Supporting Information

Metal-enhancement Study of Dual Functional Photosensitizers with

Aggregation-Induced Emission and Singlet Oxygen Generation

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Experimental Section

Materials

Poly(ethyleneimine) solution (PEI, 50%, Mw=2000), poly(sodium 4-styrene sulfonate) (PSS, Mw=70000), tannic acid, nitric acid, hydrochloric acid, potassium carbonate, silver nitrate, 9,10 anthracenediyl-bis(methylene)dimalonic acid (ABDA), Tetraethyl orthosilicate (TEOS), ethanol, ammonia solution, and sodium chloride were purchased from Sigma-Aldrich, Germany. For the synthesis of AIEgens, chemicals were all purchased from Sigma-Aldrich and used as received without further purification. Tetrahydrofuran (THF) and dichloromethane were dried by distillation using sodium or calcium hydride as the drying agent. All non-aqueous reactions were carried out under the nitrogen atmosphere in oven-dried glassware. Deuterated solvents were purchased from Cambridge Isotope Laboratories Inc.

Characterization

The extinction of AgNPs and the absorbance of ABDA was measured by a UV-Vis spectrophotometer (Shimadzu, UV-1700, Japan). The fluorescence of the samples was obtained using Infinite M-200 Microplate reader. Transmission electron microscopy (TEM) images were taken by JEOL JEM-2010F operating at 200kV. The zeta potential and hydrodynamic size of the nanoparticles were acquired on Malvern Zetasizer at room temperature. Fluorescence lifetime was measured using a FluoTime 200 TCSPC machine (Picoquant BmbH, Germany). Nuclear magnetic resonance (NMR) spectra were measured on a Bruker ARX 400 NMR spectrometer. Chemical shifts are reported in parts per million referenced with respect to residual solvent (CDCl₃ = 7.26 ppm, MeOD = 3.31 ppm) for ¹H NMR and (CDCl₃ = 77.1 ppm, MeOD = 49.0) for ¹³C NMR. The extent of reaction was monitored by thin-layer chromatography (TLC) using Merck 60 F254 pre-coated silica gel plates with fluorescent indicator UV254 after the plates were subjected to elution in the TLC chamber. Flash column chromatography was carried out using Merck silica gel (0.040-0.063).

Synthesis of AIEgens

Compound $1,1$ compound $2,2$ and AIE-R-SOG³ were synthesized according to the reported literature.

AIE-B: Trimethylamine (2 mL, 25 wt. % in H₂O) was added dropwise into compound 1 (100 mg, 0.21 mmol) in methanol (20 mL) under 20 ℃. After stirring at 20 ℃ overnight, the solvent and excess trimethylamine were removed under reduced pressure. The obtained residue was dissolved in water (20 mL) and washed by dichloromethane (50mL \times 3). The aqueous phase was dried by freeze drier to obtain AIE-B as white powder (85 mg, yield 76.2%). ¹H NMR (400 MHz, MeOD) δ 7.13–7.05 (m, 9H), 7.04–6.97 (m, 6H), 6.92 (d, *J* = 8.8 Hz, 2H), 6.69 (d, *J* = 8.8 Hz, 2H), 4.00 (t, *J* = 5.9 Hz, 2H), 3.48–3.42 (m, 2H), 3.17 (s, 9H), 2.05–1.94 (m, 2H), 1.85 (dd, *J* = 13.5, 6.1 Hz, 2H). ¹³C NMR (101 MHz, MeOD) δ 157.47, 143.96, 140.60, 140.26, 136.24, 132.21, 130.96, 127.34, 126.03, 113.35, 66.49, 66.17, 52.20, 25.75, 19.72. The purity of the sample was about 99%.

AIE-Y-SOG: Compound 2 (78 mg, 0.18 mmol) and bromoethane (110 mg, 1.1 mmol) were dissolved in DMF (5 mL) and heated at 80 °C under argon atmosphere for 14 h. After cooling to room temperature, the mixture was separated with chromatography directly (Eluent: hexane/ethyl acetate = 1/5, dichloromethane/methanol = 10/1) to get the desired product as a yellow solid (80 mg, yield 81.1 %).¹H NMR (400 MHz, CDCl3) δ 9.12 (d, *J* = 6.3 Hz, 2H), 7.93 (d, *J* = 6.3 Hz, 2H), 7.56 (d, *J* = 16.2 Hz, 1H), 7.30 (d, *J* = 8.3 Hz, 2H), 7.07 – 7.01 (m, 10H), 6.95 (dd, *J* = 5.6, 2.4 Hz, 8H), 4.79 (q, *J* = 7.2 Hz, 2H), 1.60 (t, *J* = 7.2 Hz, 3H). ¹³C NMR (101 MHz, CDCl3) δ 153.52, 147.17, 144.02, 143.30, 143.24, 143.11, 142.52, 141.94, 139.97, 132.43, 132.17, 131.28, 127.91, 127.88, 127.70, 126.94, 126.86, 124.11, 121.68, 56.24, 17.01. The purity of the sample was about 95%.

Synthesis of silver nanoparticles (AgNPs)

Aqua regia (HCl: HNO₃ with 3:1 molar ratio) was used to wash all glassware before the synthesis. Then, glassware was washed with DI-water and ethanol and, finally, with D-water.

The silver nanoparticles (AgNPs) with different sizes were synthesized using tannic acid as both reducing agent and capping agent according to the literature ⁴. Briefly, the 19.5 mL of tannic acid solution was prepared freshly, and the pH was adjusted to the desired value using K_2CO_3 solution (0.5 M or 0.05 M). Then, 500 µL of AgNO₃ solution (20×10^{-3} M) was added to the above reaction. For 25 nm and 40 nm AgNPs, the Ag/TA molar ratio was kept at 5, and the pH was adjusted at 7 and 6, respectively. For 85 nm AgNPs, the Ag/TA molar ratio increased to 20, and the pH was adjusted at 6. All synthesis experiments were carried out at 30 $°C$.

Self-assembly of polyelectrolytes on AgNPs

The polyelectrolyte coating of AgNPs was performed using layer-by-layer (LBL) approach ⁵. The PEI and PSS solutions (both 10 mM based on Mw of repeat unit) were prepared using NaCl solution (10 \times 10⁻³ M) as the solvent, and then the pH was changed to 3 using HCl solution (1M). A washing solution also was prepared using 10 mM NaCl solution and HCl (1 M) with a pH of 3. In addition to the NaCl concentration which reduces the electrostatic repulsion, it has been reported that the thickness of polyelectrolytes on the substrate is a function of the deposition pH. Selection of these parameters is important to obtain a thin layer of PEI/PSS bilayer and enables linear growth of thickness for each bilayer.⁶⁻⁸ In LBL approach, the degree of aggregation could be controlled by performing the LBL at a low concentration of AgNPs and also sonication of samples during the assembly process. For LBL, AgNPs solution (250 µL) was added to PEI solution (750 µL), vortexed for 30 sec and sonicated for 10 min. After 3 times washing, the same procedure was done with PSS solution. The AgNPs with different bilayers of PEI/PSS were prepared by repeating the previous steps.

Synthesis of silica nanoparticles

S3

Silica nanoparticles (~100 nm) were synthesized according to the modified Stöber method reported in our previous work 9, 10.

Metal-enhanced fluorescence study

To study the MEF, the same amount of fluorophore should exist in all samples. Hence, an adequate amount of AIEgen solution (0.15×10^{-3} M) was added to polymer-coated AgNPs and sonicated for 15 min. Then, the sample was centrifuged, and the supernatant was removed, and the precipitate was diluted with PBS to reach 1 mL volume. By comparing the absorbance of the supernatant and control sample without AgNPs, the amount of AIEgen in all samples was kept constant at a concentration of 10×10⁻⁶ _M. The control samples for MEF also were prepared with the same amount of AIEgen (AIE-Y-SOG and AIE-R-SOG, and AIE-B) loaded on silica nanoparticle. For the fluorescence study, 40 µL of each solution was transferred to a 394-well black microplate.

Singlet oxygen generation measurement

ABDA was used as an indicator of singlet oxygen generation. A 1 mL solution containing 10×10⁻⁶ M AIE photosensitizer (AIE-PS) adsorbed on polymer-coated AgNPs and 50 µM ABDA was prepared and exposed to the 40 mWcm⁻² white light. The degradation of ABDA was monitored by measuring the

UV-Vis absorbance spectra of ABDA. The first-order kinetic model ($\left(\frac{A}{A_0}\right) = kt$) was used to $\left(\frac{R}{A_0}\right) = kt$ determine the degradation rate of ABDA and consequently, the singlet oxygen generation rate of AIE-PS enhaced by the AgNPs in the nanohybrids.¹¹ The loaded AIE-PS molecules on silica nanoparticles (SiNP) were used as a control.

The enhancement in singlet oxygen generation was calculated based on the below equation:

Singlet oxygen generation enhancement factor
$$
(EF_{SOG}) = \frac{k_{Ag@AIE}}{k_{Si@AIE}}
$$
 eq. (S1)

where $k_{Ag@AIE}$ is the first-order kinetic constant for degradation of ABDA in the presence of AIE-PS on AgNPs with different bilayers of polyelectrolytes and the $k_{Si@AIE}$ is the same parameter for the control sample, which is aggregated AIE-PS on silica nanoparticles.

Fluorescence Lifetime measurement

2 mL of each sample containing 10×10^{-6} M AIEgen was diluted to 3 mL using DI-water, and the fluorescence lifetime was measured by excitation at 374 nm for AIE-Y-SOG samples, 374 nm for AIE-R-SOG samples and 321 nm for AIE-B samples. The emission wavelength was set according to the maximum fluorescence of each AIEgen. The biexponential model was used to describe the fluorescence decay which can be defined as below:

$$
I(t) = \sum \alpha_i exp(-t/\tau_i)
$$
eq.
(S2)

Where $^{\tau_i}$ is decay time, $^{\alpha_i}$ is amplitude, and $\sum_i \alpha_i = 1$.

The amplitude-weighted fluorescence lifetime is defined as:

$$
\langle \tau \rangle = \sum \alpha_i \tau_i \qquad \text{eq.}
$$
\n
$$
(53)
$$

Moreover, the average lifetime can be written as:

(S4)

$$
\bar{\tau} = \sum f_i \tau_i \qquad \qquad \text{eq.}
$$

S5

$$
f_i = \frac{\alpha_i \tau_i}{\sum_{i} \alpha_i \tau_i}
$$

Where the contribution fraction of each component is given as follows: eq. (S5)

The curve fitting was performed using the Curve-fitting toolbox in Matlab2014b software.

Simulation of the electric field distribution

To simulate the enhanced electric field around the silver nanoparticles, Finite element method (FEM) based simulations were carried out by the COMSOL Multiphysics software package (see www.comsol.com). The complex refractive index of Ag was taken from CRC handbook ¹². Water was used as the surrounding environment. A plane wave propagating in the *y*-direction and linearly polarized along the *z*-axis spanning the wavelength range of 300 nm to 700 nm was set as the source. Due to the symmetry of the problem only one-quarter of the sphere was modeled. To ensure high accuracy, the mesh size for all simulations was chosen as 0.5 nm. To calculate the electric field distribution a total field scattered field (TFSF) analysis was used.

In order to calculate the average surface electric field, individual Ag nanoparticles (*d* = 25, 40, 85 nm) were illuminated by a plane wave at 385 nm. The real part of the electric field was integrated over the volume of the spherical shell surrounding the nanoparticle and on the outer surface of the shell. The results were divided by the spherical shell outer surface area, respectively, to generate the average surface electric field.

Figure S1. ¹H NMR and ¹³C NMR spectra of AIE-Y-SOG in CDCl₃.

Figure S2. The hydrodynamic size distribution of AIE-Y-SOG in PBS:DMSO (99:1) as measured by the dynamic light scattering (DLS) instrument

Figure S3. a) Extinction spectra and b) DLS data of AgNPs of different sizes (25, 40, and 85 nm)

Figure S4. TEM images of AgNPs of different sizes: a) 25 nm, b) 40 nm, and c) 85 nm

Table S1- Fluorescence lifetime analysis for AIE-Y-SOG in different mediums and

Ag@AIE-Y-SOG prepared using different size of AgNPs and different number of PEI/PSS bilayers

 $\langle \tau \rangle$: Amplitude – weighted lifetime

Figure S5. Normalized surface average enhanced electric field around AgNPs with different size as a function of distance from the surface of the nanoparticle

Scheme S1. Synthetic route of AIE-B

Figure S6. ¹H NMR and ¹³C NMR spectra of AIE-B in CD₃OD.

Figure S7. a) Absorbance and fluorescence spectra of AIE-B, b) Effect of PBS fraction on fluorescence spectra of AIE-B, c) Fluorescence spectra of AIE-B in PBS solution and loaded on silica NPs. The insets show the optical photographs of these samples obtained under 365 nm light irradiation

Figure S8. a) Absorbance and fluorescence spectra of AIE-R-SOG in PBS, Inset shows optical photograph obtained under 365 nm light irradiation b) Effect of PBS fraction on fluorescence spectra of AIE-R-SOG, c) Degradation of 50 µM ABDA in the presence of 10µM AIE-R-SOG in PBS under 40 mWcm-2 white light

Table S2- Fluorescence lifetime analysis for AIE-B in different mediums

Sample	α_i	τ_i (ns)	f_i	τ (ns)	$\langle \tau \rangle$ (ns)
AIE-B in Water	0.8839	0.81	0.36627	7.059	1.954
	0.1161	10.67	0.63373		
$Si@AIE-B$	0.9244	1.494	0.70817	3.255	1.950
	0.0756	7.528	0.29183		
85Ag/1BL@AIE-B	0.9662	1.516	0.76930	4.161	1.904
	0.0338	12.98	0.23070		
85Ag/2BL@AIE-B	0.9107	1.486	0.68009	3.290	1.990
	0.0893	7.126	0.31991		
85Ag/3BL@AIE-B	0.9534	1.831	0.77710	3.822	2.247
	0.0466	10.76	0.22290		
$\bar{\tau}$: Average lifetime					
$\langle \tau \rangle$: Amplitude – weighted lifetime					

and Ag@AIE-B prepared using different size of AgNPs and different number of PEI/PSS bilayers

Table S3- Fluorescence lifetime analysis for AIE-R-SOG in different mediums and Ag@AIE-R-SOG prepared using different size of AgNPs and different number of PEI/PSS bilayers

 $\langle \tau \rangle$: Amplitude – weighted lifetime

Figure S9. Degradation of 50 µM ABDA in the presence of 10 ×10-6 M AIEgen loaded on AgNPs with different PEI/PSS bilayers and under 40 mWcm-2 white light a) 25 nm Ag@AIE-Y-SOG, b) 85 nm Ag@AIE-Y-SOG , c) 85 nm Ag@AIE-R-SOG

Figure S10. a) Metal-enhanced singlet oxygen generation of 25 nm (square) and 85 nm (sphere) Ag@AIE-Y-SOG nanohybrids with different PEI/PSS bilayers, b) Simulated scattering efficiency spectra for 25 nm and 85 nm AgNP placed in water as a medium

Figure S11. Degradation of 50 µM ABDA in the presence of 10 ×10-6 M AIE-R-SOG loaded on 85 nm AgNPs with different PEI/PSS bilayers and under 40 mWcm⁻² white light.

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