1 Supplementary materials

3	Prostate-specific Oncogene OTUD6A Promotes Prostatic Tumorigenesis via
4	Deubiquitinating and Stabilizing c-Myc
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17	Supplementary Figures. 1-9
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Supplementary figure 1. c-Myc protein overexpression in prostate cancer (PrCa) patients might be due to both the transcriptional and post-translational regulation.
(A) SPOP, a ubiquitin E3 ligase of c-Myc in PrCa setting is highly mutated in PrCa patients. (B) SPOP expression is not a prognostic marker for PrCa patients. (C) The tumor suppressive FBXW7, another ubiquitin E3 ligase of c-Myc is not frequently mutated in PrCa patients. (D) FBXW7 is not a prognostic marker for PrCa patients.





Supplementary figure 2. The USPs are not physiological DUBs for c-Myc oncoprotein in PrCa cells. (A) Genetic variations, including copy number alternation (CNA) or point mutations, of of USP22, USP28, USP36 and USP37 in human PrCa

32	patients in TCGA database. (B) Genetic variations of USP22, USP28 or USP37 did not
33	predict worse prognosis of PrCa patients. (C. D) Depletion of USP22, USP28, USP36 or
34	USP37 did not change the protein level of c-Myc in PC3 (C) and DU145 (D) PrCa cells.
35	(E) The USP37 mRNA in PrCa cells after depletion of USP37 using CRISPR/Cas9. (F)
36	Depletion of USP DUBs led to reduce in c-Myc in MCF7 breast cancer cells, NCI-H1299
37	lung cancer cells and HeLa cervical cancer cells. (G) Depletion of USP DUBs
38	compromised the tumorigenesis ability of breast and lung cancer cell lines.
39	The relevant raw data are provided in Supplemental Materials.



42 Supplementary figure 3. Screening of physiological DUBs for c-Myc oncoprotein in

43 PrCa setting. (A) Genetic variations, including copy number alternation (CNA) or point
44 mutations, of DUBs in human PrCa. (B-E) The correlation between the genetic variations

- 45 of OTU (**B**), USP (**C**), MINDY (**D**) and JAMM (**E**) DUBs and prognosis of PrCa patients.
- **(F)** Screen pitfall of physiological DUBs for c-Myc oncoprotein in PrCa setting.





57	length (FL) or N-terminal (N) and OTU-domain (OTU) of OTUD6A. (G) c-Myc binds
58	with the OTU domain of OTUD6A. (H, I) Ectopic WT, but not the C152A mutant of
59	OTUDA6 extends the protein half-life of c-Myc in 22Rv1 cells. Immunoblot (H) and
60	quantification (I) of cell lysis derived from 22Rv1 cells transient transfected with either
61	EV, OTUD6A-WT or OTUD6A-C152A, and treated with 200 $\mu g/ml$ of CHX for
62	indicated time. (J) A schematic diagram shows the deubiquitination of c-Myc by
63	OTUD6A.
64	The relevant raw data are provided in Supplemental Materials.



Supplementary figure 5. OTUD6A is a prostatic specific oncogene in PrCa. (A-C)
OTUD6A is not amplified but mutated at very low frequency in breast cancer (BRCA),
lung cancer (LUAD) and colorectal cancer (COADREAD). (D, E) Validation of IHC
staining for OTUD6A (D) and c-Myc antibody (E), in which staining without primary

71	antibody as negative control. (F) The expression patten of OTUD6A in different
72	tissue/organ in human. (G) AR and FOXA1 are predicted transcription factor of OTUD6A
73	based on the prediction of Cistrome. (H) AR binds with the promoter region of OTUD6A
74	in LNCaP PrCa cells. (I) FOXA1 binds with the promoter/enhancer region of OTUD6A
75	in VCaP PrCa cells.



Supplementary figure 6. Depletion of endogenous OTUD6A represses the proliferation and tumorigenesis of PrCa cells. (A, B) Depletion of OTUD6A leads to reduced protein levels of c-Myc in DU145 and PC3 PrCa cells. DU145 (A) and PC3 (B) cells were infected with shControl or shOTUD6A lenti-virus and selected with puromycin for 3 days, followed by IB assay for indicated proteins. (C, D) Depletion of

84	OTUD6A reduces the half-life of c-Myc protein in C4-2 cells. C4-2 Cells were infected
85	with shControl or shOTUD6A virus and selected with puromycin for 3 days, followed by
86	transient transfection with EV or shRNA resistant Flag-OTUD6A constructs. The cells
87	were treated with CHX (200 $\mu\text{g/ml})$ for indicated time before harvest, followed by IB
88	assay for indicated proteins (C) and quantification (D). (E, F) Depletion of OTUD6A
89	suppresses cell growth of DU145 (E) and PC3 cells (F). The cells as in a and b were
90	subjected to growth curve analysis. (G, H) Depletion of OTUD6A suppresses colony
91	growth of DU145 and PC3 PrCa cells. DU145 (G) and PC3 cells (H) as in A and B were
92	subjected to colony formation assay. ***: $P < 0.001$. (I) Overexpression of OTUD6A
93	leads to increased protein level of protein levels of c-Myc in 22Rv1 cells. (J)
94	Overexpression of OTUD6A leads to increased cell proliferation of 22Rv1 cells. (K)
95	Overexpression of OTUD6A leads to an increase in the tumorigenesis of 22Rv1 cells in
96	the colony formation assay. (L) The sequence of sgDNA for CRISPR knockout of
97	OTUD6A. (M, N) Sequencing validation of OTUD6A knockout in 22Rv1 (M) and C4-2
98	cell (N). (O) Depletion of OTUD6A did not compromise the tumorigenesis of MCF7
99	BRCA cells and H1299 LUAD cells. (P) A schematic diagram to show the effect of
100	OTUD6A in promoting PrCa through deubiquitinating c-Myc oncoprotein.
101	The relevant raw data are provided in Supplemental Materials.



Supplementary figure 7. Breeding of Hi-Myc mice with/without depletion of endogenous *Otud6a*. (A) A schematic diagram shows that the breeding strategy for OTUD6A null mice. (B) Representative Genotyping results of OTUD6A WT and null mice. (C) Representative Genotyping results of Hi-Myc mice with/without depletion of

108	endougenous Otud6a. (D, E) Body weight (D) and the weight of prostate tissue (E) of
109	mice as in Fig. 6B. N=7, 7, 5 and 6 for $Otud6a^{+/y}$, $Otud6a^{-/y}$, Hi -Myc; $Otud6a^{+/y}$ and
110	<i>Hi-Myc;Otud6a^{-/y}</i> group, individually. * <i>P</i> <0.05, *** <i>P</i> <0.001. One-way ANOVA.
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Supplementary figure 8. The OTUD6A/c-Myc axis regulates the prostate cancer cell metabolism. (A) Heatmap shows the genes (DEG) in prostate tissue/tumors that increased in Myc-driven mice and further decreased in *Otud6a* knockout mice. (B, C) Depletion of *OTUD6A* led to reduced mRNA expressions of *HK2* and *LDHA* in C4-2 (B) and PC3 PrCa cells (C). These PrCa cells were depleted of OTUD6A by shRNA, and the mRNA levels were measured by qPCR. (D-G) c-Myc expression level is positively

119 correlated with the mRNA levels of LDHA (**D**, **E**) and HK2 (**F**, **G**) in TCGA database.



Supplementary figure 9. Depletion of OTUD6A leads to the reduce in glycolytic
metabolism in PrCa cells. (A, B) Depletion of *OTUD6A* reduced the glycolysis level in
C4-2 (A) and PC3 cells (B). These PrCa cells were depleted of *OTUD6A* by shRNA, and

126 the cancer cell metabolism were measured by XF24 Seahorse extracellular flux analyzer.