

Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our [Editorial Policies](#) and the [Editorial Policy Checklist](#).

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.

n/a Confirmed

- The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement
- A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
- The statistical test(s) used AND whether they are one- or two-sided
Only common tests should be described solely by name; describe more complex techniques in the Methods section.
- A description of all covariates tested
- A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
- A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
- For null hypothesis testing, the test statistic (e.g. F , t , r) with confidence intervals, effect sizes, degrees of freedom and P value noted
Give P values as exact values whenever suitable.
- For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
- For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
- Estimates of effect sizes (e.g. Cohen's d , Pearson's r), indicating how they were calculated

Our web collection on [statistics for biologists](#) contains articles on many of the points above.

Software and code

Policy information about [availability of computer code](#)

Data collection

Data analysis

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio [guidelines for submitting code & software](#) for further information.

Data

Policy information about [availability of data](#)

All manuscripts must include a [data availability statement](#). This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our [policy](#)

Field-specific reporting

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Life sciences Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see [nature.com/documents/nr-reporting-summary-flat.pdf](https://www.nature.com/documents/nr-reporting-summary-flat.pdf)

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	For mouse infection model assays, the mice (n=17, per group) were intraperitoneally infected with <i>E. coli</i> W3110 (pUC19-mcr-1) to cause a systemic infection. The mouse (n=16, per group) intraperitoneal infection model <i>A. baumannii</i> ATCC19606 was also used to confirm the synergistic effect of colistin and artemether injection. For <i>S. typhimurium</i> infection analysis, the mice (n=15, per group) were housed for 5 days with water containing streptomycin (5 mg/mL) before infection and were orally gavaged with <i>S. typhimurium</i> HYM2. For bacterial loading analysis, weight analysis and morbidity assays, the mice (n=10, per group) were intraperitoneally infected with <i>E. coli</i> W3110 (pUC19-mcr-1).
Data exclusions	No data were excluded from the analyses.
Replication	The experiments were repeated at least three times independently, and representative data are displayed.
Randomization	All samples were randomly assigned to different groups. Such as the experimental mice were first mixed together then randomly grabbed and divided into different groups before treatment.
Blinding	The investigators were blinded to group allocation during data collection and analysis.

Reporting for specific materials, systems and methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> Antibodies
<input type="checkbox"/>	<input checked="" type="checkbox"/> Eukaryotic cell lines
<input checked="" type="checkbox"/>	<input type="checkbox"/> Palaeontology and archaeology
<input type="checkbox"/>	<input checked="" type="checkbox"/> Animals and other organisms
<input checked="" type="checkbox"/>	<input type="checkbox"/> Human research participants
<input checked="" type="checkbox"/>	<input type="checkbox"/> Clinical data
<input checked="" type="checkbox"/>	<input type="checkbox"/> Dual use research of concern

Methods

n/a	Involved in the study
<input checked="" type="checkbox"/>	<input type="checkbox"/> ChIP-seq
<input checked="" type="checkbox"/>	<input type="checkbox"/> Flow cytometry
<input checked="" type="checkbox"/>	<input type="checkbox"/> MRI-based neuroimaging

Eukaryotic cell lines

Policy information about [cell lines](#)

Cell line source(s)	J774 cells, Vero cells and HeLa cells in current study were obtained from the American Type Culture Collection (ATCC).
Authentication	Our laboratory uses the gold standard STR method for cell identification.
Mycoplasma contamination	Confirmed free from mycoplasma.
Commonly misidentified lines (See ICLAC register)	HeLa cells, for the invasion ability of <i>S. typhimurium</i> .

Animals and other organisms

Policy information about [studies involving animals](#); [ARRIVE guidelines](#) recommended for reporting animal research

Laboratory animals	6-8-week old female BALB/c mice (20 ± 2 g) were obtained from the Liaoning Changsheng Biotechnology Co., Ltd.
Wild animals	Our study did not involve wild animals.

Field-collected samples

Our study did not involve samples collected from Field.

Ethics oversight

Animal experiments were approved by the Animal Care and Use Committee of Jilin University (No. ALKT202102003) and were operated following the guidelines of this committee.

Note that full information on the approval of the study protocol must also be provided in the manuscript.