Supplementary Figures



Figure S1. Unique GIST dependencies. A. Plot showing the top 18 significantly enriched gene ontology terms among genes uniquely essential to GIST. B, Relative reads for the top 8 sgRNAs targeting the indicated genes in GIST-T1 or GIST430, normalized to baseline plasmid library (n=2 per sgRNA). Data were analyzed by one-way ANOVA with Dunnett's multiple comparisons test, compared to Baseline; ***, P<0.001; **, P<0.01. C, Plot of rank in screen and β-score highlighting MOZ complex members and KAT6B as comparator; statistically significant β-scores are indicated in red, and non-significant in orange. **D**, Plot of rank in screen and βscore highlighting Menin-MLL complex members. E, Plot of rank in screen and β-score highlighting INO80 complex members. **F**, Plot of rank in screen and β-score highlighting NuA4 histone acetyltransferase complex members. G-I, Ranked Sensitivity Score from Project Drive cell lines (n=387) for select members of the INO80 and NuA4 complexes, with GIST-T1 highlighted in red. J, Plot of rank in screen and β -score highlighting FACT complex members. K-L, Ranked Sensitivity Score from Project Drive cell lines (n=387) for members of the FACT complex, with GIST-T1 highlighted in red. M. Plot of rank in screen and β-score highlighting PAF1 complex members. N-O. Ranked Sensitivity Score from Project Drive cell lines (n=387) for select members of the PAF1 complex, with GIST-T1 highlighted in red.



Figure S2. PRC2 complex dependency in GIST. A, Plot of rank in screen and β -score highlighting core PRC2 complex members. **B-C,** Ranked Sensitivity Score from Project Drive cell lines (*n*=387) for select members of the PRC2 complex, with GIST-T1 highlighted in red. **D,** Plot of rank and CERES dependency score for EZH2 complex members across DepMap cell lines (*n*=726), with the dotted line at -1 indicating significant dependency. **E,** Top 70 gene dependency correlations of EZH2 in DepMap. Co-dependent chromatin modifying enzymes and complex members are labeled.



Figure S3. Menin-MLL and MOZ complex localization in GIST. **A**, Overlap of enriched regions between Menin and BRPF1, with select GIST-associated genes indicated. **B-F**, Tracks showing regions of genomic occupancy of the TF HAND1, MOZ complex members BRPF1 and KAT6A, Menin-MLL complex members Menin and MLL1n and histone marks H3K4me3, H3K9ac and H3K27ac at the *OSR1*, *KIT*, *MEIS1*, *HAND1* and *USP1* loci.



Figure S4. KAT6A, Menin and BRPF1 inhibition. A, Growth over time assay following treatment of GIST-T1 or GIST48B with 50 nM imatinib (*n*=5 per condition). Data were analyzed by two-way ANOVA, compared to GIST48B; ***,*P*<0.0001. **B**, DMSO-normalized cell count after the first passage of slowly growing GIST cell lines GIST430 (day 6), GIST882 (day 12) and GIST48 (day 12) in comparison to GIST-T1 (day 4). Cells were treated with VTP-50469 at 0.5 μ M, WM-1119 at 1 μ M or the combination with each drug at 0.1 μ M (*n*=5-6 per condition). The first passage cell count was used to demonstrate on-target drug toxicity comparable to GIST-T1, as GIST882 and GIST48 cell lines replate poorly in this assay. Data were analyzed by one-way ANOVA with Dunnett's multiple comparisons test; compared to DMSO control ***,*P*<0.001; *,*P*<0.05. **C**, Growth over time assay treating GIST-T1 or GIST48B cells with selective BRPF1 inhibitors GSK6853 or PFI-4 at the indicated doses. No significant differences were observed by two-way ANOVA compared to DMSO control.



Figure S5. Transcriptional effects of MOZ and Menin disruption. A, Heatmap of controlnormalized expression of GIST-associated TFs (10) in response to drug or sgRNA treatment. **B**, DMSO-normalized expression of 18 GIST-associated TFs in the indicated drug treatment (*n*=4 per condition). Data in were analyzed by one-way ANOVA with Tukey's multiple comparisons test, compared to VTP-50469; ***,*P*<0.001. **C-F**, Relative mRNA level by qRT-PCR of negative regulator of KIT signaling DUSP6 and HAND1- and SE-associated gene NPR3 in the indicated GIST cell lines treated for 5 days with VTP-50469 at 0.5 µM, WM-1119 at 1 µM or the combination with each drug at 0.1 µM (*n*=3-4 per group). Data in were analyzed by one-way ANOVA with Dunnett's multiple comparisons test, compared to DMSO; ***,*P*<0.001; **,*P*<0.01; *,*P*<0.05. **G**, Heatmap of GSEA data indicating the NES of Reactome translation-associated gene sets in each drug or sgRNA treatment condition. Only gene sets with significant FDRs are displayed using the color scale, with those bearing non-significant FDRs indicated in gray. **H**, Control-normalized expression of all translation-associated genes (*n*=48) in each sgRNA and drug treatment condition. Data were analyzed by Welch's *t* test, compared to All Genes; ***,*P*<0.001; **,*P*<0.01.



Figure S6. ChIP-seg of DOT1L, H3K79me2 and MEAF6 and effects of VTP-50469 and WM-**1119. A**, Top 70 gene dependency correlations of DOT1L in DepMap, with members of Menin-MLL, MOZ and PRC2 complex indicated. **B**, Enriched genomic regions of DOT1L binding. Select GIST-associated genes are indicated. C, DMSO-normalized signal for DOT1L in regions showing enriched (n=1,343) or typical (n=45,256) signal for DOT1L. Data were compared by Welch's *t* test; ***,*P*<0.001. **D**, Western blot showing DOT1L signal following 5 days of treatment with the indicated inhibitors. E, Levels of DOT1L expression by RNA-seg following 5 days of treatment with the indicated drugs; no significant changes were observed by one-way ANOVA with Dunnett's multiple comparisons test. F, Heat maps demonstrating spike-in normalized signal of MEAF6 at MACS-defined peaks (n=22,581) in GIST-T1 cells treated with DMSO, VTP-50469 or WM-1119. Scaled read densities ± 1.25 kb from the peak center are shown in rows. G-H, Enriched genomic regions of H3K79me2 and MEAF6 binding. Select GIST-associated genes are labeled. I, Heat maps demonstrating genomic localization in GIST-T1 of DOT1L, H3K79me2 and MEAF6 by ChIP-seq. Scaled read densities ± 10 kb from the TSS, H3K27ac-defined enhancers or ATAC-defined peaks are shown in rows. J, Tracks showing regions of genomic occupancy of spike-in normalized DOT1L under the indicated treatments, H3K79me2, MEAF6 and H3K27ac at the GPR20 locus. K, Day 21 GIST-T1 cell count in a growth over time assay comparing sqRNAs targeting two DOT1L exons or Luc or RPS19 as control (*n*=5 per sqRNAs). Data were analyzed by one-way ANOVA with Dunnett's multiple comparisons test, compared to Luc control; ***,*P*<0.001; **,*P*<0.01.



Figure S7. Effects of sgRNA or drug treatment in vivo. A. GIST-T1/Cas9 cells were stably transduced with the indicated sgRNA, selected with puromycin in vitro for 14 days, and identical numbers of cells implanted and monitored for growth (*n*=10 tumors per group). Data were analyzed by two-way ANOVA, compared to sgLuc; ***, P<0.001. B, Weight of mice engrafted with the GIST-T1 cell line and treated for 28 days with WM-1119 (50 mg/kg gavage three times per day, 7d/week; n=6), VTP-50469 (0.1% in chow; n=5), combination WM-1119 and VTP-50469 (n=6) or vehicle control (n=5). C. Weight of mice engrafted with the GIST-T1 cell line and treated for 28 days with imatinib (50 mg/kg gavage 5d/week; n=5), VTP-50469 (0.1% in chow; n=4), combination imatinib and VTP-50469 (n=5) or vehicle control (n=5). **D**, Control-normalized expression of all expressed genes (n=7,434) or those whose expression is upregulated by HAND1 (*n*=438) in each treatment group. Data were analyzed by Welch's *t* test, compared All Genes; ***, P<0.001; **, P<0.01; *, P<0.05. E, Weight of mice engrafted with the PG27 PDX and treated for 18 days with imatinib (50 mg/kg gavage 5d/week; n=5), VTP-50469 (0.1% in chow; n=5), combination imatinib and VTP-50469 (n=5) or vehicle control (n=5). Data were analyzed by two-way ANOVA, compared to vehicle; *, P<0.05. F, PG27 tumors were harvested at the end of the treatment period and fixed tissues sectioned and evaluated for Ki-67 (top row) and cleaved caspase-3 (bottom row); scale bar = 25μ m.

Table S1. Genome-scale CRISPR screen. Columns indicate gene name, β -score, *P*-value and FDR.

Table S2. MEAF6 proximal proteins identified by BioID.

 Table S3. Oligonucleotide sequences.

Table S4. Tumor Measurements.