Supplementary material

N-terminal and mid-region tau fragments as fluid biomarkers in neurological diseases

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Supplementary methods

T-tau immunoassay development and validation for CSF

For NTA t-tau and MR t-tau, mouse monoclonal antibody targeting aa 159-163 (HT7, #MN1000, Thermo Scientific) and for NTB t-tau mouse monoclonal antibody targeting aa 6-18 (1-100, #816601, BioLegend) were conjugated to magnetic homebrew carboxylated beads (#103207, Quanterix). All detector antibodies were biotinylated in-house with EZ-LinkTM NHS-PEG4-Biotin (Thermo Scientific, USA). NTA and MR t-tau were measured using a 2-step protocol, where the assay beads, detector antibody and the analyte are first co-incubated, followed by washing of the beads, and incubation with the enzyme. NTB t-tau was measured using a 3-step protocol, where the beads were initially incubated with the analyte, then with the detector, and finally with the enzyme. After another wash, the substrate resorufin β -D-galactopyranoside (RGP; #103159, Quanterix) was added to the beads to gain the measured fluorescent signal. Recombinant non-phosphorylated full-length Tau-441 (#T08-54N, SignalChem) was used as a calibrator in all the inhouse t-tau assays. Three-fold diluted calibrator series in assay diluent (Tau 2.0 diluent, #101556, Quanterix) ranging from 500 to 0.7 pg/ml (MR and NTB t-tau), 56 to 0.1 pg/ml (CSF NTA t-tau) or 18.5 to 0.03 pg/ml (plasma NTA t-tau) were included in each sample plate.

Limits of detection (LOD) and lower limits of quantification (LLOQ) were defined by running 16 blank samples in duplicates and adding three or ten standard deviations to the measured mean value, respectively. Upper limits of quantification (ULOQ) were set as the concentration of the highest calibration accounting for the used dilution factor. For dilution linearity, two human CSF samples (pooled Alzheimer's disease and pooled controls) were measured with 2-fold dilution series (1:2, 1:4, 1:8) as duplicates. For spike recovery, two CSF samples were diluted 1:2 and spiked 10 pg/ml and 40 pg/ml (NTA t-tau) or 50 pg/ml and 150 pg/ml (NTB and MR t-tau) of exogenous non-P Tau441 (the assay calibrator). Spiked samples were then measured on a same plate with non-spiked CSF and spiked assay diluent. Spike recovery was calculated as: % Recovery = Concentration of spiked sample / (Concentration of non-spiked sample + Concentration of spiked buffer). Precision and accuracy were tested by measuring five replicates of two human CSF samples as duplicates in three consecutive days and calculating the intra-assay precision (variation within run, CV_r (%)) and inter-assay precision (variation between runs, CV_{rw} (%)).

Supplementary results

T-tau assay development and validation for CSF

Analytical performance of all the in-house assays were acceptable. Mean spike recovery was 79.3% (range 76-84%) for NTA t-tau, 145% (range 133-157%) for NTB t-tau, and 123% (range 118-126%) for MR t-tau assays (**Supplementary Table 4**).

Recovery % with the used sample dilution (1:4) was 78-79% with all assays (**Supplementary Figure 1**). With dilution, measured concentrations became lower in all in-house assays, implying that the sample matrix has some interference with the obtained signal. However, when comparing samples with the same dilution, this should not be problematic. Due to extremely high t-tau concentrations, some of the Creutzfeldt-Jakob disease and acute neurological disorders samples were further diluted 1:10, and these measurements were above the ULOQ (2000 pg/ml) and all extrapolated from the standard curve. This should be kept in mind and the concentrations gained from these analyses are likely underestimations and should be interpreted with caution.

T-tau measurements for CSF

All pilot CSF cohort samples measured above the quantification limits of each t-tau assay. From the clinical cohorts, 26/292 (8.9%) CSF samples were below the LOD of NTA t-tau (six controls, nine A β - subjects with mild cognitive impairment (MCI), one A β + MCI, three non-Alzheimer's disease dementia, four progressive supranuclear palsy, and three Alzheimer's disease), and 5/292 (1.7%) of NTB t-tau (two controls, one A β - MCI, one A β + MCI, and one non-Alzheimer's disease dementia). One CSF sample measured with NTB (A β - MCI subject) and one sample measured with MR t-tau (A β - MCI subject) was excluded due to technical error (no reading). In addition, one CSF sample was finished before measurement with NTB (A β - MCI).

All t-tau levels were extremely high in Creutzfeldt-Jakob disease and acute neurological disorders, and most of the samples gave readings above the defined upper limit of quantification (ULOQ) even with the additional 1:10 dilution. Thus, majority of the concentrations were extrapolated from the calibration curve (>2000g/ml) and results should be interpreted with caution.

Supplementary tables

Supplementary Table 1 Quantification limits for NTA, NTB and MR t-tau assays for CSF

	NTA t-tau	NTB t-tau	MR t-tau
LOD (pg/ml)	0.098	0.29	0.26
ULOQ (pg/ml)	224	2000	2000
LLOQ (pg/ml)	0.28	1.0	1.2

Abbreviations: LOD, limit of detection; LLOQ, lower limit of quantification; ULOQ, Upper level of quantification.

Supplementary Table 2 Repeatability and intermediate precision of NTA, NTB and MR ttau in CSF (results from validation experiments)

	Sample	Conc (pg/ml)	Repeatability (CVr) %	Intermediate precision (CVRw) %
	CSFI	8	1.5	16.9
NTA t-tau	CSF2	36	7.2	16.7
	Mean		4.4	16.8
	CSFI	86	8.1	27.4
NTB t-tau	CSF2	122	4.1	29.6
	Mean		6.1	28.5
	CSFI	244	2.7	9.6
MR t-tau	CSF2	280	3.6	15.3
	Mean		3.2	12.5

Supplementary Table 3 Repeatability and intermediate precision of NTA, NTB and MR ttau in CSF (results from clinical cohorts 1 and 2)

	Sample	Conc (pg/ml)	Repeatability (CVr) %	Intermediate precision (CVRw) %
	LowQC	9	6.5	15.8
NTA t-tau	HighQC	35	4.0	15.0
	Mean		5.3	15.4
	LowQC	50	П	15
NTB t-tau	HighQC	67	10	15.5
	Mean		10.5	15.3
	LowQC	9	6.8	14.4
MR t-tau	HighQC	263	8.0	15.0
	Mean		7.4	14.7

Supplementary Table 4 Spike recovery of NTA, NTB and MR t-tau in CSF

	Sai	mple	Measured (pg/ml)	Mean (pg/ml)	%CV conc	Expected conc. (pg/mL)	Spike recovery (%)
		Neat	6.69 6.70	6.69	0		
	CSFI	High spike	39.8 40.0	39.9	0.2	46.7	77.5
_		Low spike	10.4 11.2	10.8	5.1	16.7	76.0
NTA t-tau		Neat	10.1 10.6	10.3	3.5		
NTA	CSF2	High spike	47.3 45.6	46.4	2.7	50.3	84.2
		Low spike	14.3 14.1	14.2	1.2	20.3	79.5
	Diluent	High spike	46.2 43.4	44.8	4.5	40	
		Low spike	7.68	7.54	2.6	10	
		Neat	48.0 47.4	47.7	0.9		
	CSFI	High spike	312 296	304	3.8	198	157
-		Low spike	135 144 69.6	139	4.6	97.7	133
NTB t-tau		Neat	69.8 322	69.7	0.2		
LN	CSF2	High spike	343 167	332	4.3	220	155
		Low spike	172	170	2.2	120	134
	Diluent	High spike	139 54.7	145	6.2	150	
		Low spike	58.9 226	56.8	5.3	50	
	CSFI	Neat High spike	227 474	226 483	0.2 3.4	376	126
		Low spike	493 316	320	2.1	276	118
au		Neat	324 234	231	1.8	_,,	
MR t-tau	CSF2	High spike	228 494	503	2.7	381	130
-		Low spike	513 323 328	325	I	281	118
		High spike	328 316 313	157	0.8	150	
	Diluent	Low spike	109 73.8	45.7	27.2	50	

Supplementary Table 5 Innotest, MR, NTA and NTB t-tau levels within the non-Alzheimer's disease dementia and acute neurological disorders groups (clinical cohorts 1 and 2)

		No	n-AD dementia			Acute I	neurological	diseases (an	d AD)
	Alcohol related dementia	Vascular dementia	Mixed vascular/non- AD cortical dementia	Unspecified dementia	Ρ	lschemic stroke	Status epilepticus	AD	Ρ
N	3	4	7	8		7	9	112	
CSF Aβ42 (pg/ml)	925 (773- 1249)	1084 (1029- 1302)	5 (663- 92)	1096 (936- 1248)	0.89	984 (645- 1220)*	773 (578- 1287)*	555 (6491- 631)	<0.000
CSF Innotest ® t-tau (pg/ml)	292 (220- 362)	254 (185- 299)	201 (161- 236)#	322 (256- 373)	0.03	2270 (2039- 2500)*	1972 (1446- 2326)*	777 (610- 1020)	<0.000
CSF p-tau 8 (pg/ml)	34.0 (31.0- 45.0)	41.5 (31.0- 46.0)	33.0 (31.0- 34.0)#	43.5 (57.5- 49.0)	0.046	61.0 (56.0- 83.0)*	48.0 (43.5- 66.0)*	103 (85.0- 133)	<0.000
CSF MR t-tau (pg/ml)	37 (3 - 65)	23 (65.2- 35)	77.0 (60.0- 117)#	163 (132- 202)	0.0098	1156 (881- 3219)*	1676 (1234- 2906)*	412 (278- 587)	<0.000
CSF NTA t- tau (pg/ml)	0.73 (0.23- 1.77)	0.87 (0.61- 1.32)	1.48 (0.64-4.9)	2.8 (0.69-5.8)	0.37	112 (67.9- 834)*	128 (69.0- 207)*	9.95 (5.9- 16.2)	<0.000
CSF NTB t- tau (pg/ml)	6.3 (4.7-11.6)	5.68 (5.4- 6.1)#	8.5 (4.5-16.8)	15.6 (8.8- 31.8)	0.033	647 (558- 8265)*	1050 (498- 1881)*	47.7 (22.9- 72.2)	<0.000

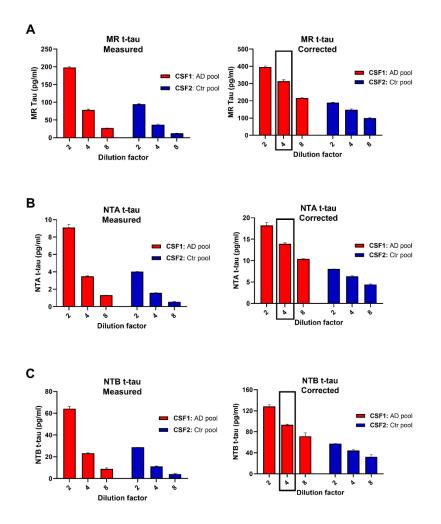
Data presents as median (interquartile range). Significant differences in pairwise comparisons to unspecified dementia (#) and AD (*) are presented. Hepatic encephalopathy and limbic encephalitis were not included in the analysis (n = 1 for both).

	Innotest	MR t-tau	NTA t-tau	NTB t-tau	Р
A β- control	1.0 (0.8-1.2)	0.9 (0.6-1.2)	0.7 (0.5-1.0)	0.7 (0.5-1.1)	0.11
Αβ- ΜΟΙ	1.2 (0.9-1.4)	0.9 (0.7-1.3)	0.9 (0.4-1.3)*	0.7 (0.4-1.2)*	0.0004
Αβ+ ΜCΙ	1.4 (1.3-1.7)	1.2 (1.0-1.3)	3.3 (2.3-4.0)*	2.2 (1.6-2.9)*	< 0.0001
NADD	1.1 (0.9-1.4)	1.0 (0.6-1.2)	0.5 (0.2-1.4)*	0.5 (0.4-1.0)*	0.0048
AD	3.4 (2.7-4.5)	3.3 (2.2-4.7)	3.7 (1.9-6.2)	3.1 (1.5-4.6)	0.064
CJD	29 (11-51)	42 (21-89)	57 (32-102)*, #	133 (56-341)*, #	< 0.0001
AND	9.5 (7.4-10)	11 (7.5-20)	45 (25-84)*, #	61 (33-136)*, #	< 0.0001
PSP	0.9 (0.8-1.2)	0.6 (0.4-1.0)	0.8 (0.3-1.1)	0.6 (0.4-1.1)	0.076

Supplementary Table 6 Fold changes to controls for Innotest, MR, NTA and NTB t-tau (clinical cohorts 1 and 2)

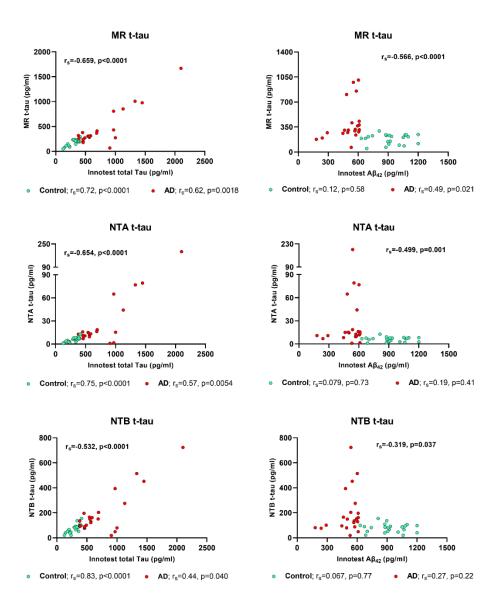
Data presents as median (interquartile range). Significant differences in pairwise comparisons to Innotest t-tau (*) and MR t-tau (#) are presented.

Supplementary figures



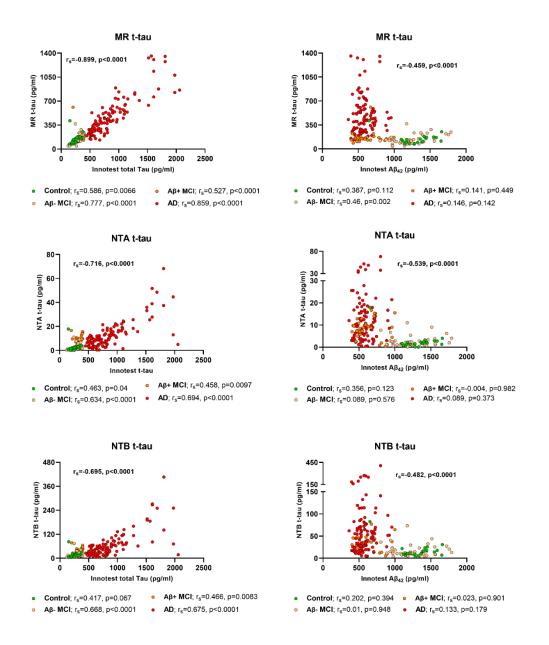
Supplementary Figure 1 Dilution linearity of MR, NTA, and NTB t-tau in CSF

CSF pooled from different Alzheimer's disease (CSF1: AD pool) or control (CSF2: Ctr pool) samples were measured diluted two-, four- and eight-fold. Measured concentrations and concentrations corrected for dilution factors are presented for (**A**) MR t-tau, (**B**) NTA t-tau and (**C**) NTB- t-tau. Sample dilution used for further CSF analysis (1:4) is highlighted in the figure.



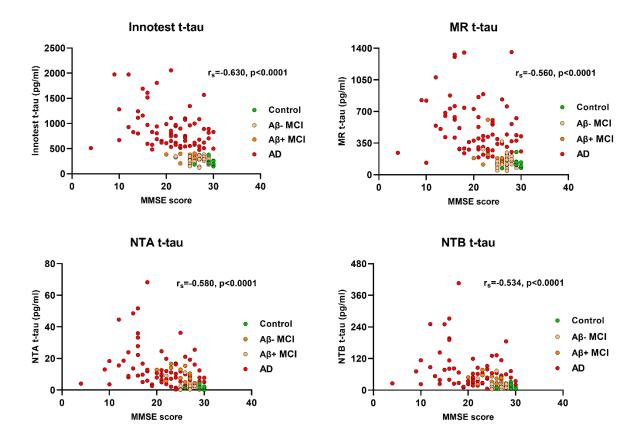
Supplementary Figure 2 Correlation of CSF MR, NTA, and NTB t-tau with Innotest CSF t-tau and CSF β -amyloid₁₋₄₂ (CSF pilot cohort).

Association between developed in-house CSF t-tau biomarkers with Innotest t-tau and Innotest amyloid β_{42} in the CSF pilot cohort was evaluated using Spearman's correlation both in the whole cohort and within diagnostic groups.



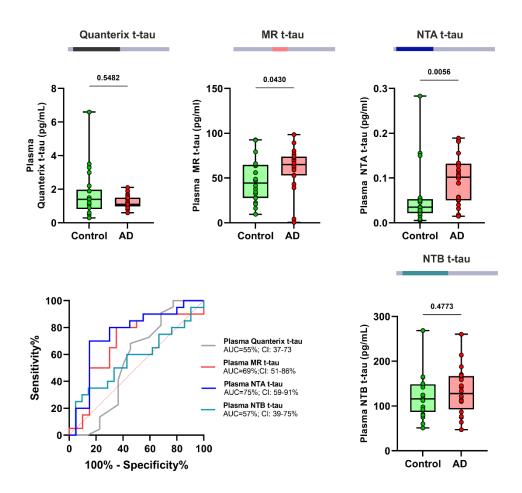
Supplementary Figure 3 Correlation of CSF MR, NTA, and NTB t-tau with Innotest CSF ttau and CSF β-amyloid₁₋₄₂ (CSF clinical cohort 1)

Association between developed in-house CSF t-tau biomarkers with Innotest t-tau and Innotest amyloid β_{42} in the CSF clinical cohort 1 was evaluated using Spearman's correlation both in the whole cohort and within diagnostic groups.



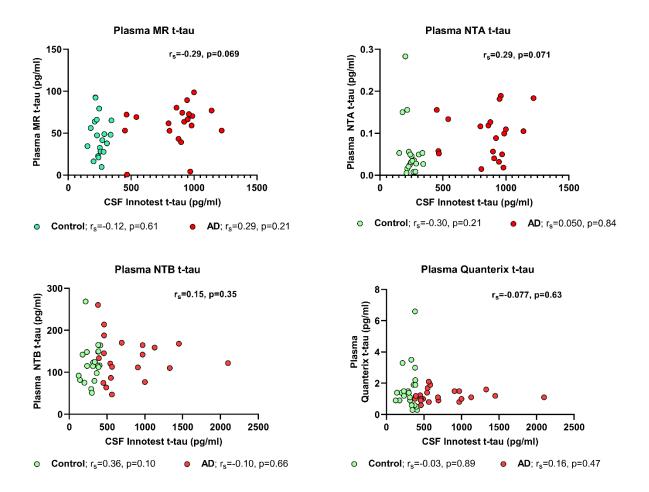
Supplementary Figure 4 Correlation of CSF t-tau biomarkers with Mini-Mental State Examination score (CSF clinical cohort 1)

Association between CSF t-tau biomarkers and Mini-mental state examination (MMSE) score was evaluated using Spearman's correlation in a sub-group of participants from the clinical CSF cohort 1. MMSE scores were available from 127/276 participants (Alzheimer's disease (AD), n = 72; amyloid-positive mild cognitive impairment (A β + MCI), n = 16; amyloid negative (A β -) MCI, n = 28; controls, n = 11).



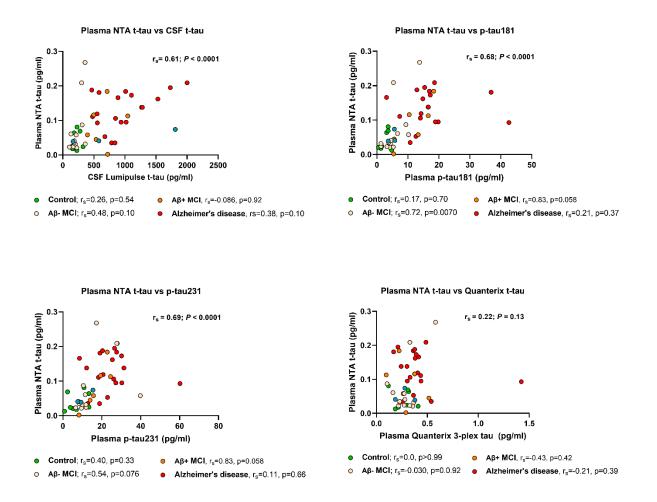
Supplementary Figure 5 Plasma concentrations and diagnostic performance of MR, NTA and NTB t-tau (Plasma pilot cohort)

Boxplots showing plasma MR t-tau, NTA t-tau and Quanterix t-tau concentrations in plasma pilot cohort including samples from biomarker-positive Alzheimer's disease patients (n = 20 for MR and NTA t-tau; n = 22 for NTB and Qanterix t-tau) and control patients (n = 20 for MR and NTA t-tau; n = 22 for NTB and Quanterix t-tau) assessed in the Sahlgrenska University Hospital, Gothenburg, Sweden. Due to limited sample availability, same set of samples was analysed with MR and NTA t-tau, and another set of samples with NTB and Quanterix t-tau (analyzed with single-analyte Tau 2.0 kit, #101552).



Supplementary Figure 6 Correlation of plasma MR, NTA, and NTB t-tau with CSF t-tau (Plasma pilot cohort).

Plasma pilot cohort included samples from biomarker-positive Alzheimer's disease patients (n = 20 for MR and NTA t-tau; n = 22 for NTB and Qanterix t-tau) and control patients (n = 20 for MR and NTA t-tau; n = 22 for NTB and Quanterix t-tau) assessed in the Sahlgrenska University Hospital, Gothenburg, Sweden. Groups were defined as Alzheimer's disease or control patients according to CSF biomarker profiles as described for the pilot CSF cohort. Association between plasma t-tau biomarkers and CSF t-tau (Innotest) was evaluated using Spearman's correlation.



Supplementary Figure 7 Correlation of plasma NTA t-tau with CSF t-tau, plasma p-tau181, plasma p-tau231 and plasma Quanterix t-tau (Plasma clinical cohort).

The cohort included patients with Alzheimer's disease dementia (n = 19), amyloid positive (A β +) mild cognitive impairment (MCI) (n = 6), amyloid negative (A β -) MCI (n = 13), non-Alzheimer's disease dementia (n = 3) and controls (n = 8). Groups were defined according to their CSF biomarker profiles and neuropsychological assessment. Association between plasma t-tau biomarkers and CSF t-tau (Lumipulse) was evaluated using Spearman's correlation.