Supplemental Online Content

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This supplemental material has been provided by the authors to give readers additional information about their work.

eMethods. Whole-exome sequencing and haplotype analysis

For whole-exome sequencing, we fragmented 1 μg of DNA with sonication technology (Bioruptor, diagenode, Liège, Belgium). The fragments were end-repaired and adaptor-ligated, including incorporation of sample index barcodes. After size selection and pooling, we subjected all libraries to an enrichment process with the SureSelect Human All Exon V6 or V7 kits following manufacturer's procedures (Agilent, Santa Clara, CA, USA). The final libraries were sequenced on an Illumina HiSeq 4000 or an NovaSeq 6000 sequencing instrument (Illumina, San Diego, CA, USA) with a paired-end 2×75 bp or 2x101 bp protocol. The reads were then mapped to the Genome Reference Consortium human genome build 38 (UCSC version hg20) using the Burrows-Wheeler Alignment tool (BWA-mem) (unpublished data, H.L.). The 30X coverage reached 68 - 88% of the target sequences. The Genome Analysis Toolkit (GATK) v.3.8¹ was used to mark duplicated reads, perform local realignment around short insertions and deletions (INDELs), recalibrate the base quality scores, and call SNVs and INDELs. Samtools $v.1.6²$ and Platypus v.0.8.1³ were used as independent short read variation caller. The Varbank2 pipeline v.3.3/v.3.10/v.3.14 and interface, which were developed in-house at the Cologne Center for Genomics, were used for data analysis and filtering (https://varbank.ccg.uni-koeln.de/varbank2, unpublished data, H.T., J.A., and P.N.). All variation calls were filtered for rare, high-quality variants in either a homozygous (AF>0.75) or heterozygous (AF>0.25) state that were predicted to modify a protein sequence or impair splicing. Criteria used to determine high quality GATK UnifiedGenotyper calls were for SNVs: passed VQSR filter, QD>5; MQ>50; FS<40; MQRankSum>−5; ReadPosRankSum>−5 and for INDELs: passed VQSR filter; QD>4; FS<100; ReadPosRankSum>-5. Platypus and Mpileup calls were filtered for QD>5 and MQ>50. Rare variant status was assigned on the basis of an MAF of ≤ 0.01 , according to the highest value in any given population of an in-house GRCh38 lifted version of the gnomAD v.2.1 database⁴.

. Genotyping for haplotype analysis was carried out using GSAMD24v2-0 chips (Ilumina) according to manufacturer's instructions (Infinium HTS Assay protocol, Ilumina). Phasing of the samples genotypes was performed with Eagle 2.4.1⁵ using 1000 Genomes Phase 3 data⁶ as a reference panel. Haplotypes were reconstructed in a 10 megabase region around PADI3 on chromosome 1. Of the 40 individuals, 16 were carriers for a pair of the four analyzed variants, 11 were homozygous carriers for one of the variants and 13 carried one of the four variants in heterozygosity (either unaffected parents or affected individuals carrying an additional PADI3 pathogenic variant that was not assessed by haplotype analysis). In detail, haplotypes of 22 individuals (of whom 2 were homozygous carriers) were analyzed for c.335T>A (p.Leu112His), haplotypes of 26 individuals (of whom 8 were homozygous carriers) were analyzed for c.881C>T (p.Ala294Val), haplotypes of 4 heterozygous variant carriers were analyzed for c.1813C>A (p.Pro605Thr), and haplotypes of two individuals (of whom one was a homozygous carrier) were analyzed for c.505C>T (p.Gln169^{*}).

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eFigure 1. Clinical manifestation of UHS in additional individuals and the respective underlying genotypes

Additional individuals with UHS carrying bi-allelic pathogenic variants in PADI3. Their respective genotypes are (a) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (b) c.335T>A (p.Leu112His) homozygous, (c) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (d) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (e) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (f) c.881C>T (p.Ala294Val) & c.505C>T (p.Gln169*), (g) c.881C>T (p.Ala294Val) homozygous, (h) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (i) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (j) c.335T>A (p.Leu112His) & c.652G>A (p.Gly218Ser), (k) c.881C>T (p.Ala294Val) homozygous, (l) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (m) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (n) c.335T>A (p.Leu112His) & c.1813C>A (p.Pro605Thr), (o) c.881C>T (p.Ala294Val) homozygous, (p) c.881C>T (p.Ala294Val) homozygous, (q) c.335T>A (p.Leu112His) & c.652G>A (p.Gly218Ser), (r) c.505C>T (p.Gln169*) homozygous, (s) c.335T>A (p.Leu112His) & c.881C>T (p.Ala294Val), (t) c.335T>A (p.Leu112His) & c.881C>T $(p.A$ la294 V al).

eFigure 2. 3-Dimensional protein modeling of the pathogenic stop and frameshift variants identified in PADI3

The wild type protein (left) in comparison to the previously identified mutant Lys578* and the newly identified mutant proteins Glu395Ansfs*7 and Gln169*, respectively. It should, however, be noted that the mutant proteins, particularly the latter two with earlier premature stop codons, may not be translated at all due to a nonsense mediated mRNA decay.

eFigure 3. 3-Dimensional protein modeling of the three most common PADI3 variants

The top panel shows the wild type protein at the respective site of the substitution p.Leu112His (left), p.Ala294Val (middle) and p.Pro605Thr (right). The bottom panel shows the site of substitution in the respective mutant proteins.

eFigure 4. Genetic screening of two pedigrees suggesting an autosomal dominant inheritance

Mutation analysis revealed in both of the cases that affected parents also carry two pathogenic PADI3 variants.

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eTable 1. Pathogenic PADI3 variants underlying UHS

The newly identified pathogenic variants since our previous report are depicted in bold. Note that the countries of individuals who carry two different variants are represented twice (e.g. only one individual from Chile participated in the study who carried c.881C>T (p.Ala294Val) and c.1114A>G (p.Arg372Gly)). Exception to this are individuals with parents of different countries of origin and for whom we could link the inheritance of the two mutations to the respective parent by genetic screening of the whole family (e.g. c.1115G>T (p.Arg372Met)).

*c.60_66delinsTGCTTGG (p.Gly22Trp) may also represent a rare haplotype composed of three single nucleotide variants, namely, rs1360902614, rs1221559173 and rs1344213588 which have been observed in the same single individual (minor allele frequency=6.5e-6) in gnomAD.

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eTable 2. The variable extent of sharing across individual haplotypes carrying c.505C>T (p.Gln169*)

The start and end positions are based on hg19.

eTable 3. The variable extent of sharing across individual haplotypes carrying c.335T>A (p.Leu112His)

The start and end positions are based on hg19. *These two haplotypes are from an individual born to consanguineous parents and are thus inherently representing the same information.

The start and end positions are based on hg19.

eTable 5. The variable extent of sharing across individual haplotypes carrying c.1813C>A (p.Pro605Thr)

The start and end positions are based on hg19.

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