

Supplementary Materials for  
**A neurogenetic mechanism of experience-dependent suppression  
of aggression**

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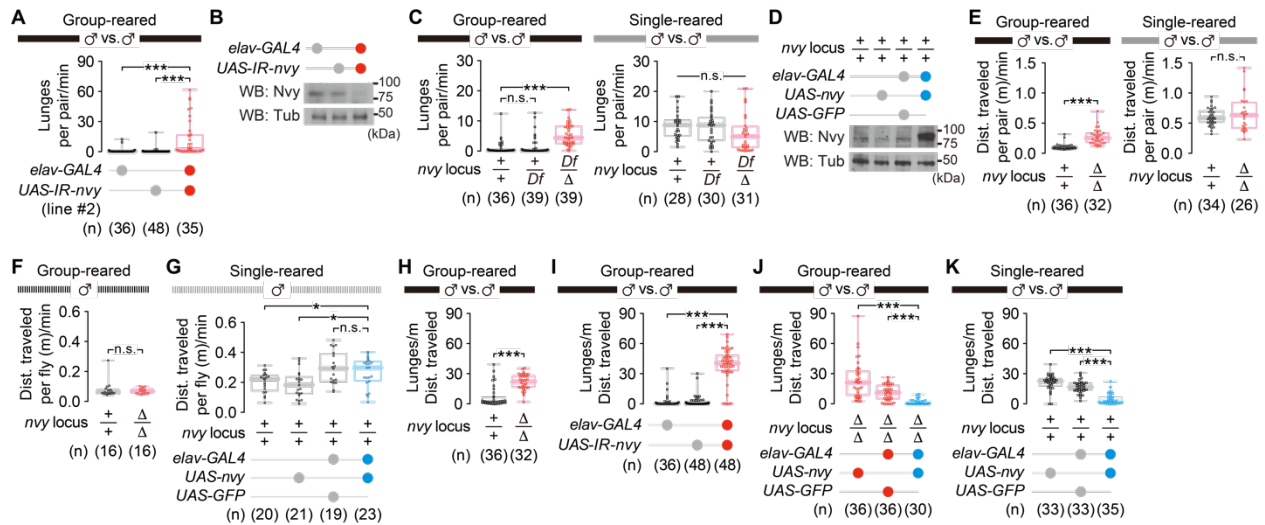
**The PDF file includes:**

Figs. S1 to S13  
Table S3  
Legends for tables S1 and S2, S4 to S8  
Legends for movies S1 to S10  
Legends for data S1 and S2  
References

**Other Supplementary Material for this manuscript includes the following:**

Tables S1 and S2, S4 to S8  
Movies S1 to S10  
Data S1 and S2  
Source data

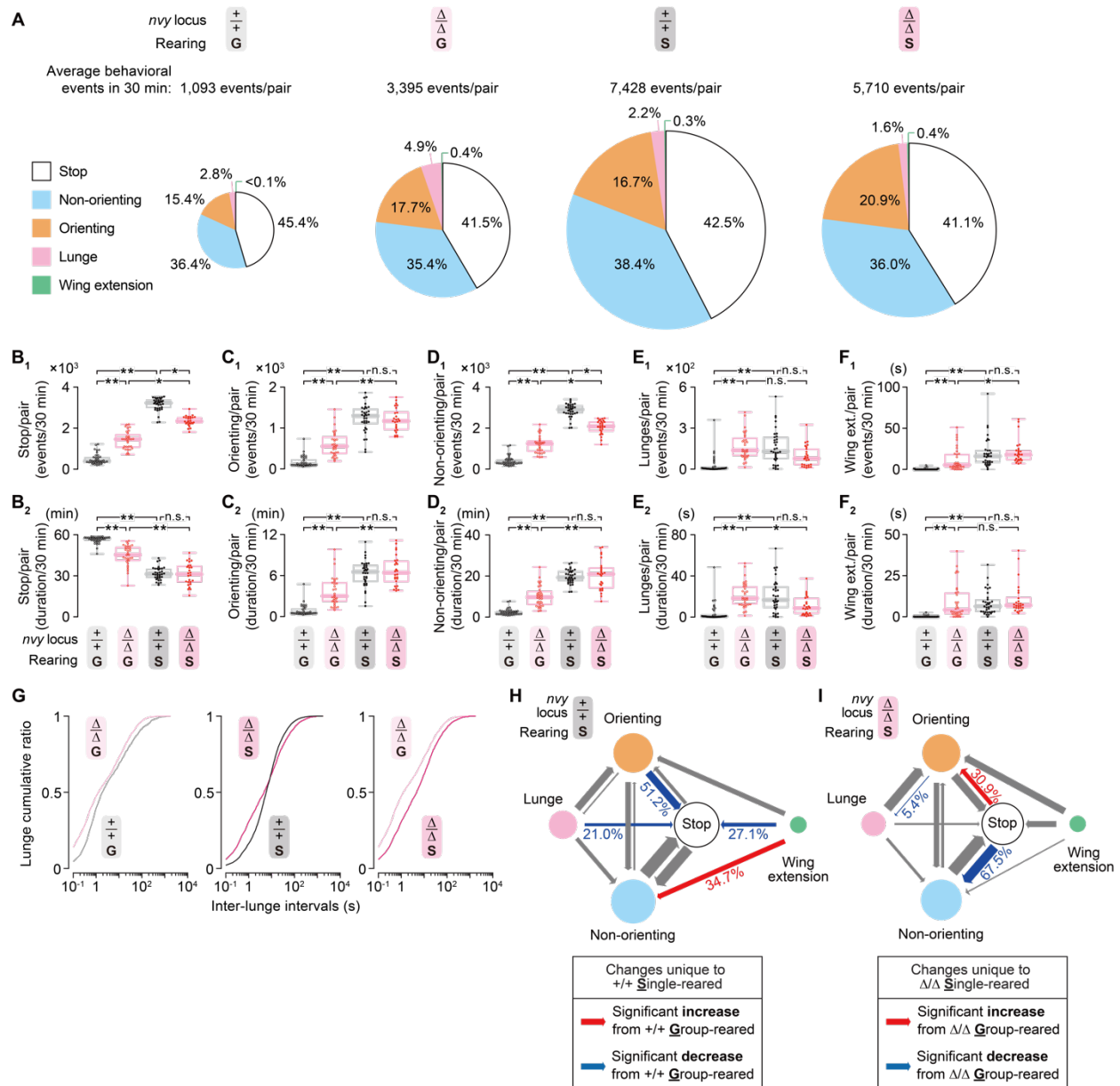
## Supplementary Figure S1.



**Fig. S1. Additional behavioral characterization of *nvyl* mutants.**

(A) Increased lunges performed by males following pan-neuronal *nvyl* knockdown by another *UAS-IR* strain (JF03349). (B) Reduced expression of Nvy protein in fly heads following the pan-neuronal *nvyl* knockdown, verified by Western blot.  $\alpha$ -Tubulin (Tub) was used as an internal control. (C) Transheterozygous *nvyl* mutation increased lunges in group-reared (left) but not in single-reared (right) males. *Df*, *Df(2R)Exel6082* (a deficiency that covers the *nvyl* locus, see Fig. 1C);  $\Delta$ ,  $\Delta nvyl$ . (D) Pan-neuronal Nvy overexpression in fly heads verified by Western blot. (E) Distance traveled in pairs of  $\Delta nvyl$  males showed an increase after group-rearing (left) but not in single-rearing (right), similarly to lunge numbers. (F) Locomotion of solitary  $\Delta nvyl$  males (i.e., not as a pair) was comparable to the wild-type control. (G) Pan-neuronal *nvyl* overexpression failed to impair the locomotion of solitary males. (H to K) Lunge numbers normalized by distance traveled in male pairs of  $\Delta nvyl$  (H), pan-neuronal *nvyl* RNAi (I), neuron-specific rescue of  $\Delta nvyl$  (J), and *nvyl* overexpression (K) showed significant changes consistently with their respective phenotypes in raw lunge counts (see Fig. 1 for comparison). \*\*\*  $p < 0.0005$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(A,C,E,F-K), Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction.]

### Supplementary Figure S2.

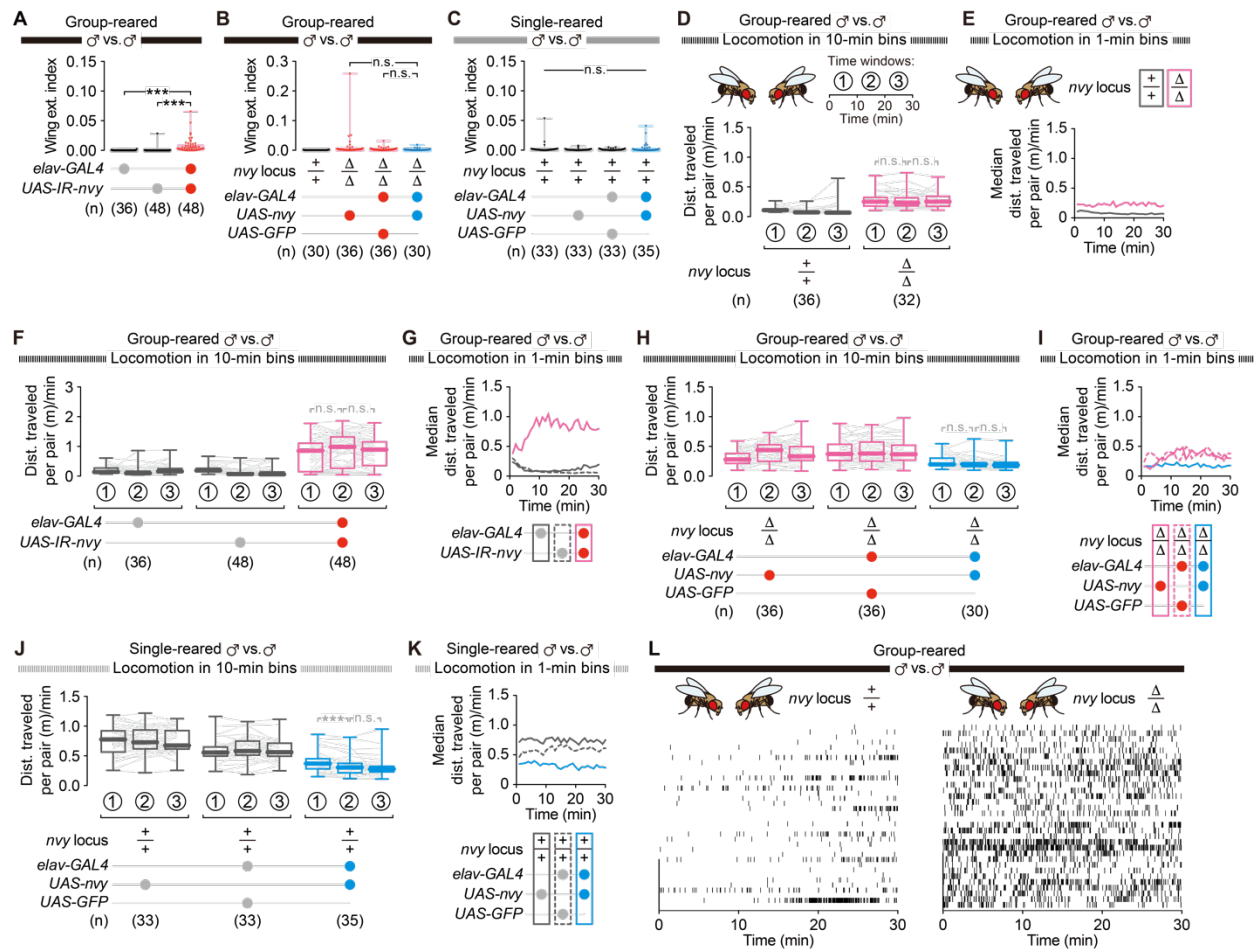


**Fig. S2. Additional analysis related to behavioral transitions in wild-type and  $\Delta nvyl$  males.**

(A) Breakdown of 5 classified behaviors performed by (from left) group-reared  $\Delta nvyl$  males, group-reared wild type males, single-reared  $\Delta nvyl$  males, and single-reared wild-type males. The diameter of pie charts is proportional to the square root of behavioral events per pair. (B to F) Quantification of the number (B<sub>1</sub> to F<sub>1</sub>) and the total duration (B<sub>2</sub> to F<sub>2</sub>) of stopping (B), orienting (C), non-orienting (D), lunge (E), and wing extension (F) of group-reared wild-type males, group-reared  $\Delta nvyl$  males, single-reared wild-type males, and single-reared  $\Delta nvyl$  males. Note that the maximum value of “duration/30 min” in B<sub>2</sub>–F<sub>2</sub> is 60 minutes per pair to account for the combined values from both flies. (G) Cumulative plots of inter-lunge intervals in group-reared wild type males (gray), group-reared  $\Delta nvyl$  males (light pink), single-reared wild-type males (black), and single-reared  $\Delta nvyl$  males (dark pink). Note that the x-axis (intervals) is scaled

in logarithm. **(H, I)** Ethograms between 5 classified behaviors for single-reared wild-type (H) (replot of Fig. 2C) and single-reared  $\Delta nvy$  (I) males. Numbers represent transition probabilities from the source of arrows. \*\*  $p < 0.005$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(B,D,F,G) Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction, comparing all combinations]

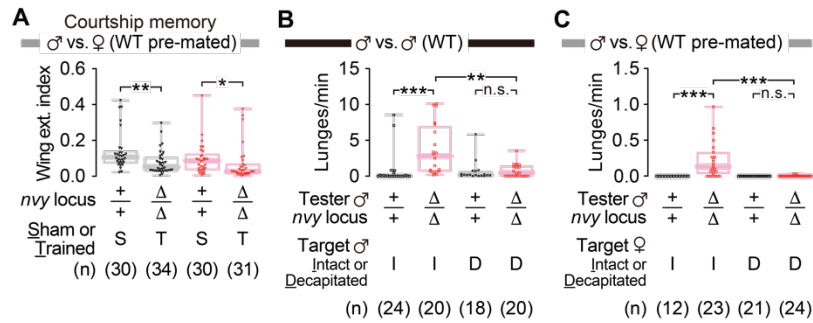
### Supplementary Figure S3.



**Fig. S3. Temporal dynamics of locomotor and aggressive behaviors in *nvy* mutants.**

(A to C) Male-to-male wing extensions analyzed from the movies used in Fig. 1. RNAi of *nvy* increased the wing extension duration (A), whereas  $\Delta nvy$  rescue (B) and *nvy* overexpression did not (C). (D to K) Temporal changes of locomotor activities in fly pairs during aggression assays in Fig. 1. Locomotion was analyzed in detail for  $\Delta nvy$  mutants (D and E), *nvy* RNAi (F and G),  $\Delta nvy$  rescue (H and I), and *nvy* overexpression (J and K), each in 10-min (D, F, H, J) or 1-min (E, G, I, K) bins. (L) Raster plots of lunges performed by either wild-type (left) or  $\Delta nvy$  male pairs shown in Fig. 1D. \*\*\*  $p < 0.0005$ , n.s.  $p \geq 0.05$  [(A), (B), (C), in black: Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction; (D), (F), (H), (J), in gray: Kruskal–Wallis one-way ANOVA and post-hoc Wilcoxon signed rank test.]

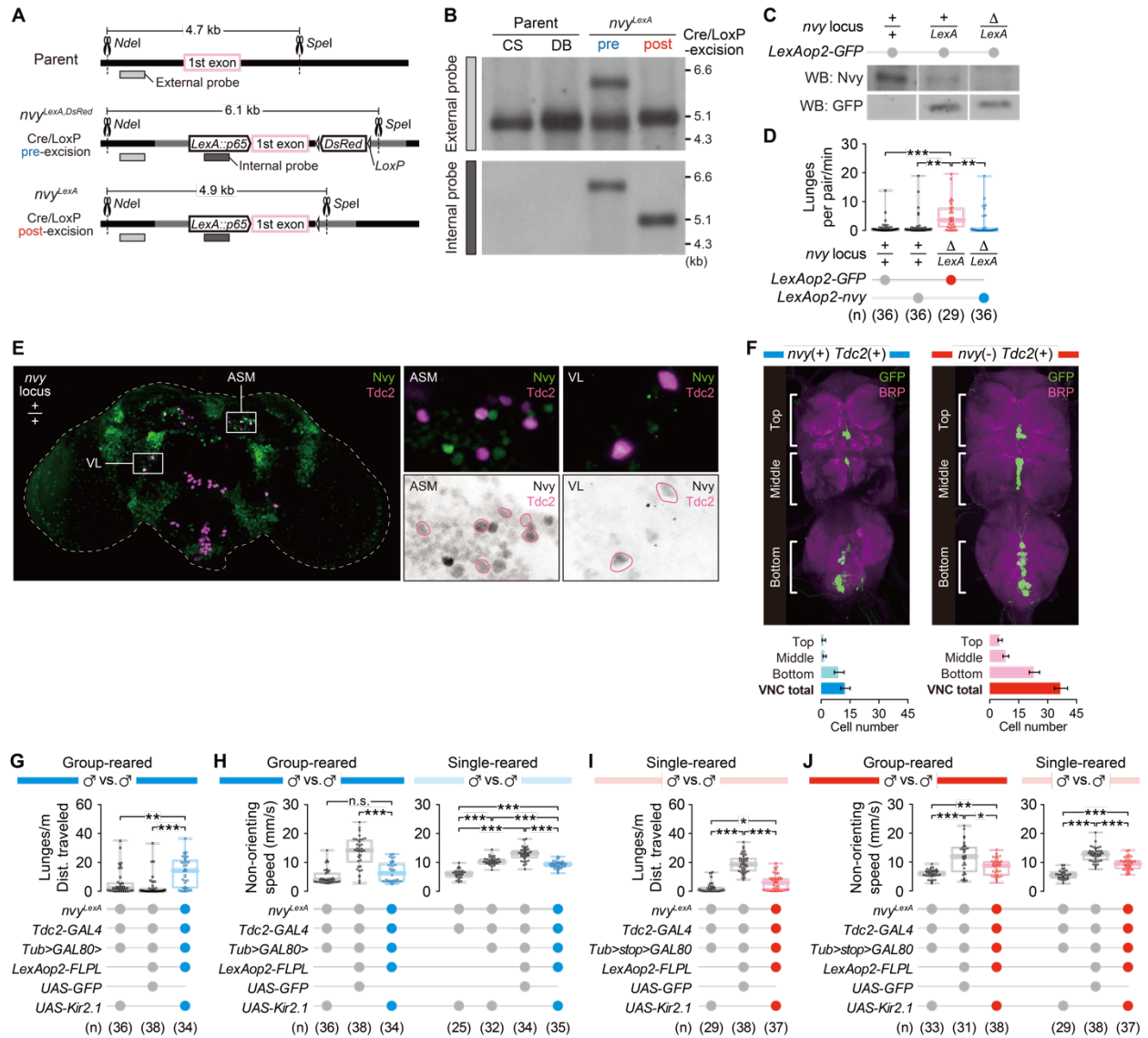
## Supplementary Figure S4.



**Fig. S4. Additional characterization of the behavior of  $\Delta nvy$  males toward a female**

(A) Wing extension indices of males during the test session of courtship memory assay. Males were previously either trained with pre-mated females (“T”) or sham-trained (“S”). (B, C) Aggression by  $\Delta nvy$  males requires behavioral feedback from the opponent. Lunges in 30 min by either wild-type or  $\Delta nvy$  tester males, toward intact (“I”) or decapitated (“D”) wild-type target males (B) or pre-mated females (C). \*\*\*  $p < 0.0005$ , \*\*  $p < 0.005$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(A), (B), (C), Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction.]

## Supplementary Figure S5.



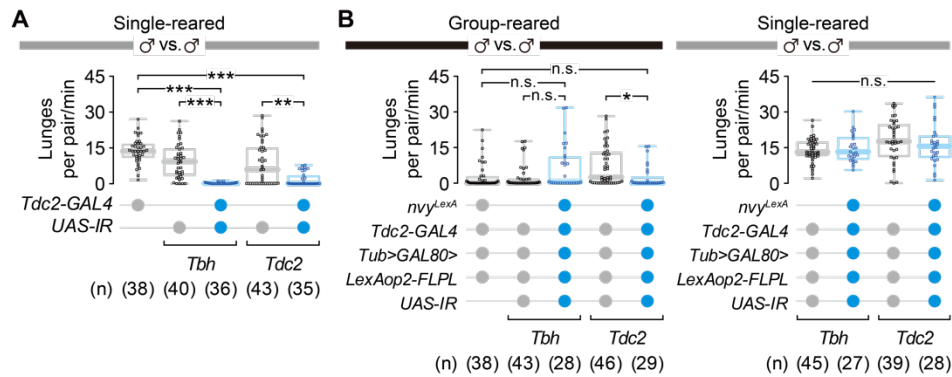
**Fig. S5. Generation of the *nvy<sup>LexA</sup>* knock-in lines, and additional histological/behavioral characterization of *nvy*-expressing neurons.**

(A) Genome schematics of the *nvy<sup>LexA</sup>* knock-in alleles. The *nvy* locus of the parental line (top) was targeted by CRISPR/Cas9-mediated cleavage, leading to homologous recombination with the plasmid harboring the coding sequences of *LexA:p65* and the eye-specific genetic marker *3XP3-DsRed* (middle). After backcrossing with the wild-type strain, the *DsRed* marker flanked by *LoxP* was excised by crossing with an *hs-Cre* line (bottom). Predicted distances between the recognition sites of two restriction enzymes, *NdeI* and *SpeI*, are shown for each genotype. For the following Southern blot analysis, one region outside the *nvy* exon and another region inside the *LexA:p65* coding sequence were targeted by “external” and “internal” probes, respectively. (B) Southern blot analysis of the parental and *nvy<sup>LexA</sup>* knock-in lines. *NdeI/SpeI*-digested genomic DNA from each line was hybridized with either the external (top) or internal (bottom) probe. For parental lines, the wild-type Canton-S (CS) used for backcrossing and the “double-balancer”

(DB: *w*; *Bl/CyO*; *TM2/TM6B*) used to establish the knock-in lines are shown. Note that the *nvy<sup>LexA</sup>* knock-in lines were maintained with the second chromosome balancer *CyO* derived from the parental DB line. (C) Western blot analysis of Nvy protein extracted from the *nvy<sup>LexA</sup>* fly heads. (D) Hyperaggressive phenotype induced by trans-heterozygosity of *nvy<sup>LexA</sup>* and  $\Delta nvy$ , and its rescue by *nvy* expression. (E) Immunohistochemistry using anti-Nvy antibody reveals a broad expression pattern in the fly brain with signals colocalized with *Tdc2-GAL4*-driven tdTomato. Representative areas containing two OA/TA subpopulations (ASM and VL) mainly co-labeled by *nvy<sup>LexA</sup>* and *Tdc2-GAL4* (see Fig. 4C and 4D) are shown at high magnification. (F) GFP expression in *nvy*-positive (left) or *nvy*-negative (right) *Tdc2* neurons in the VNC is visualized by immunohistochemistry. VNC was divided into three regions (“top” being closest to the head) and cell counts for each area were shown as means  $\pm$  S.D. of 5 brains. (G to J) Locomotion changes followed by *Kir2.1*-mediated silencing of *Tdc2* subpopulations, analyzed from the movies used in Fig. 4E and 4G. Changes in lunge numbers normalized by distance traveled in group-reared *nvy*-positive *Tdc2* neurons (G) and single-reared *nvy*-negative *Tdc2* cells (I) were consistent to the raw lunge results shown in Fig. 4. Speed during non-orienting locomotion changed irrespectively to lunge patterns in flies expressing *Kir2.1* in *nvy*-positive *Tdc2* neurons (H), whereas both correlated with each other when *Kir2.1* was expressed in *nvy*-negative *Tdc2* neurons (J). \*\*\*  $p < 0.0005$ , \*\*  $p < 0.005$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(D) Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction.]



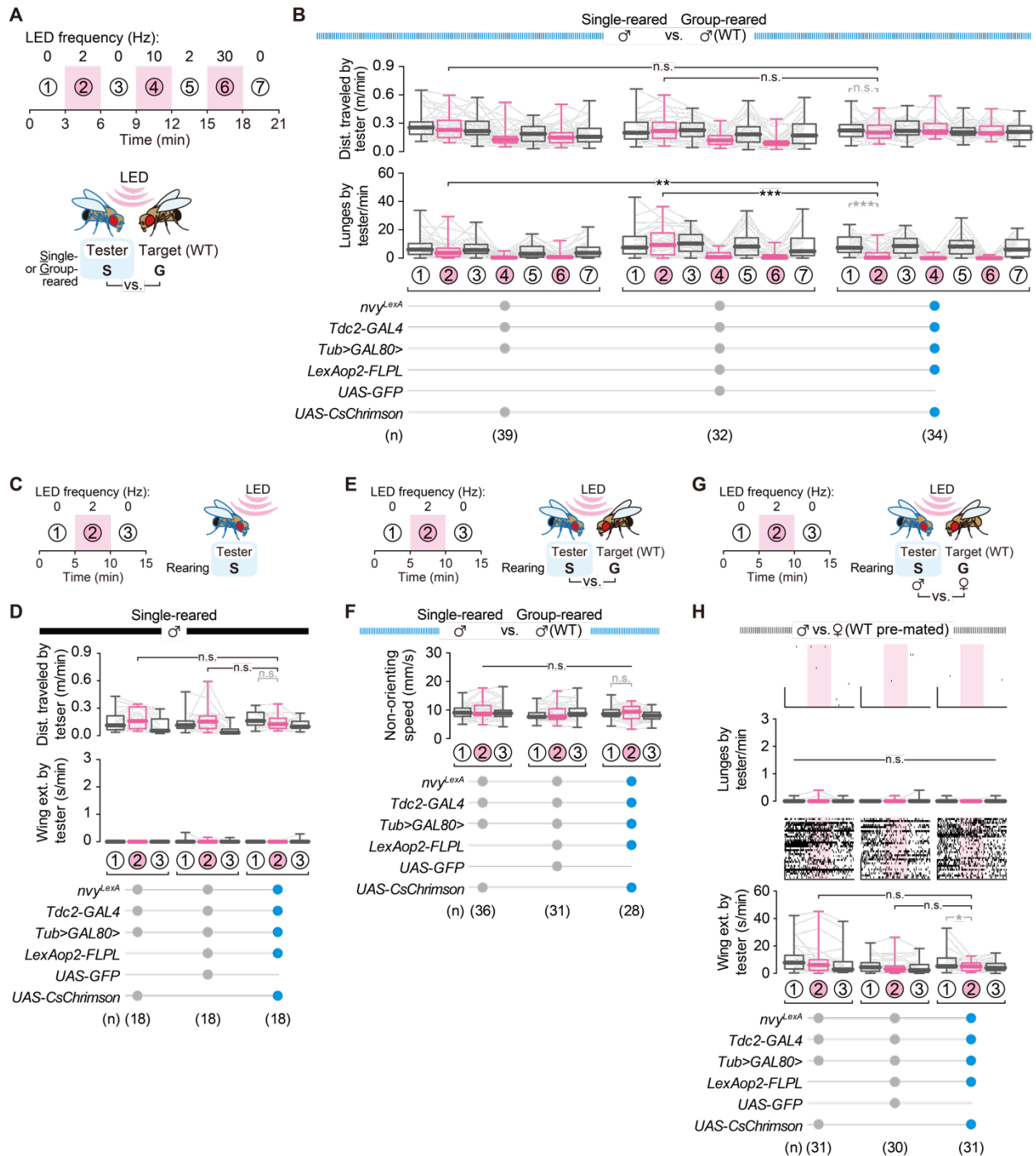
## Supplementary Figure S6.



**Fig. S6. Knock-down of octopamine/tyramine biosynthesis genes in the *nvyl*-positive *Tdc2* neurons.**

(A) Decrease in lunges by single-reared males after *Tbh* and *Tdc2* RNAi in the entire *Tdc2* neurons. (B) Neither RNAi of *Tbh* nor *Tdc2* in the *nvyl*-positive *Tdc2* neurons significantly affected aggressiveness of group- (left) or single-reared (right) males. \*\*\*  $p < 0.0005$ , \*\*  $p < 0.005$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(A), (B), Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction.]

## Supplementary Figure S7.

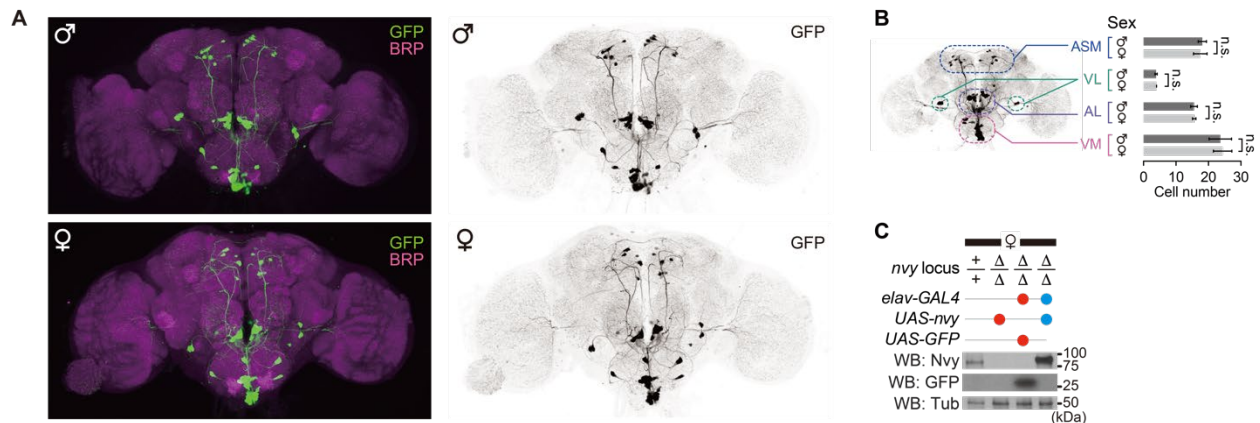


**Fig. S7. Additional behavioral data obtained during optogenetic stimulation of *navy*-positive *Tdc2* neurons.**

(A, B), Optogenetic stimulation of *navy*-positive *Tdc2* neurons at various LED frequencies. The stimulation was performed at 2, 10, and 30 Hz for 3 min, each separated by a 3-min interval (A). Distance traveled (B; top) and lunges (B; bottom) performed by tester males during each 3-min time window are shown in box plots. (C, D) Optogenetic stimulation of *navy*-positive *Tdc2*

neurons in solitary testers. Stimulation was performed at 2 Hz for 5 min in the absence of a target fly (C). Distance traveled (D; top) and wing extensions (D; bottom) performed by tester males during each 5-min time window. (E, F) Speed of the “non-orienting” locomotion during optogenetic stimulation of *nvγ*-positive *Tdc2* neurons. The original movies used in Fig. 4I were reanalyzed. (G, H) Male-to-female lunges and wing extensions during optogenetic stimulation of *nvγ*-positive *Tdc2* neurons. Male testers were paired with wild-type pre-mated females, and the stimulation was performed at 2 Hz for 5 min (G). Lunges (H, top) and wing extensions (H, bottom) performed by target males during each 5-min window. The pink area within each raster plot indicates the stimulation period (time window “2” in G). \*\*\*  $p < 0.0005$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(B), (D), (F), (H), in black: Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction; in gray: Kruskal–Wallis one-way ANOVA and post-hoc Wilcoxon signed rank test.]

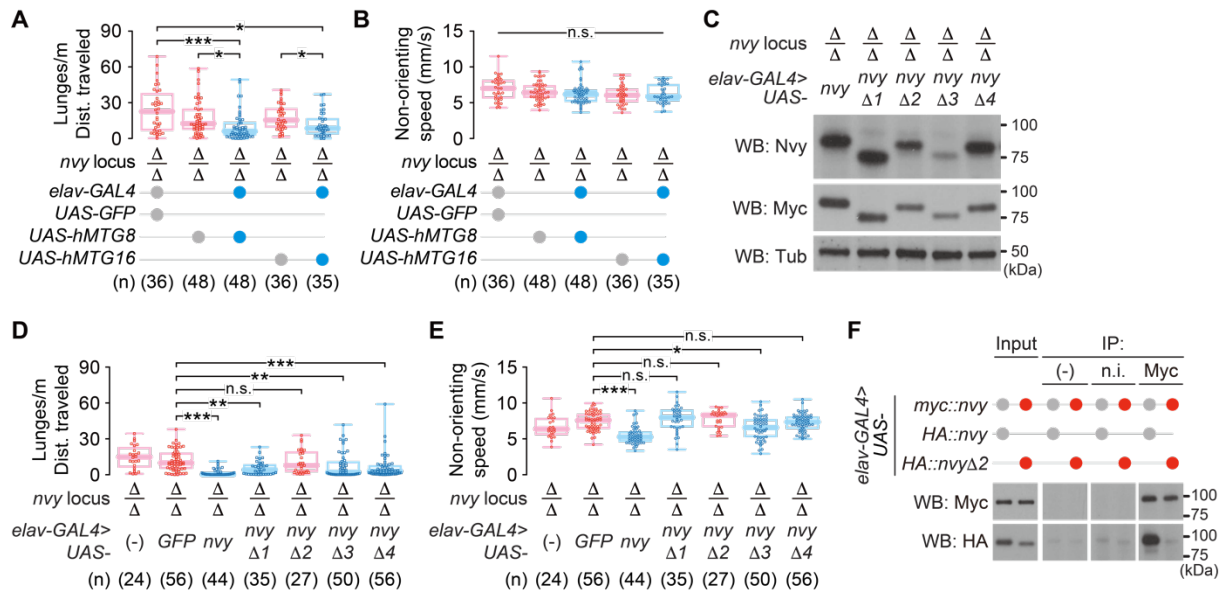
## Supplementary Figure S8.



**Fig. S8. Gross morphology of *Tdc2* neurons is similar across sexes.**

(A) Neuronal morphology of *Tdc2* neurons in male and female brains. Left: GFP expressed under the control of *Tdc2-GAL4*, along with the neuropil marker Bruchpilot (BRP), were visualized by immunohistochemistry using male (top) or female (bottom) brains. Right: same images as left with GFP signals visualized in gray scale. (B) Cell counts of *Tdc2* neurons in males and females. Subtypes of *Tdc2-GAL4* neurons were classified according to a previous anatomical study (65). (C) Pan-neuronal expression of Nvy in  $\Delta nvy$  females verified by Western blot.  $\alpha$ -Tubulin (Tub) was detected as an internal control. n.s.  $p \geq 0.05$  [(B), unpaired t-test; error bars indicate means  $\pm$  S.D. of 8–9 brains.]

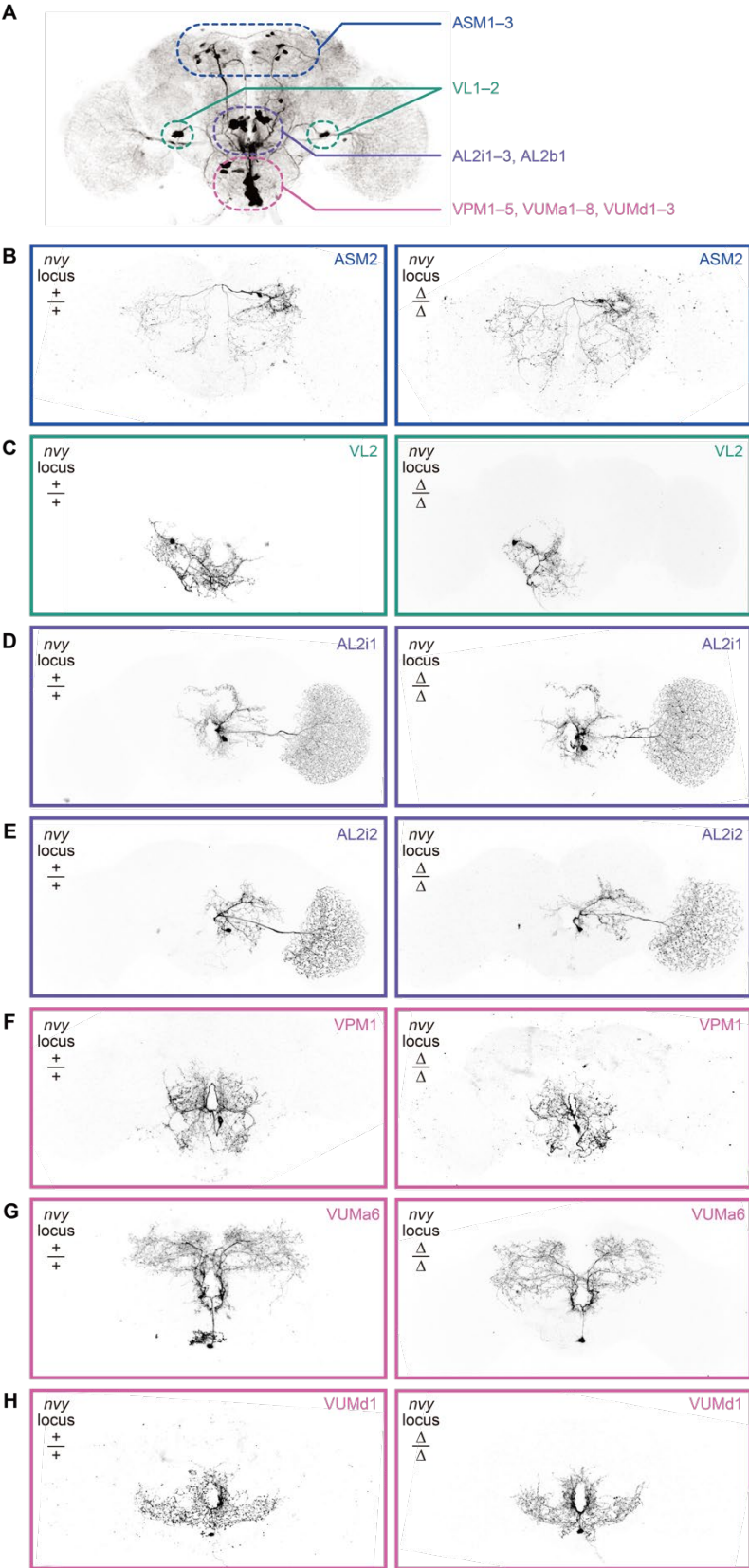
## Supplementary Figure S9.



**Fig. S9. Additional biochemical and behavioral data for human MTGs and truncated versions of Nvy.**

(A, B) Locomotion by *MTGs*-expressing male pairs analyzed in Fig. 6C. Pan-neuronal expression of human *MTGs* in the  $\Delta navy$  background significantly reduced lunge numbers even after normalization by the locomoted distance per pair (B), whereas walking speed during the “non-orienting” state was barely affected (B). (C) Pan-neuronal expression of mutated *navy* transgenes lacking one of the NHR1–4 domains in the  $\Delta navy$  background. All *UAS-navy* constructs contain 3xMyc tags at the N-terminus.  $\alpha$ -Tubulin (Tub) was used as an internal control. (D, E) Locomotion by male pairs expressing truncated *UAS-navy* constructs analyzed in Fig. 6E. Only *UAS-navy* lacking the NHR2 domain (*navy* $\Delta 2$ ) failed to rescue the  $\Delta navy$  phenotype for lunge numbers normalized by distance traveled per pair (D), which is in agreement with the raw lunge result in Fig. 6E. The *UAS-navy* $\Delta 2$  expression did not have a significant impact on the non-orienting walking speed of the same pairs (E). (F) Homo-multimer formation of Nvy protein mediated by the NHR2 domain. Myc-tagged Nvy was co-immunoprecipitated with either HA-tagged intact Nvy or the mutated version lacking NHR2. Input: 7.5% of lysate used for the precipitation. IP, (-): samples precipitated with no antibody. IP, n.i.: samples precipitated with normal IgG. IP, Myc: samples precipitated with an anti-Myc antibody. \*\*\*  $p < 0.0005$ , \*\*  $p < 0.005$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(A,B,D,E), Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction.]

**Supplementary Figure S10.**

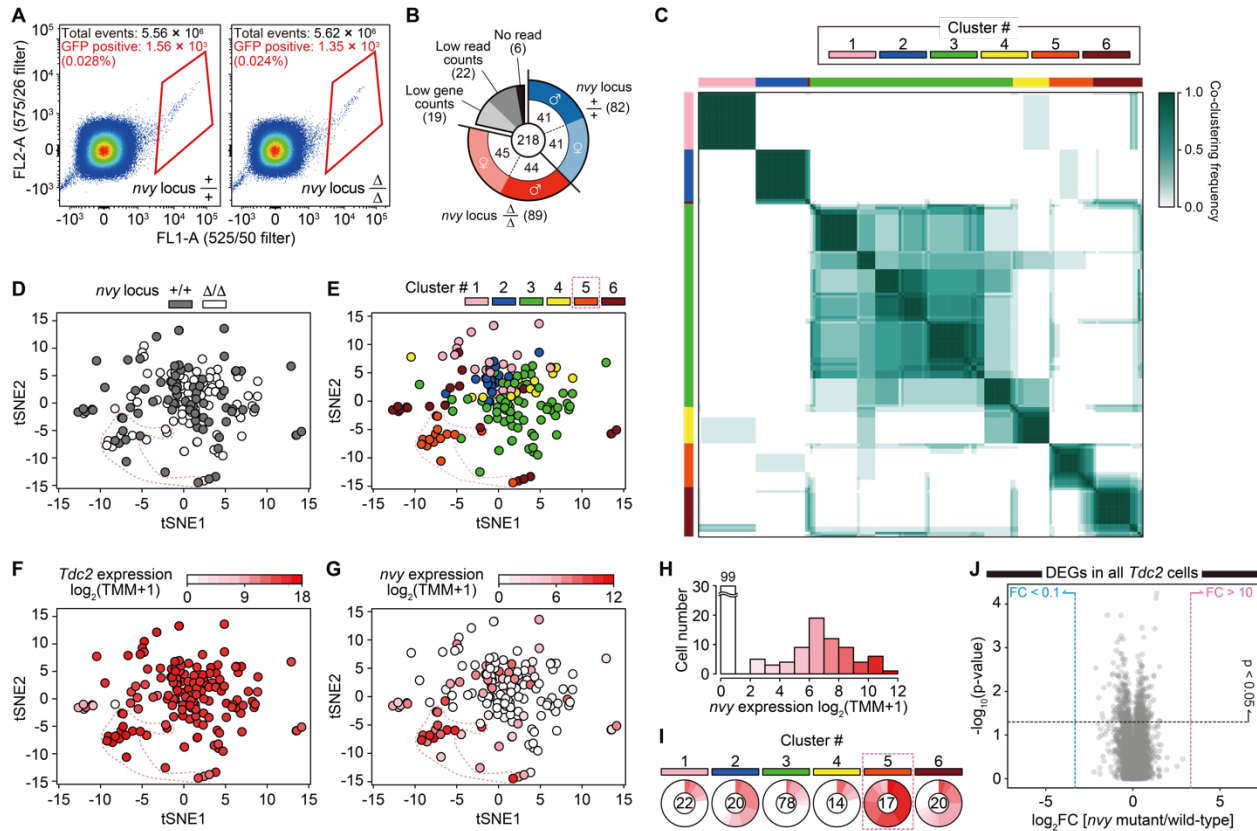


**Fig. S10. *Tdc2* neurons in  $\Delta nvy$  males largely retain their anatomical characteristics.**

(A) Locations of *Tdc2* neuronal clones identified in both wild-type and  $\Delta nvy$  males.

Nomenclature is based on (65). The image is reproduced from Fig. 4C. (B to H) Images of single *Tdc2* neuronal clones produced by MultiColor FlpOut, from wild type (left) and  $\Delta nvy$  (right) males. Names of cell types are indicated at the top right corner.

## Supplementary Figure S11.

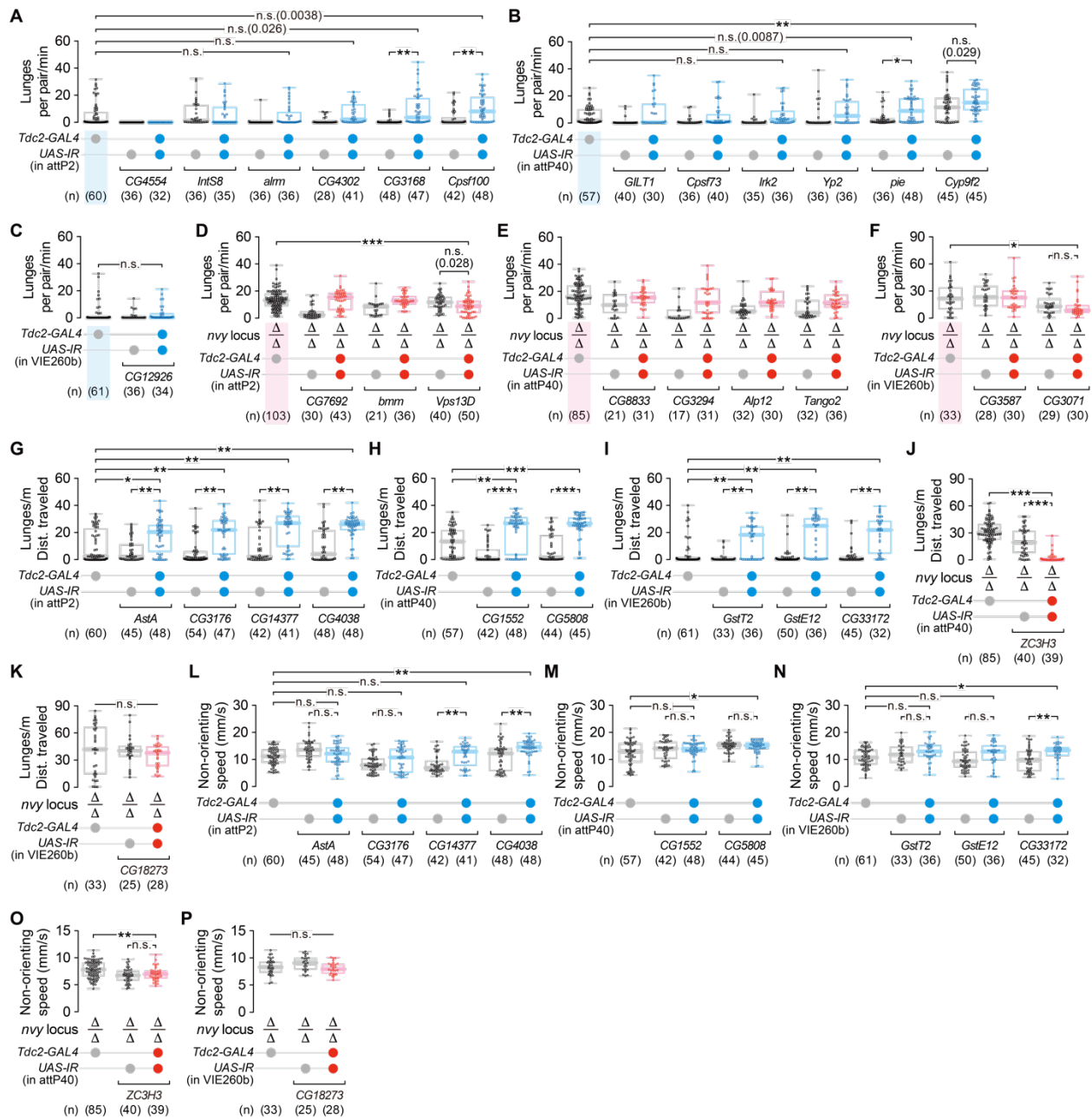


**Fig. S11. Additional analyses of cell clusters and DEGs from single-cell RNA-sequencing of *Tdc2* neurons.**

(A) FACS results for GFP-labeled *Tdc2* cells. GFP-positive cells inside the red lines were collected for sequencing. (B) Number of cells used in the sequencing analysis. (C) Co-clustering frequency matrix from the iterative clustering analysis with 100 random samplings. The plot shows the probability of co-occurrence in the same cluster for given pairs of cells. (D to G) tSNE plots of sequenced *Tdc2* cells, color-coded for the *nvy* locus genotypes (D; wild-type in black,  $\Delta nvy$  in white), cell clusters (E), and expression levels of *Tdc2* (F) or *nvy* (G). (H) Histogram of *Tdc2* cells according to the expression level of *nvy*. (I) Ratio of *nvy*-expressing cells within each cluster. Red intensity corresponds to the level of *nvy* expression shown in G and H. Total cell numbers for each cluster are shown at the center. (J) A volcano plot of DEGs analyzed in all *Tdc2* cells. Dots are plotted according to the fold change (FC) and the p-value (by Mann-Whitney U-test) of each gene when the  $\Delta nvy$  mutant cells were compared against the wild-type cells.



## Supplementary Figure S12.

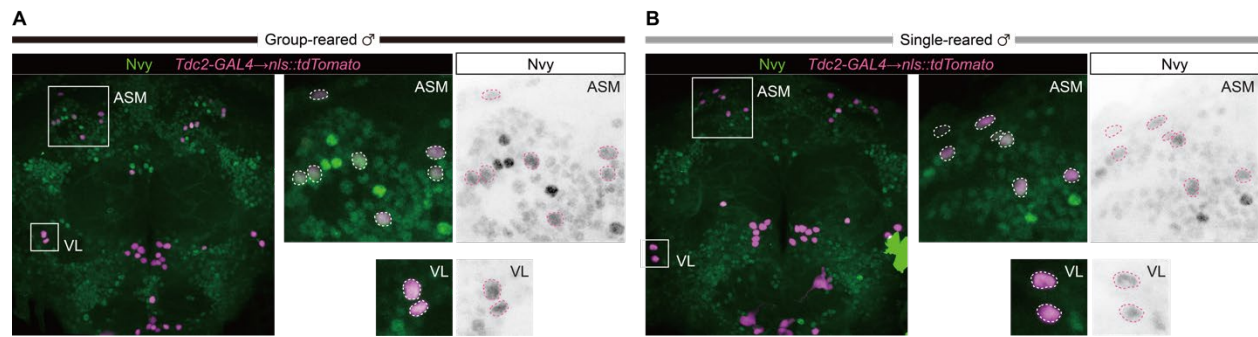


**Fig. S12. Additional behavioral data related to RNAi of DEGs in *Tdc2* neurons.**

(A to C) Lunges performed by males with *Tdc2-GAL4* driving RNAi constructs of down-regulated DEGs in cluster #5 that did not elevate aggression compared to at least one genetic control. *UAS-IR* constructs were inserted either in attP2 (A), attP40 (B), or VIE260b (C). (D to F) Lunges performed by  $\Delta nvy$  males with *Tdc2-GAL4* driving RNAi constructs of up-regulated DEGs in cluster #5 that did not reverse high levels of aggression in the homozygous  $\Delta nvy$  background compared to at least one genetic control. *UAS-IR* constructs were inserted either in attP2 (D), attP40 (E), or VIE260b (F). Colored boxes on the genotypes indicate that the genetic control results (i.e., *Tdc2-GAL4*-only controls with empty vectors inserted in each landing site)

were replotted in Fig. 7 as a part of the same experiment. **(G to K)** Lunge numbers normalized by distance traveled in the male pairs following *Tdc2-GAL4*-driven RNAi against up- (G to I) and down-regulated (J to K) DEGs, of which lunges were analyzed in Fig. 7F–J. Values were mostly consistent with the raw lunge results, except for *CG182723* RNAi in  $\Delta nvy$  background (K). **(L to P)** Locomotion speed during non-orienting by the male pairs analyzed in Fig. 7F–J. \*\*\*  $p < 0.0005$ , \*\*  $p < 0.01$ , \*  $p < 0.05$ , n.s.  $p \geq 0.05$  [(A) to (P), Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test with Bonferroni correction.]

### Supplementary Figure S13.



**Fig. S13. Additional expression data related to *nvy* functions in *Tdc2* neurons.**

Nvy immunoreactivities (green) in ASM and VL clusters of *Tdc2* neurons (labeled by nuclear-localizing tdTomato, magenta) from group-reared (A) and single-reared (B) male brains.

**Table S3.** Results of the *navy*-RNAi GAL4 screen.

**Supplementary Table S3. Results of the *navy*-RNAi GAL4 screen**

I.	II.	III.	IV.	V.	VI.	VII.
GAL4	25th percentile	Median	75th percentile	n	P-value vs GAL4-only control	P-value vs UAS-only control
<i>Tdc2</i>	12.00	363.0	844.0	35	<0.0001 (<0.0002)	0.0007 (0.0014)
<i>Fru</i>	10.50	48.50	667.5	42	<0.0001 (<0.0004)	<0.0001 (<0.0004)
<i>Mip</i>	4.250	42.00	311.8	24	0.9552	0.0043 (0.0344)
<i>Orc</i>	8.750	41.00	731.3	34	0.1368	0.0720
<b><i>R84H09</i></b>	<b>3.000</b>	<b>40.00</b>	<b>209.8</b>	<b>36</b>	<b>&lt;0.0001 (&lt;0.0008)</b>	<b>0.0002 (0.0016)</b>
<i>R20G01</i>	4.250	25.50	407.0	36	0.0090 (0.0720)	<0.0001 (<0.0008)
<i>Ey(4-8)</i>	3.000	25.00	319.3	36	0.0344 (0.1376)	0.1892
<b><i>NP2631</i></b>	<b>3.500</b>	<b>17.00</b>	<b>128.5</b>	<b>49</b>	<b>&lt;0.0001 (&lt;0.0002)</b>	<b>&lt;0.0001 (&lt;0.0002)</b>
<i>Capa</i>	1.250	15.50	329.8	36	0.2471	0.0326 (0.1956)
<i>GH146</i>	1.250	11.00	46.50	30	0.2280	0.0043 (0.0172)
<i>Npf</i>	1.000	10.00	314.5	46	0.8135	0.0143 (0.0572)
<i>5HTR1B(II)</i>	1.750	9.500	311.3	30	0.0581	0.0006 (0.0024)
<i>Ey(3-8)</i>	1.000	9.500	430.3	36	0.9337	0.9671
<i>Ato(10)</i>	1.000	8.500	40.00	36	0.8730	0.5607
<i>R38G08</i>	2.000	7.500	20.00	22	0.2481	0.0413 (0.2478)
<i>Trh</i>	2.000	7.000	91.75	36	0.6056	0.6070
<i>R93G12</i>	3.000	7.000	18.00	27	0.3155	0.0579
<i>Dsk</i>	1.250	6.000	41.00	24	0.8817	0.1495
<i>Akh</i>	2.000	5.000	10.25	30	0.5929	0.0026 (0.0208)
<i>Eth</i>	1.250	4.500	29.75	36	<0.0001 (<0.0006)	0.1495
<i>Dh31</i>	3.000	4.500	25.00	24	0.9304	0.1990
<i>Pdf</i>	1.250	4.000	20.25	36	0.4062	0.0228 (0.0912)
<i>R27G01</i>	1.000	4.000	6.000	30	0.1254	0.0003 (0.0018)
<i>AstA1</i>	2.250	3.500	9.000	24	0.4076	0.9058
<i>AstC</i>	1.000	3.500	9.250	36	0.7080	0.0407 (0.3256)
<i>Dh44</i>	0.250	3.500	9.750	24	0.8409	0.9712
<i>Burs</i>	1.250	3.000	24.25	24	0.0109 (0.0654)	0.8333
<i>R70B01</i>	1.000	3.000	6.000	30	0.0186 (0.1116)	0.7708
<i>R11H09</i>	1.000	3.000	14.25	30	0.9912	0.5981
<i>Poxn</i>	0.7500	2.500	5.000	30	0.5119	0.7420
<i>Crz</i>	0.7500	2.000	23.75	30	0.9026	0.4760
<i>R17C11</i>	1.000	2.000	6.000	30	0.2980	0.0012 (0.0072)
<i>R15F02</i>	0.0	2.000	4.250	30	0.1968	0.4381
<i>Ppk23</i>	0.0	2.000	8.000	24	0.3787	0.5499
<i>Ddc(II)</i>	0.0	2.000	23.00	23	0.9701	0.4673
<i>AstA2</i>	0.2500	1.500	6.000	24	0.5866	0.4224
<i>FMRFa</i>	0.0	1.000	5.500	24	0.9958	0.0513
<i>Droc</i>	0.0	1.000	6.500	30	0.6060	0.5257
<i>5HTR1B(III)</i>	0.0	1.000	4.250	30	0.9673	0.2972
<i>R16F12</i>	0.0	1.000	6.000	27	0.8409	0.0301 (0.1806)
<i>Ppk25</i>	0.0	1.000	2.750	24	0.0870	0.5398
<i>Dsx</i>	0.0	1.000	3.750	30	0.0009 (0.0036)	0.4744
<i>Amon</i>	0.0	1.000	23.00	24	0.0004 (0.0024)	0.8937
<i>Ddc(III)</i>	0.0	1.000	2.000	24	0.2483	0.4869

Notes:

Group-reared males harboring either *XX-GAL4*, *UAS-IR-navy*, or both *XX-GAL4* and *UAS-IR-navy* were paired each with same genotypes. Lunge counts and pair numbers tested for the knockdown mutants are shown in II–V. Statistical significances between the knockdown mutants and two genetic controls were analyzed by Kruskal–Wallis one-way ANOVA and post-hoc Mann–Whitney U-test. P-values between knockdown mutant pairs and GAL4-only (VI) or UAS-only (VII) pairs are shown. Adjusted p-values by Bonferroni correction within each experiment are shown in parenthesis. Data in bold indicate 4 out of 44 tested *GAL4s* where knockdown mutants showed significant increase in lunges compared to two genetic controls, with corrected p-values less than 0.05.

**Table S1. (separate file)**

Results of the primary RNAi screen (.xlsx file).

**Table S2. (separate file)**

Results of the secondary RNAi screen (.xlsx file).

**Table S4. (separate file)**

The list of the complete genotypes used in this study (.xlsx file).

**Table S5. (separate file)**

Sequences of primers used in this study (.xlsx file).

**Table S6. (separate file)**

The list of antibodies, along with concentration and incubation conditions for Western blotting experiments (.xlsx file).

**Table S7. (separate file)**

Sequences data from scRNAseq experiments (.txt file). Data were filtered and normalized as described in Materials and Methods.

**Table S8. (separate file)**

The list of software used in this study (.xlsx file).

**Movie S1.**

An example of interactions between a pair of group-reared wild-type males (a sample from the dataset used in Fig. 1D).

**Movie S2.**

An example of interactions between a pair of group-reared  $\Delta nvy$  males (a sample from the dataset used in Fig. 1D).

**Movie S3.**

An example of interactions between a group-reared wild-type male and a pre-mated female (a sample from the dataset used in Fig. 3C).

**Movie S4.**

An example of interactions between a group-reared  $\Delta nvy$  male and a pre-mated female (a sample from the dataset used in Fig. 3C).

**Movie S5.**

Three-dimensional rendering of *nvy*-positive *Tdc2* neurons (a sample from Fig. 4D).

**Movie S6.**

Three-dimensional rendering of *nvy*-negative *Tdc2* neurons (a sample from Fig. 4F).

**Movie S7.**

An example of interactions between a group-reared wild-type male and a group-reared male in which CsChrimson was expressed in *nvy*-positive *Tdc2* neurons, before and during the LED stimulation (a sample from the dataset used in Fig. 4I).

**Movie S8.**

An example of interactions between a group-reared wild-type male and a male from one of genetic controls (which lacks *LexAop2-FLP*) of the optogenetic experiment in Fig. 4I, before and during the LED stimulation.

**Movie S9.**

Three-dimensional rendering of *Tdc2* neurons in a wild-type male (a sample from Fig. 7A).

**Movie S10.**

Three-dimensional rendering of *Tdc2* neurons in a  $\Delta nvy$  male (a sample from Fig. 7A).

**Data S1. (separate file)**

The complete statistical results for all applicable figures, and coding sequences of DNA constructs created in this study.

**Data S2. (separate file)**

Custom R codes for scRNAseq data analysis.

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