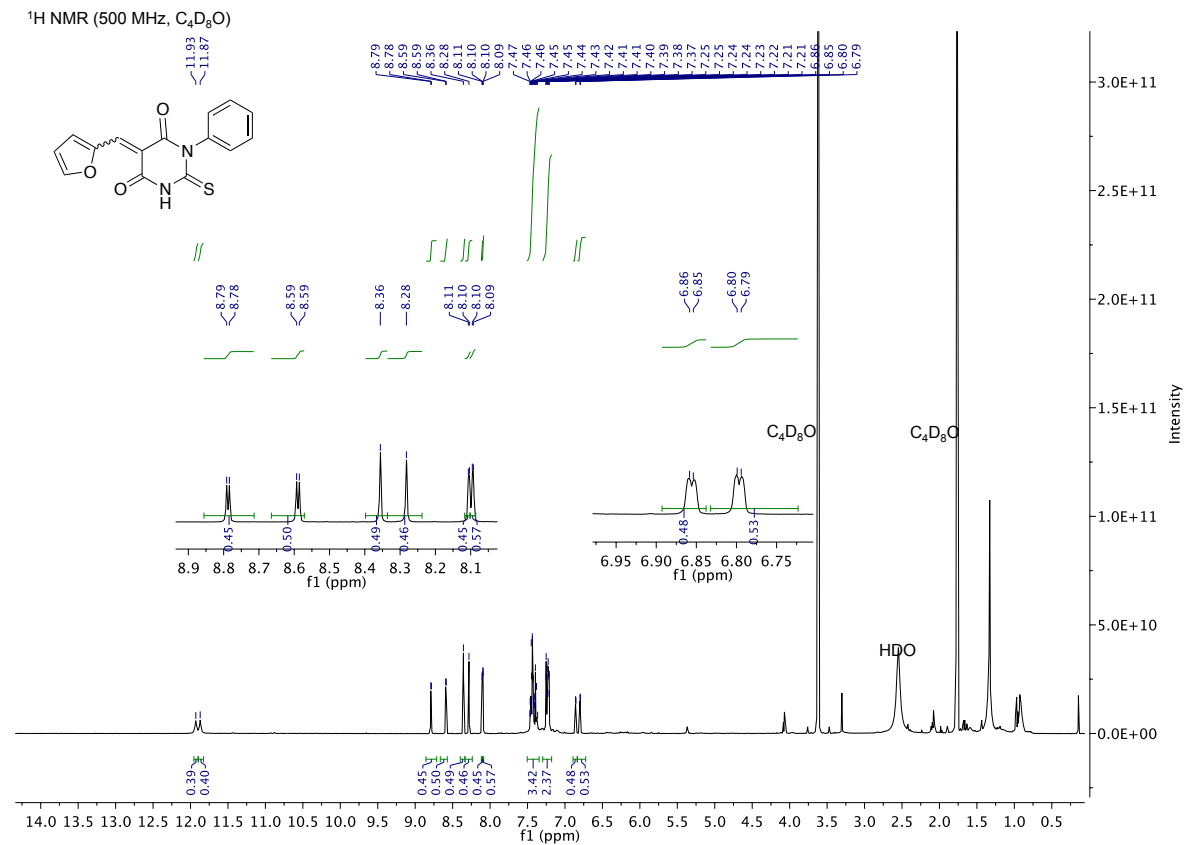


¹³C NMR (126 MHz, C₄D₈O)

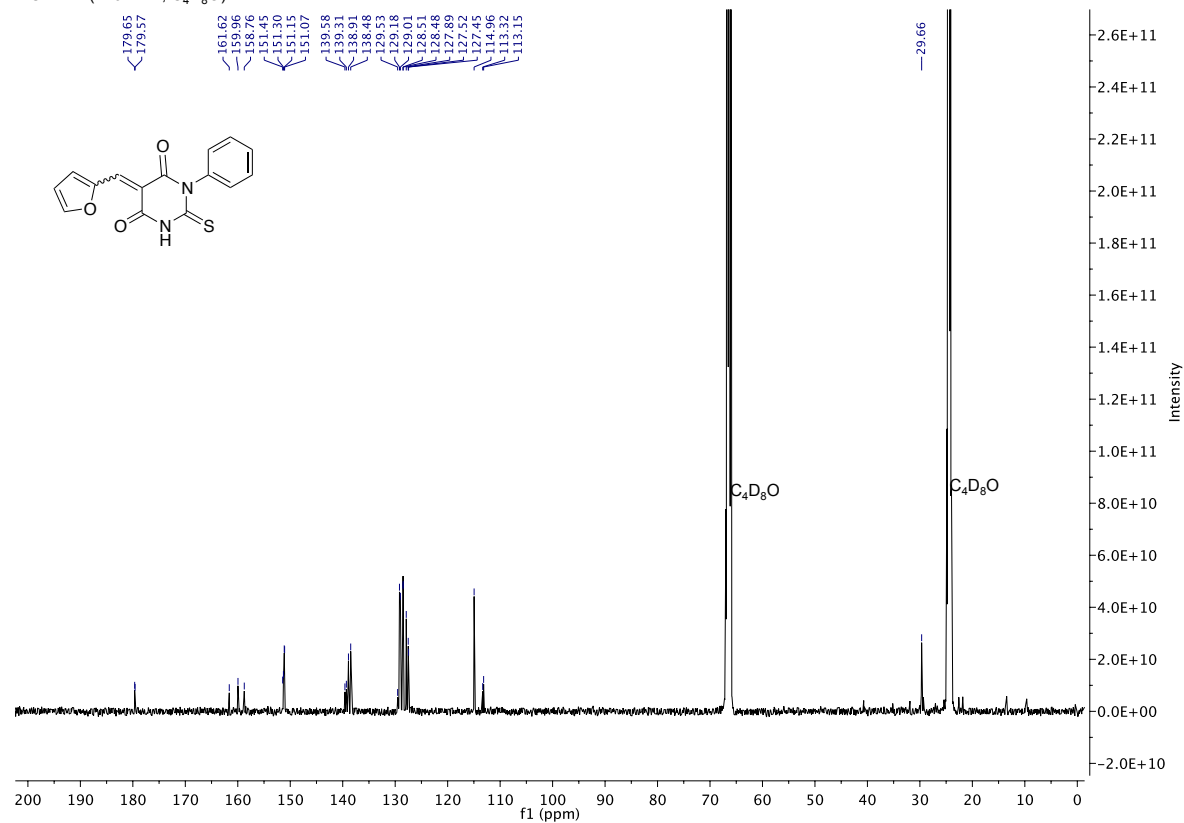




Compound 6k

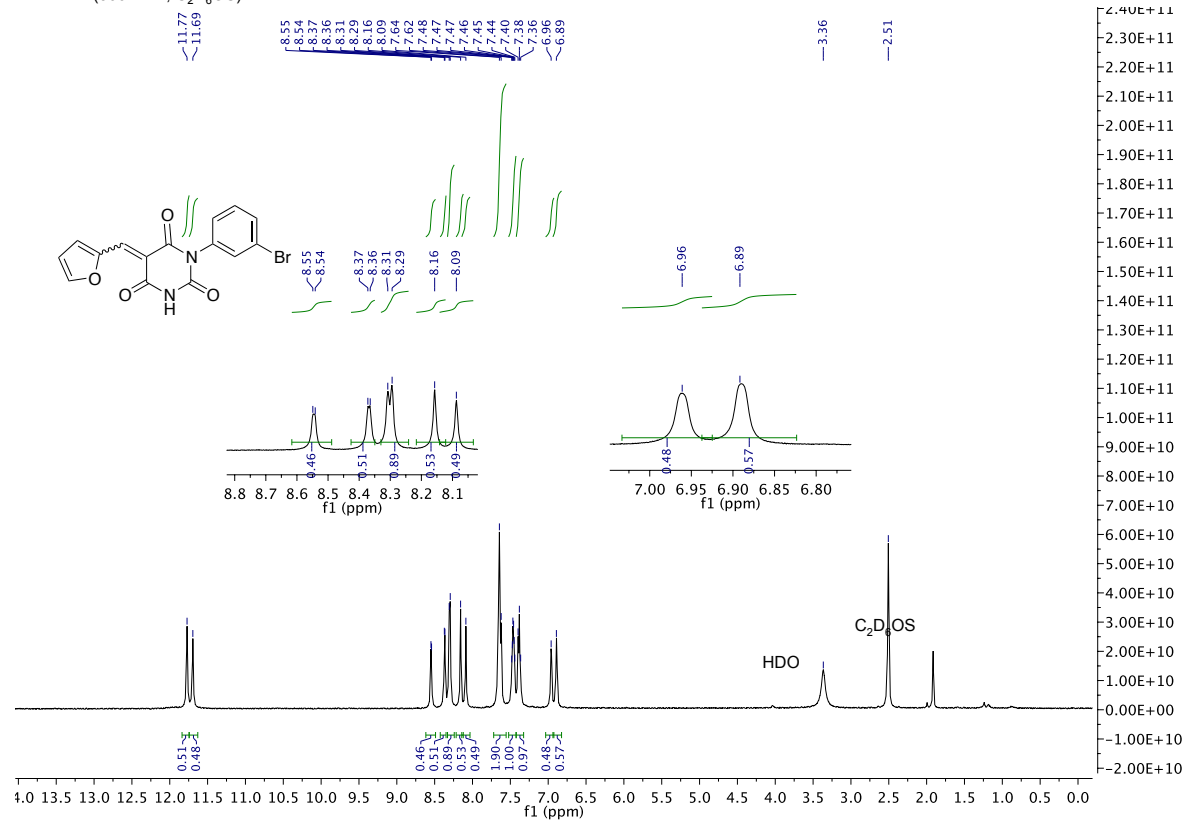


^{13}C NMR (126 MHz, $\text{C}_4\text{D}_8\text{O}$)

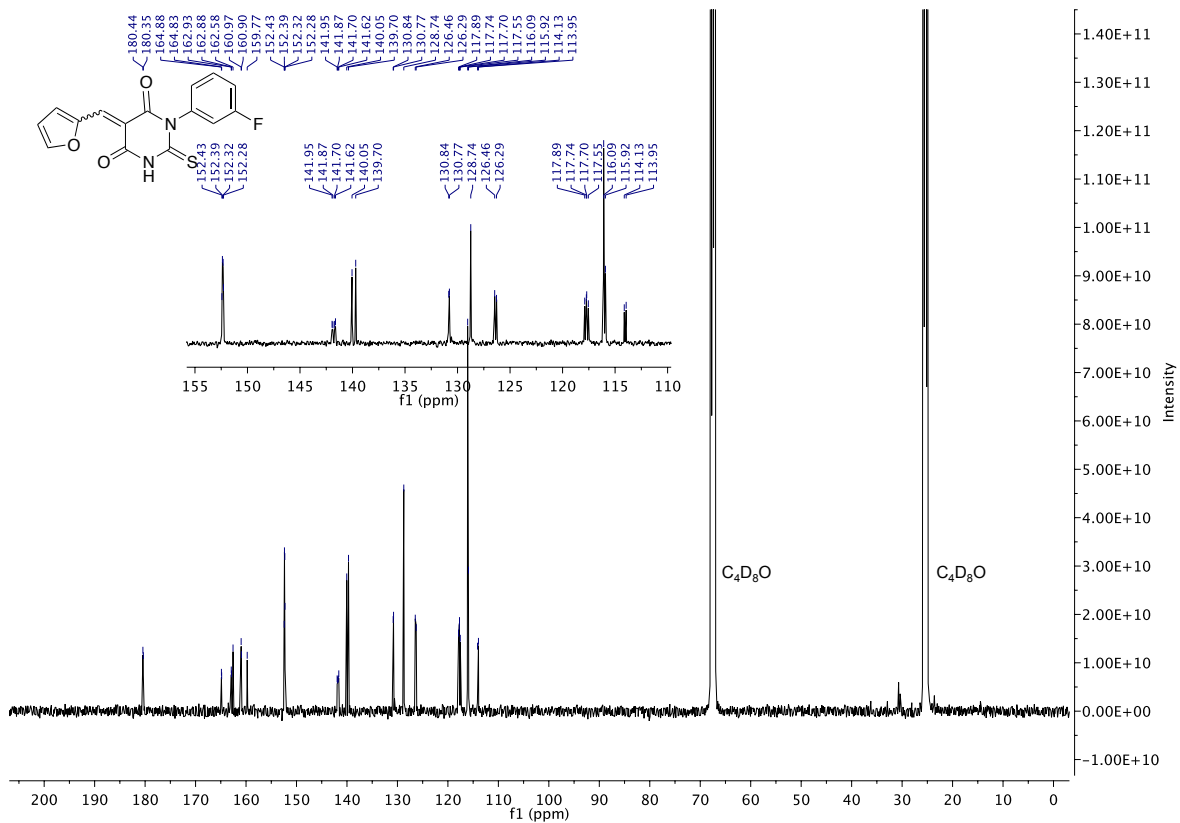


Compound 6I

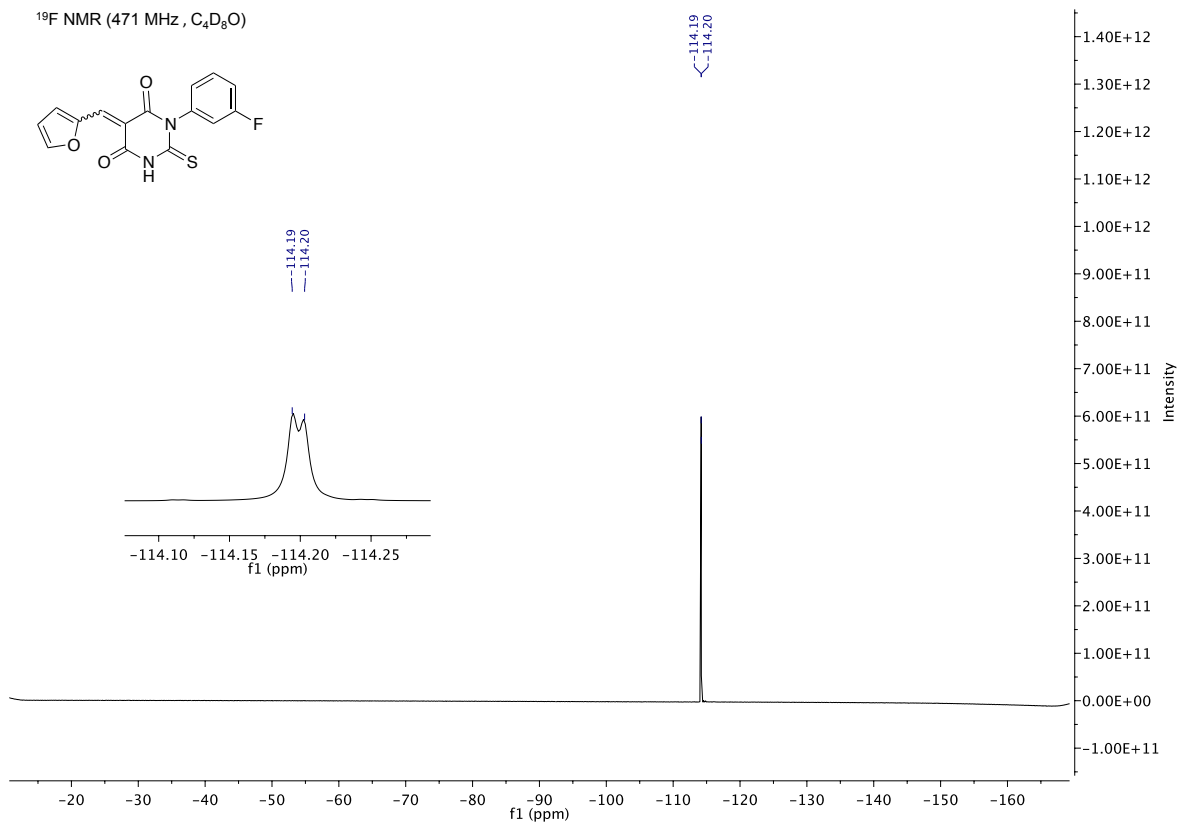
^1H NMR (500 MHz, $\text{C}_2\text{D}_6\text{OS}$)



¹³C NMR (126 MHz, C₄D₈O)

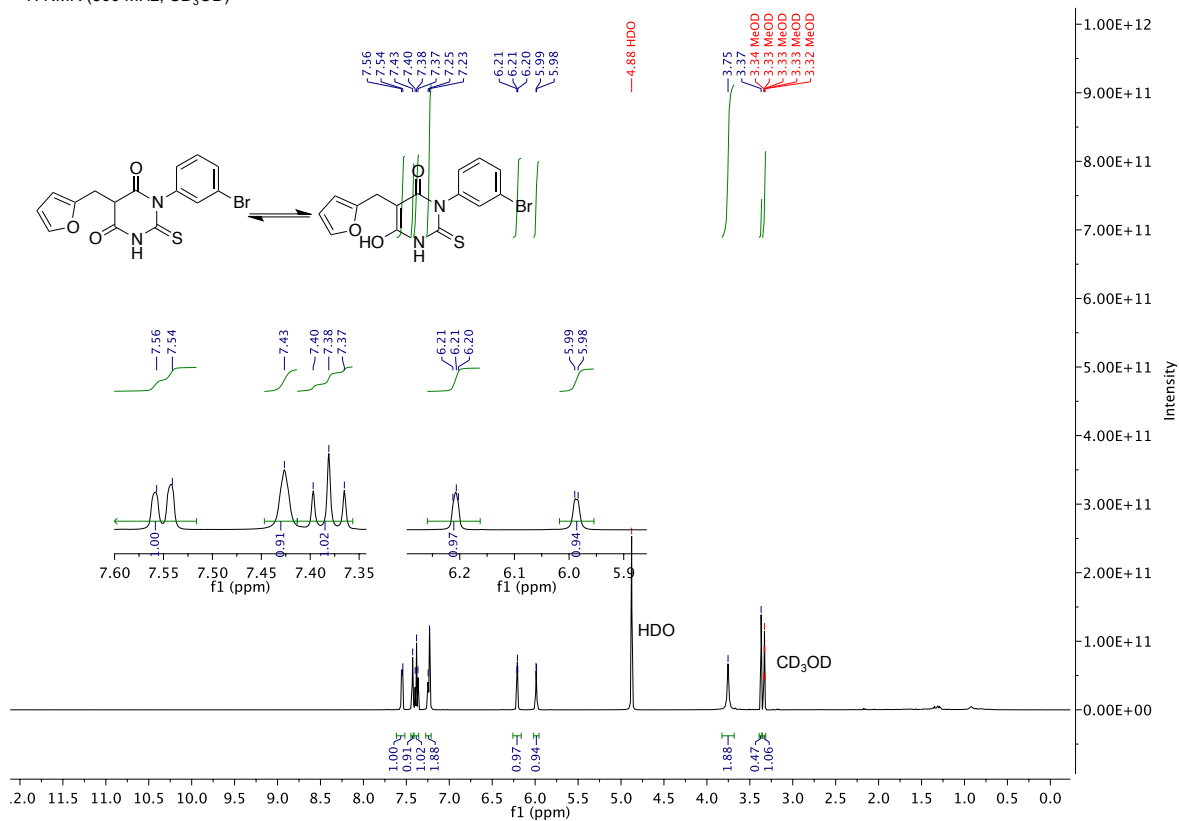


¹⁹F NMR (471 MHz, C₄D₈O)

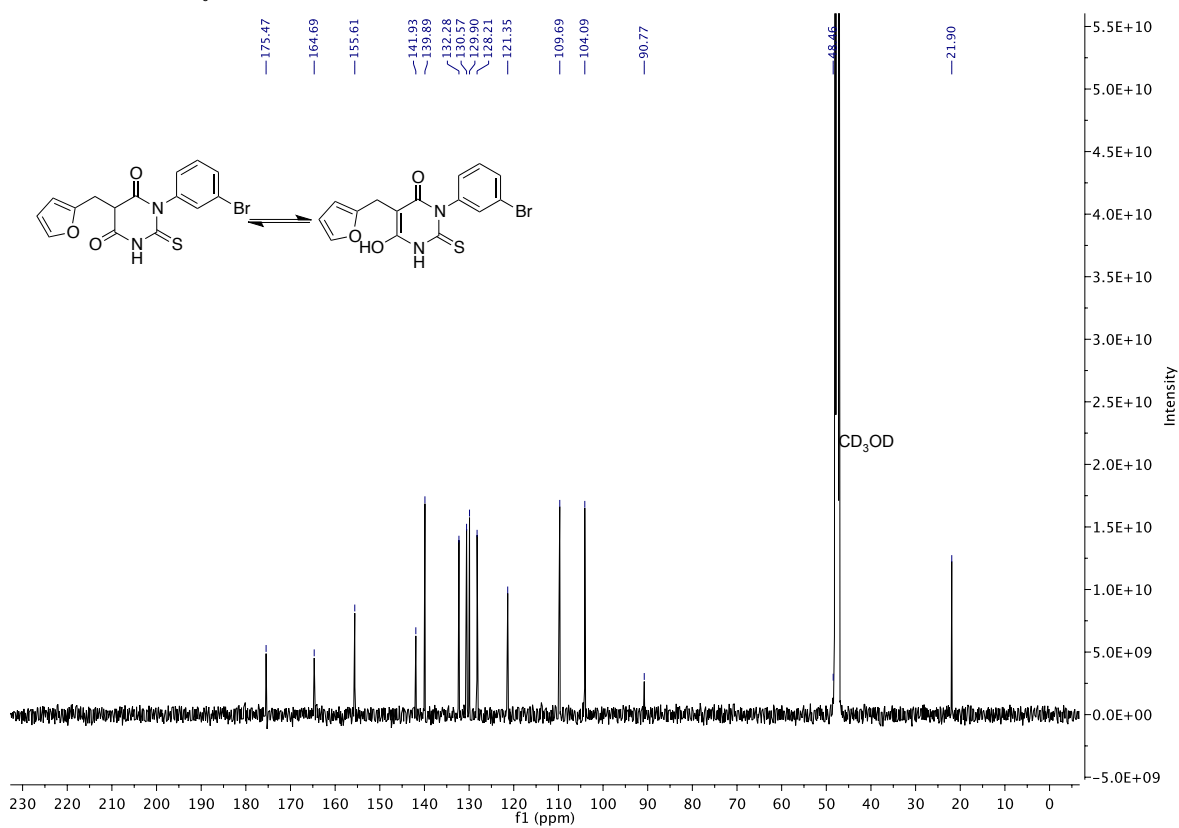


Compound 7a

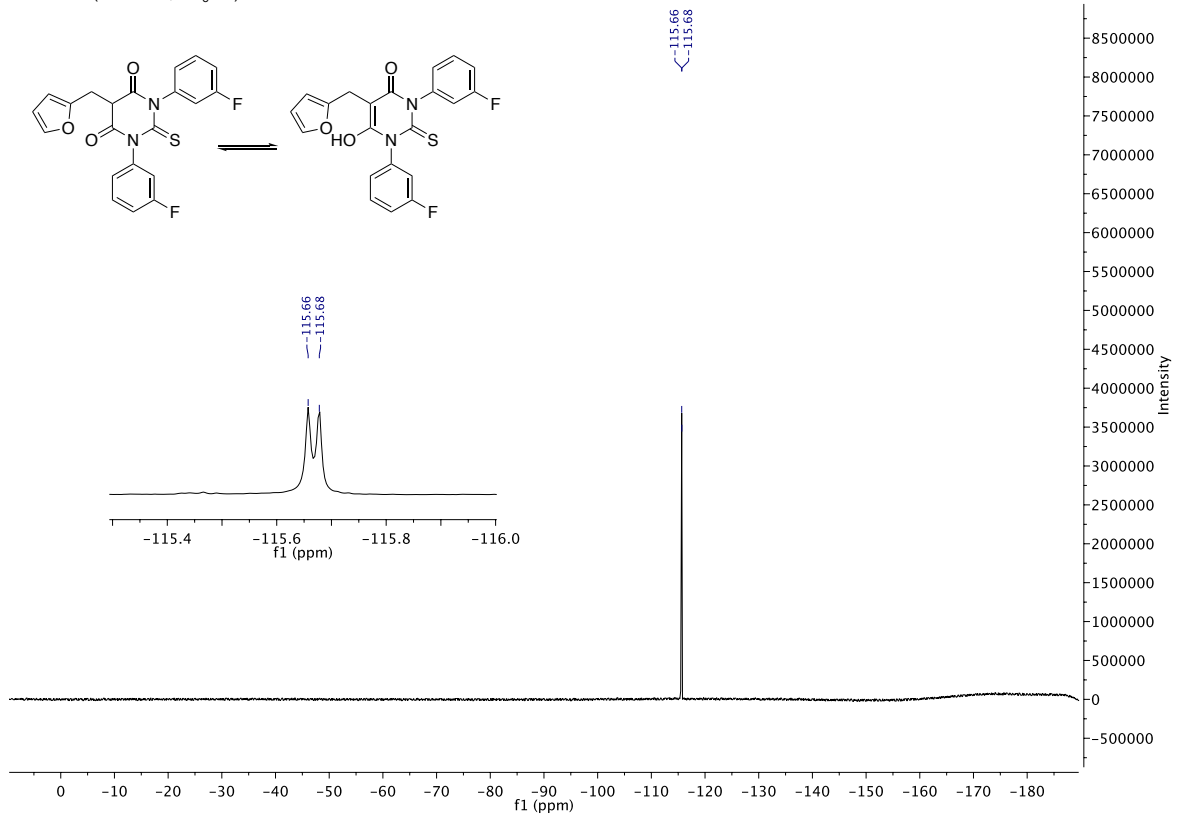
¹H NMR (500 MHz, CD₃OD)



¹³C NMR (126 MHz, CD₃OD)

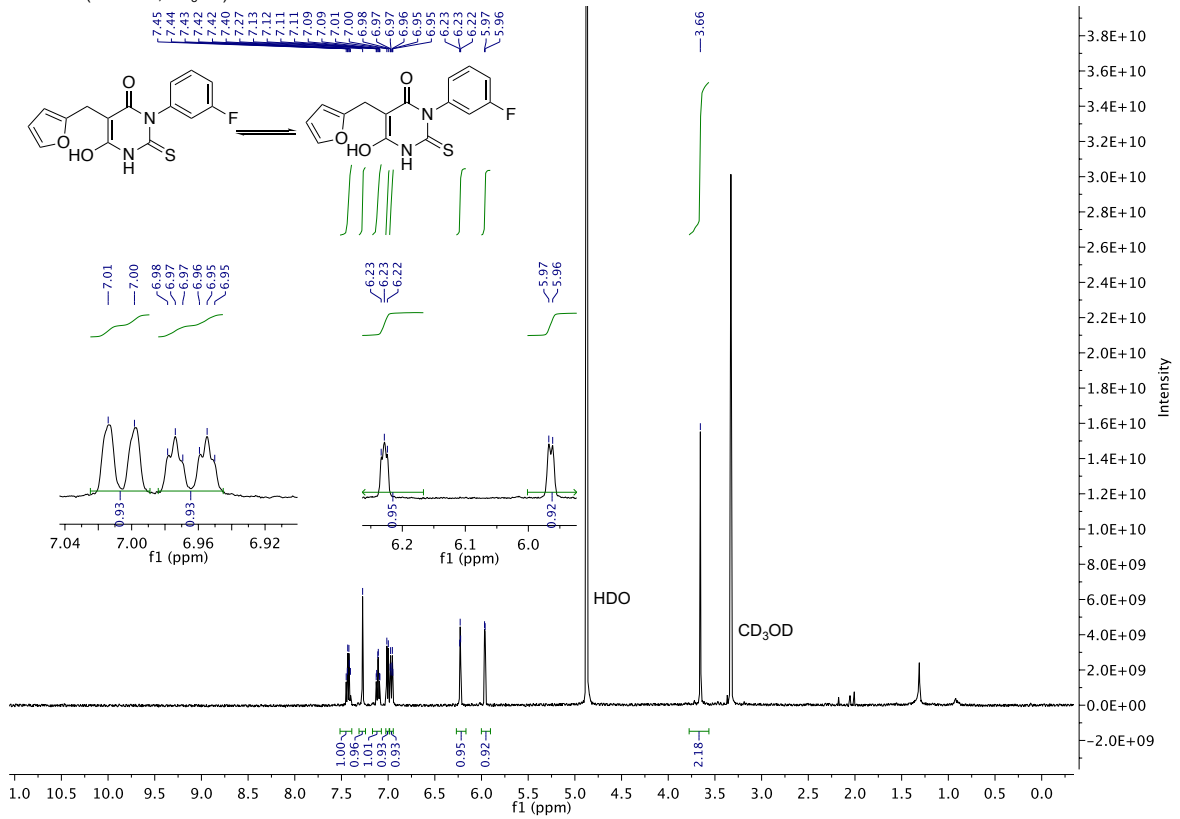


¹⁹F NMR (471 MHz, CD₃OD)

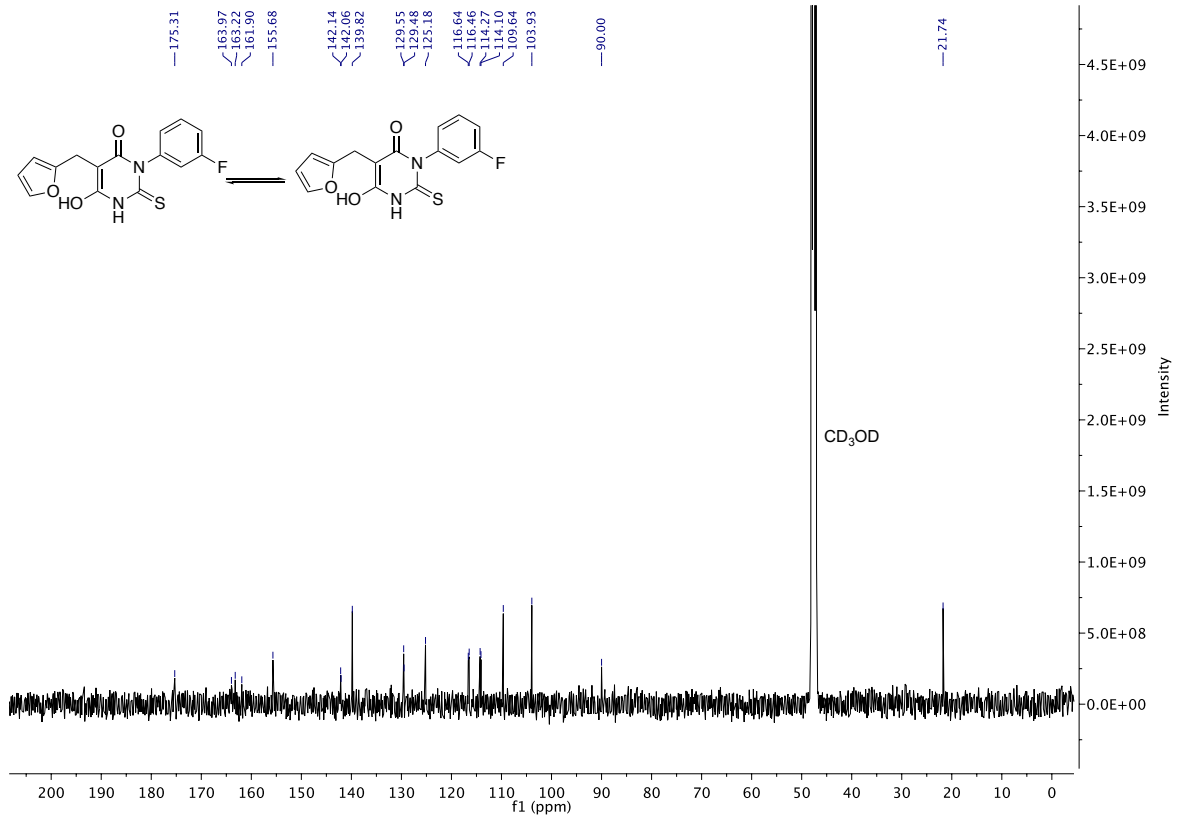


Compound 7c

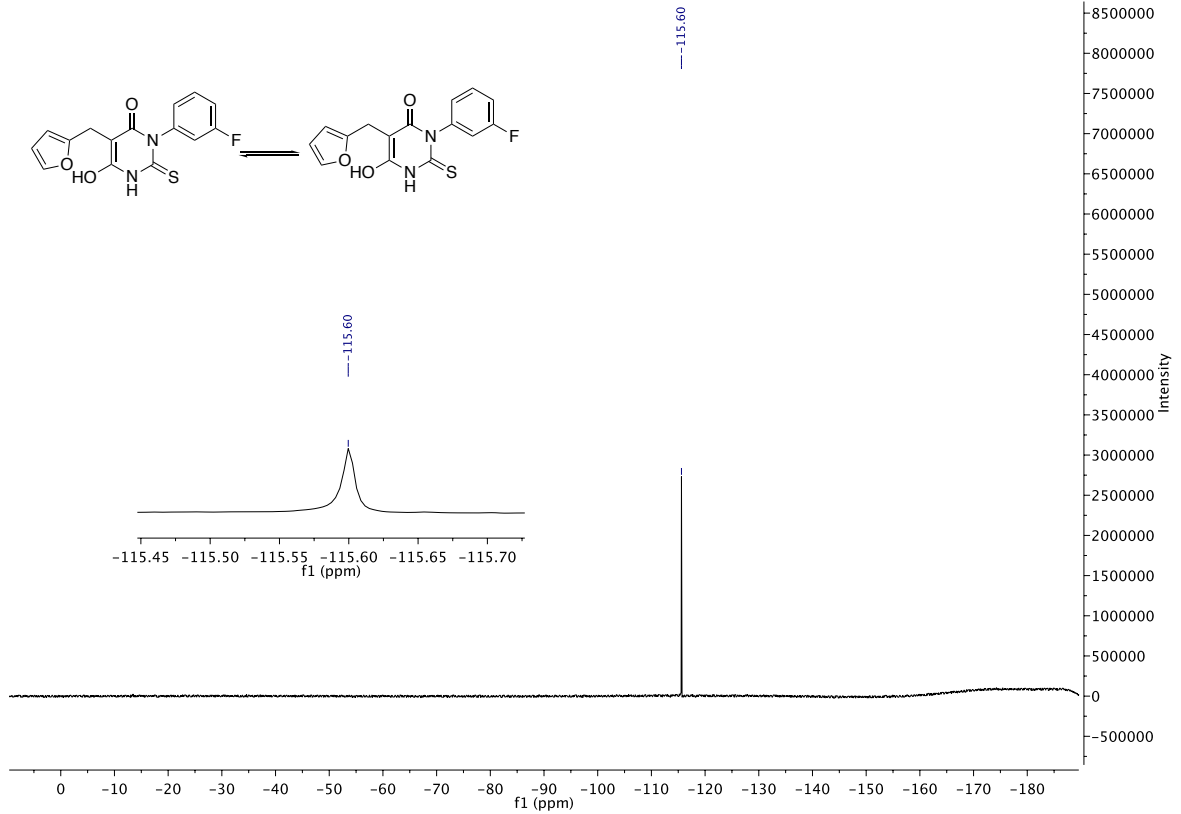
¹H NMR (500 MHz, CD₃OD)



¹³C NMR (126 MHz, CD₃OD)

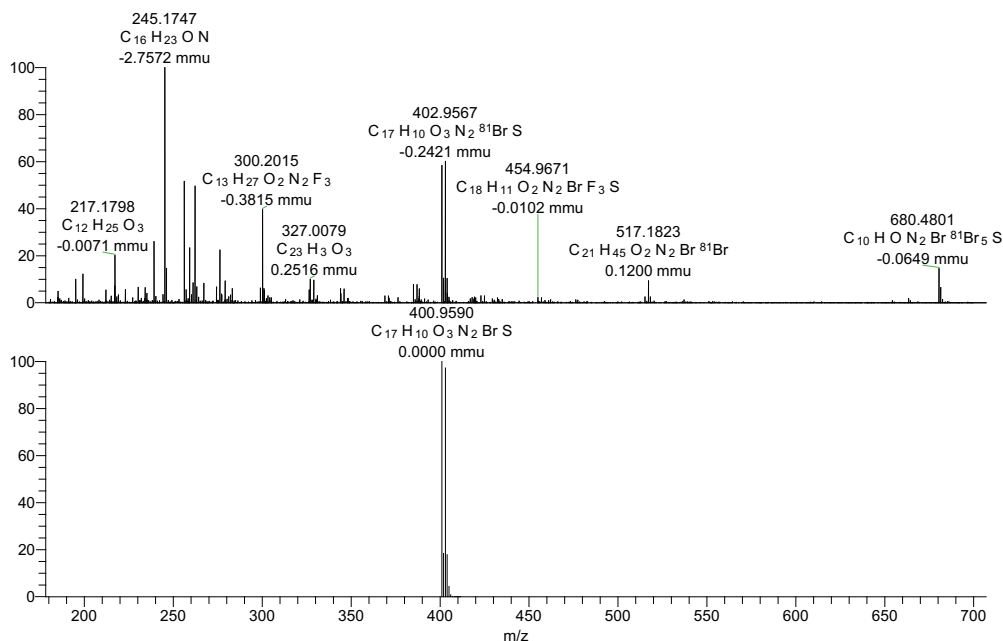


¹⁹F NMR (471 MHz, CD₃OD)



HRMS spectra and UPLC-MS chromatogram of final compounds

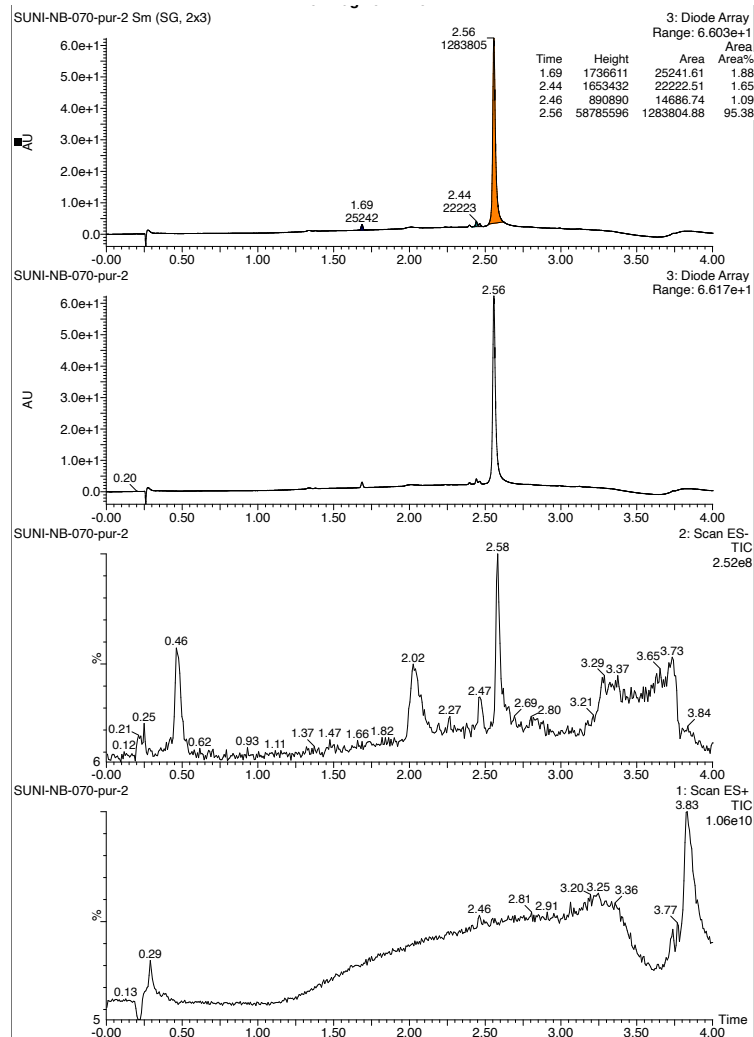
Compound 5a



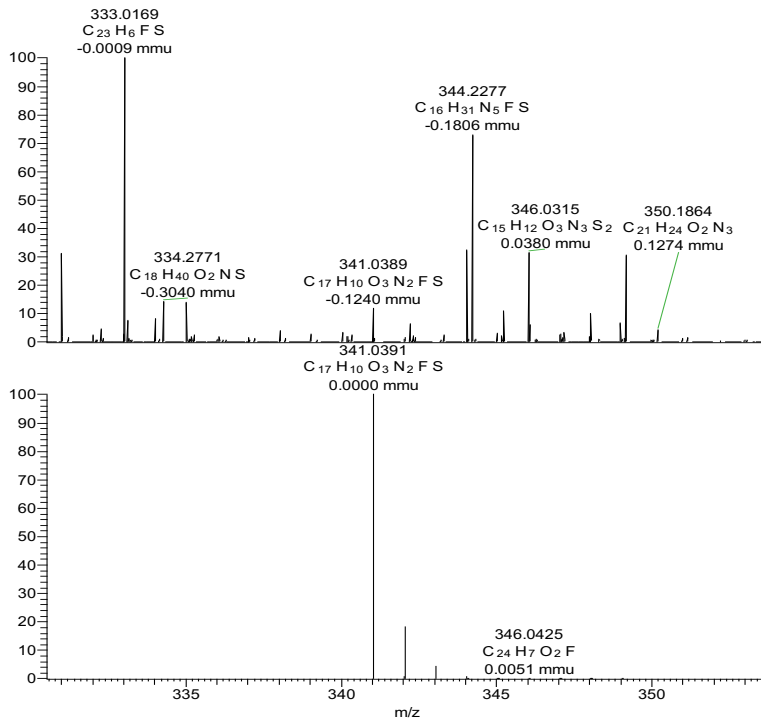
NL:
5.09E7
210423_100_sclick_20u
m#1-28 RT: 0.02-0.50
AV: 28 T: FTMS + p NSI
Full ms
[180.0000-700.0000]

NL:
3.94E5
C₁₇H₁₀O₃N₂Br S:
C₁₇H₁₀O₃N₂Br₁S₁
pa Chrg 1

UPLC-MS



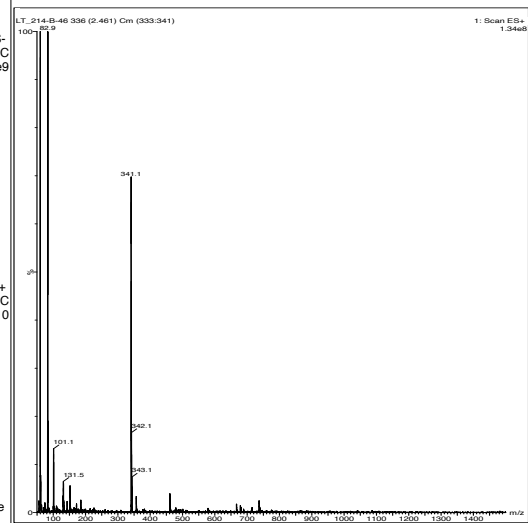
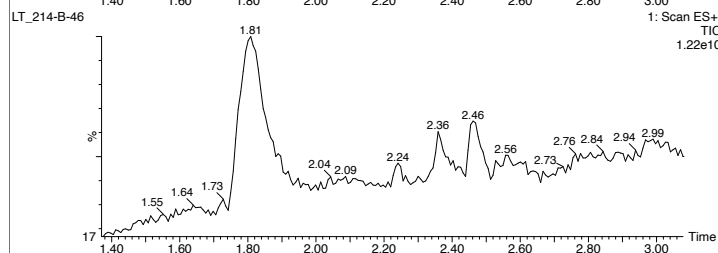
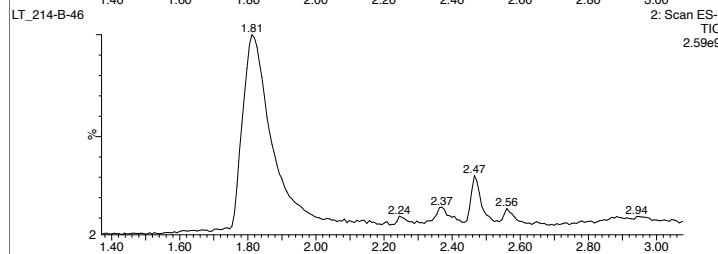
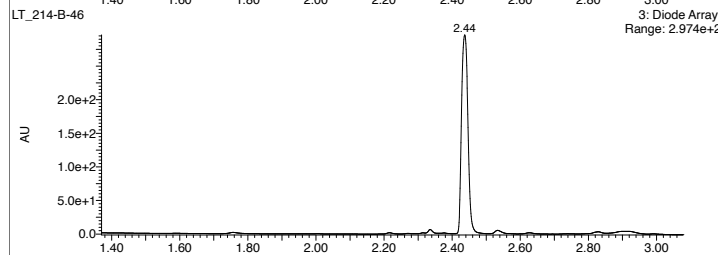
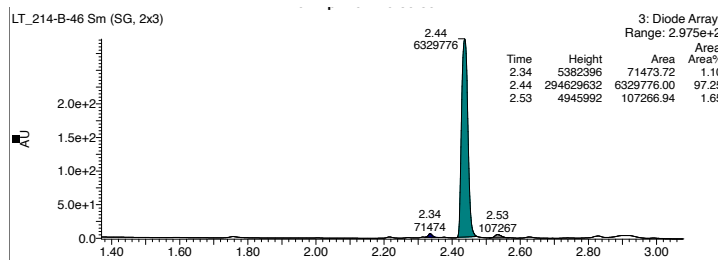
Compound 5b



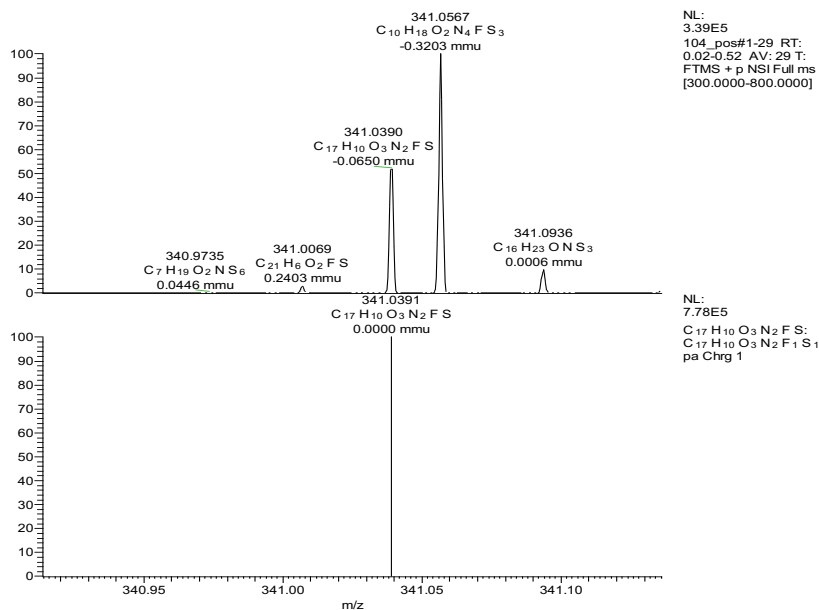
NL:
2.69E6
101_pos#1-29 RT:
0.02-0.52 AV: 29 T:
FTMS + p NSI Full ms
[300.0000-800.0000]

NL:
7.78E5
C₁₇H₁₀O₃N₂F S:
C₁₇H₁₀O₃N₂F₁S₁
pa Chrg 1

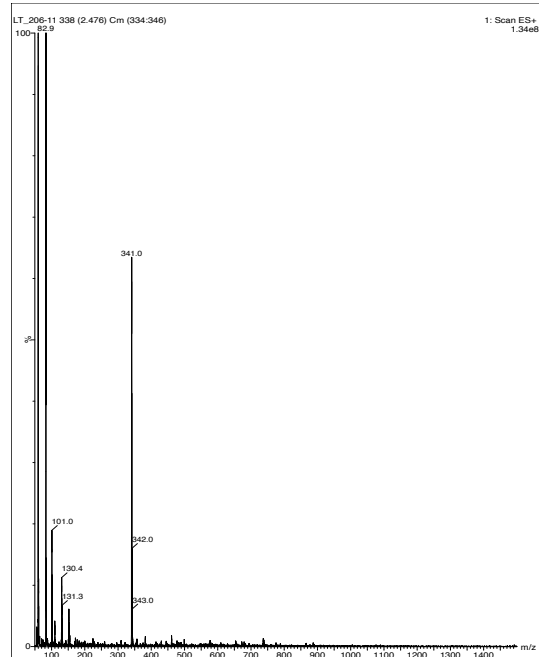
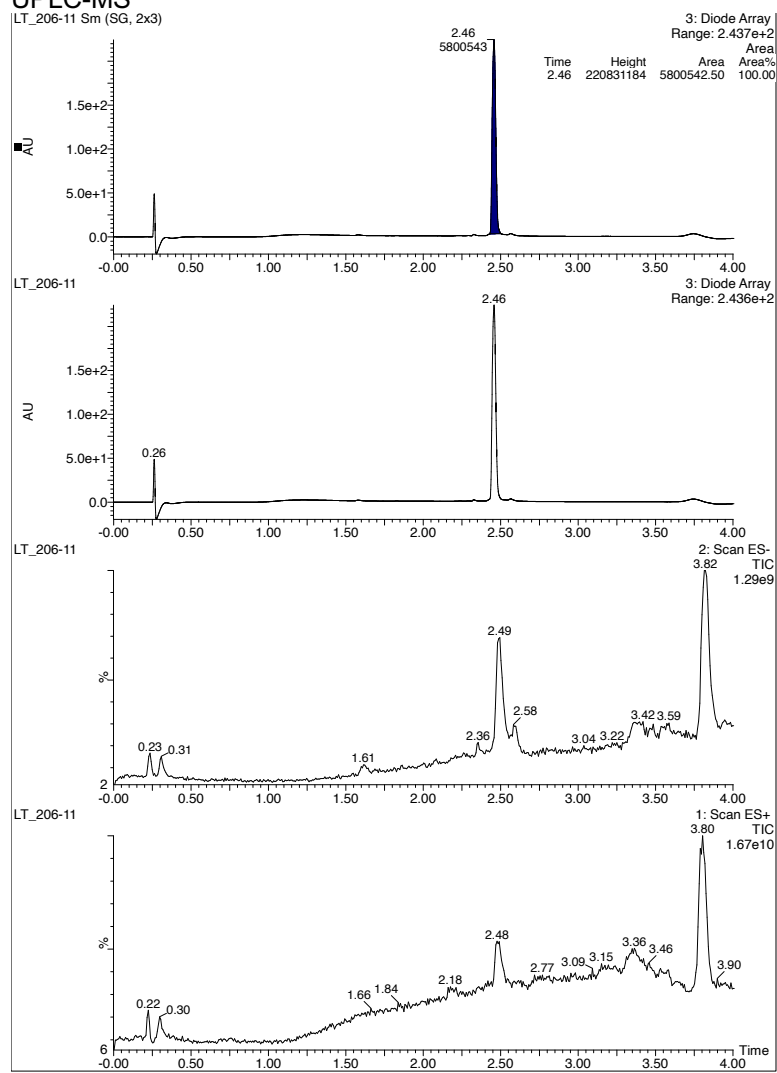
UPLC-MS



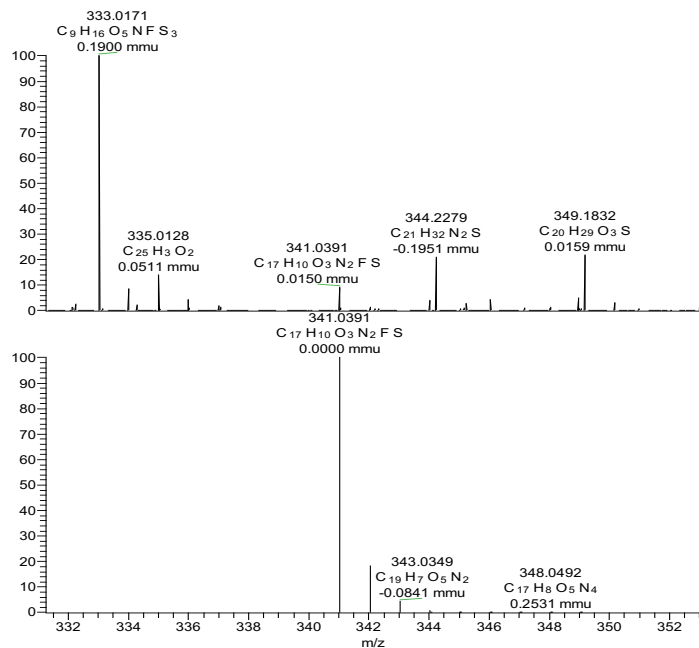
Compound 5c



UPLC-MS



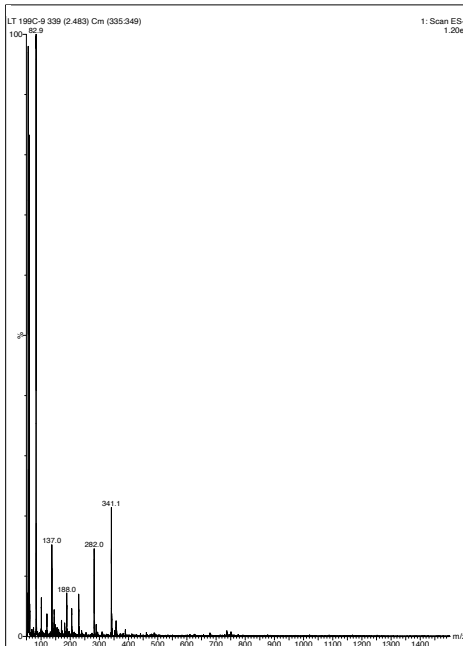
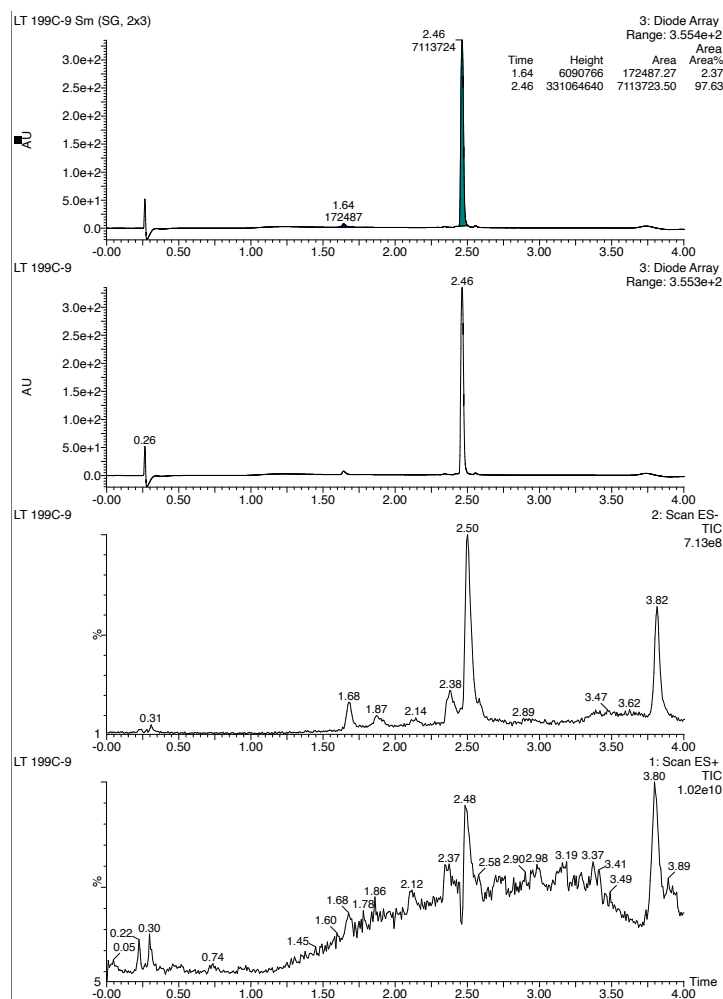
Compound 5d



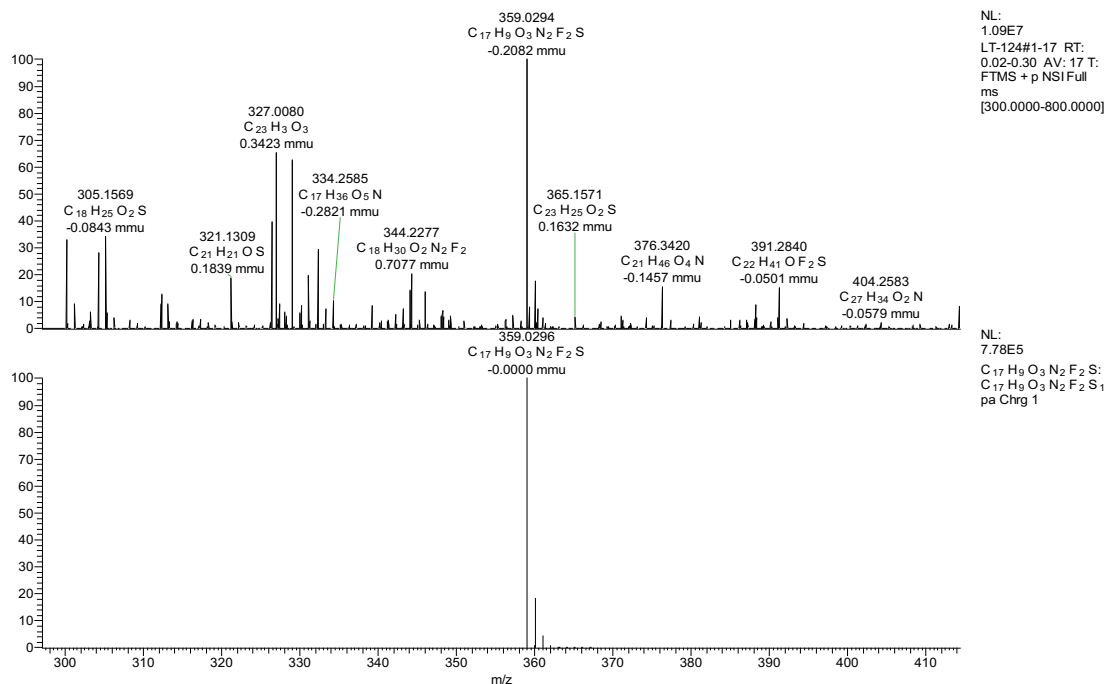
NL:
9.93E6
103_pos#1-29 RT:
0.02-0.52 AV: 29 T:
FTMS + p NSI Full ms
[300.0000-800.0000]

NL:
7.78E5
C17H10O3N2FS:
C17H10O3N2F1S1
pa Chrg 1

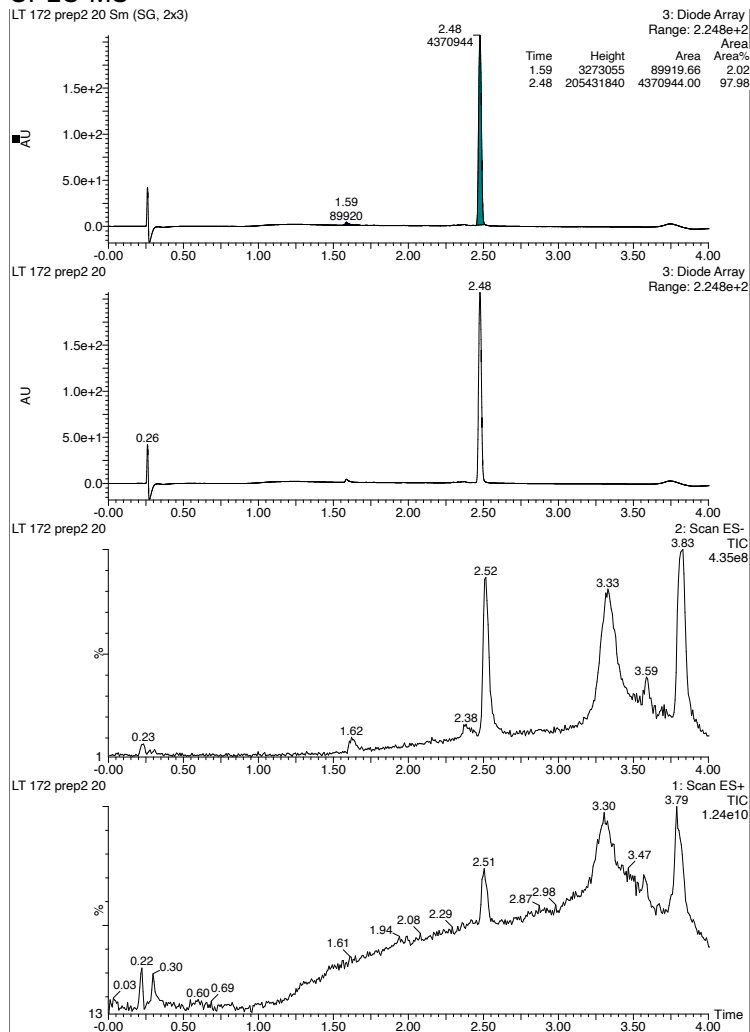
UPLC-MS



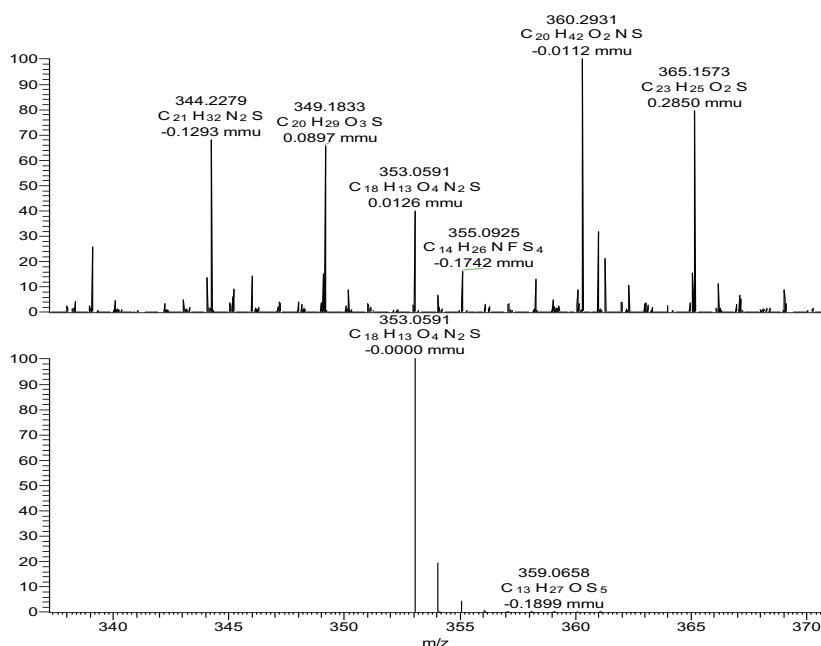
Compound 5e



UPLC-MS



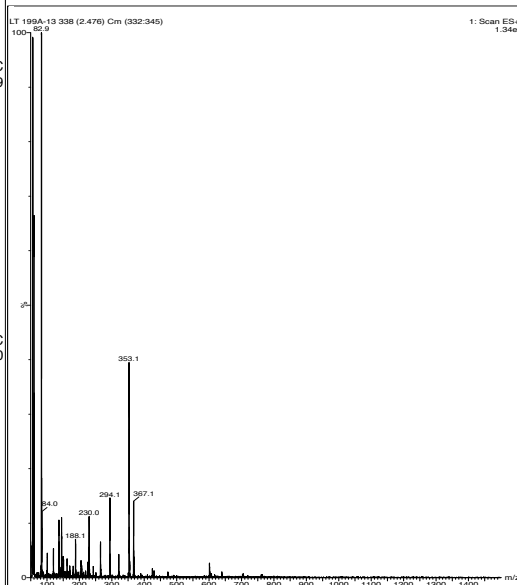
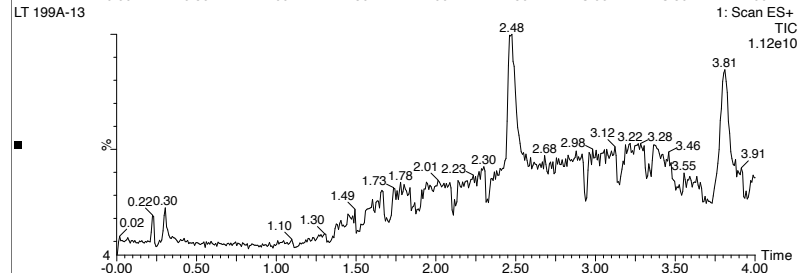
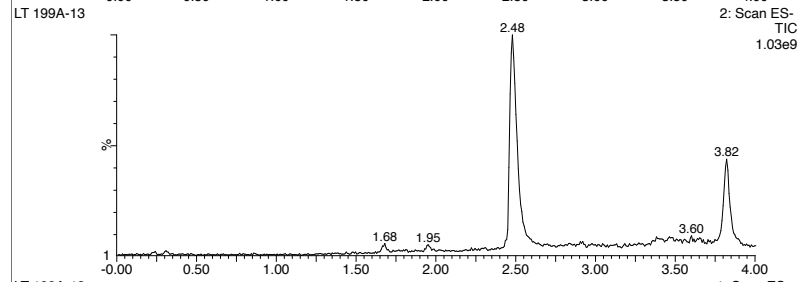
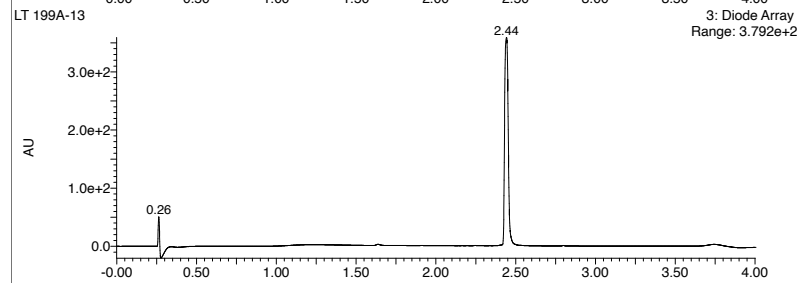
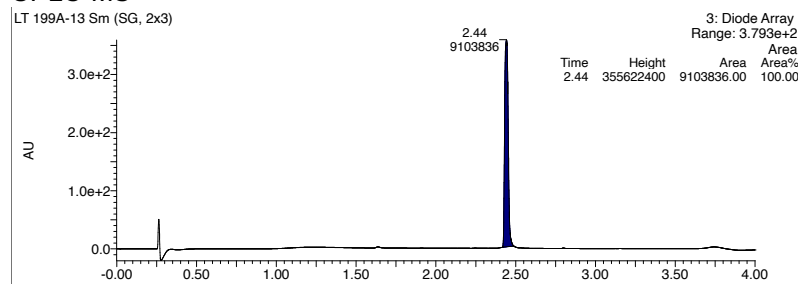
Compound 5f



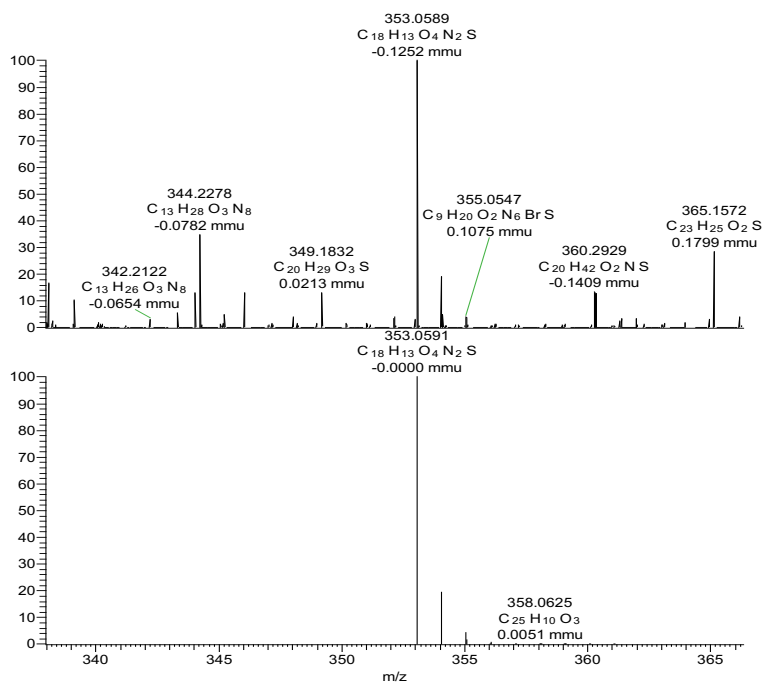
NL:
1.67E6
105_pos#1-29 RT:
0.02-0.52 AV: 29 T:
FTMS + p NSI Full
ms
[300.0000-
800.0000]

NL:
7.68E5
C₁₈H₁₃O₄N₂S:
C₁₈H₁₃O₄N₂S 1
pa Chrg 1

UPLC-MS



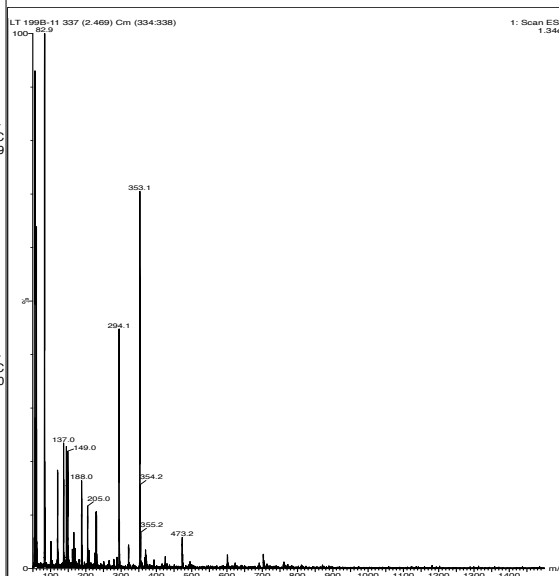
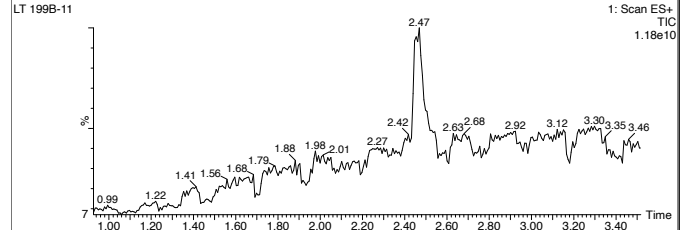
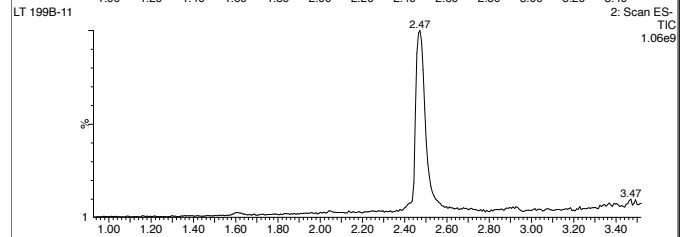
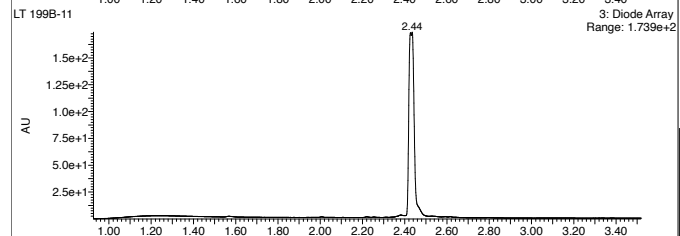
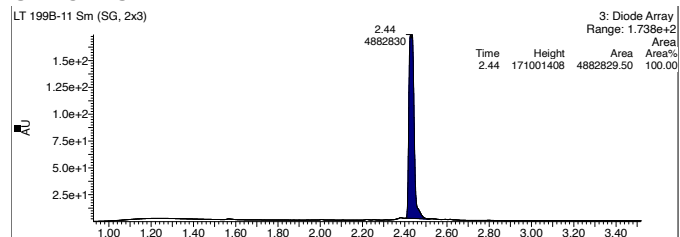
Compound 5g



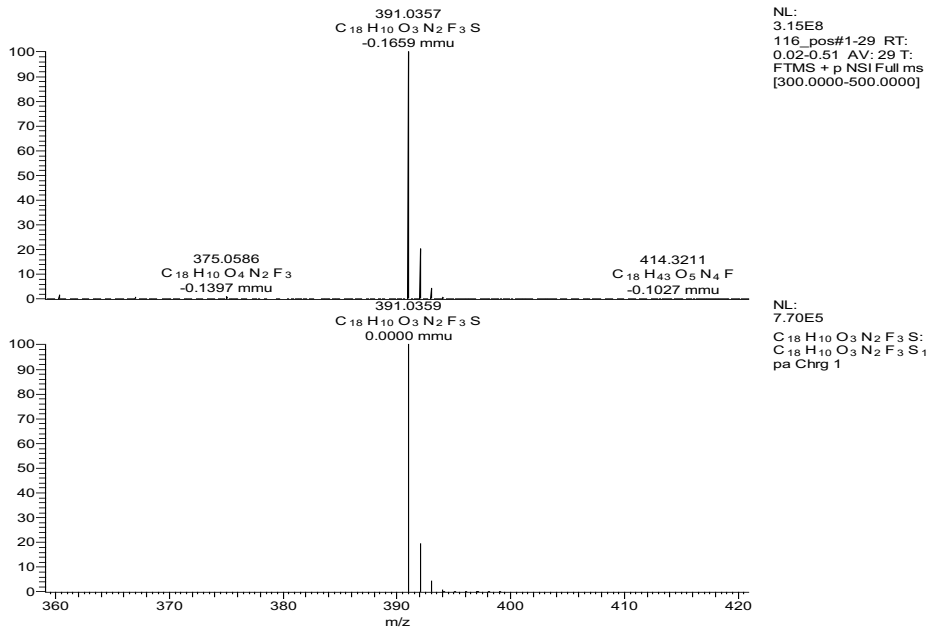
NL:
3.16E6
109_pos#1-29 RT:
0.02-0.52 AV: 29 T:
FTMS + p NSI Full
ms
[300.0000-
800.0000]

NL:
7.68E5
C₁₈H₁₃O₄N₂S:
C₁₈H₁₃O₄N₂S 1
pa Chrg 1

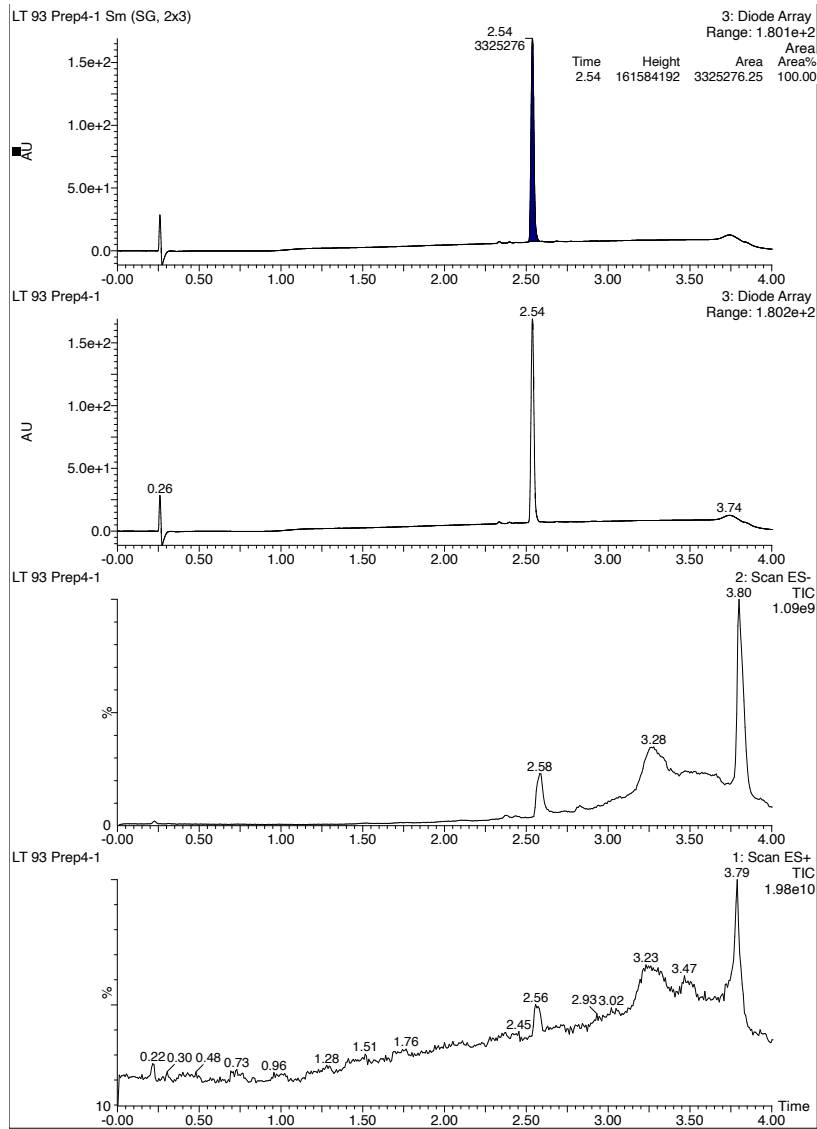
UPLC-MS



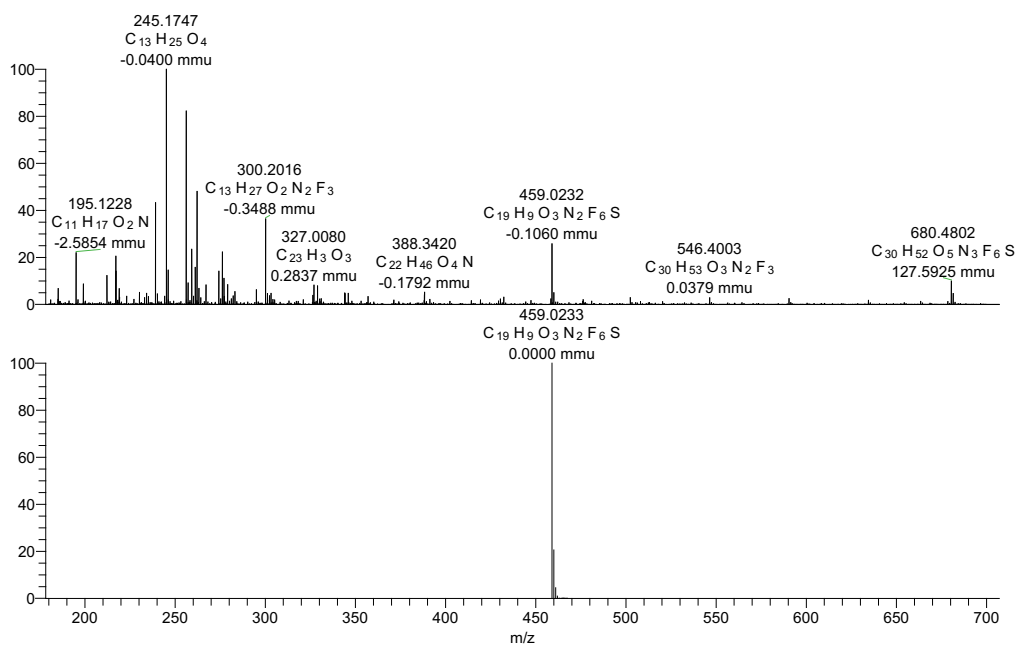
Compound 5h



UPLC-MS



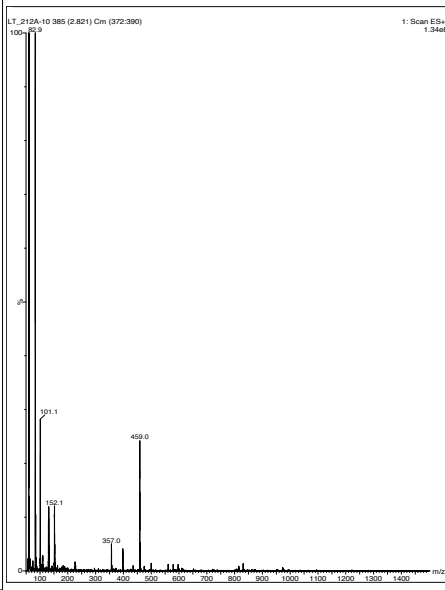
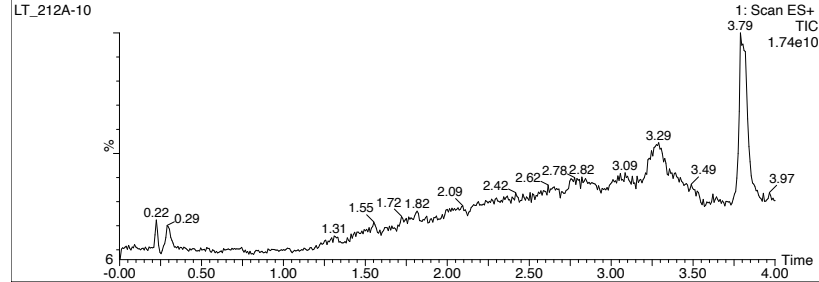
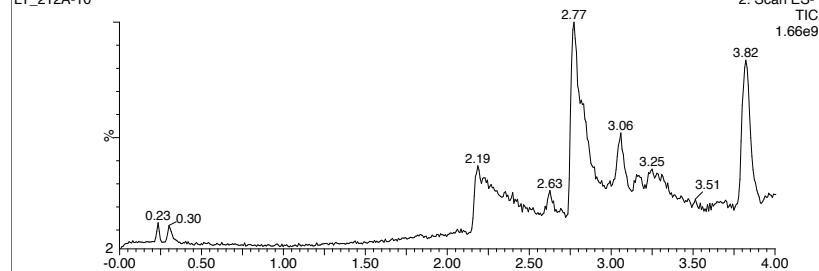
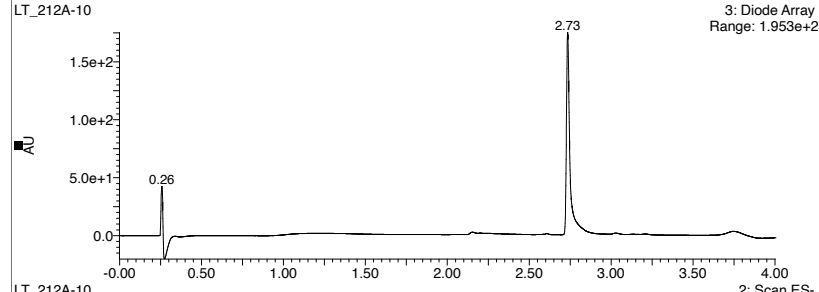
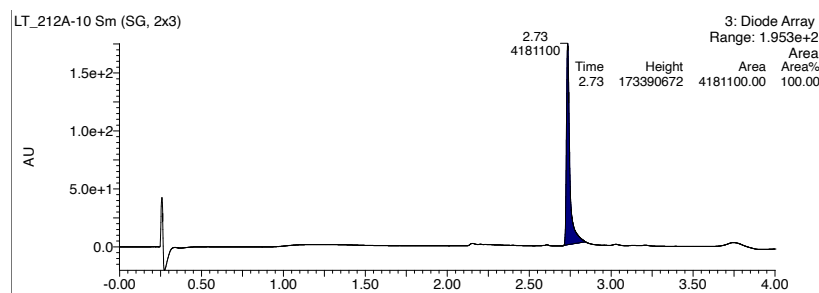
Compound 5i



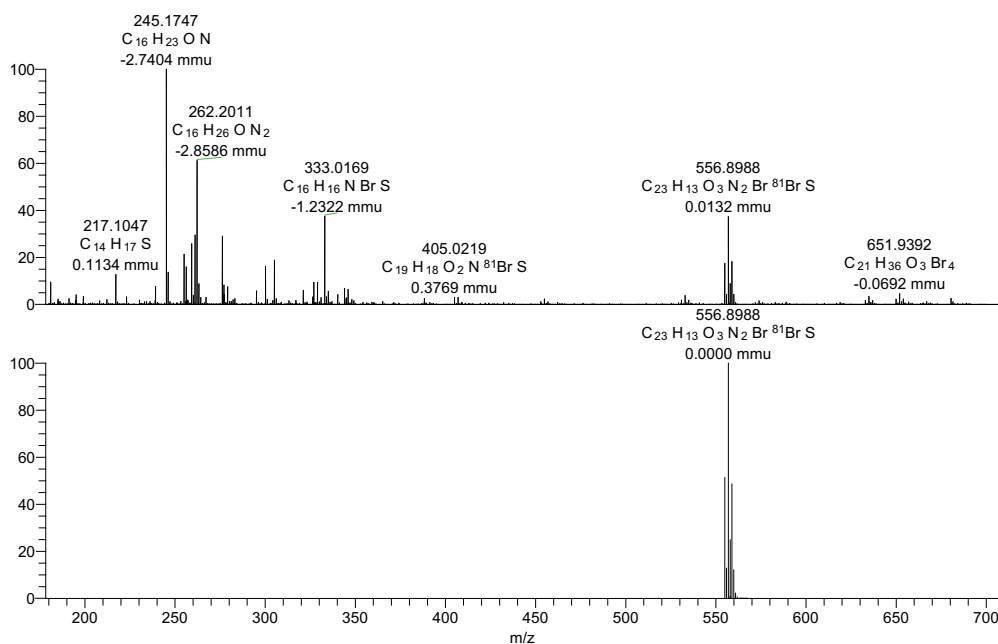
NL:
8.94E7
210423_126_10um#1-
28 RT: 0.02-0.50 AV:
28 T: FTMS + p NSI
Full ms
[180.0000-700.0000]

NL:
7.62E5
C₁₉H₉O₃N₂F₆S:
C₁₉H₉O₃N₂F₆S₁
pa Chrg 1

UPLC-MS



Compound 5j

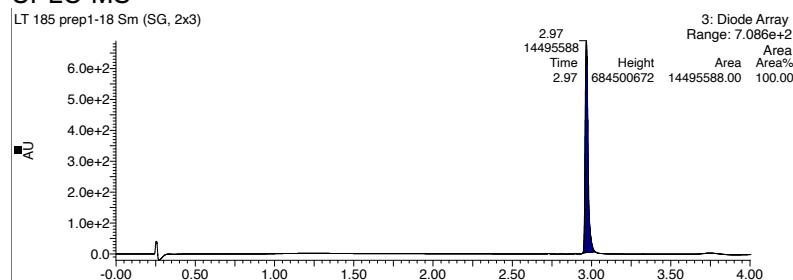


NL:
3.26E7
210423_128_20um#1-
28 RT: 0.02-0.50 AV:
28 T: FTMS + p NSI
Full ms
[180.0000-700.0000]

NL:
3.65E5
C₂₃H₁₃O₃N₂Br₂S:
C₂₃H₁₃O₃N₂Br₂S₁
pa Chrg 1

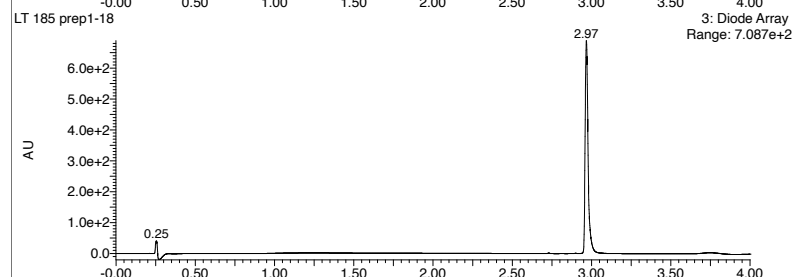
UPLC-MS

LT 185 prep1-18 Sm (SG, 2x3)

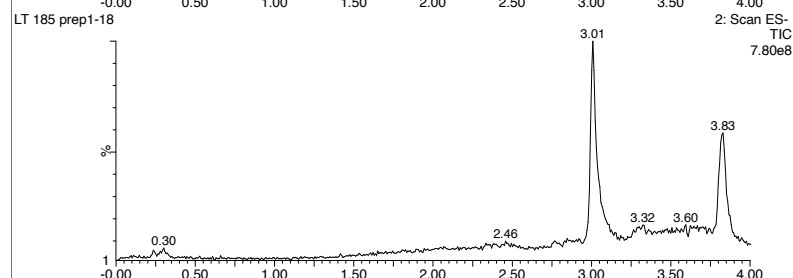


Compound 5k

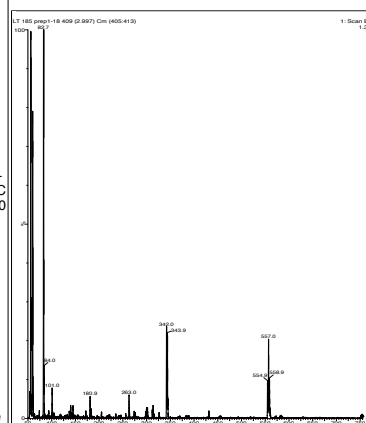
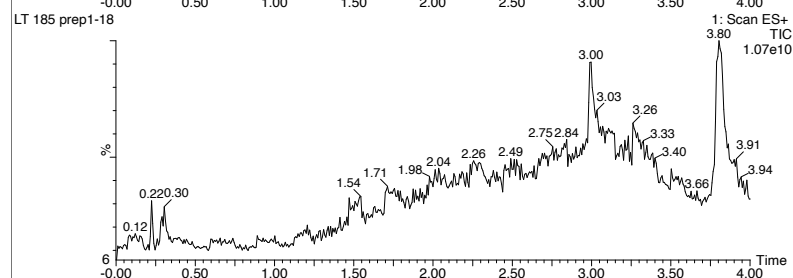
LT 185 prep1-18

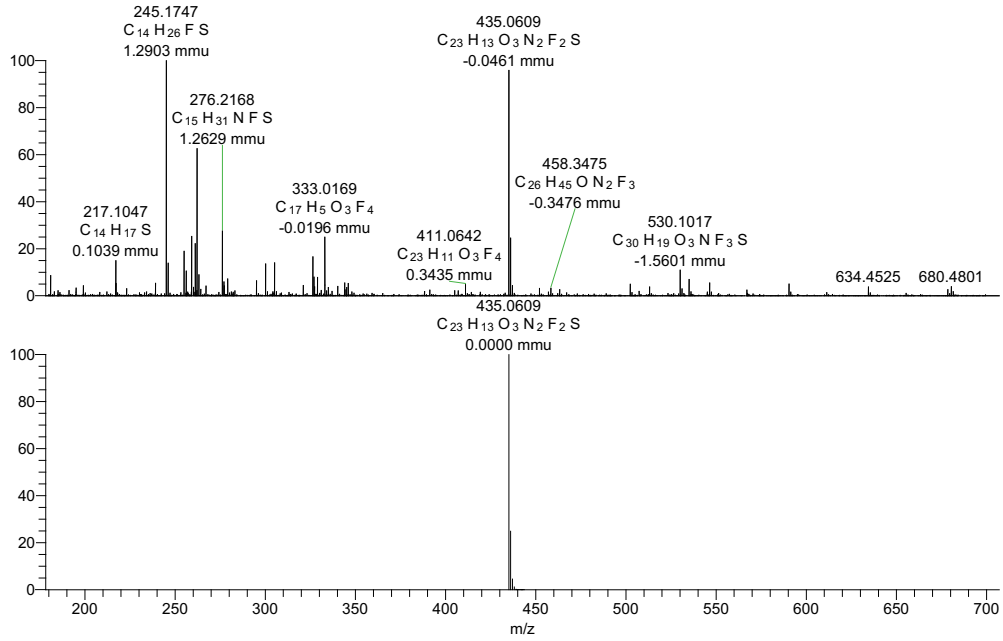


LT 185 prep1-18



LT 185 prep1-18

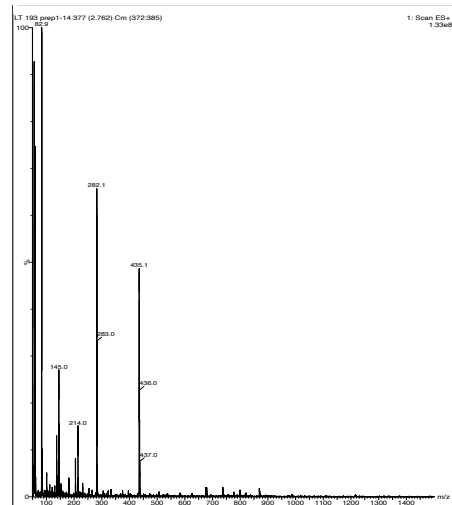
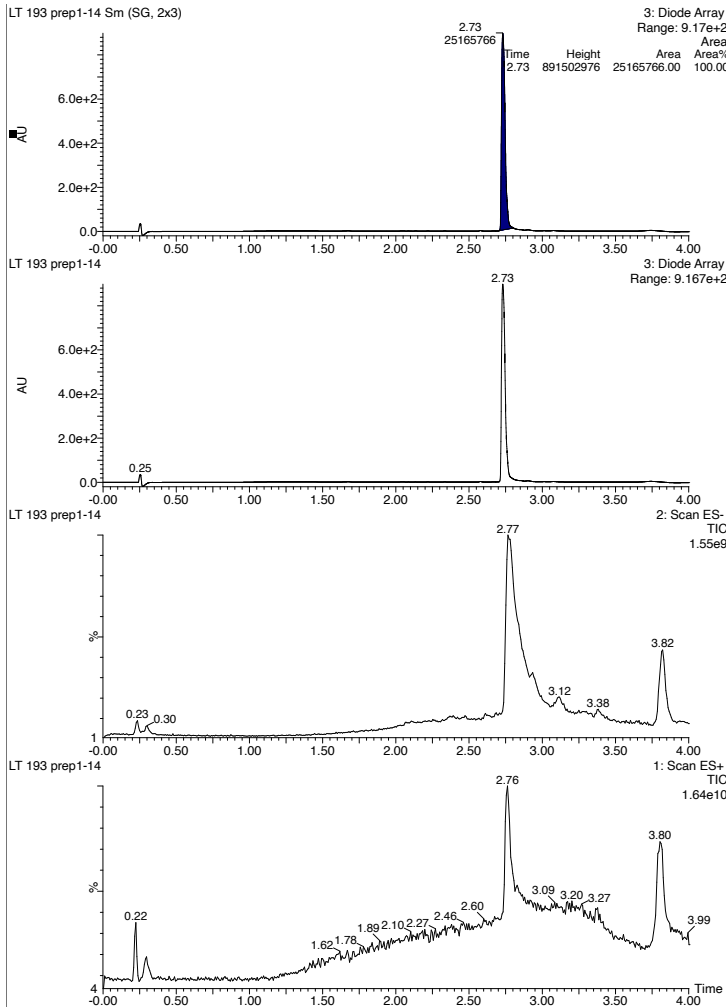




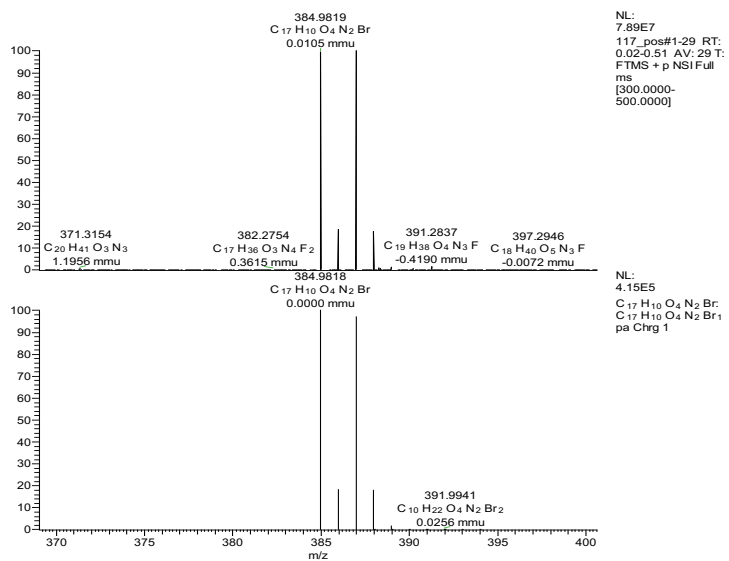
NL:
3.99E7
210423_129_10um#1-
29 RT: 0.02-0.51 AV:
29 T: FTMS + p NSI
Full ms
[180.0000-700.0000]

NL:
7.29E5
C₂₃H₁₃O₃N₂F₂S:
C₂₃H₁₃O₃N₂F₂S₁
pa Chrg 1

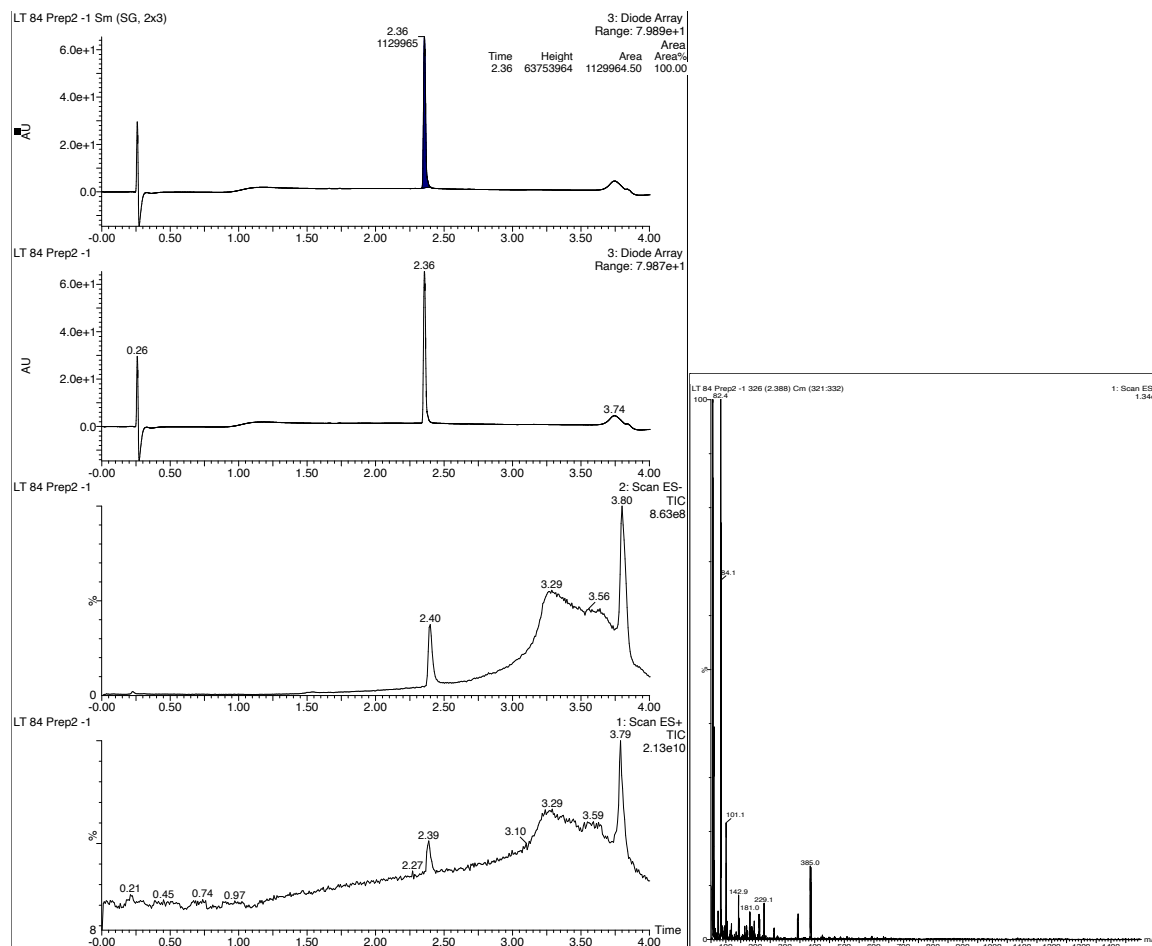
UPLC-MS



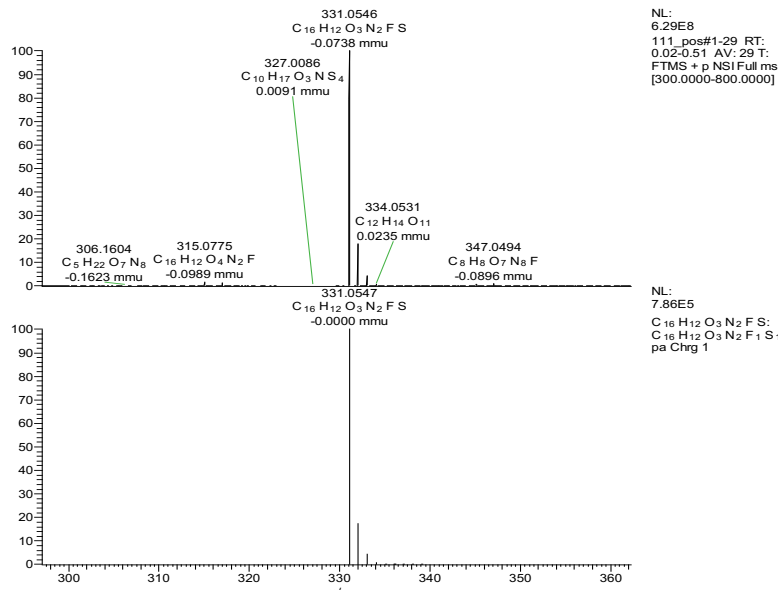
Compound 51



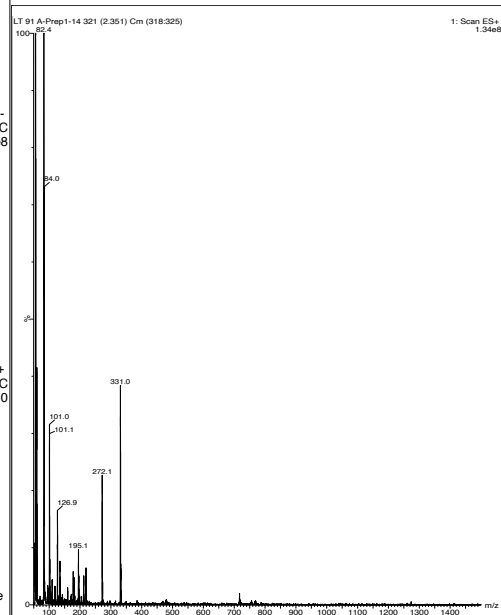
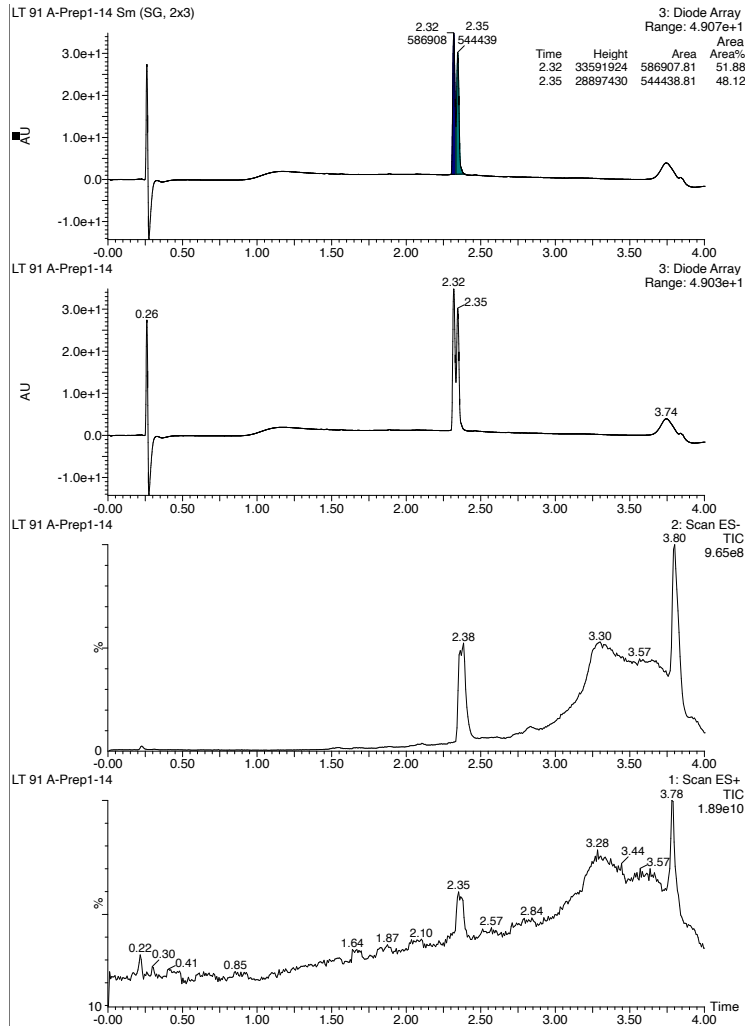
UPLC-MS



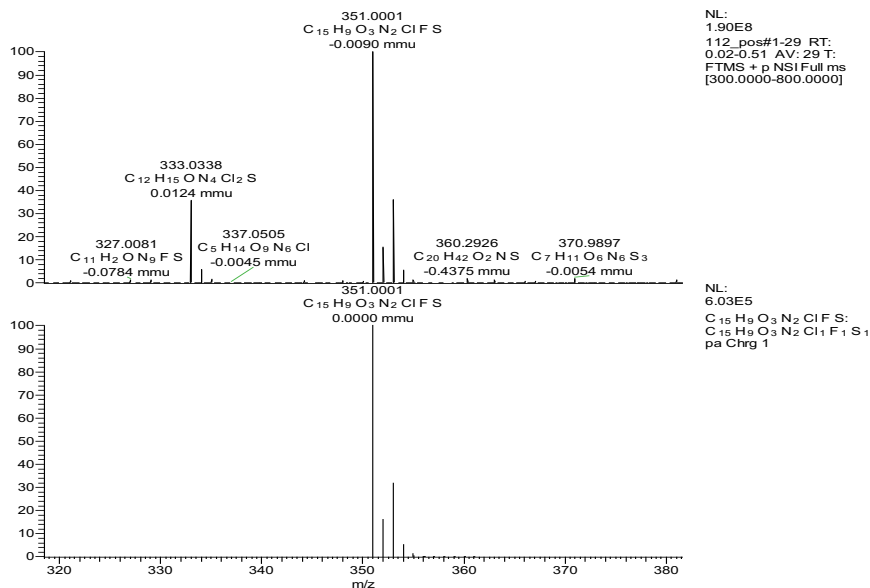
Compound 6a



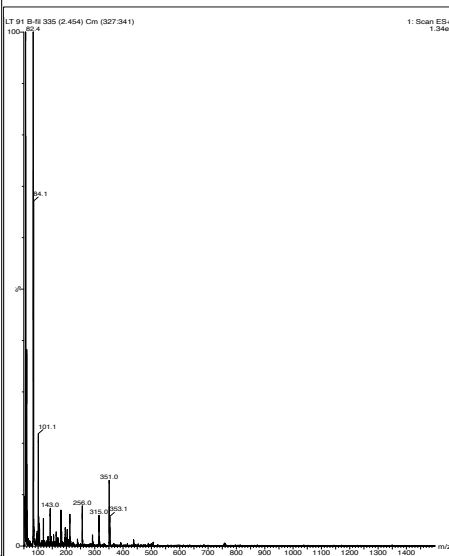
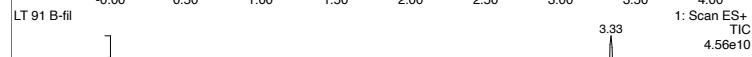
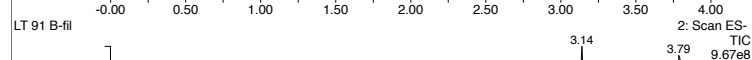
UPLC-MS



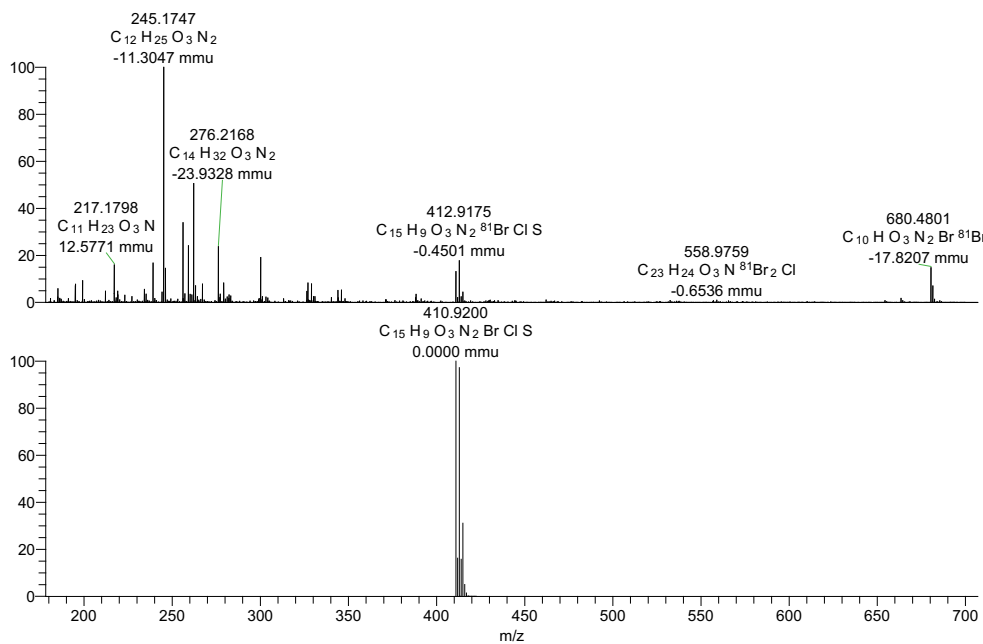
Compound 6b



UPLC-MS



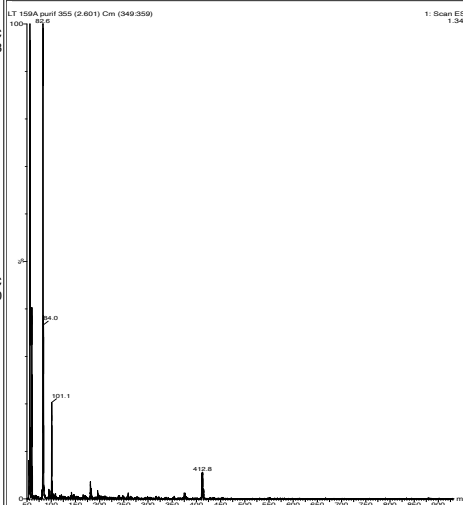
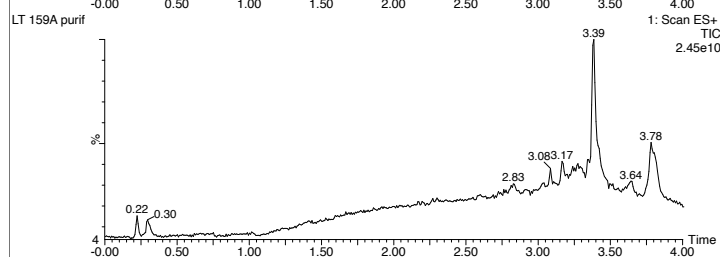
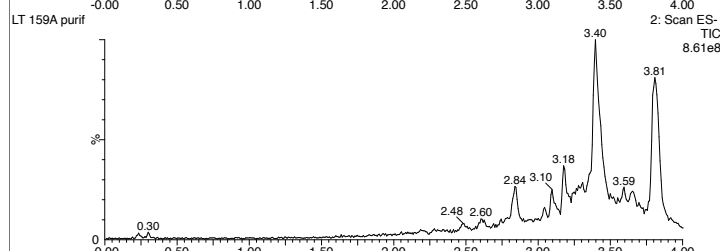
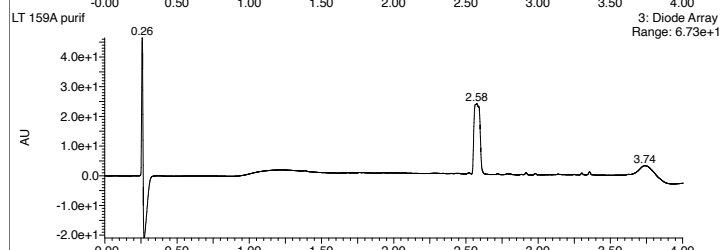
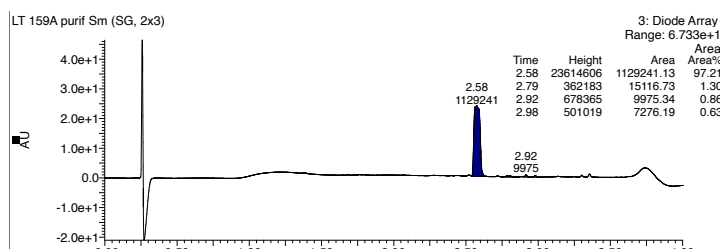
Compound 6c



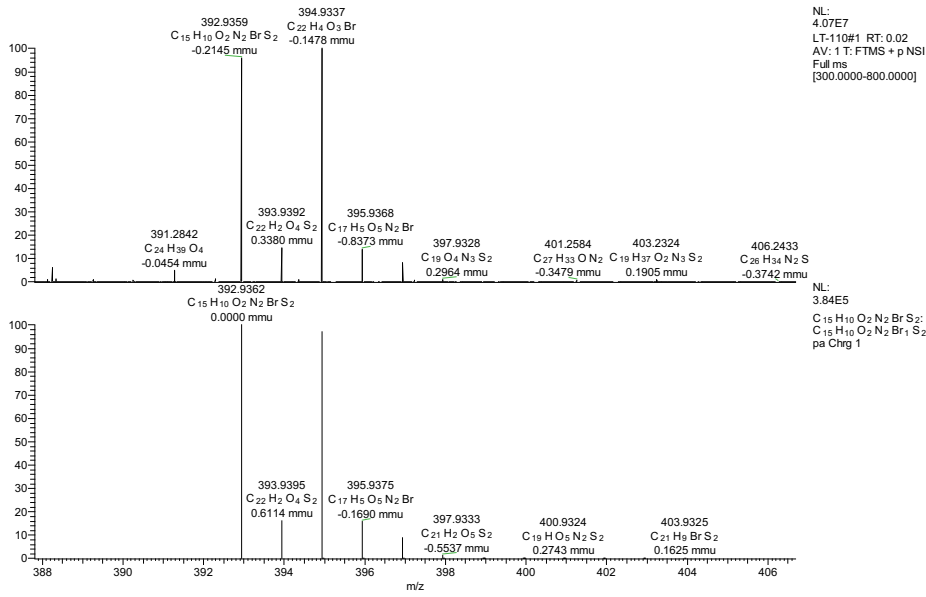
NL:
6.61E7
210423_125_30um#1-28
RT: 0.02-0.50 AV: 28 T:
FTMS + p NSI Full ms
[180.0000-700.0000]

NL:
3.05E5
 $C_{15}H_9O_3N_2BrClS$:
 $C_{15}H_9O_3N_2Br_1Cl_1S_1$
pa Chrg 1

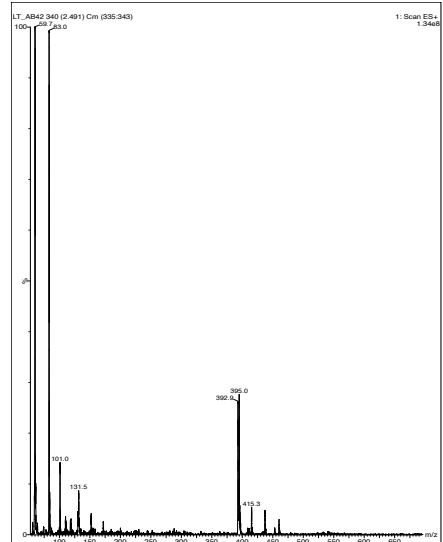
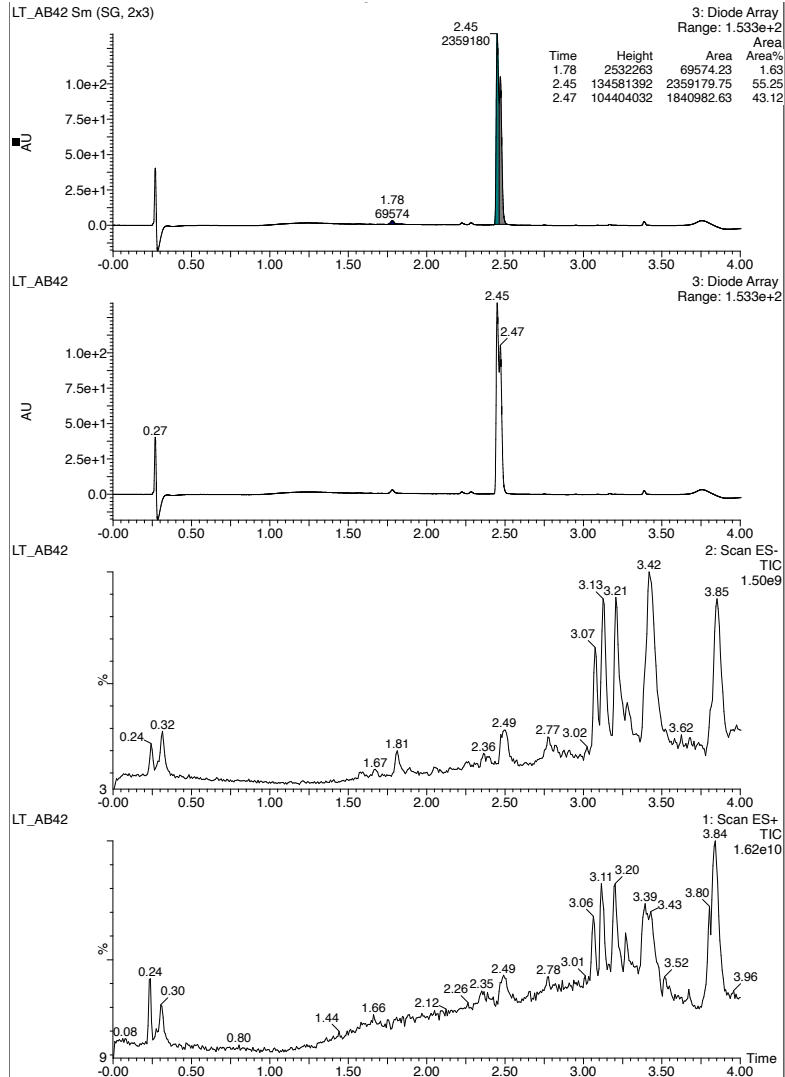
UPLC-MS



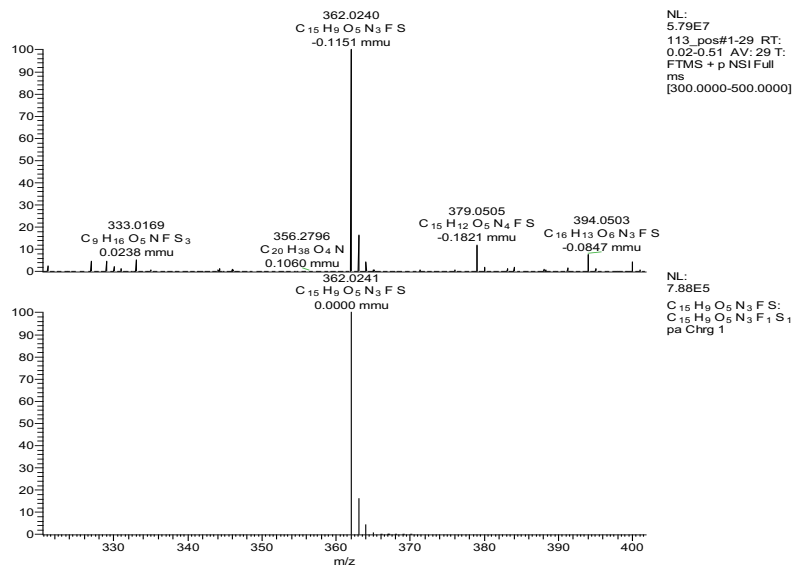
Compound 6d



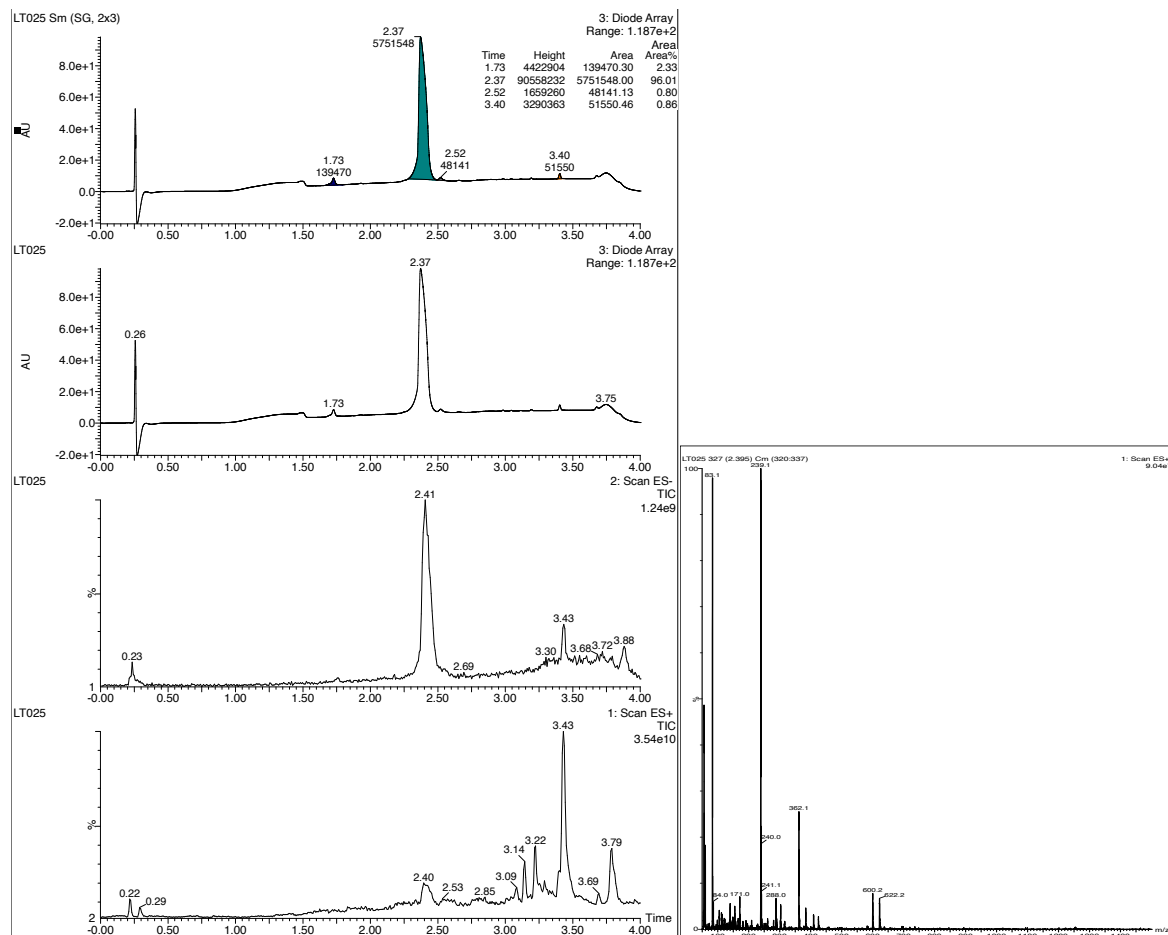
UPLC-MS



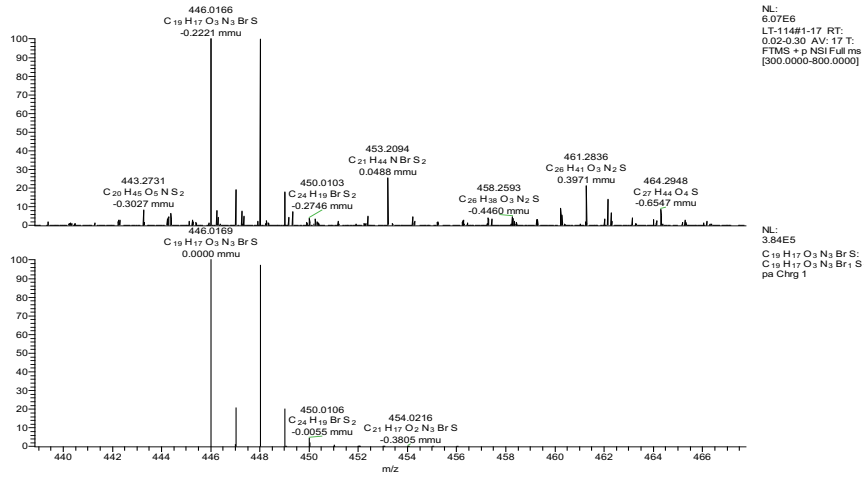
Compound 6e



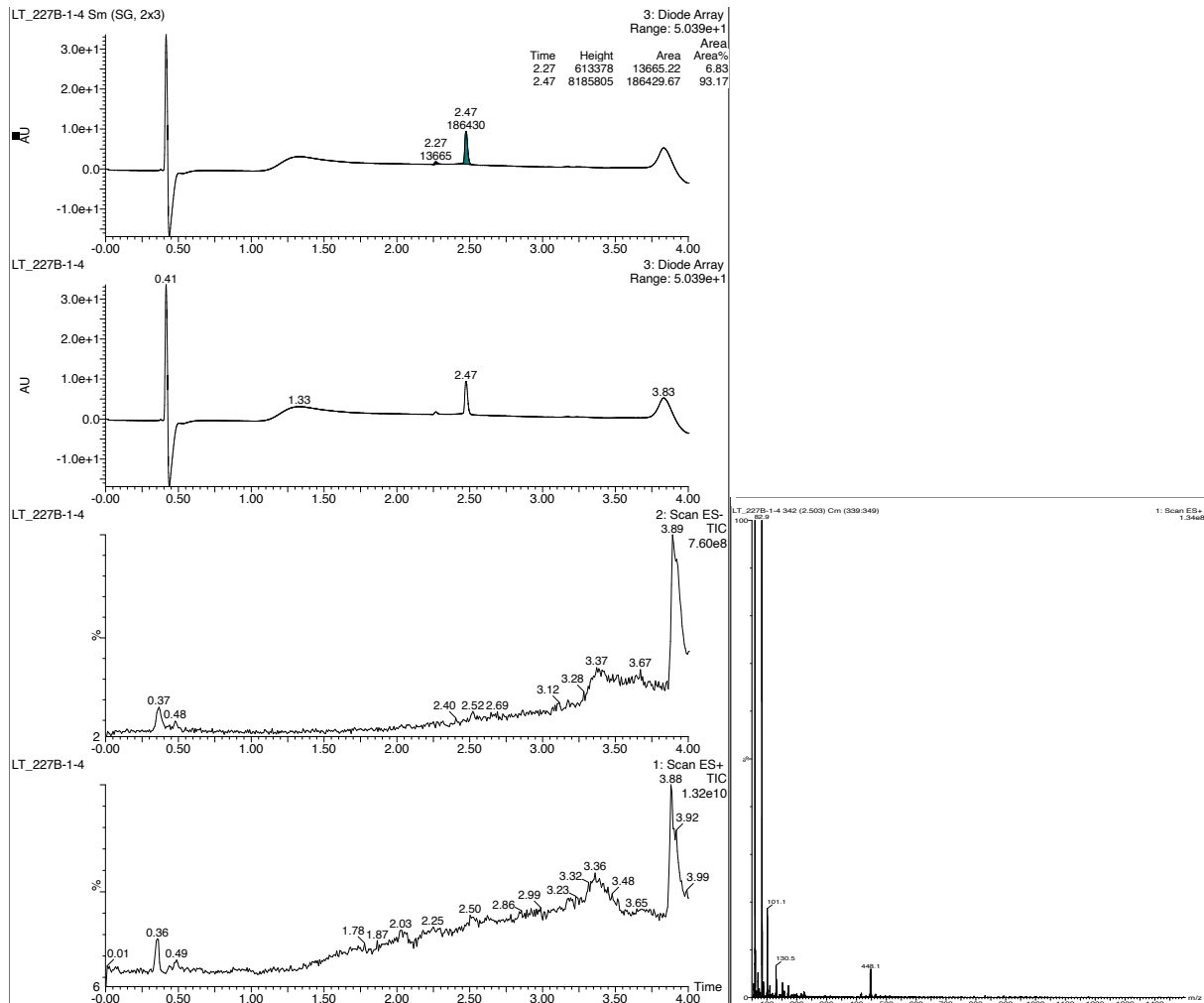
UPLC-MS



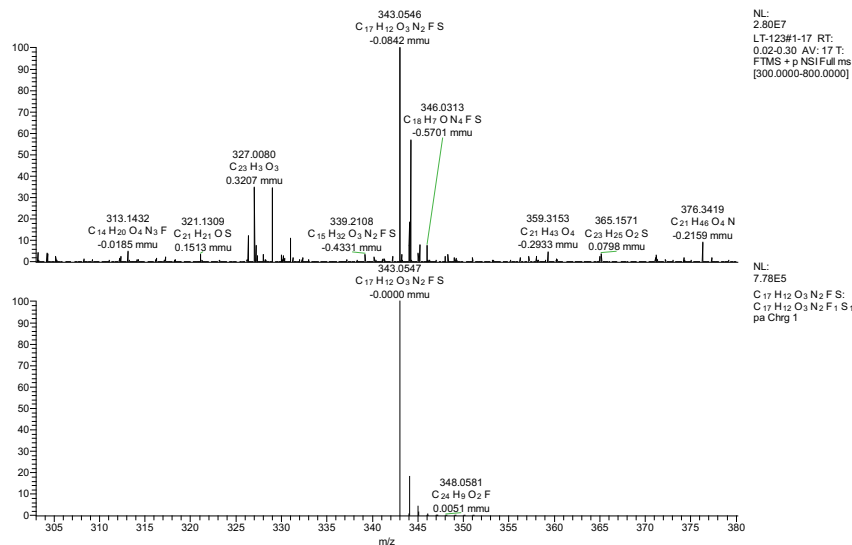
Compound 6f



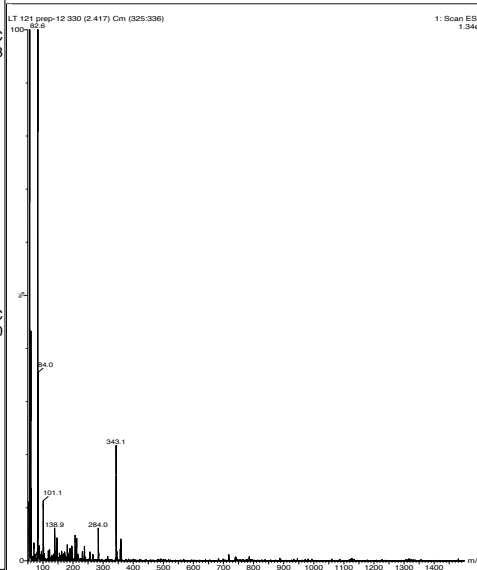
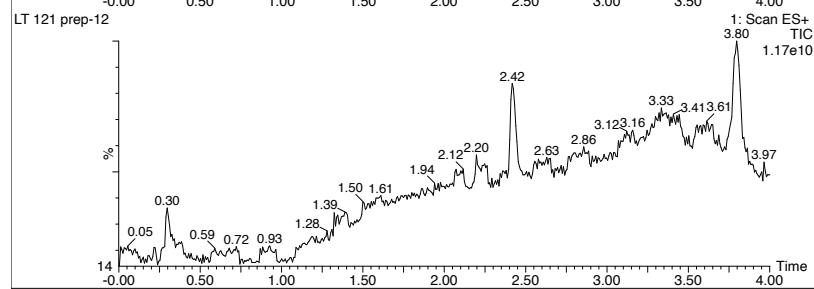
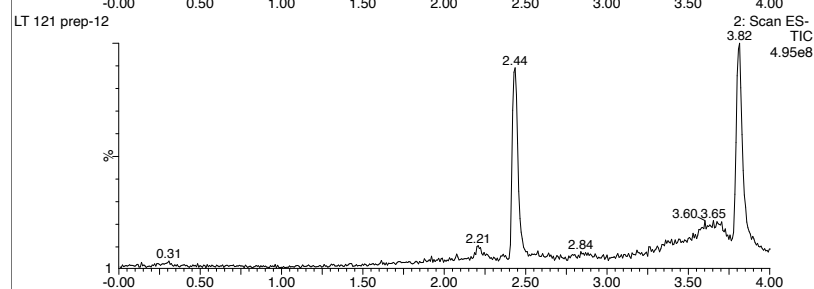
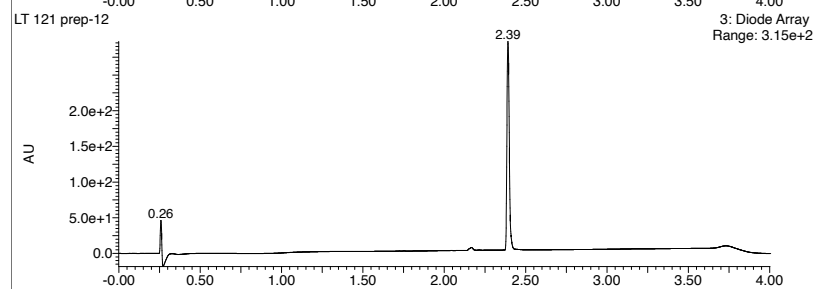
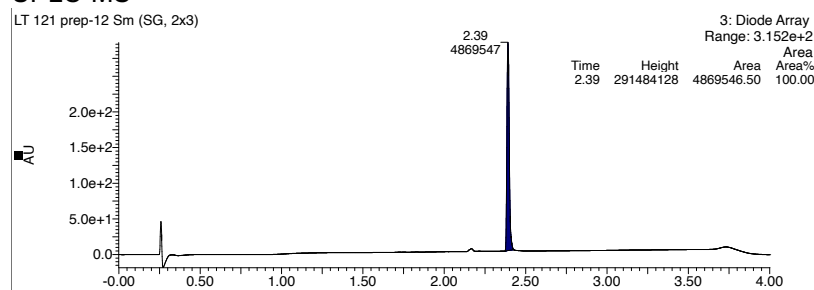
UPLC-MS



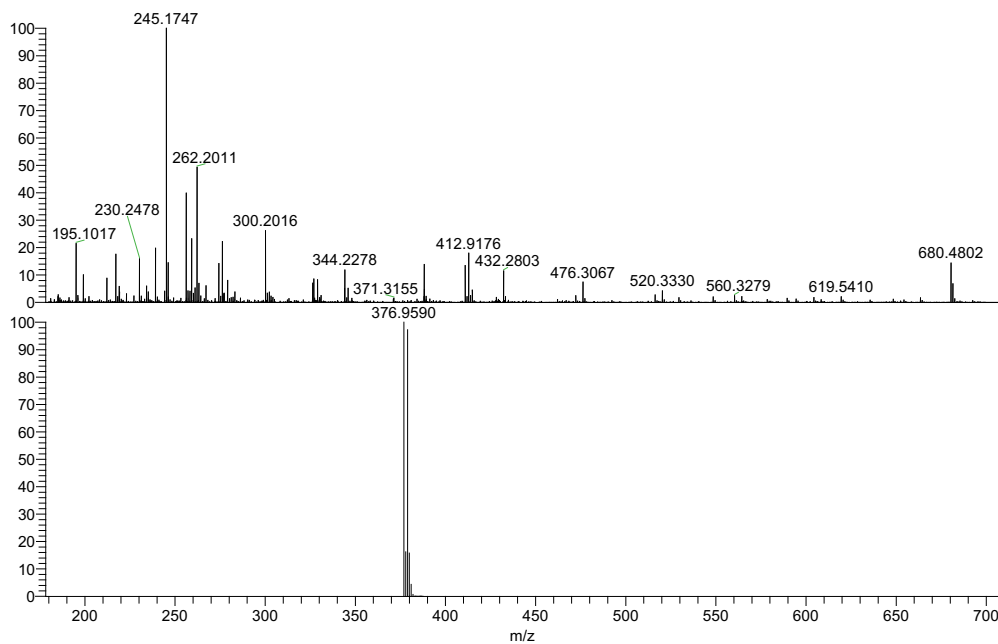
Compound 6g



UPLC-MS



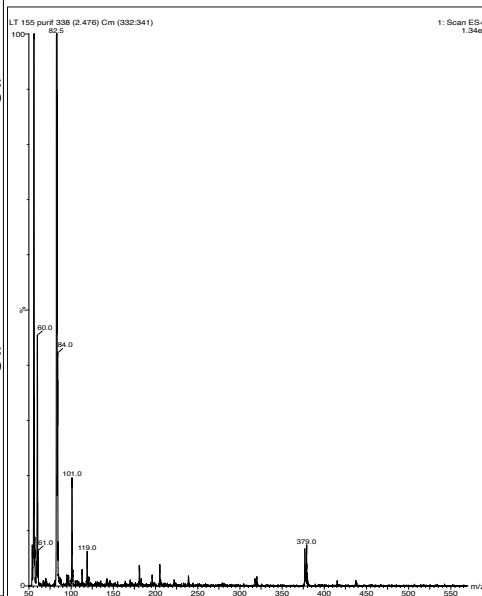
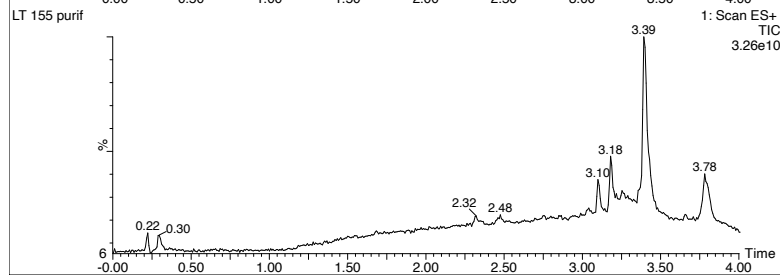
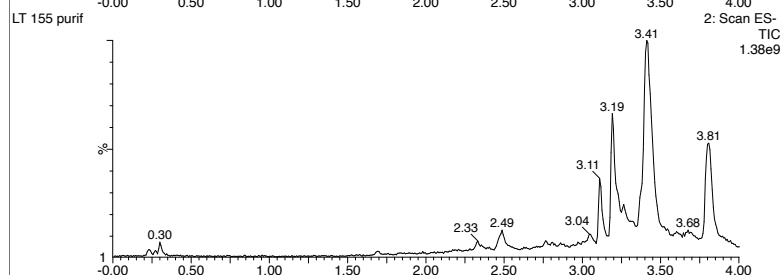
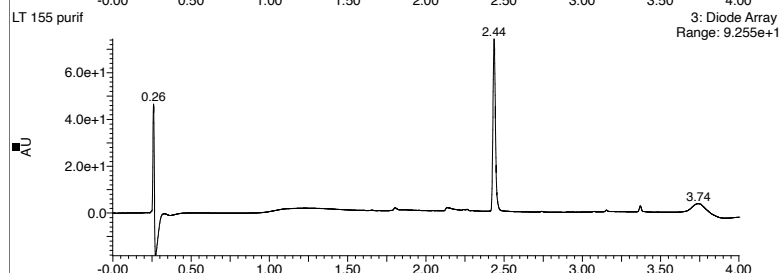
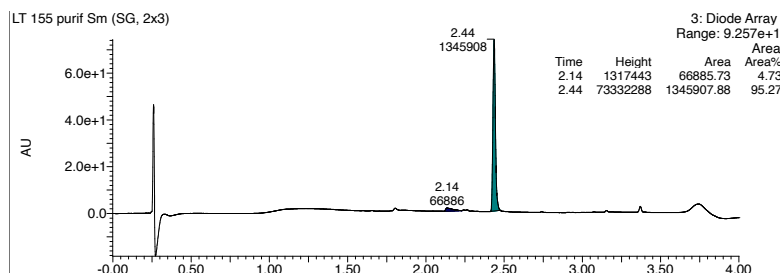
Compound 6h



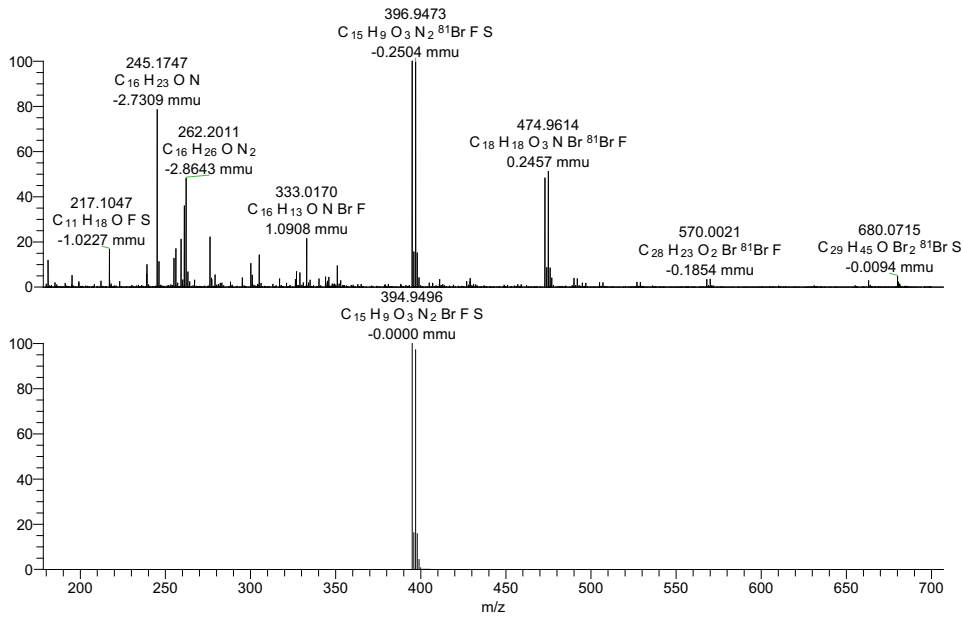
NL:
5.88E7
210423_127_30um#1-
29 RT: 0.02-0.51 AV:
29 T: FTMS + p NSI
Full ms
[180.0000-700.0000]

NL:
4.03E5
C₁₅H₁₀O₃N₂S Br:
C₁₅H₁₀O₃N₂S₁Br₁
pa Chrg 1

UPLC-MS



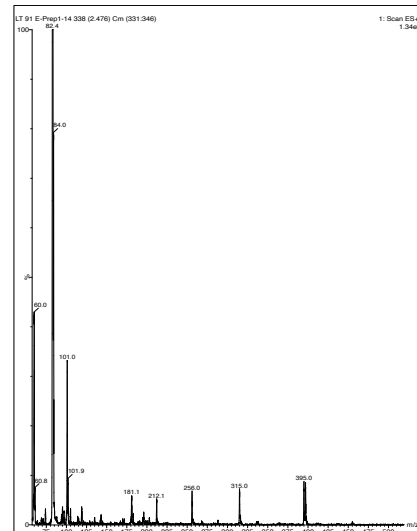
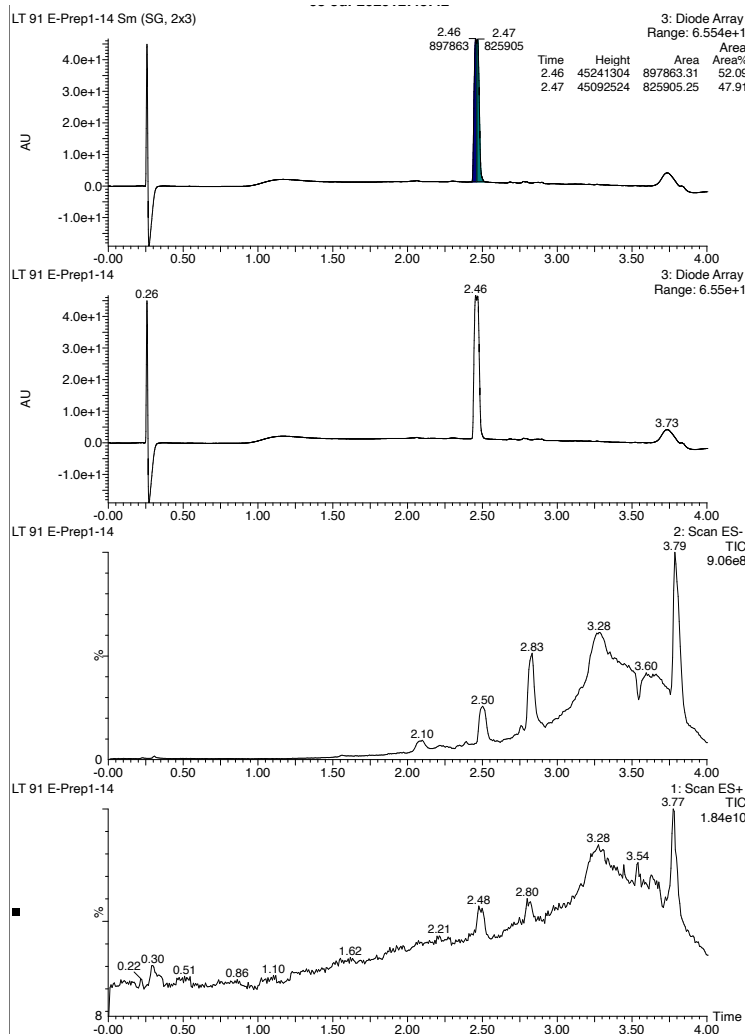
Compound 6i



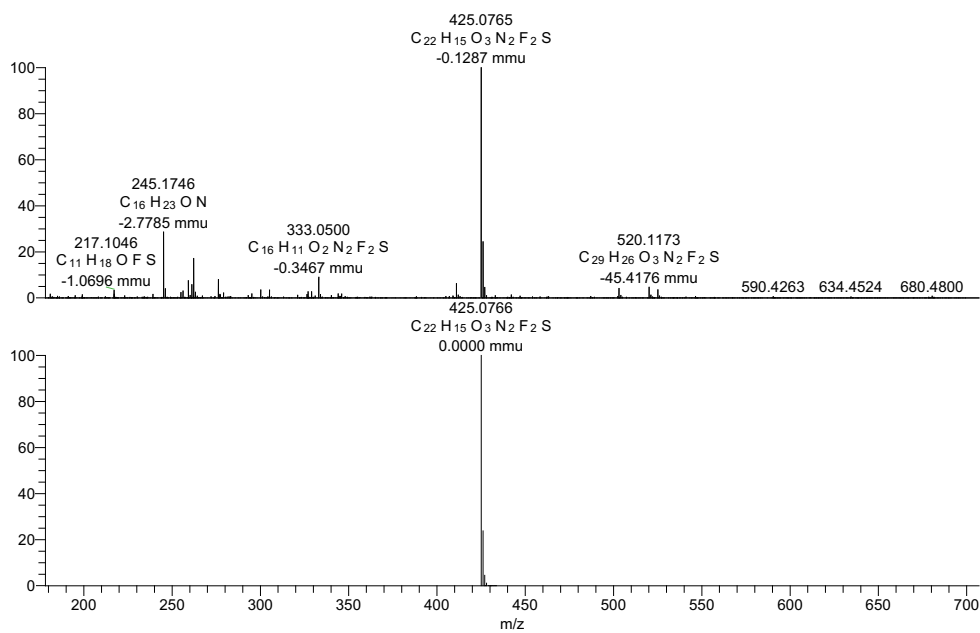
NL:
5.85E7
210423_115_20um#2-28
RT: 0.04-0.50 AV: 27 T:
FTMS + p NSI Full ms
[180.0000-700.0000]

NL:
4.03E5
C₁₅H₉O₃N₂BrFS:
C₁₅H₉O₃N₂Br₁F₁S₁
pa Chrg 1

UPLC-MS



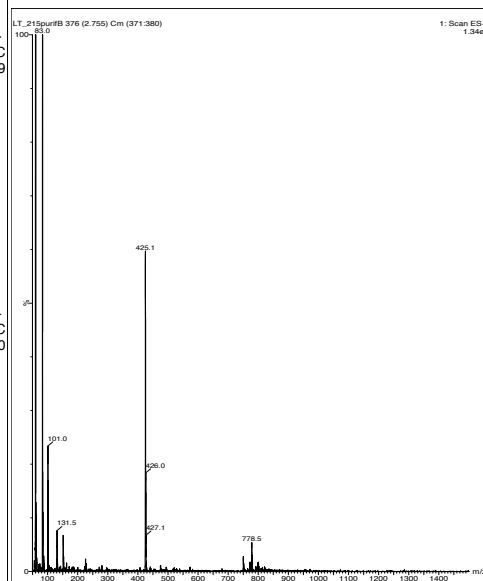
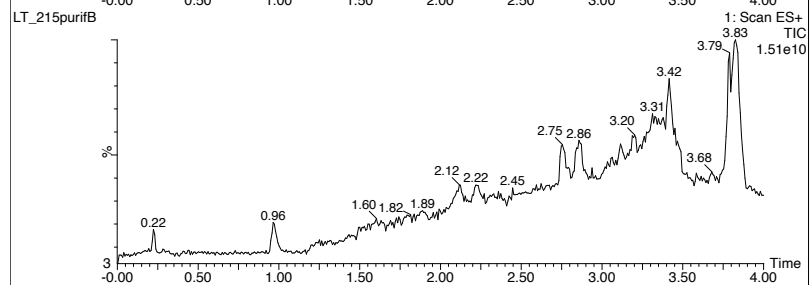
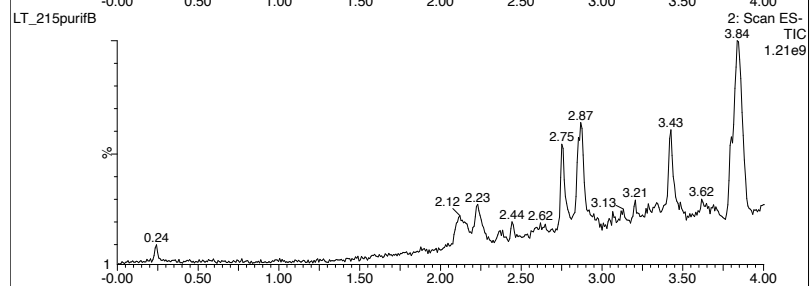
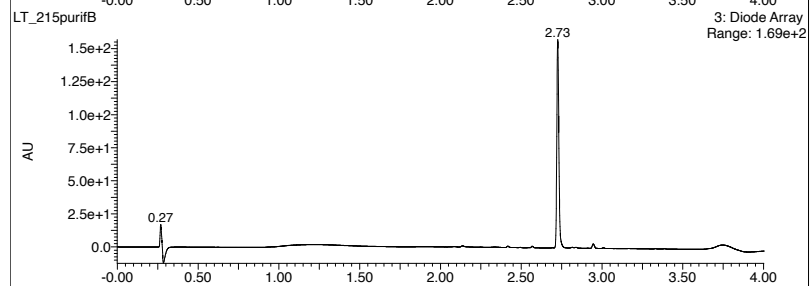
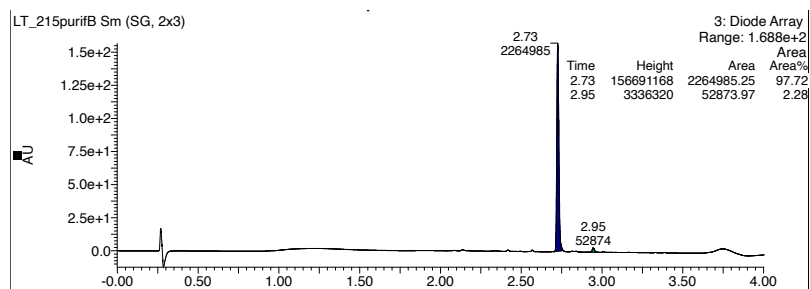
Compound 6j



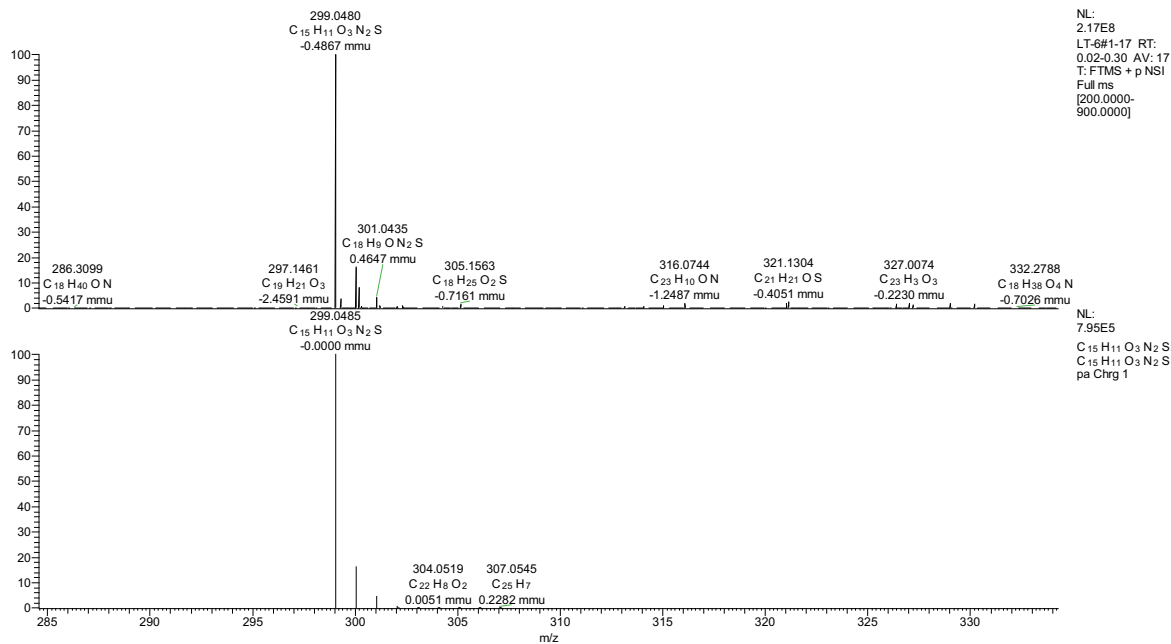
NL:
1.69E8
210423_118_10um#1-
28 RT: 0.02-0.50 AV:
28 T: FTMS + p NSI
Full ms
[180.0000-700.0000]

NL:
7.37E5
C₂₂H₁₅O₃N₂F₂S:
C₂₂H₁₅O₃N₂F₂S₁
pa Chrg 1

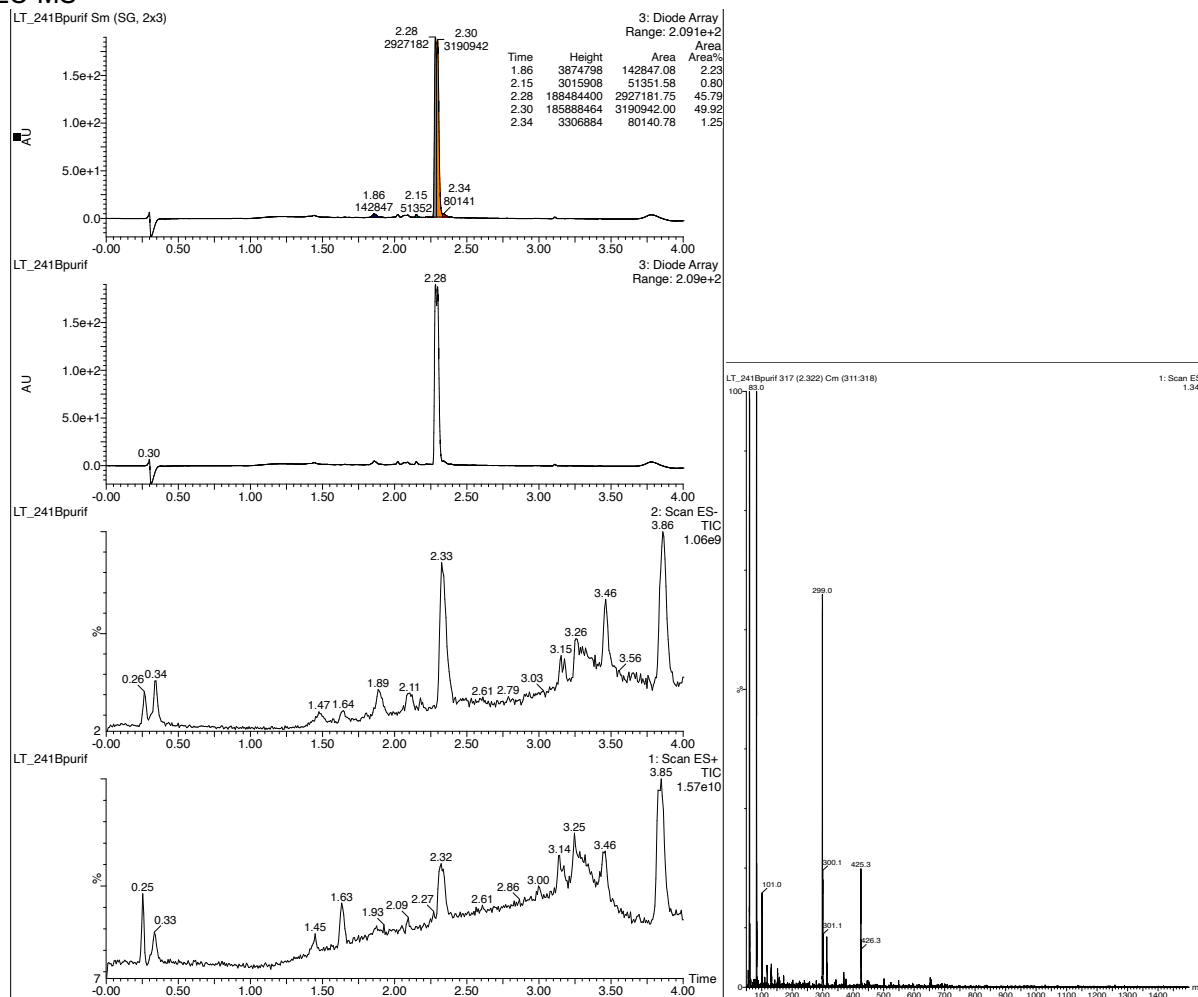
UPLC-MS



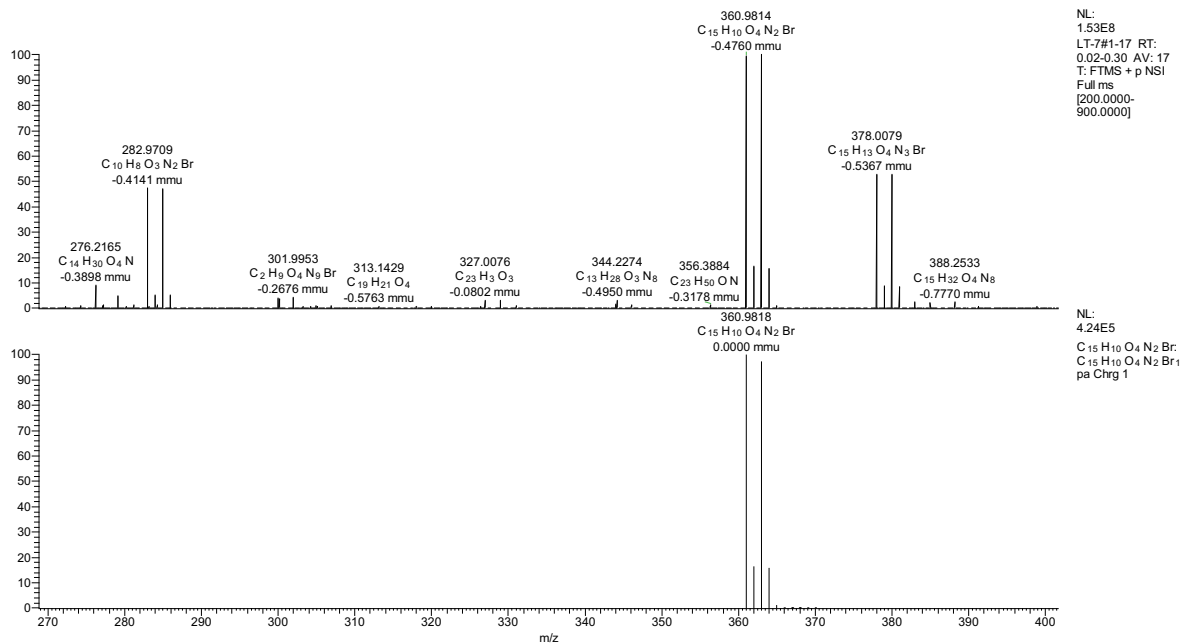
Compound 6k



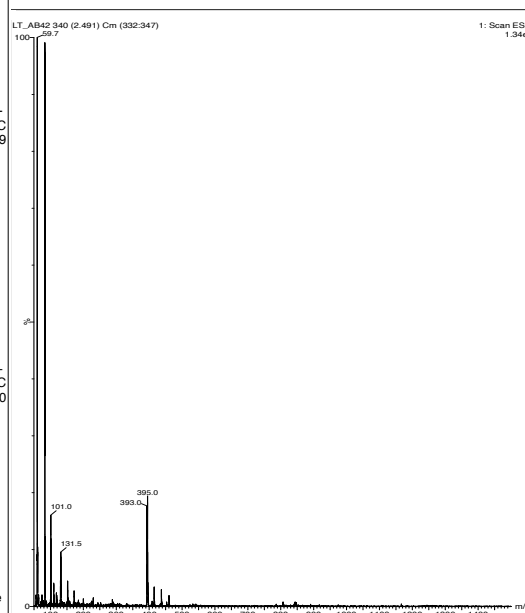
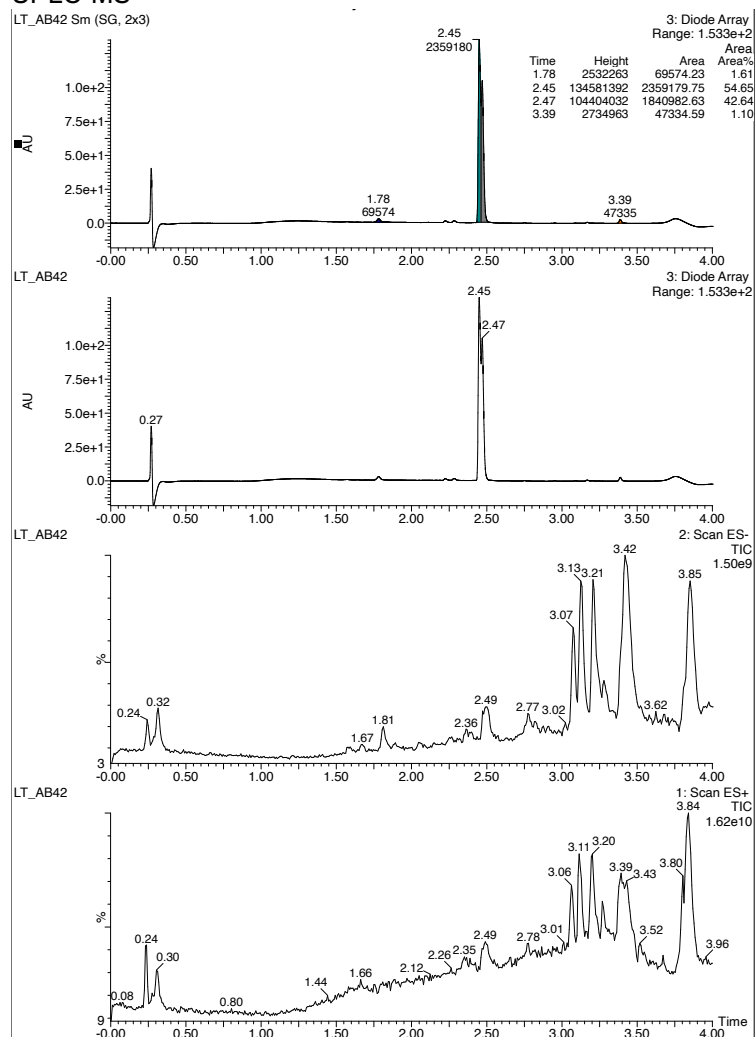
UPLC-MS



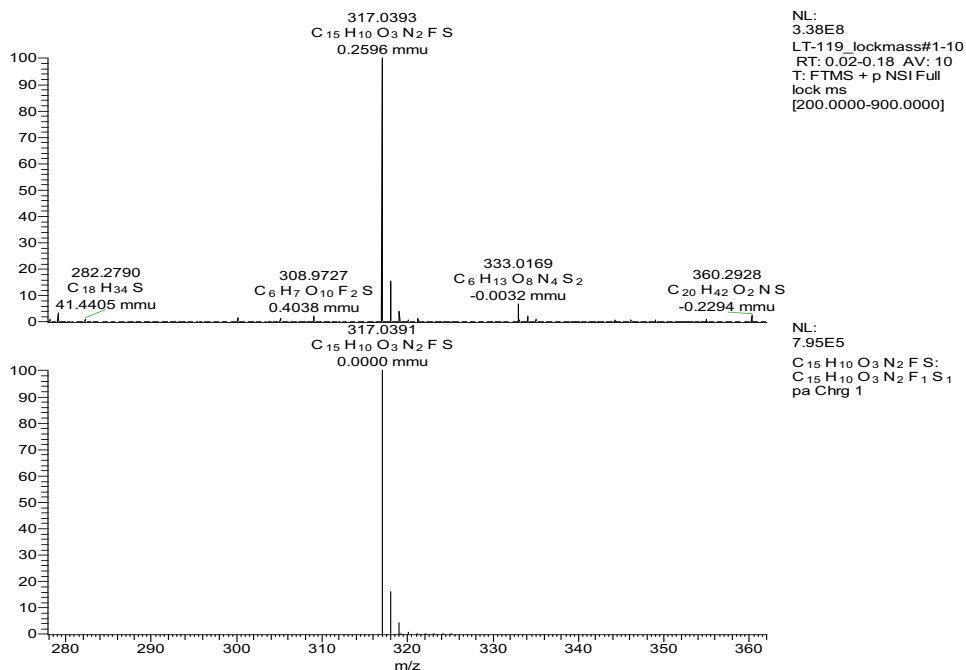
Compound 61



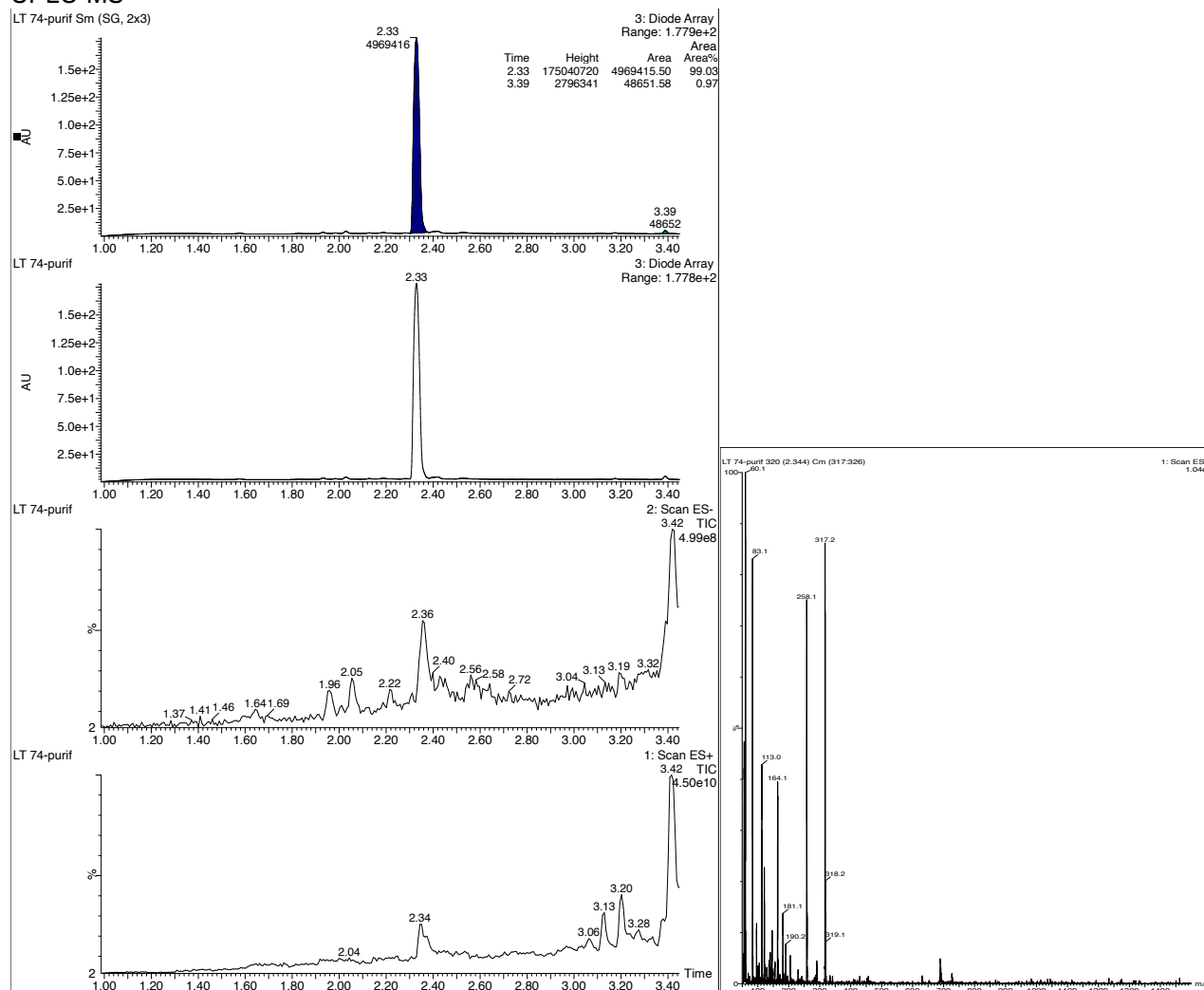
UPLC-MS



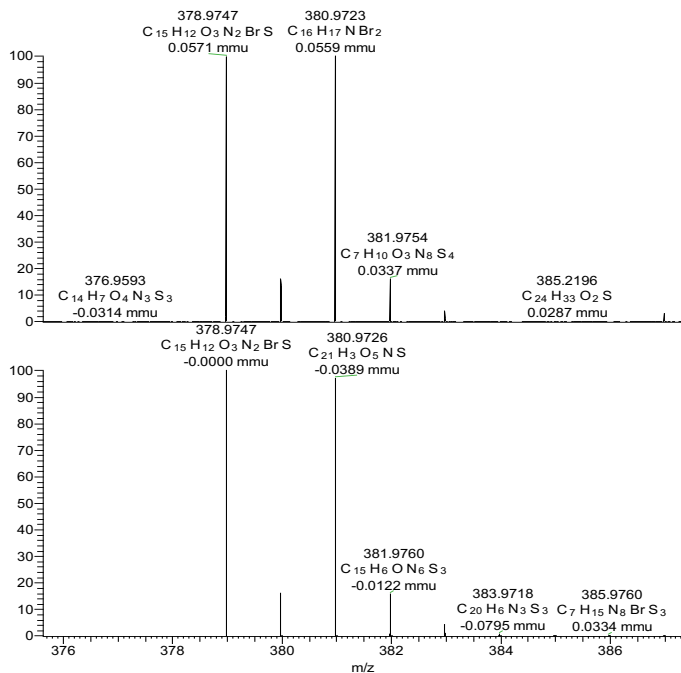
Compound 6m



UPLC-MS



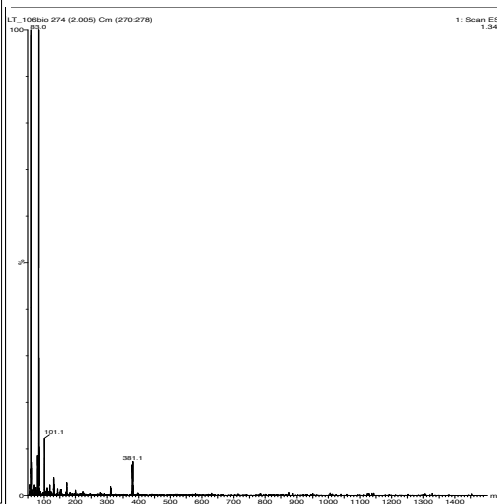
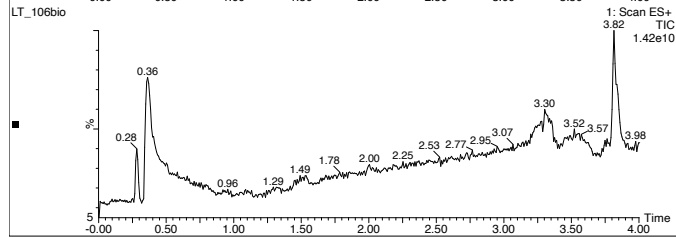
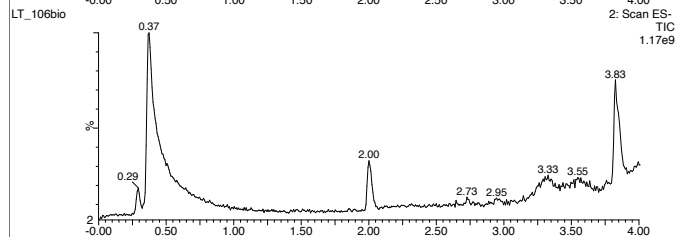
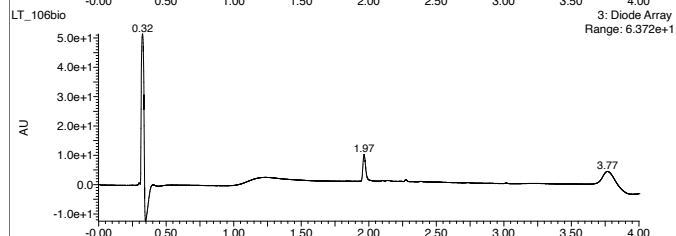
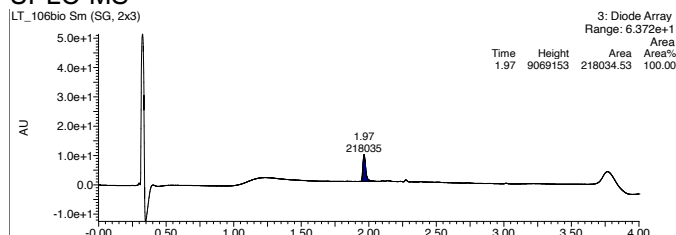
Compound 7a



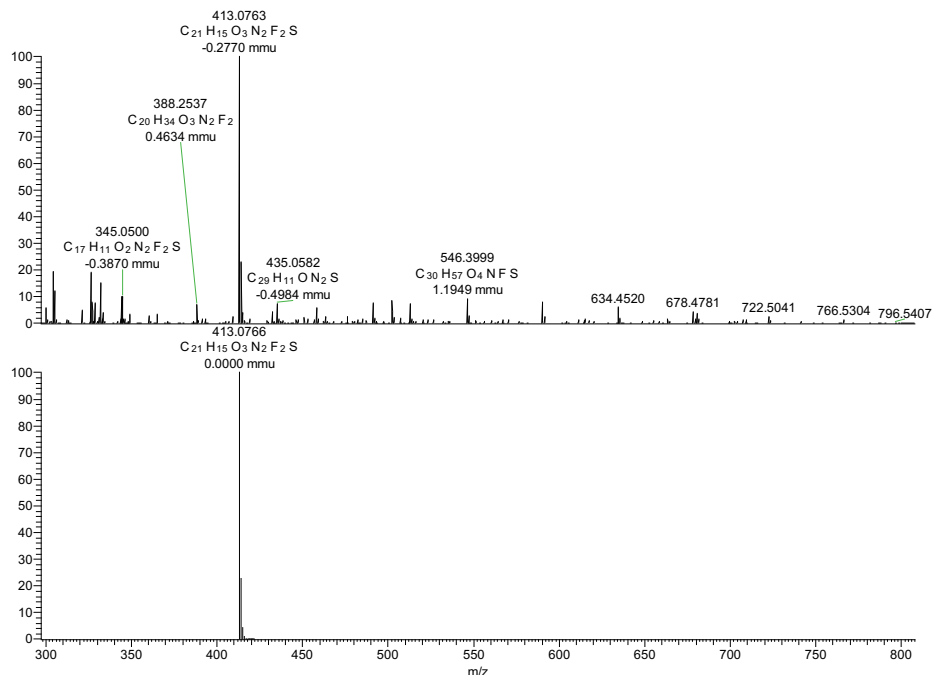
NL:
4.18E7
106_pos#1-29 RT:
0.02-0.51 AV: 29 T:
FTMS + p NSI Full ms
[300.0000-800.0000]

NL:
4.03E5
C₁₅H₁₂O₃N₂BrS:
C₁₅H₁₂O₃N₂Br₁S₁
pa Chrg 1

UPLC-MS



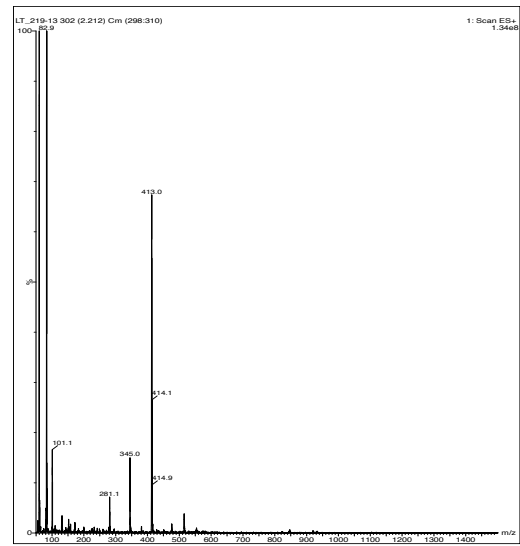
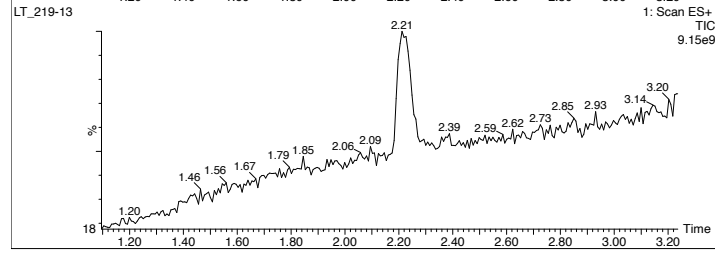
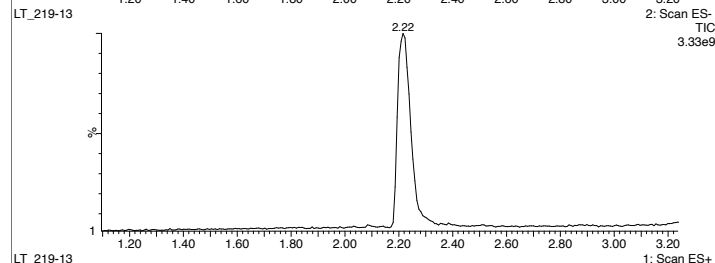
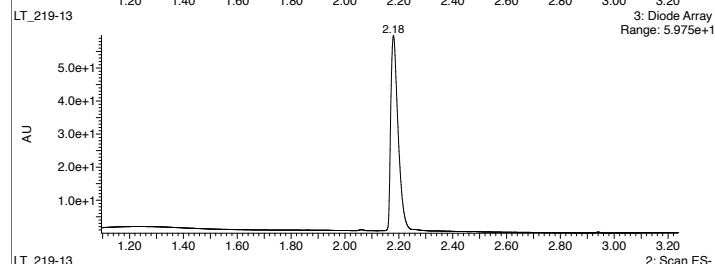
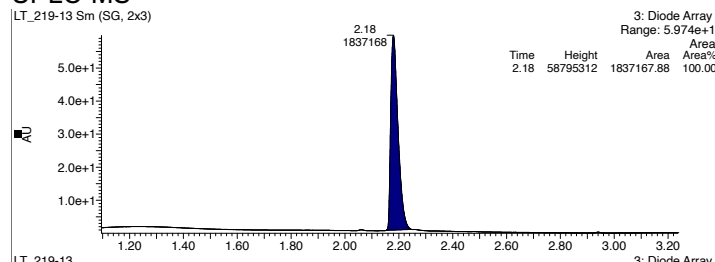
Compound 7b



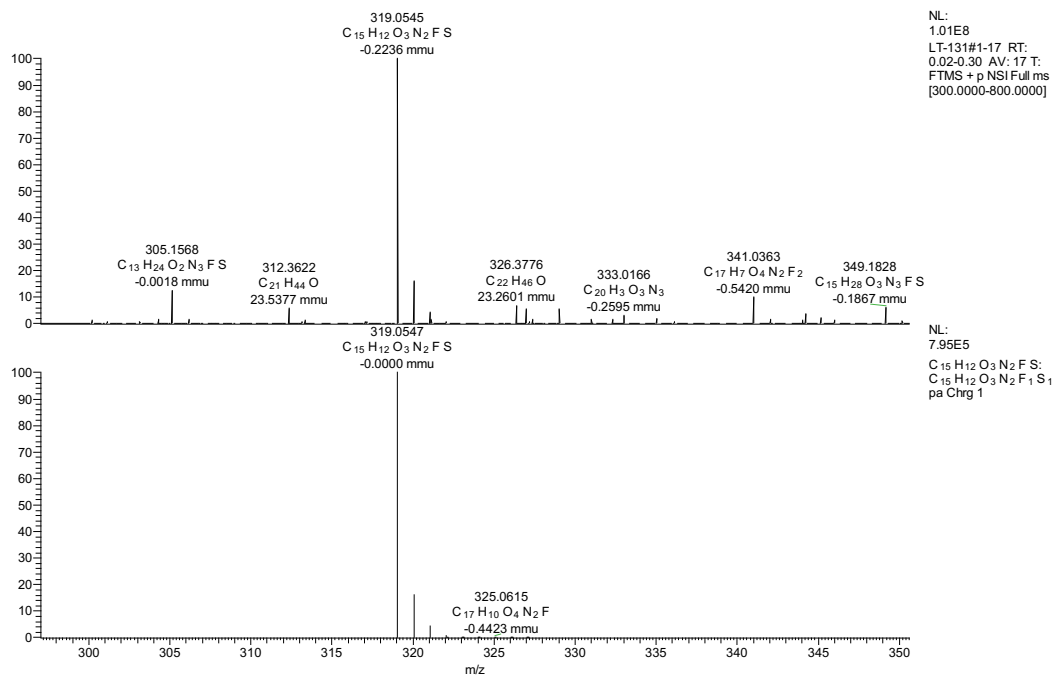
NL:
8.85E7
LT-130#1-17 RT:
0.02-0.30 AV: 17 T:
FTMS + p NSI Full ms
[300.0000-800.0000]

NL:
7.45E5
C₂₁H₁₅O₃N₂F₂S:
C₂₁H₁₅O₃N₂F₂S 1
pa Chrg 1

UPLC-MS



Compound 7c



UPLC-MS

