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**Supplementary information**

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**Characterization of interactions of dietary cholesterol with the murine and human gut microbiome**

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## **Supplementary Information**

### **Characterization of interactions of dietary cholesterol with the murine and human gut microbiome**

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## Supplemental Tables

### Supplementary Table 1 - Primers

Name/Use	Sequence
pET28_BT0416_fwd	gtgccgcgcgccagccatagGGATTACTTGAATTTAATAAACTTCCGATTAATACGTTG
pET28_BT0416_rev	gttatgctagtattgctcagcTTATAAATTATAGTTCCATTGTTTCAAAGCAAAATCCC
pET28_BT0468_fwd	ggtgccgcgcgccagccatagATAAAAAATAAAAAAATCAGAGGATTGTTGTGGATGTTC
pET28_BT0468_rev	ctagtattgctcagcTTATTGGAGTTTTCTTAATACTCTTTTAATAATTTGTTTCACTA
pET28_BT0487_fwd	gcccctggtgccgcgcgccagccatagAATAAATTGCCGAAAGAATCAGAATCAAAG
pET28_BT0487_rev	ggttatgctagtattgctcagcTTATTTGCTATGGTAATAGTCTATTGTTTCTACAGTC
pET28_BT0969_fwd	gcctggtgccgcgcgccagccatagAATAATCTTCGAATCTTATTATACGTGGTTGTCC
pET28_BT0969_rev	cccaaggggttatgctagtattgctcagcCTACAATGATTTGATGTAGTCTTCCAGTG
pET28_BT1511_fwd	ggtgccgcgcgccagccatagAAAATAAGAAATTTATTAGTAGCCATACTTGC AATGAG
pET28_BT1511_rev	aaggggttatgctagtattgctcagcTCATTCTGCTATACAGATACAAACACGATT TTC
pET28_BT1712_fwd	agcggcctggtgccgcgcgccagccatagAAAAGGTAGATATAATTCATGGTGGATGG
pET28_BT1712_rev	gttatgctagtattgctcagcTCATATAGCGTTCAAATATTAGCATAAGATTCAATCA
pEx_frag1_0416_fwd	CGAGTCGACGGTATCGATAAGCTTGATATCAAGTGGCATCGGGTATTGTCCGCAAGGGAG
frag1_0416_fwd	AAGTGGCATCGGGTATTGTCCGCAAGGGAGATACAGTGAT
frag1_0416_rev	TTTCTTAATCATCAATACTTATCCTTTCTTCGTCTACATTTTATTTAATCACTTTTATG
frag2_0416_fwd	CATAAAAGTGATTAAATAAAATGTAGAACGAAGAAAGGATAAGTATTGATGATTAAGAAA
frag2_0416_rev	AAGTCACCAACGTTCCGATACGCTTGGCATAAACATTTCC
pEx_frag2_0416_rev	GGAATTCCTCCACC CGCGGTGGCGGCCGCAAGTCACCAACGTTCCGATACGCTTGGCAT
qPCR_BT0416_fwd	CCAGCCTCTGAAACTGGAGG
qPCR_BT0416_rev	TTCTCCGTCATTCCCATCGC
qPCR_BT0412_fwd	CCGTTTCCCCGGTATCAACA
qPCR_BT0412_rev	CGTATCATCCGCCATCACCA
16s_qPCR_fwd	GGTAGTCCACACAGTAAACGATGAA
16s_qPCR_rev	CCCGTCAAATTCCTTTGAGTTTC
515F	AATGATACGGCGACCACCGAGATCTACACTATGGTAATTGTGTGCCAGCMGCCGCGGTAA
806R	CAAGCAGAAGACGGCATAACGAGATXXXXXXXXXXXXXAGTCAGTCA GCCGGACTACHVGGGTWTCTAAT

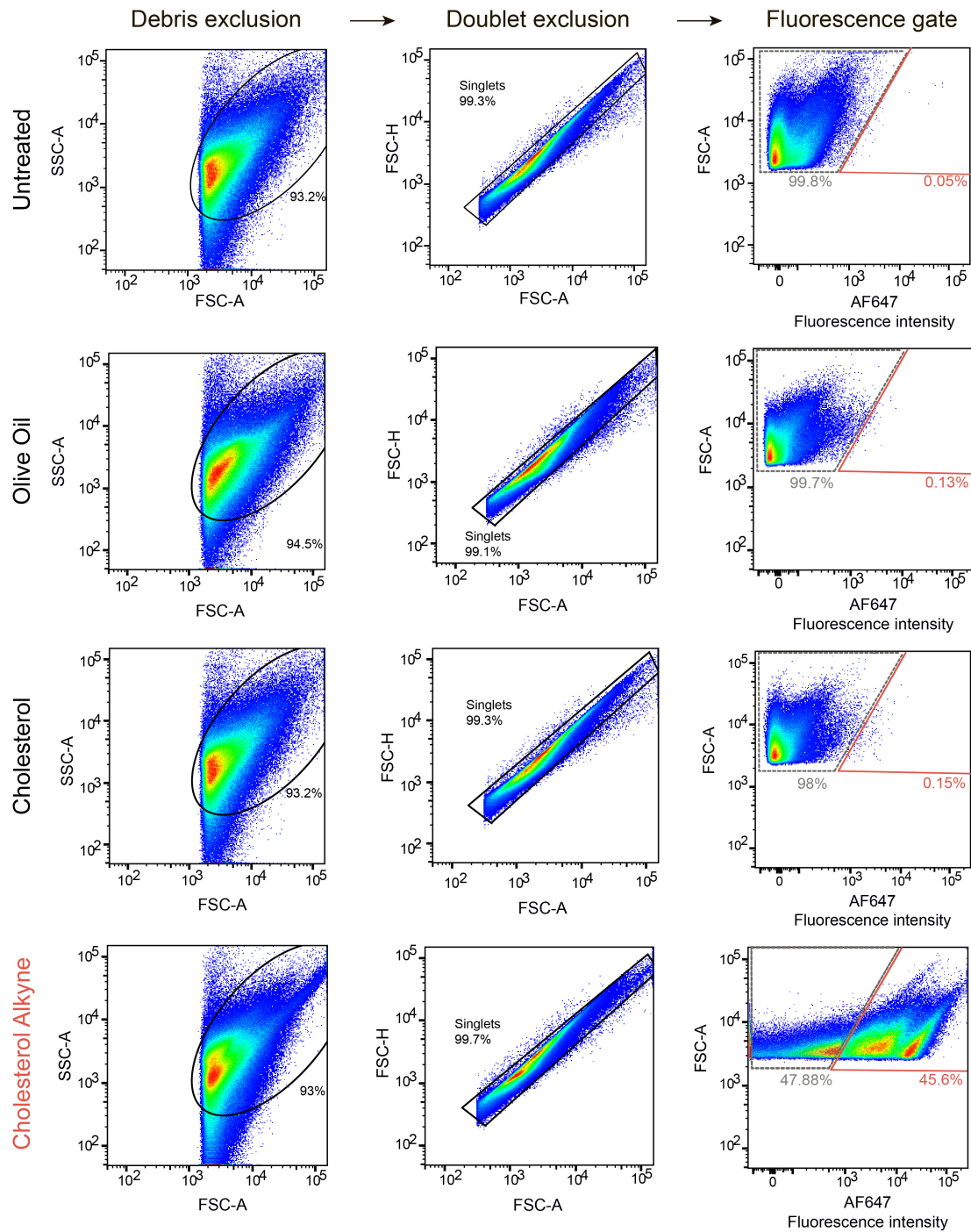
**Supplementary Table 2 - Strain sources**

Strain	Source
<i>A. caccae</i>	DSM 14662
<i>B. caccae</i>	ATCC 43185
<i>B. ovatus</i>	ATCC 8483
<i>B. fragilis</i>	ATCC 25285
<i>B. uniformis</i>	ATCC 8492
<i>B. vulgatus</i>	ATCC 8482
<i>B. thetaiotaomicron</i>	DSM 2079
<i>B. thetaiotaomicron</i> (sphingolipid-deficient)	Professor Andrew Goodman, Yale University
<i>B. thetaiotaomicron</i> VPI-5482 <i>tdk</i>	Professor Andrew Goodman, Yale University
<i>B. thetaiotaomicron</i> VPI-5482 <i>tdk</i> $\Delta$ 0416	This study
<i>B. longum</i> subsp. <i>infantis</i>	ATCC 15697
<i>B. pseudolongum</i> subsp. <i>pseudolongum</i>	ATCC 25526
<i>E. coli</i> (BL21)	New England Biolabs C2530H
<i>E. coli</i> (TOP10)	Invitrogen C404010
<i>E. coli</i> (S17-1 $\lambda$ pir)	Biomedal
<i>E. coprostanoligenes</i>	ATCC 51222
<i>L. amylovorus</i> Nakamura	ATCC 33620
<i>Ruminococcaceae</i> species	ATCC TSD-27
American Type Culture Collection	ATCC
German Collection of Microorganisms	DSM

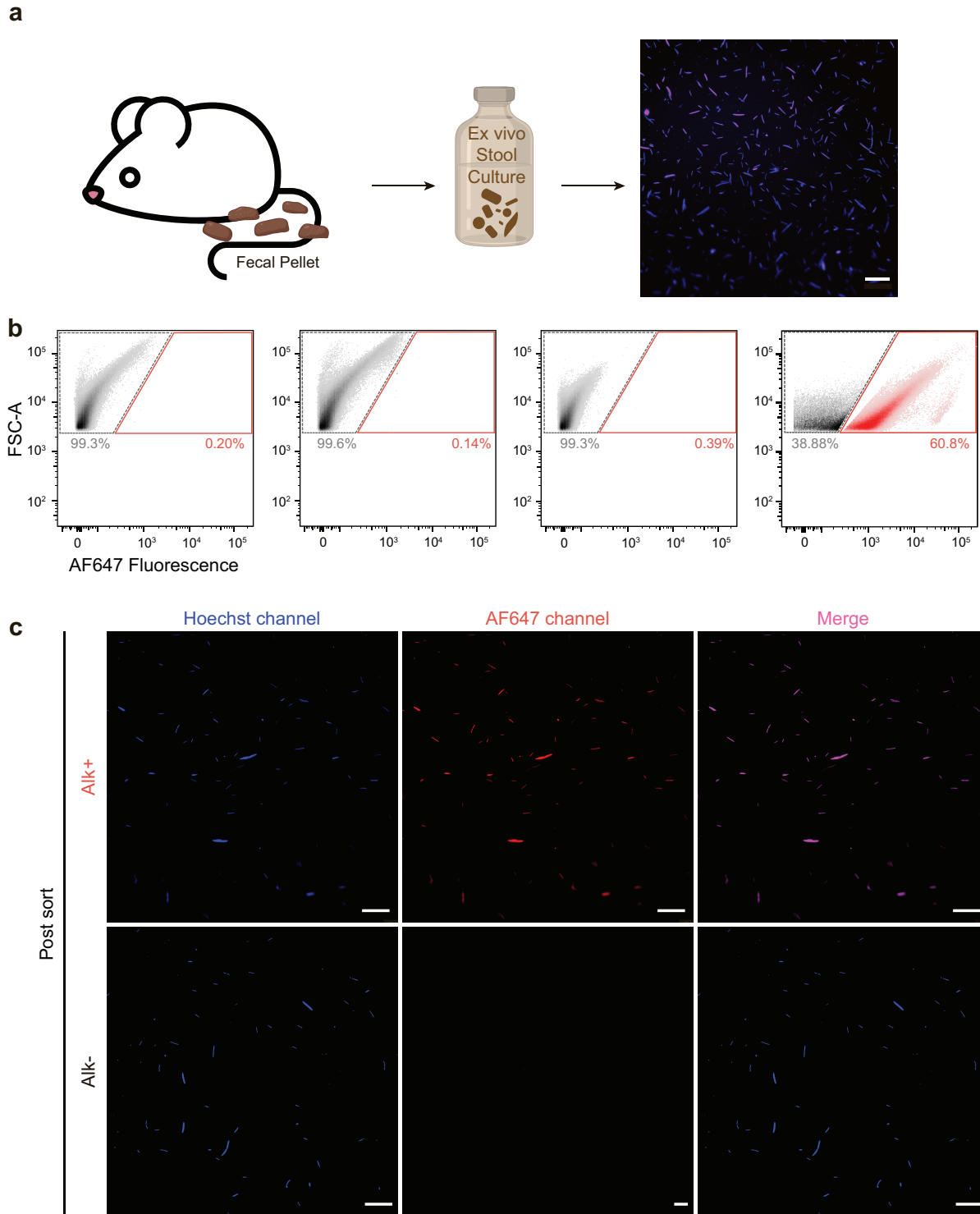
All unique plasmids and strains available upon request.

**Supplementary Table 3 – Sort efficiency**

Replicate	# of Alk+ events collected	# of Alk- events collected	% of parent population <sup>a</sup> in Alk+ gate	% of parent population <sup>a</sup> in Alk- gate	% Sort Efficiency in Alk+ gate	% Sort Efficiency in Alk- gate
Mouse 1	1,152,000	1,152,000	36.17%	63.70%	95%	94%
Mouse 2	1,152,000	1,152,000	40.38%	59.42%	93%	93%
Mouse 3	1,152,000	1,152,000	34.87%	65.12%	98%	97%
exvivo 1	1,103,000	1,103,000	37.61%	62.37%	95%	96%
exvivo 2	1,152,000	1,152,000	36.12%	63.27%	96%	96%
exvivo 3	1,018,000	1,023,010	38.12%	60.33%	98%	97%

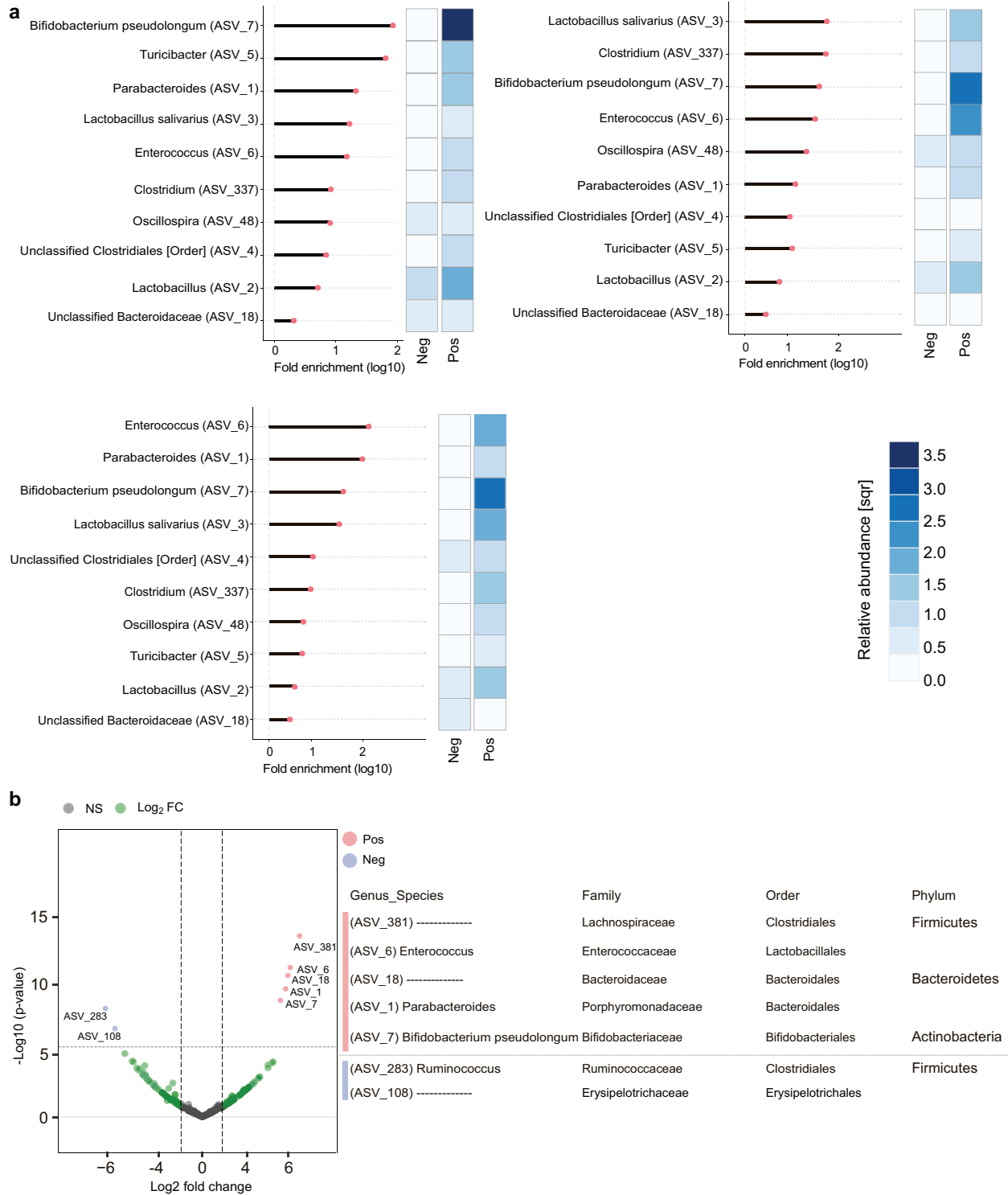


**Supplementary Fig. 1 | Representative density plots from successive FACS gating steps applied to cecal microbial samples from each dietary treatment group.** This gating strategy demonstrates the steps used to isolate Alk+ and Alk- populations. The definition of each step is outlined on the top of the panel, and the treatment group is shown on the left side of each row.

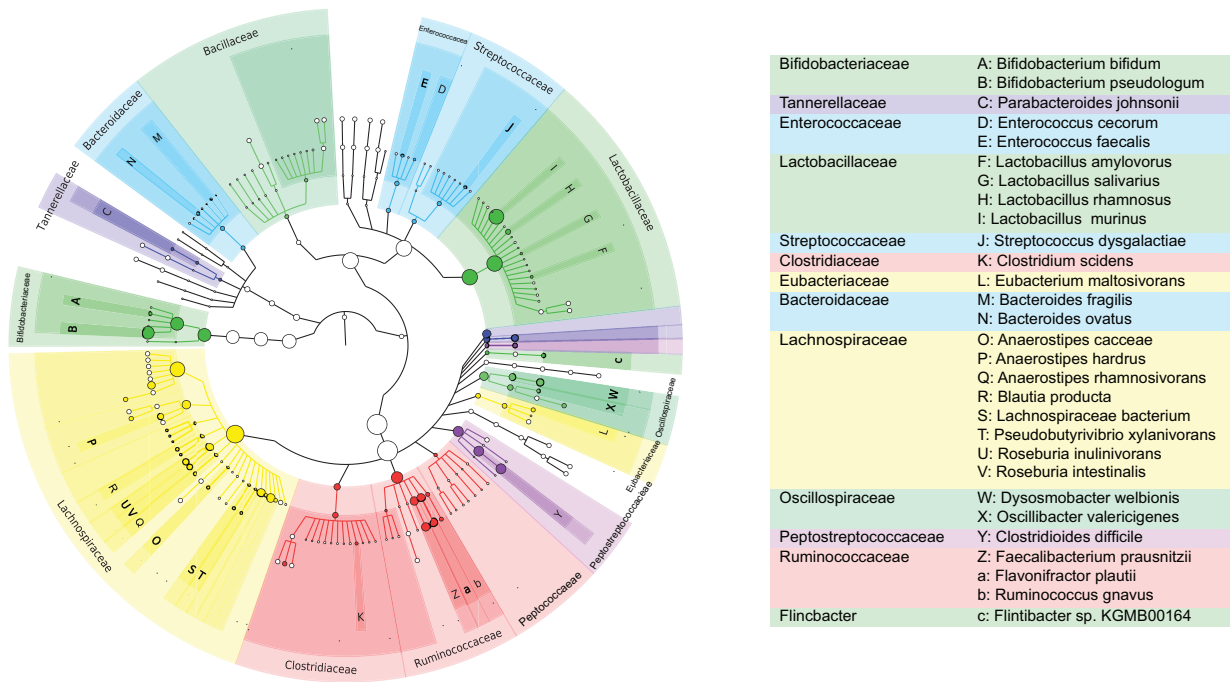


**Supplementary Fig. 2 | FACS enrichment of AF647-azide positive microbes arising from *ex vivo* culturing system.** **a**, Fecal microbes were extracted from murine fecal pellets and cultured with Chol<sup>Alk</sup>. Bacterial uptake was confirmed by fluorescent microscopy and the merged image is shown. Chol<sup>Alk</sup>-interacting microbes are red (AF647-azide) while the nucleic acids stain marks all microbes in blue using Hoechst 33342. (Scale bar: 20  $\mu$ m) **b**, FACS density plots

demonstrating isolation of Chol<sup>Alk</sup>-interacting organisms. **c**, Fluorescence microscopy demonstrating staining of Chol<sup>Alk</sup>-interacting or non-interacting organisms post-FACS. (Scale bar: 20  $\mu$ m).

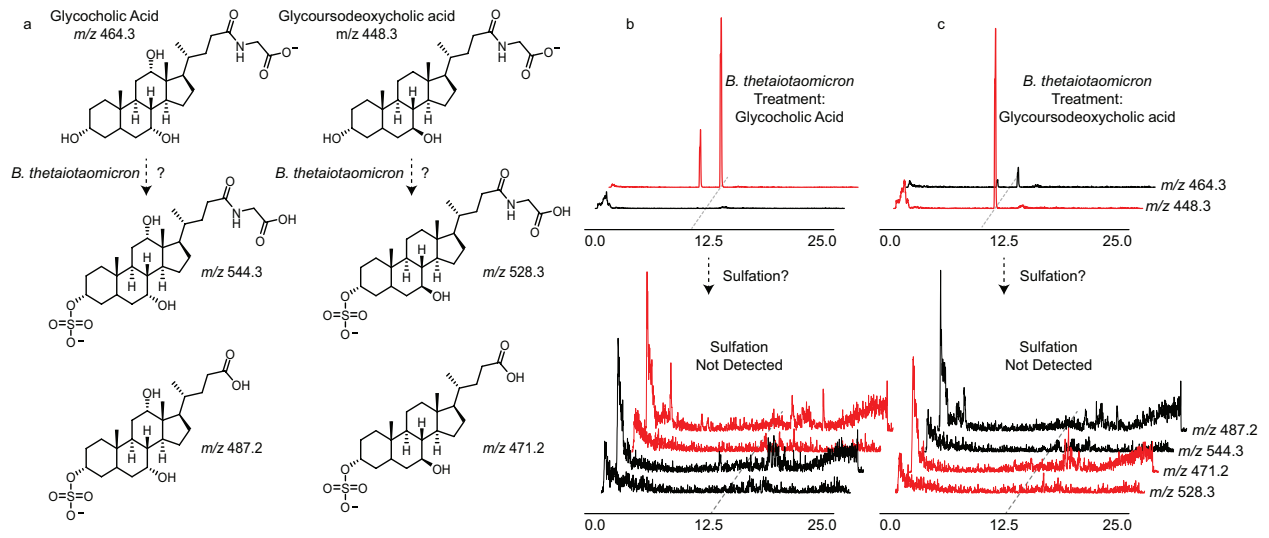


**Supplementary Fig. 3 | BioOrthogonal-labeling Sort-Sequence- (BOSS-) reveals the taxonomic identities of cholesterol-interacting gut microbes.** **a**, Enrichment results from 16S amplicon sequencing determined taxonomic classifications of cholesterol-interacting microbes separated using FACS. Heat maps indicate the magnitude of representation of the listed taxa in the neg (Alk-, cholesterol non-interacting) and pos (Alk+, cholesterol-interacting) sorted fractions. (n = 3 for each sorted population). Red dots indicate the value of the fold enrichment. **b**, Volcano plot showing taxonomic differences between Alk+ and Alk- using DESeq2 analysis. The pink dots represent those ASVs that have a log<sub>2</sub> fold change greater than 2 enriched in Alk+ and are significantly different (Wald test, Benjamin Hochberg correction, adjusted *P* < 0.05) from the Alk- fraction. (n = 3 for each sorted population, Pos: Alk+ and Neg: Alk-).

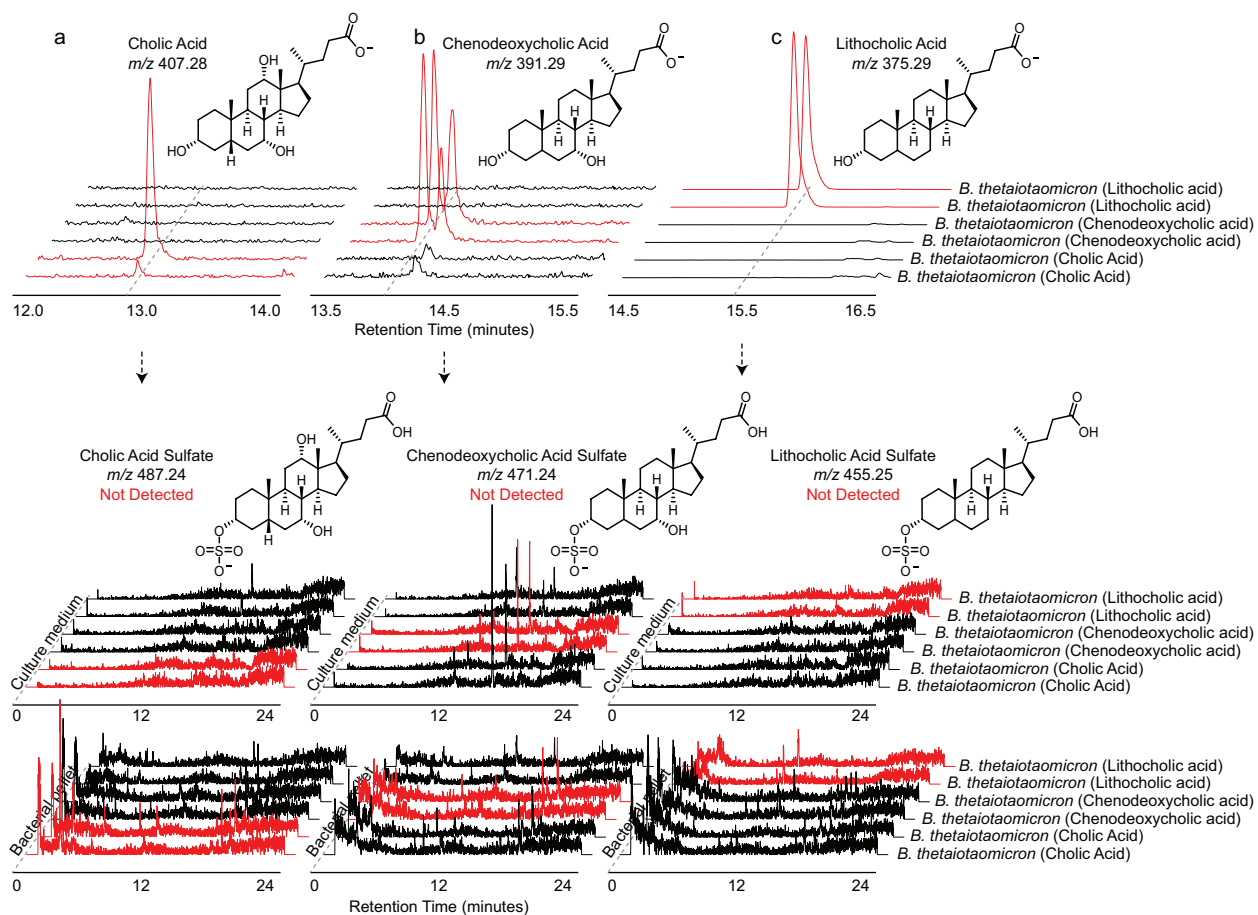


**Supplementary Fig. 4 | Shotgun metagenomic sequencing analysis identifies various taxa enriched in the Alk+ fraction.** Each dot represents a node in the phylogenetic tree, and its size indicates the prevalence of representatives of the corresponding node in the sample. Family-level taxonomic classification is colored for differentiation. The letters represent those microbes identified with relatively high abundance with species-level resolution.

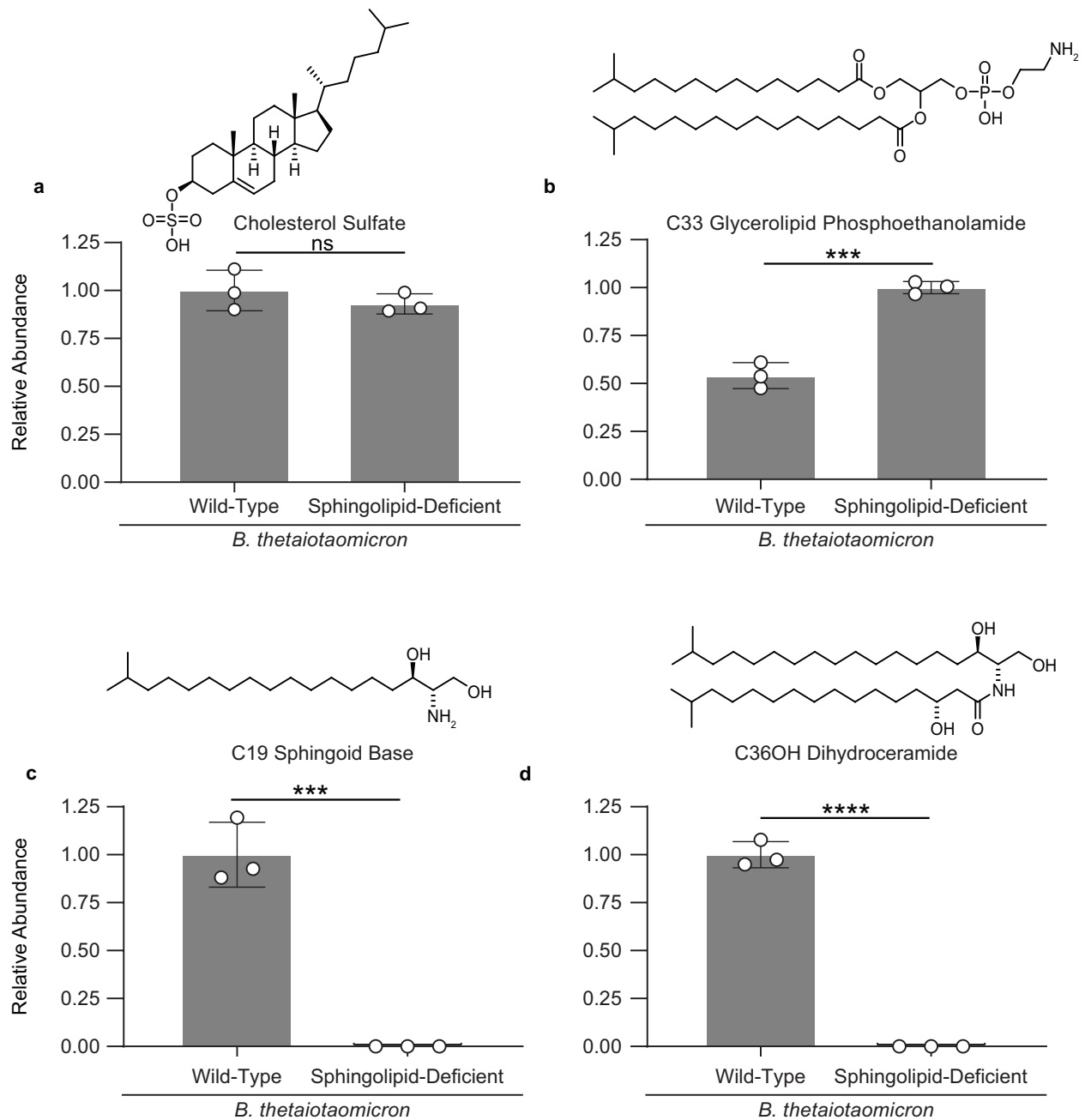




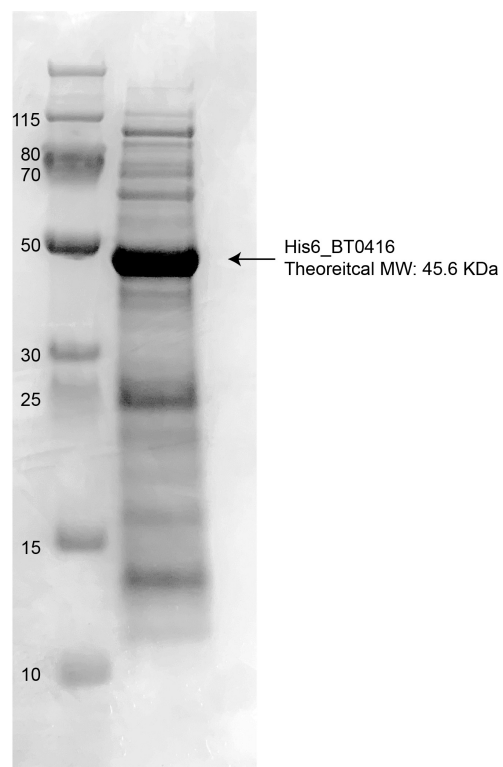
**Supplementary Fig. 5 | Conjugated bile acids are not substrates for sulfotransferase in *B. thetaiotaomicron*.** **a**, Schematic of conjugated bile acids tentatively undergoing conversion to their corresponding sulfates via *B. thetaiotaomicron*. Ion chromatograms demonstrating that **b**, glycocholic acid, and **c**, glycoursodeoxycholic acid, are not converted to their corresponding sulfates by *B. thetaiotaomicron*. Red lines indicate the chromatogram and information corresponding to the denoted bile acid.



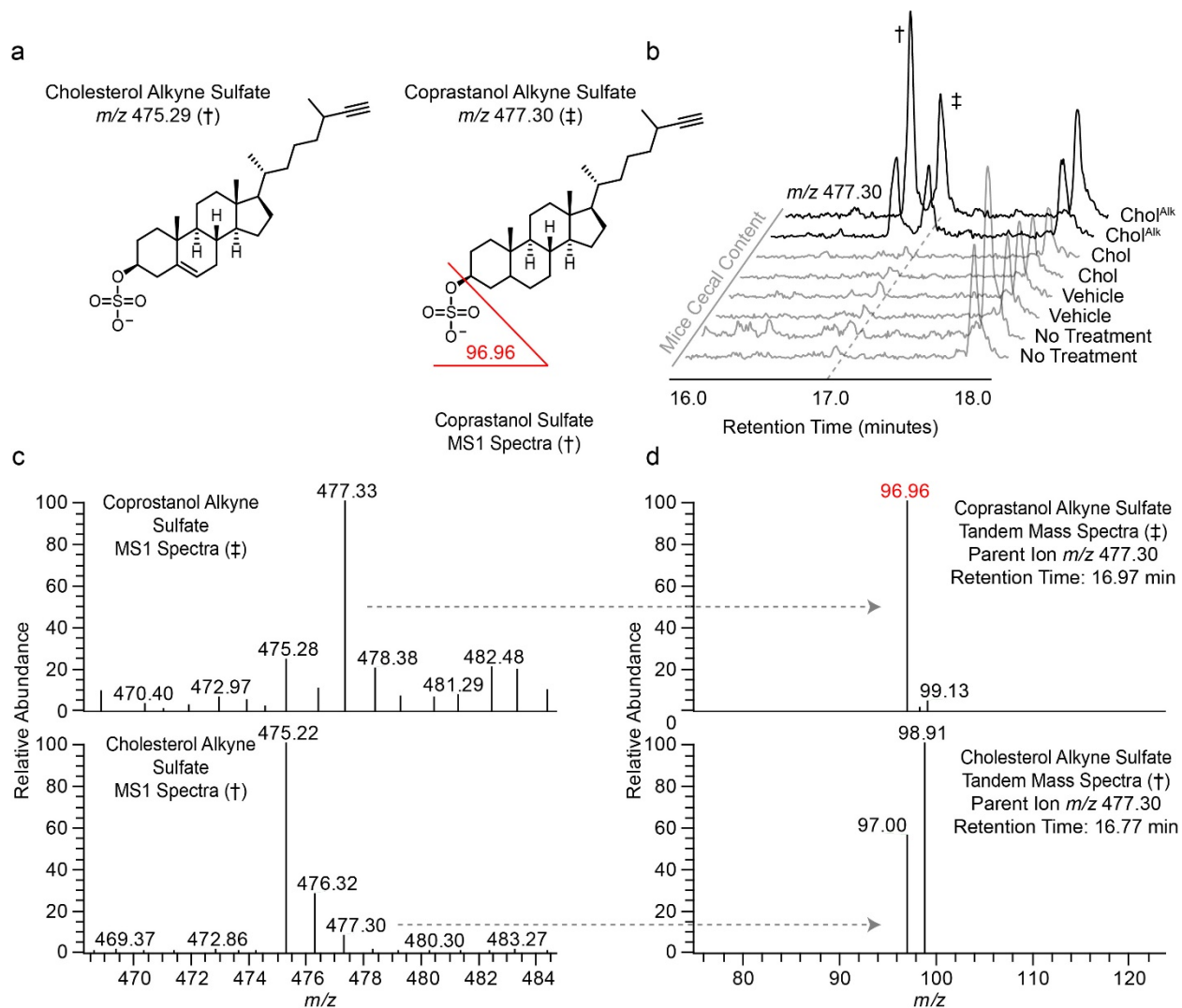
**Supplementary Fig. 6 | Primary and secondary bile acids are not substrates for sulfotransferase in *B. thetaiotaomicron*.** **a**, Cholic acid, **b**, chenodeoxycholic acid, and **c**, lithocholic acid are not converted to their corresponding sulfates by *B. thetaiotaomicron*. Red lines indicate the chromatogram and information corresponding to the denoted bile acid.



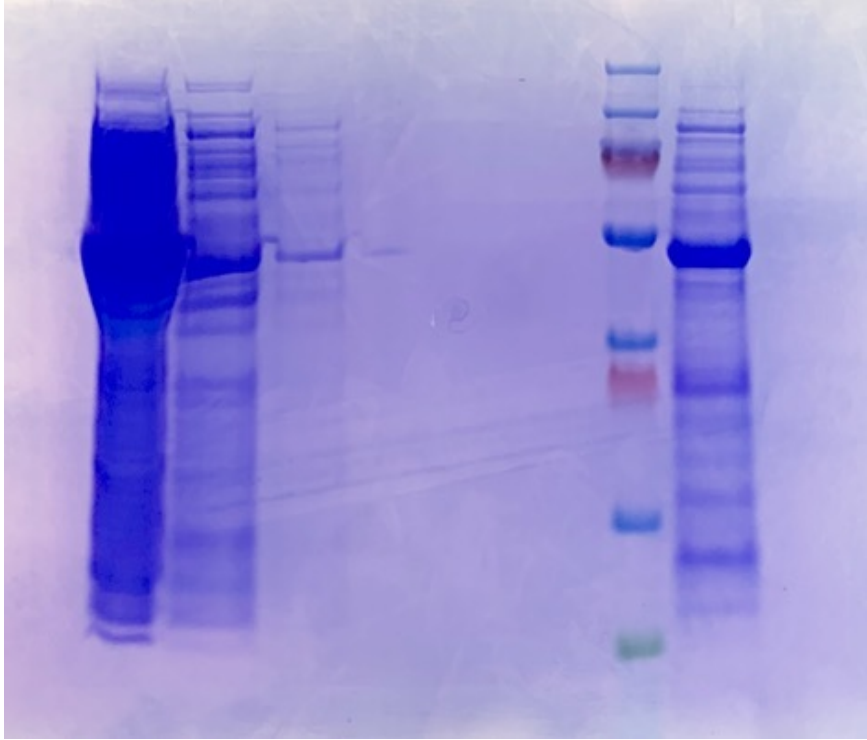
**Supplementary Fig. 7 | Cholesterol sulfate production is independent from sphingolipid production in *Bacteroides thetaiotaomicron*.** Relative quantification of **a**, cholesterol sulfate, **b**, C33 glycerolipid phosphoethanolamide, **c**, C19 sphingoid base, and **d**, C36OH dihydroceramide from either wild-type or sphingolipid-deficient *B. thetaiotaomicron* cultures. Bar chart values are the mean  $\pm$  SD. (n = 3 biologically independent cultures per condition).



**Supplementary Fig. 8 | Coomassie stained SDS-PAGE of partially isolated His6-BT0416 from expression in BL21 *E. coli*.**



**Supplementary Fig. 9** | **a**, Representative structures, **b**, ion chromatograms, and tandem mass spectra fragmentation patterns of, **c**, coprastanol alkyne sulfate and, **d**, cholesterol alkyne sulfate from cecal content of mice treated with cholesterol alkyne.



Uncropped blot of Supplementary Figure 8