Supplemental Information

GENOMIC LANDSCAPE OF PATIENTS WITH *FLT3*-MUTATED ACUTE MYELOID LEUKEMIA (AML) TREATED WITHIN THE CALGB 10603/RATIFY TRIAL

AUTHORS

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SUPPLEMENTAL METHODS

Library Enrichment and Sequencing

The library enrichment was performed using the commercially available SureSelectXT in-solution capture technology from Agilent Technologies (Santa Clara, CA, USA). The library comprised the entire coding region of 262 candidate genes involved in hematological malignancies including 20 kinases targeted by midostaurin. For library design Agilent's online tool SureDesign was used. Only exons with a consensus annotation in the RefSeq, Ensembl, CCDS, Gencode, and SNP databases were considered relevant. The UCSC human genome 19 (H. sapiens, hg19, GRCh37, February 2009) served as reference genome for design and determination of genomic coordinates.

Genomic DNA (200ng per sample) extracted from pre-treatment bone marrow (409, 86%) or peripheral blood (66, 14%) specimens was used for molecular screening. SureSelect library preparation and indexing were performed following the manufacturer's instructions for Illumina paired-end sequencing. Samples were then transferred to a cBot (Illumina, San Diego, CA, USA) to create clonal clusters on a flow cell by bridge amplification (Illumina reagent kit: TruSeq PE Cluster Kit v3-cBot-HS). Finally, 2x 100 bp paired-end sequencing by synthesis was carried out on a HiSeq2000 (Illumina, San Diego, CA, USA) using Illumina's TruSeq SBS Kit v3-HS reagents.

Variant calling

The sequencing quality of each sample was assessed using the NGS QC toolkit (2.3.3) and, where necessary, adapter and read end trimming were performed using cutadapt (1.8.3) and in-house scripting respectively.

Paired-end reads were then aligned to the hg19 reference using BWA-MEM (0.7.10). Alignments are sorted and indexed by Picard (1.138) and locally realigned using GATK (3.4.46). For each sample, coverage statistics were calculated using BEDTools (2.24.0) and processed by SAMtools (0.1.19). VarScan2 (2.3.9) was then used for variant calling within the target regions sequenced. All variants were annotated by Annovar (release 22Mar2015) but only non-synonymous mutations affecting exons or splice sites were retained. These were further filtered to remove calls within known regions of segmental duplication, variants annotated in dbSNP (138) but not COSMIC (70) and variants with a minor allele frequency (MAF) above 0.01 in either the 1000 Genomes Project or the Exome Sequencing Project (ESP 6500).

Curation of oncogenic variants

All calls yielded by the computational annotation workflow were subject to further curation. Only variants considered oncogenic were included in the subsequent analyses. The algorithm for mutation reporting is as follows:

 a) Removal of all variants that are annotated in SNP databases and occur with a minor allele frequency (MAF) >0.001 in the 1000 Genomes Project, dbSNP150 or the Exome Sequencing Project (ESP 6500).

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- b) Removal of variants present within regions prone to sequence context specific artifacts, including regions of high depth, enriched for reads of low mapping quality that harbor multiple mismatches
- c) Removal of all one bp insertions or deletions present adjacent to regions of more than 5 homopolymer bases (for example insG adjacent to GGGGG) and a variant allele frequency of ≤0.1
- d) Removal of all missense variants with a variant allele frequency between ≥0.45 and ≤0.55 or ≥0.9 and
 1.0, indicative of polymorphisms, unless they are present with ≥5 counts in COSMIC database (v85) and with ≥1 confirmed somatic.
- e) Retention of all frameshift, nonsense or splicing variants with a variant allele frequency ≥0.03
- f) Retention of all missense variants with a variant allele frequency between 0.03 and <0.45 or >0.55 and <0.9, indicative of (likely) oncogenic variants.</p>

SUPPLEMENTAL TABLES

Supplemental Table S1: Comparison of clinical characteristics of patients included and excluded into this analysis of entire CALGB 10603/RATIFY trial cohort (N=717).

Characteristic	Excluded, N = 242 ¹	Included, N = 475 ¹	p-value ²
Age at registration	48 (40, 53)	48 (39, 54)	1
WBC count at baseline (10E9/L)	34 (13, 89)	35 (12, 72)	0.5
Unknown	7	3	
Sex			0.6
Male	111 (46%)	208 (44%)	
Female	131 (54%)	267 (56%)	
ECOG performance category			0.5
0-1	211 (87%)	422 (89%)	
2	31 (13%)	53 (11%)	
FLT3 mutation type			0.073
ТКД	47 (20%)	116 (24%)	
ITD allelic ratio <0.5	82 (34%)	126 (27%)	
ITD allelic ratio >=0.5	109 (46%)	232 (49%)	
Unknown	4	1	
Treatment			0.022
Midostaurin	107 (44%)	253 (53%)	
Placebo	135 (56%)	222 (47%)	
¹ Median (IQR); n (%)			

² Wilcoxon rank sum test; Pearson's Chi-squared test

Supplemental Table S2: List of all genes targeted by custom sequencing panel

ABCA12	CDKN2B	DNMT3B	GNB1	MAP3K4	NXF1	RASGRF1	STAG1
ABL1	CDKN2C	DYNC1H1	H3F3A	MAP3K9	OBSCN	RB1	STAG2
ACIN1	CEBPA	EED	H3F3B	MGA	OMG	RBBP5	STAT3
ACSS3	CHEK2	EEFSEC	HAX1	MLL3/KMT2C	PAX5	RBBP6	STAT5A
ADGRV1	CLTCL1	EGFR	HCN1	MLL5/KMT2E	PDGFB	RBMX	SUZ12
ALK	CNNM2	ELANE	НІРК2	MN1	PDGFRB	RET	SYNE1
ANKRD26	COPRS	EP300	HNRNPK	MPL	PDPK1	RHOA	TCIRG1
ARHGEF10	CREBBP	EPHA6	HRAS	MST1	PHF6	RMI1	TERC
ARID1A	CSF1R	ETNK1	IDH1	МҮС	PHIP	ROBO1	TERT
ARID2	CSF2RB	ETV6	IDH2	MYH9	PHKG1	ROBO2	TET1
AS3MT	CSF3R	EVI2A	IKZF2	MYLK2	РІКЗСА	RPS6KA2	TET2
ASXL1	CSMD1	EVI2B	INPP5D	MYO1F	PKN2	RPS6KA3	TINF2
ASXL2	CSMD2	EWSR1	IRF1	NCOA7	PLEKHH1	RPS6KA6	TNK1
ATRX	CSNK1A1	EZH1	IRF4	NDE1	PLEKHS1	RRAS	TNK2
BAP1	CTC1	EZH2	IRF8	NEK2	PPM1D	RUNX1	TP53

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Genomic Landscape of Patients with *FLT3*-mutated AML CALGB 10603/RATIFY Trial Supplemental Information

					Ju	spicificitia	mormation
BCL10	CTCF	FAM175A	JAK1	NF1	PRKAG2	RYR2	TTC39A
BCL2	CTNNB1	FAM5C	JAK2	NF2	PRKG2	SAMHD1	U2AF1
BCOR	CUX1	FAT4	JAK3	NFE2	PRPF40A	SETBP1	U2AF2
BCORL1	DCC	FBXW7	JARID2	NFE2L1	PRPF40B	SETD2	UBQLN1
BCR	DDX23	FGFR2	KAT6A	NFE2L2	PRPF8	SETDB1	UBXN11
BRAF	DDX4	FLG	KDM5C	NIPBL	PTEN	SF1	WAC
BRCC3	DDX41	FLT3	KDM6A	NOTCH1	PTPN11	SF3A1	WHSC1
С6	DDX54	FOXP1	KDR	NOTCH2	PTPRF	SF3B1	WRAP53
C9orf103	DHX15	FRMD3	KIF27	NPM1	PTPRT	SH2B3	WT1
CALR	DHX33	G6PC3	KIT	NRAS	PXDN	SMARCB1	YLPM1
CBL	DICER1	GALNT11	KMT2A	NRXN1	RAB11FIP4	SMC1A	ZBTB33
CCDC26	DIS3	GALNTL5	KMT2D	NRXN3	RAC1	SMC3	ZBTB7A
CCND1	DKC1	GATA1	KRAS	NSD1	RAD21	SMG1	ZMYM3
CCND2	DNAH9	GATA2	LAMA1	NT5C2	RAD50	SPI1	ZNF318
CDHR1	DNAJB8	GFI1	LAMC3	NTRK1	RAD51	SPRED2	ZNF687
CDK4	DND1	GIGYF2	LUC7L2	NTRK3	RASA2	SRCAP	ZRSR2
CDKN1B	DNM2	GKAP1	MAP3K10	NUMA1	RASA3	SRP72	
CDKN2A	DNMT3A	GNAS	MAP3K11	NUP98	RASEF	SRSF2	

Supplemental Table S3: Genes targeted by midostaurin (Midostaurin kinome)

JAK3	KDR	KIT	MAP3K10	MAP3K11	МАРЗК9	MST1	NTRK1	NTRK3	PDGFRB
PDPK1	PHKG1	PKN2	PRKG2	RET	RPS6KA2	RPS6KA3	RPS6KA6	TNK1	TNK2

Supplemental Table S4: Frequency of gene mutations overall and by FLT3 mutational subgroups

Characteristic	Overall, N = 475 ¹	TKD, N = 116 ¹	ITD, N = 359 ¹	p ²
NPM1	291 (61%)	71 (61%)	220 (61%)	1.00
DNMT3A	187 (39%)	44 (38%)	143 (40%)	0.74
WT1	100 (21%)	16 (14%)	84 (23%)	0.03
TET2	55 (12%)	10 (8.6%)	45 (13%)	0.32
RUNX1	53 (11%)	10 (8.6%)	43 (12%)	0.40
NRAS	53 (11%)	28 (24%)	25 (7.0%)	<0.001
PTPN11	45 (9.5%)	16 (14%)	29 (8.1%)	0.10
IDH1	39 (8.2%)	9 (7.8%)	30 (8.4%)	1.00
ASXL1	38 (8.0%)	5 (4.3%)	33 (9.2%)	0.12
<i>IDH2</i> (R140)	34 (7.2%)	7 (6.0%)	27 (7.5%)	0.68
SMC1A	28 (5.9%)	12 (10%)	16 (4.5%)	0.04
CEBPA	26 (5.5%)	2 (1.7%)	24 (6.7%)	0.06
SMC3	24 (5.1%)	9 (7.8%)	15 (4.2%)	0.14
RAD21	23 (4.8%)	8 (6.9%)	15 (4.2%)	0.22
BCOR	20 (4.2%)	3 (2.6%)	17 (4.7%)	0.43
KMT2D	18 (3.8%)	1 (0.9%)	17 (4.7%)	0.09
STAG2	18 (3.8%)	6 (5.2%)	12 (3.3%)	0.40
BCORL1	16 (3.4%)	5 (4.3%)	11 (3.1%)	0.56

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			Suppleme	ntal mormation
ZBTB7A	15 (3.2%)	6 (5.2%)	9 (2.5%)	0.22
МҮС	15 (3.2%)	5 (4.3%)	10 (2.8%)	0.38
GATA2	14 (2.9%)	2 (1.7%)	12 (3.3%)	0.53
OBSCN	14 (2.9%)	3 (2.6%)	11 (3.1%)	1.00
KRAS	13 (2.7%)	6 (5.2%)	7 (1.9%)	0.10
NFE2	13 (2.7%)	1 (0.9%)	12 (3.3%)	0.20
CSMD1	11 (2.3%)	3 (2.6%)	8 (2.2%)	0.73
KDM6A	11 (2.3%)	4 (3.4%)	7 (1.9%)	0.48
MGA	11 (2.3%)	3 (2.6%)	8 (2.2%)	0.73
SETD2	10 (2.1%)	2 (1.7%)	8 (2.2%)	1.00
SYNE1	10 (2.1%)	1 (0.9%)	9 (2.5%)	0.46
CREBBP	10 (2.1%)	4 (3.4%)	6 (1.7%)	0.27
ARID2	9 (1.9%)	4 (3.4%)	5 (1.4%)	0.23
MN1	9 (1.9%)	4 (3.4%)	5 (1.4%)	0.23
SF3B1	9 (1.9%)	2 (1.7%)	7 (1.9%)	1.00
SRSF2	8 (1.7%)	3 (2.6%)	5 (1.4%)	0.41
KIT	8 (1.7%)	5 (4.3%)	3 (0.8%)	0.02
ZBTB33	7 (1.5%)	2 (1.7%)	5 (1.4%)	0.68
HNRNPK	7 (1.5%)	3 (2.6%)	4 (1.1%)	0.37
NF1	7 (1.5%)	1 (0.9%)	6 (1.7%)	1.00
PTPRF	6 (1.3%)	0 (0%)	6 (1.7%)	0.34
ZNF318	6 (1.3%)	1 (0.9%)	5 (1.4%)	1.00
NOTCH1	6 (1.3%)	2 (1.7%)	4 (1.1%)	0.64
PHF6	6 (1.3%)	0 (0%)	6 (1.7%)	0.34
CSMD2	6 (1.3%)	4 (3.4%)	2 (0.6%)	0.03
CBL	6 (1.3%)	0 (0%)	6 (1.7%)	0.34
DNAH9	6 (1.3%)	2 (1.7%)	4 (1.1%)	0.64
GNAS	6 (1.3%)	1 (0.9%)	5 (1.4%)	1.00
KMT2A	6 (1.3%)	1 (0.9%)	5 (1.4%)	1.00
FOXP1	6 (1.3%)	4 (3.4%)	2 (0.6%)	0.03
CSF3R	6 (1.3%)	1 (0.9%)	5 (1.4%)	1.00
EP300	6 (1.3%)	1 (0.9%)	5 (1.4%)	1.00
ACIN1	5 (1.1%)	0 (0%)	5 (1.4%)	0.34
RYR2	5 (1.1%)	0 (0%)	5 (1.4%)	0.34
ARID1A	5 (1.1%)	0 (0%)	5 (1.4%)	0.34
ASXL2	5 (1.1%)	1 (0.9%)	4 (1.1%)	1.00
LAMC3	5 (1.1%)	2 (1.7%)	3 (0.8%)	0.60
UBXN11	5 (1.1%)	0 (0%)	5 (1.4%)	0.34
ROBO1	5 (1.1%)	1 (0.9%)	4 (1.1%)	1.00
GPR98	5 (1.1%)	1 (0.9%)	4 (1.1%)	1.00
SRCAP	5 (1.1%)	2 (1.7%)	3 (0.8%)	0.60
EZH2	5 (1.1%)	1 (0.9%)	4 (1.1%)	1.00
FBXW7	5 (1.1%)	2 (1.7%)	3 (0.8%)	0.60
BCR	4 (0.8%)	1 (0.9%)	3 (0.8%)	
MYH9	4 (0.8%)	2 (1.7%)	2 (0.6%)	
ETV6	4 (0.8%)	1 (0.9%)	3 (0.8%)	
DNMT3B	4 (0.8%)	1 (0.9%)	3 (0.8%)	
LAMA1	4 (0.8%)	2 (1.7%)	2 (0.6%)	
U2AF1	4 (0.8%)	1 (0.9%)	3 (0.8%)	
NTRK3	4 (0.8%)	0 (0%)	4 (1.1%)	
CTCF	4 (0.8%)	0 (0%)	4 (1.1%)	
MAP3K11	4 (0.8%)	1 (0.9%)	3 (0.8%)	

Genomic Landscape of Patients with *FLT3*-mutated AML CALGB 10603/RATIFY Trial

			Supplemen	ntal Information
CNNM2	4 (0.8%)	0 (0%)	4 (1.1%)	
NSD1	4 (0.8%)	2 (1.7%)	2 (0.6%)	
SH2B3	4 (0.8%)	0 (0%)	4 (1.1%)	
KMT2E	4 (0.8%)	1 (0.9%)	3 (0.8%)	
KDM5C	4 (0.8%)	2 (1.7%)	2 (0.6%)	
BRCC3	4 (0.8%)	0 (0%)	4 (1.1%)	
KMT2C	4 (0.8%)	1 (0.9%)	3 (0.8%)	
CUX1	4 (0.8%)	3 (2.6%)	1 (0.3%)	
TERT	4 (0.8%)	0 (0%)	4 (1.1%)	
EPHA6	4 (0.8%)	1 (0.9%)	3 (0.8%)	
FAT4	4 (0.8%)	0 (0%)	4 (1.1%)	
INPP5D	4 (0.8%)	1 (0.9%)	3 (0.8%)	
TP53	4 (0.8%)	2 (0.3%)	2 (0.6%)	
ROBO2	3 (0.6%)	1 (0.9%)	2 (0.6%)	
SETRD1	3 (0.6%)	1 (0.9%)	2 (0.6%)	
	3 (0.6%)	0.0%)	2 (0.0%)	
CDTIN1 SE2A1	2 (0.6%)	1 (0.0%)	2 (0.6%)	
	3 (0.0%) 3 (0.6%)	I (0.9%)	2 (0.0%)	
	3 (0.0%)	0 (0%)	2 (0.0%)	
SEIDBI	3 (0.6%)	0 (0%)	3 (0.8%)	
EGFK	3 (0.6%)	2 (1.7%)	1 (0.3%)	
KBBP6	3 (0.6%)	2 (1.7%)	1 (0.3%)	
AIRX	3 (0.6%)	1 (0.9%)	2 (0.6%)	
PDGFRB	3 (0.6%)	1 (0.9%)	2 (0.6%)	
RPS6KA6	3 (0.6%)	2 (1.7%)	1 (0.3%)	
NTRK1	3 (0.6%)	0 (0%)	3 (0.8%)	
YLPM1	3 (0.6%)	0 (0%)	3 (0.8%)	
RRAS	3 (0.6%)	1 (0.9%)	2 (0.6%)	
KAT6A	3 (0.6%)	3 (2.6%)	0 (0%)	
PPM1D	3 (0.6%)	2 (1.7%)	1 (0.3%)	
НІРК2	3 (0.6%)	0 (0%)	3 (0.8%)	
MAP3K10	2 (0.4%)	0 (0%)	2 (0.6%)	
TCIRG1	2 (0.4%)	0 (0%)	2 (0.6%)	
MPL	2 (0.4%)	1 (0.9%)	1 (0.3%)	
DDX41	2 (0.4%)	0 (0%)	2 (0.6%)	
RAD50	2 (0.4%)	0 (0%)	2 (0.6%)	
WRAP53	2 (0.4%)	0 (0%)	2 (0.6%)	
CCND2	2 (0.4%)	1 (0.9%)	1 (0.3%)	
CSF1R	2 (0.4%)	0 (0%)	2 (0.6%)	
CSF2RB	2 (0.4%)	1 (0.9%)	1 (0.3%)	
DHX33	2 (0.4%)	0 (0%)	2 (0.6%)	
WHSC1	2 (0.4%)	1 (0.9%)	1 (0.3%)	
RPS6KA2	2 (0.4%)	1 (0.9%)	1 (0.3%)	
MAP3K9	2 (0.4%)	1 (0.9%)	1 (0.3%)	
FIG	2 (0.4%)	2 (1.7%)	0 (0%)	
ABCA12	2 (0.4%)	0 (0%)	2 (0.6%)	
CTC1	2 (0.4%)	0 (0%)	2 (0.6%)	
HCN1	2 (0.4%)	1 (0.9%)	1 (0 3%)	
IARID2	2 (0.4%)	0 (0%)	2 (0.6%)	
	2 (0.4%)	0 (0%)	2 (0.6%)	
SF1	2 (0.4%)	0 (0%)	2 (0.6%)	
	2 (0.4%)		1 (0.2%)	
PDGER	2 (0.470)	2 (1 7%)	L (0.378)	
	2 (0.470)	~ (1.770)	0 (0/0)	

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			Supplemen	tal information
RASA2	2 (0.4%)	0 (0%)	2 (0.6%)	
GATA1	2 (0.4%)	0 (0%)	2 (0.6%)	
TNK2	2 (0.4%)	0 (0%)	2 (0.6%)	
MAP3K4	2 (0.4%)	0 (0%)	2 (0.6%)	
JAK3	2 (0.4%)	1 (0.9%)	1 (0.3%)	
PRPF40B	2 (0.4%)	1 (0.9%)	1 (0.3%)	
DCC	2 (0.4%)	0 (0%)	2 (0.6%)	
MYO1F	2 (0.4%)	1 (0.9%)	1 (0.3%)	
DIS3	2 (0.4%)	0 (0%)	2 (0.6%)	
TINF2	2 (0.4%)	0 (0%)	2 (0.6%)	
ZMYM3	2 (0.4%)	0 (0%)	2 (0.6%)	
ALK	2 (0.4%)	1 (0.9%)	1 (0.3%)	
NUP98	2 (0.4%)	1 (0.9%)	1 (0.3%)	
RMI1	2 (0.4%)	0 (0%)	2 (0.6%)	
GFI1	2 (0.4%)	0 (0%)	2 (0.6%)	
DYNC1H1	1 (0.2%)	1 (0.9%)	0 (0%)	
PTPRT	1 (0.2%)	0 (0%)	1 (0.3%)	
RET	1 (0.2%)	0 (0%)	1 (0.3%)	
DDX54	1 (0.2%)	1 (0.9%)	0 (0%)	
ACSS3	1 (0.2%)	0 (0%)	1 (0.3%)	
EVI2B	1 (0.2%)	1 (0.9%)	0 (0%)	
JAK1	1 (0.2%)	0 (0%)	1 (0.3%)	
KDR	1 (0.2%)	0 (0%)	1 (0.3%)	
DHX15	1 (0.2%)	0 (0%)	1 (0.3%)	
STAT5A	1 (0.2%)	1 (0.9%)	0 (0%)	
BRINP3	1 (0.2%)	1 (0.9%)	0 (0%)	
BRAF	1 (0.2%)	1 (0.9%)	0 (0%)	
IRF4	1 (0.2%)	1 (0.9%)	0 (0%)	
COPRS	1 (0.2%)	0 (0%)	1 (0.3%)	
PAX5	1 (0.2%)	0 (0%)	1 (0.3%)	
PHIP	1 (0.2%)	0 (0%)	1 (0.3%)	
RPS6KA3	1 (0.2%)	0 (0%)	1 (0.3%)	
SMG1	1 (0.2%)	0 (0%)	1 (0.3%)	
EEFSEC	1 (0.2%)	1 (0.9%)	0 (0%)	
С6	1 (0.2%)	0 (0%)	1 (0.3%)	
NFE2L1	1 (0.2%)	0 (0%)	1 (0.3%)	
RASEF	1 (0.2%)	0 (0%)	1 (0.3%)	
RASA3	1 (0.2%)	0 (0%)	1 (0.3%)	
HRAS	1 (0.2%)	0 (0%)	1 (0.3%)	
MYLK2	1 (0.2%)	0 (0%)	1 (0.3%)	
RAB11FIP4	1 (0.2%)	0 (0%)	1 (0.3%)	
DICER1	1 (0.2%)	1 (0.9%)	0 (0%)	
TET1	1 (0.2%)	0 (0%)	1 (0.3%)	
SPRED2	1 (0.2%)	0 (0%)	1 (0.3%)	
GALNT11	1 (0.2%)	0 (0%)	1 (0.3%)	
ZRSR2	1 (0.2%)	0 (0%)	1 (0.3%)	
PRKAG2	1 (0.2%)	0 (0%)	1 (0.3%)	
JAK2	1 (0.2%)	0 (0%)	1 (0.3%)	
BAP1	1 (0.2%)	1 (0.9%)	0 (0%)	
NF2	1 (0.2%)	0 (0%)	1 (0.3%)	
RASGRF1	1 (0.2%)	0 (0%)	1 (0.3%)	
TNK1	1 (0.2%)	1 (0.9%)	0 (0%)	

			Supplemen	
IDNK	1 (0.2%)	0 (0%)	1 (0.3%)	
INSRR.NTRK1	1 (0.2%)	1 (0.9%)	0 (0%)	
NRXN1	1 (0.2%)	0 (0%)	1 (0.3%)	
NXF1	1 (0.2%)	0 (0%)	1 (0.3%)	
ANKRD26	1 (0.2%)	0 (0%)	1 (0.3%)	
BCL10	1 (0.2%)	0 (0%)	1 (0.3%)	
PRPF40A	1 (0.2%)	0 (0%)	1 (0.3%)	
SUZ12	1 (0.2%)	0 (0%)	1 (0.3%)	
PTEN	1 (0.2%)	0 (0%)	1 (0.3%)	
RAD51	1 (0.2%)	0 (0%)	1 (0.3%)	
PRPF8	1 (0.2%)	1 (0.9%)	0 (0%)	
PLEKHS1	1 (0.2%)	1 (0.9%)	0 (0%)	
SPI1	1 (0.2%)	1 (0.9%)	0 (0%)	
GALNTL5	1 (0.2%)	1 (0.9%)	0 (0%)	
SAMHD1	1 (0.2%)	0 (0%)	1 (0.3%)	
NRXN3	1 (0.2%)	1 (0.9%)	0 (0%)	
HAX1	1 (0.2%)	0 (0%)	1 (0.3%)	
TTC39A	1 (0.2%)	0 (0%)	1 (0.3%)	
DKC1	1 (0.2%)	0 (0%)	1 (0.3%)	
PHKG1	1 (0.2%)	1 (0.9%)	0 (0%)	
WAC	1 (0.2%)	0 (0%)	1 (0.3%)	
NUMA1	1 (0.2%)	0 (0%)	1 (0.3%)	
FGFR2	1 (0.2%)	0 (0%)	1 (0.3%)	
GIGYF2	1 (0.2%)	0 (0%)	1 (0.3%)	
RBBP5	1 (0.2%)	0 (0%)	1 (0.3%)	
NCOA7	1 (0.2%)	0 (0%)	1 (0.3%)	
$^{1}n(\%)$				

¹ n (%)

² Fisher's exact test (p-value not adjusted for multiple testing)

Supplemental Table S5: Functional categorization of recurrently mutated genes (>1%).

Methylation	Chromatin	Cohesin	Splicing	Signaling	Transcription	Other	Tumor suppressor
DNMT3A	ACIN1	RAD21	HNRNPK	CBL	ARID1A	CSMD2	CSMD1
IDH1	ARID2	SMC1A	SF1	CSF3R	CEBPA	DNAH9	TP53
IDH2	ASXL1	SMC3	SF3A1	FLT3	ETV6	FBXW7	
TET2	ASXL2	STAG2	SF3B1	GNAS	FOXP1	LAMC3	
	BCOR		SRSF2	GPR98	GATA2	NPM1	
	BCORL1		U2AF1	KIT	MGA	OBSCN	
	CREBBP		ZRSR2	KRAS	MN1	ROBO1	
	EP300			NF1	МҮС	RYR2	
	EZH2			NOTCH1	NFE2	SYNE1	
	KDM6A			NRAS	PHF6		
	KMT2A			PTPN11	RUNX1		
	KMT2D			PTPRF	WT1		
	KMT2E			UBXN11	ZBTB33		
	KMT2C				ZBTB7A		
	SETD2				ZNF318		
	SRCAP						

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Supplemental Table S6: Genomic classes according to Papaemmanuil E, Gerstung M et al NEJM 2016.

Genomic subgroup (long)	Genomic subgroup (short)
AML with inv(16)(p13.1q22) or t(16;16)(p13.1;q22); CBFB–MYH11	inv(16)
AML with t(8;21)(q22;q22); RUNX1–RUNX1T1	t(8;21)
AML with MLL fusion genes; t(x;11)(x;q23)	t(11q23;x)
AML with inv(3)(q21q26.2) or t(3;3)(q21;q26.2); GATA2, MECOM(EVI1)	inv(3)
AML with t(6;9)(p23;q34); DEK–NUP214	t(6;9)
AML with t(15;17)(q22;q12); PML–RARA	t(15;17)
AML with NPM1 mutation	NPM1
AML with biallelic CEBPA mutations*	CEBPA ^{biallelic}
AML with TP53 mutations, chromosomal aneuploidy, or both [#]	TP53-aneuploidy
AML with mutated chromatin, RNA-splicing genes, or both $^{\scriptscriptstyle +}$	Chromatin-Spliceosome
AML with IDH2R172 mutations and no other class-defining lesions	IDH2 ^{R172}
AML meeting criteria for ≥2 genomic subgroups	2 classes
AML with driver mutations but no detected class-defining lesions	No class
AML with no detected driver mutations	No drivers detected

* Patients with two different mutations in CEBPA

Classification in this subgroup requires *TP53* mutation, complex karyotype [3 or more abnormalities, in the absence of 1 of the WHO-designated recurring translocations or inversions, that is, t(8;21), inv(16) or t(16;16), t(9;11),t(v;11)(v;q23.3), t(6;9), inv(3) or t(3;3); AML with t(9;22)], or in the absence of other class-defining lesions, one or more of the following: -7/7q, -5/5q, -4/4q, -9q, -12/12p, -17/-17p, -18/18q, -20/20q, +11/11q, +13, +21, or +22.

+ Classification in this subgroup requires one or more driver mutations in *RUNX1*, *ASXL1*, *BCOR*, *STAG2*, *EZH2*, *SRSF2*, *SF3B1*, *U2AF1*, *ZRSR2*, or *MLL*-PTD. In the presence of other class-defining lesions — namely, inv(16), t(15;17), t(8;21), t(6;9), inv(3), MLL fusion genes, or complex karyotype or driver mutations in *TP53*, *NPM1*, or *CEBPA*^{biallelic} — two or more chromatin–spliceosome mutations are required.

Supplemental Table S7: Baseline characteristics of genomic AML classes in the cohort of 451 of 475 patients, in which subcategorization into genomic AML classes was possible.

	<i>NPM1</i> (N=287)	CBF (N=18)	TP53- aneuploidy (N=18)	Chromatin- spliceosome (N=68)	No Class (N=60)	Overall (N=451)
Age (years)						
Mean (SD)	46.9 (10.1)	44.3 (10.4)	46.3 (9.80)	47.2 (10.5)	39.6 (12.2)	45.9 (10.7)
Median [Min,Max]	48.8 [18.0,59.9]	44.3 [23.4,57.5]	48.4 [22.0,57.4]	50.2 [19.5 <i>,</i> 59.8]	38.8 [19.4,59.1]	48.1 [18.0,59.9]
Sex						
Male	109 (38.0%)	10 (55.6%)	7 (38.9%)	36 (52.9%)	31 (51.7%)	193 (42.8%)
Female	178 (62.0%)	8 (44.4%)	11 (61.1%)	32 (47.1%)	29 (48.3%)	258 (57.2%)
FLT3 mutation type						
TKD	70 (24.4%)	12 (66.7%)	7 (38.9%)	12 (17.6%)	8 (13.3%)	109 (24.2%)
ITD <0.5 allelic ratio	74 (25.8%)	3 (16.7%)	5 (27.8%)	21 (30.9%)	16 (26.7%)	119 (26.4%)
ITD ≥0.5 allelic ratio	143 (49.8%)	3 (16.7%)	6 (33.3%)	35 (51.5%)	35 (58.3%)	222 (49.2%)
Missing	0 (0%)	0 (0%)	0 (0%)	0 (0%)	1 (1.7%)	1 (0.2%)
WBC count at baseline (10E9/L)						
Mean (SD)	54.8 (55.8)	41.6 (43.7)	40.9 (45.3)	43.8 (47.3)	47.1 (45.7)	51.1 (52.5)
Median	39.6	26.0	25.9	28.6	30.2	35.5
[Min,Max]	[1.40,330]	[1.40,159]	[2.70,154]	[0.600,236]	[1.20,207]	[0.600,330]
Missing	1 (0.3%)	0 (0%)	1 (5.6%)	1 (1.5%)	0 (0%)	3 (0.7%)
ECOG performance status						
0-1	255	17	17	64	50	403
01	(88.9%)	(94.4%)	(94.4%)	(94.1%)	(83.3%)	(89.4%)
2	32 (11.1%)	1 (5.6%)	1 (5.6%)	4 (5.9%)	10 (16.7%)	48 (10.6%)
Treatment	100		2		26	242
Placebo	136 (47.4%)	9 (50.0%)	8 (44.4%)	31 (45.6%)	26 (43.3%)	210 (46.6%)
Midostaurin	151 (52.6%)	9 (50.0%)	10 (55.6%)	37 (54.4%)	34 (56.7%)	241 (53.4%)
Allogeneic HCT in CR1						
No	211 (73.5%)	13 (72.2%)	16 (88.9%)	48 (70.6%)	44 (73.3%)	332 (73.6%)
Yes	76 (26.5%)	5 (27.8%)	2 (11.1%)	20 (29.4%)	16 (26.7%)	119 (26.4%)

Supplemental Table S8: Impact of 12 most frequent gene mutations on overall and event-free survival. Log rank test p-values from the univariate tests are indicated without (raw) and with adjustment (adj) for multiple testing via the Bonferroni-Holm procedure (FDR). See corresponding Kaplan Meier estimates in Supplemental Figure 2 and 3). Abbreviations: CI, confidence interval; HR, hazard ratio.

	Overall Surival				Event-free Survival					
	Logrank p-value (raw)	Logrank p-value (adj)	HR	Lower Cl	Upper Cl	Logrank p-value (raw)	Logrank p-value (adj)	HR	Lower Cl	Upper Cl
NPM1	<0.001	<0.001	0.60	0.46	0.77	<0.001	<0.001	0.60	0.48	0.74
WT1	<0.001	<0.001	1.83	1.38	2.43	0.018	0.195	1.36	1.05	1.75
ASXL1	0.013	0.125	1.67	1.11	2.52	0.100	0.903	0.72	0.48	1.07
IDH2	0.130	1	0.65	0.37	1.14	0.075	0.748	0.64	0.38	1.05
NRAS	0.097	0.875	0.68	0.43	1.08	0.793	1	1.05	0.71	1.55
DNMT3A	0.984	1	1.00	0.77	1.30	0.275	1	0.88	0.71	1.10
IDH1	0.773	1	1.07	0.68	1.69	0.649	1	0.91	0.61	1.35
PTPN11	0.287	1	0.77	0.48	1.25	0.204	1	0.75	0.48	1.17
RUNX1	0.141	1	1.32	0.91	1.90	0.310	1	0.83	0.58	1.19
SMC1A	0.219	1	0.69	0.37	1.26	0.183	1	1.25	0.90	1.73
SMC3	0.841	1	0.94	0.50	1.77	0.433	1	0.81	0.47	1.38
TET2	0.460	1	0.85	0.56	1.30	0.600	1	1.09	0.79	1.52

Supplemental Table S9: 4-year overall survival rates by genomic AML classes in the cohort of 451 of 475 patients, in which subcategorization into genomic AML classes was possible.

	Ν	Number at risk	Events	Survival	Lower 95% Cl	Higher 95% Cl
NPM1	287	136	120	56.9%	0.51	0.63
CBF	18	11	5	72.2%	0.54	0.96
TP53-aneuploidy	18	5	11	35.3%	0.19	0.67
Chromatin-spliceosome	68	18	45	32.9%	0.23	0.46
No class	60	16	36	36.5%	0.26	0.52

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Supplemental Table S10: 4-year event-free survival rates by genomic AML classes in the cohort of 451 of 475 patients, in which subcategorization into genomic AML classes was possible.

	Ν	Number at risk	Events	Survival	Lower 95% Cl	Higher 95% Cl
NPM1	287	87	177	37.7%	0.32	0.44
CBF	18	7	9	50.0%	0.32	0.79
TP53-aneuploidy	18	1	17	5.6%	0.01	0.37
Chromatin-spliceosome	68	8	56	16.3%	0.09	0.28
No class	60	3	53	9.5%	0.04	0.22

Supplemental Table S11: Cox proportional hazard model for predictive impact of *FLT3* mutation type on hazard of death or event after treatment with midostaurin in cohort of 451 of 475 patients, in which subcategorization into genomic AML classes was possible. A hazard ratio of >1 indicates a higher and a hazard ratio of <1 a lower risk of death, respectively. Abbreviations: CBF, Core-binding factor AML; CI.95, 95% confidence interval; CR1, first complete remission; ECOG, Eastern Cooperative Oncology Group; HR, hazard ratio; HCT, hematopoietic cell transplantation; WBC, white blood cell count.

	Overall survival		Event-free su	rvival
Variable	HR [CI.95]	p-value	HR [CI.95]	p-value
Genomic classes				
CBF vs NPM1	0.67 [0.27;1.67]	0.392	0.80 [0.40;1.58]	0.522
TP53-aneuploidy vs NPM1	1.94 [1.04;3.61]	0.037	3.67 [2.21;6.09]	<0.001
Chromatin-Spliceosome vs NPM1	2.27 [1.61;3.21]	<0.001	1.89 [1.38;2.57]	<0.001
No Class vs NPM1	2.13 [1.43;3.17]	<0.001	2.51 [1.79;3.50]	<0.001
Age	1.01 [0.99;1.02]	0.338	1.00 [0.99;1.01]	0.849
Sex				
Female vs male	1.00 [0.76;1.32]	0.995	1.33 [1.06;1.68]	0.015
ECOG (0-1 vs 2)	1.28 [0.85;1.92]	0.236	0.87 [0.60;1.27]	0.475
log2WBC	1.08 [1.00;1.17]	0.045	1.09 [1.02;1.16]	0.014
Allogeneic HCT in CR1	0.58 [0.40;0.83]	0.003	0.72 [0.49;1.06]	0.093
FLT3 TKD				
Midostaurin vs Placebo	0.59 [0.32;1.11]	0.100	0.70 [0.43;1.17]	0.174
FLT3 ITD <0.5 allelic ratio				
Midostaurin vs Placebo	0.56 [0.32;1.00]	0.049	0.61 [0.39;0.97]	0.035
<i>FLT3</i> ITD \geq 0.5 allelic ratio				
Midostaurin vs Placebo	0.66 [0.47;0.94]	0.019	0.67 [0.49;0.91]	0.009
Placebo				
ITD <0.5 allelic ratio vs TKD	1.13 [0.64;2.01]	0.665	1.40 [0.87;2.25]	0.163
ITD \geq 0.5 allelic ratio vs TKD	1.49 [0.90;2.47]	0.126	1.52 [1.00;2.32]	0.052

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Midostaurin				
ITD <0.5 allelic ratio vs TKD	1.08 [0.58;2.03]	0.811	1.22 [0.74;2.00]	0.430
ITD ≥0.5 allelic ratio vs TKD	1.65 [0.98;2.78]	0.058	1.44 [0.94;2.20]	0.090

SUPPLEMENTAL FIGURES

В

Supplemental Figure S1: Mutational exclusivity and co-occurrence of recurrently (>5% of cases) mutated genes. A) Gene pairs that co-occurred more frequently are indicated by blue colors, gene pairs that co-occurred seldom by orange colors. The top 12 genes were tested for mutual exclusivity (66 possible combinations) resulting in 21 significant pairs before adjustment for multiple testing and 5 significant pairs after FDR adjustment: *NPM1-RUNX1* (p<.001), *DNMT3A-WT1* (p<.001), *DNMT3A-RUNX1* (p=.003), *NPM1-WT1* (p=.008) and *IDH2-TET2* (p=.021). Similarly, all combinations were tested for co-occurrence resulting in 2 significant pairs before adjustment for multiple testing and *IDH1-PTPN11*, which were not significant after FDR adjustment B) Width of bands reflects the number of cases in which mutations of corresponding genes co-occurred.





Supplemental Figure S2: Impact of gene mutations on overall survival. Log rank test p-values from the univariate tests are indicated without adjustment for multiple testing. Adjusted p-values are given in Supplemental Table S8.



Supplemental Figure S3: Impact of gene mutations on event-free survival. Log rank test p-values from the univariate tests are indicated without adjustment for multiple testing. Adjusted p-values are given in Supplemental Table S8.





Supplemental Figure S4: Impact of WT1 mutations on overall survival stratified by ELN2017 risk groups.

Supplemental Figure S5: Impact of WT1 mutations on event-free survival stratified by ELN2017 risk groups.



Supplemental Figure S6: Prognostic and possibly predictive impact of pairwise interactions of clinical and/or genetic variables in 475 patients on A) overall and B) event-free survival using random survival forests. The prognostic impact of a variable is measured via "vimp" (variable importance). This measure determines the loss in prediction accuracy using a permuted/noisy version of each variable for model fitting and predicting out-of-bag samples. The assessment of pairwise interactions between variables was based on the comparison of the joint ('paired') VIMP to the sum of their individual VIMPs (called 'additive' importance). Fitting 1000 trees per forest, we constructed 100 forests with different seeds. The following graphic depicts the variable importance for all variables across these 100 runs in terms of variability (box plots). Higher positive or negative difference between additive and paired vimp values indicate that a variable combination may have prognostic or predictive impact on the survival endpoint.



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Supplemental Figure S7: The top 10 most interesting interactions regarding overall survival as determined by random survival forests were selected for further inspection. The following Kaplan Meier curves depict the marginal distribution of first variable (left) and the second variable (middle) as well as the combination of the two (right).











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Supplemental Figure S8: The top 10 most interesting interactions regarding event-free survival as determined by random survival forests were selected for further inspection. The following Kaplan Meier curves depict the marginal distribution of first variable (left) and the second variable (middle) as well as the combination of the two (right).



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Supplemental Figure S9: Kaplan Meier estimates of impact of *NPM1*, *DNMT3A*, and combined genotypes on overall (A) and event-free (B) survival and according to *FLT3* mutation type.



Supplemental Figure S10: Kaplan-Meier estimated A) overall and B) event-free survival curves, and number of events by genomic AML classes and log rank test p-values in cohort of 451 of 475 patients, in which subcategorization into genomic AML classes was possible. Abbreviations: OS, overall survival; EFS, event-free survival.



Supplemental Figure S11: Kaplan-Meier plots for the marginal overall survival (OS) distribution in the corresponding genomic AML classes (pCat) and treatment (trt) subgroups. Abbreviations: C-S, chromatin-spliceosome; Mdst, midostaurin; Plcb, placebo; TP53an, TP53-aneuploidy.





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Supplemental Figure S12: Kaplan-Meier plots for the marginal event-free survival (EFS) distribution in the corresponding genomic AML classes (pCat) and treatment (trt) subgroups. Abbreviations: C-S, chromatin-spliceosome; Mdst, midostaurin; Plcb, placebo; TP53an, TP53-aneuploidy.





Supplemental Figure S13: Kaplan-Meier estimates for A) overall and B) event-free survival according to *FLT3* mutation type and treatment of 451 patients included into the Cox proportional hazard model.



Supplemental Figure S14: Kaplan Meier estimates for A) overall and C) event-free survival by midostaurin kinome mutation status in entire cohort. Effect of treatment on B) overall and D) event-free survival in subgroup of patients harboring midostaurin kinome mutations.



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Supplemental Figure S15: Kaplan Meier estimator for overall (OS) and event-free survival (EFS) according to *FLT3* exon 16 mutation status.



