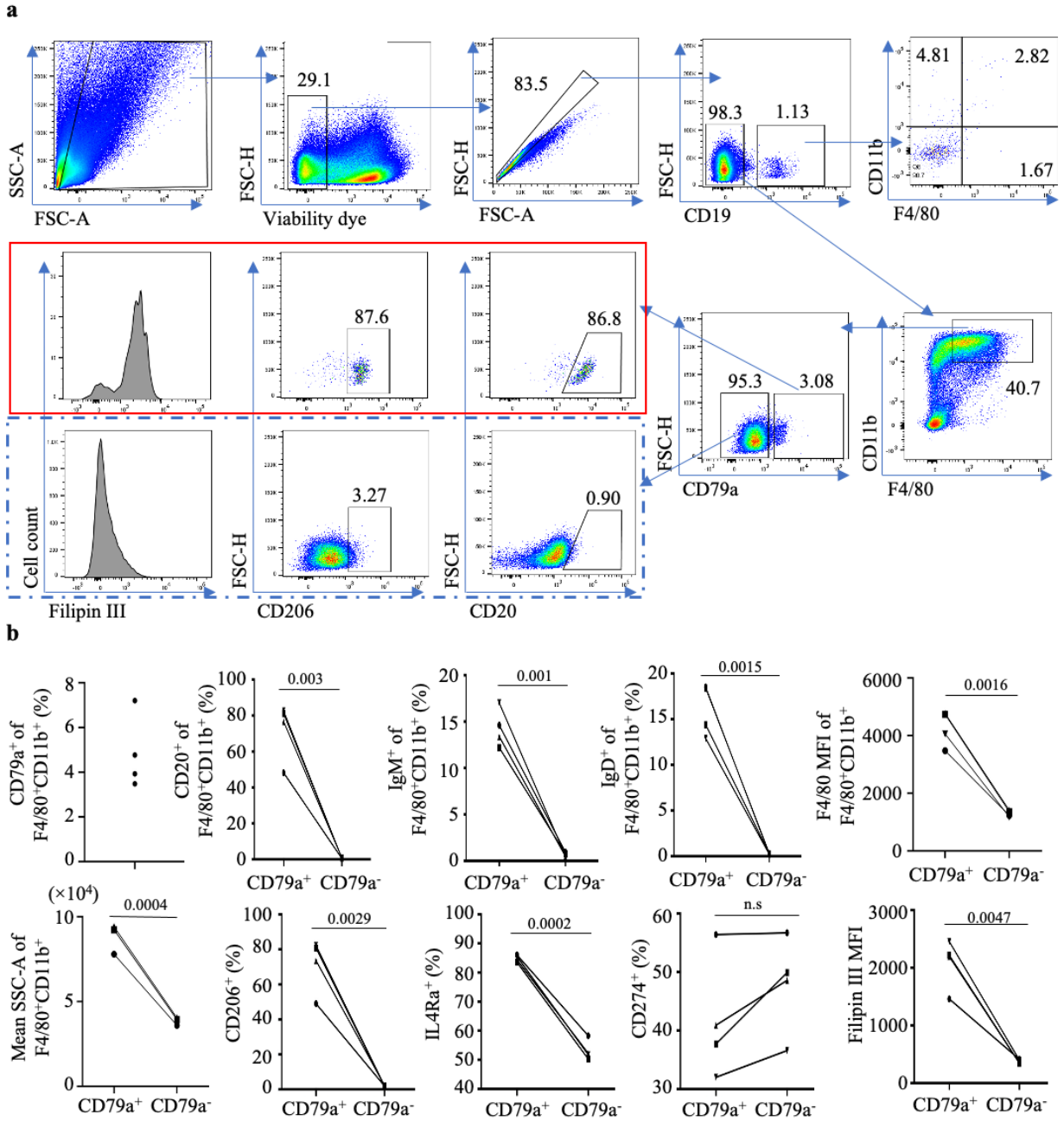
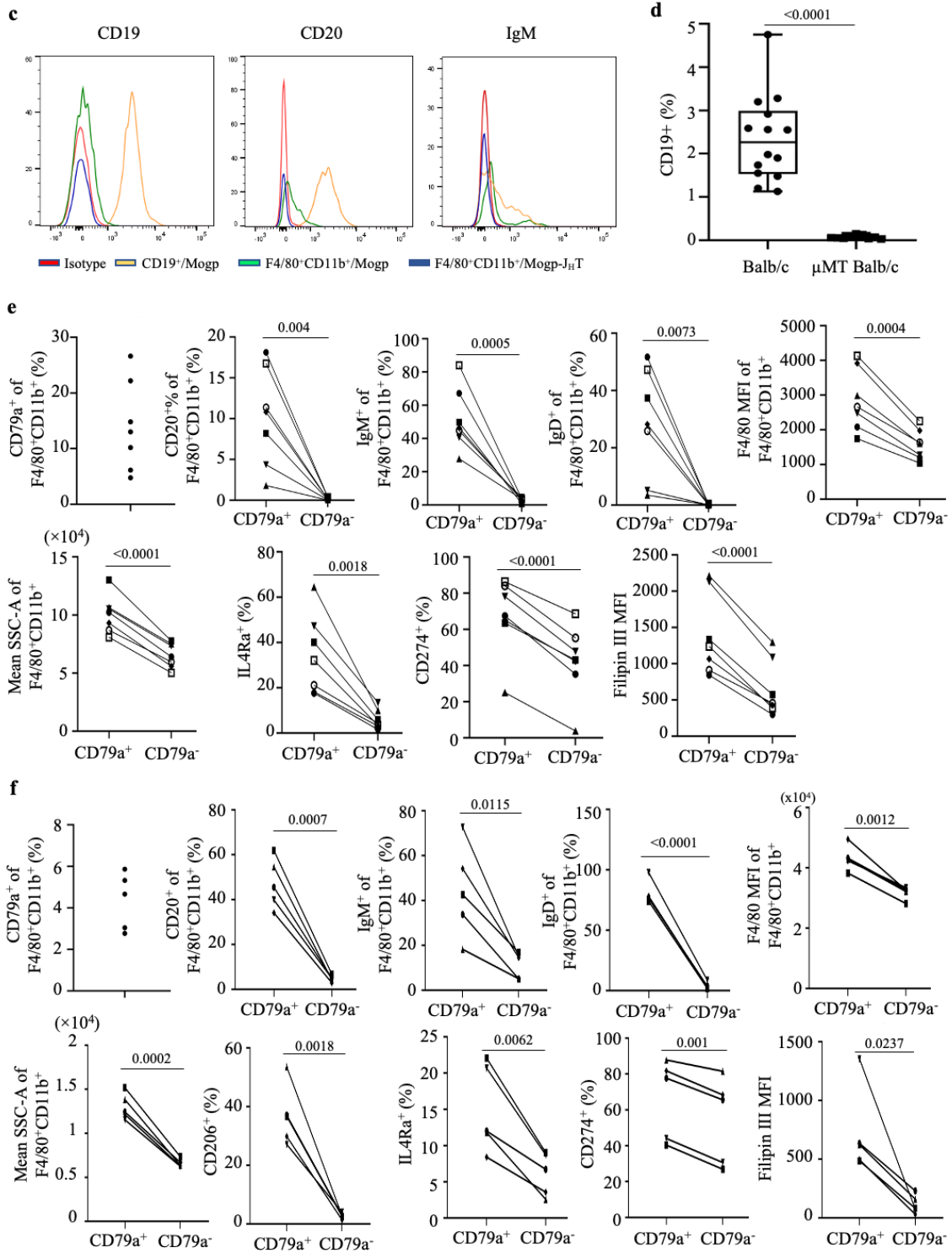


SUPPLEMENTARY FIGURES

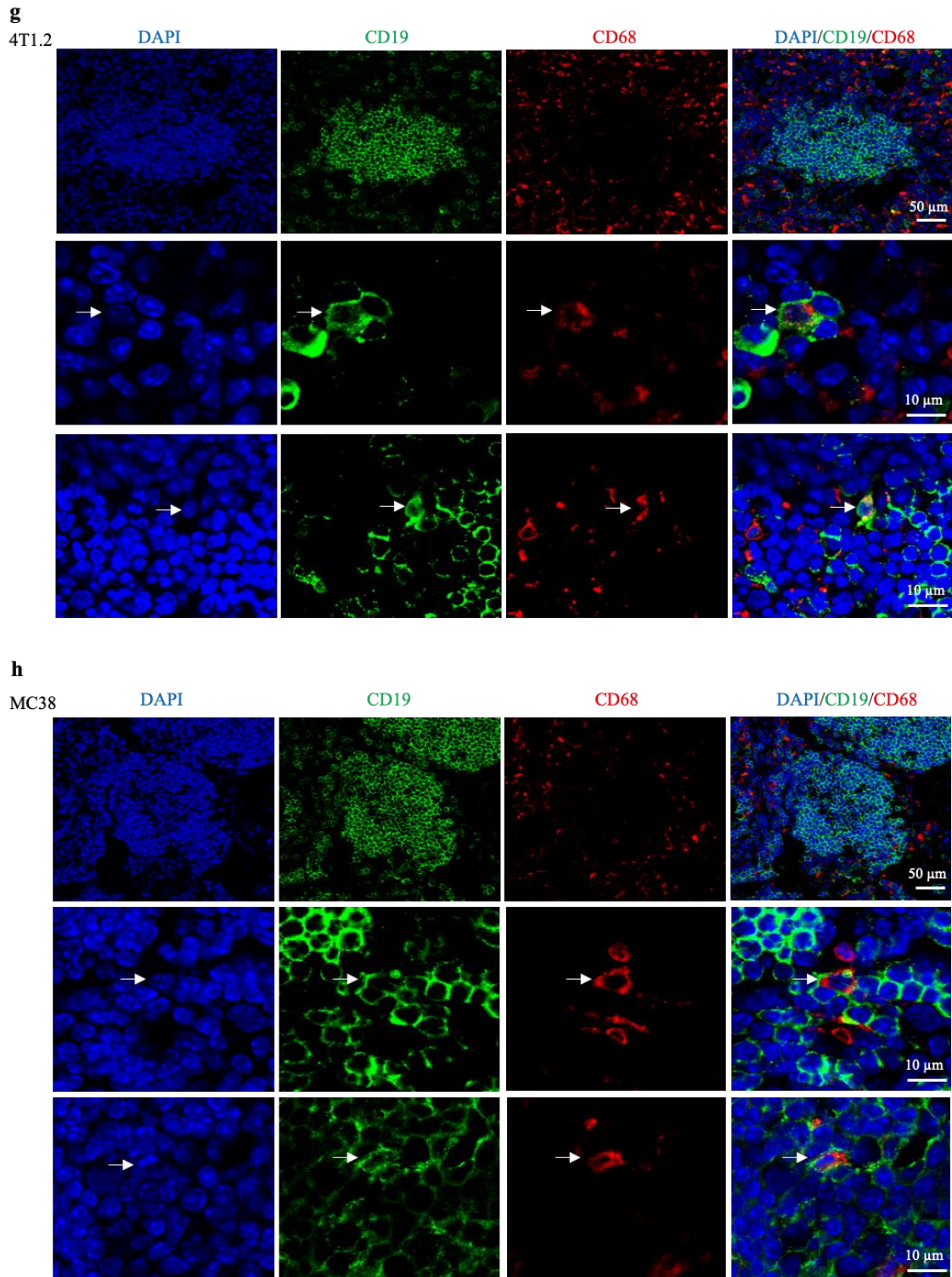
Chen C, et al. “Cancer coopts differentiation of B-cell precursors into macrophage-like cells”



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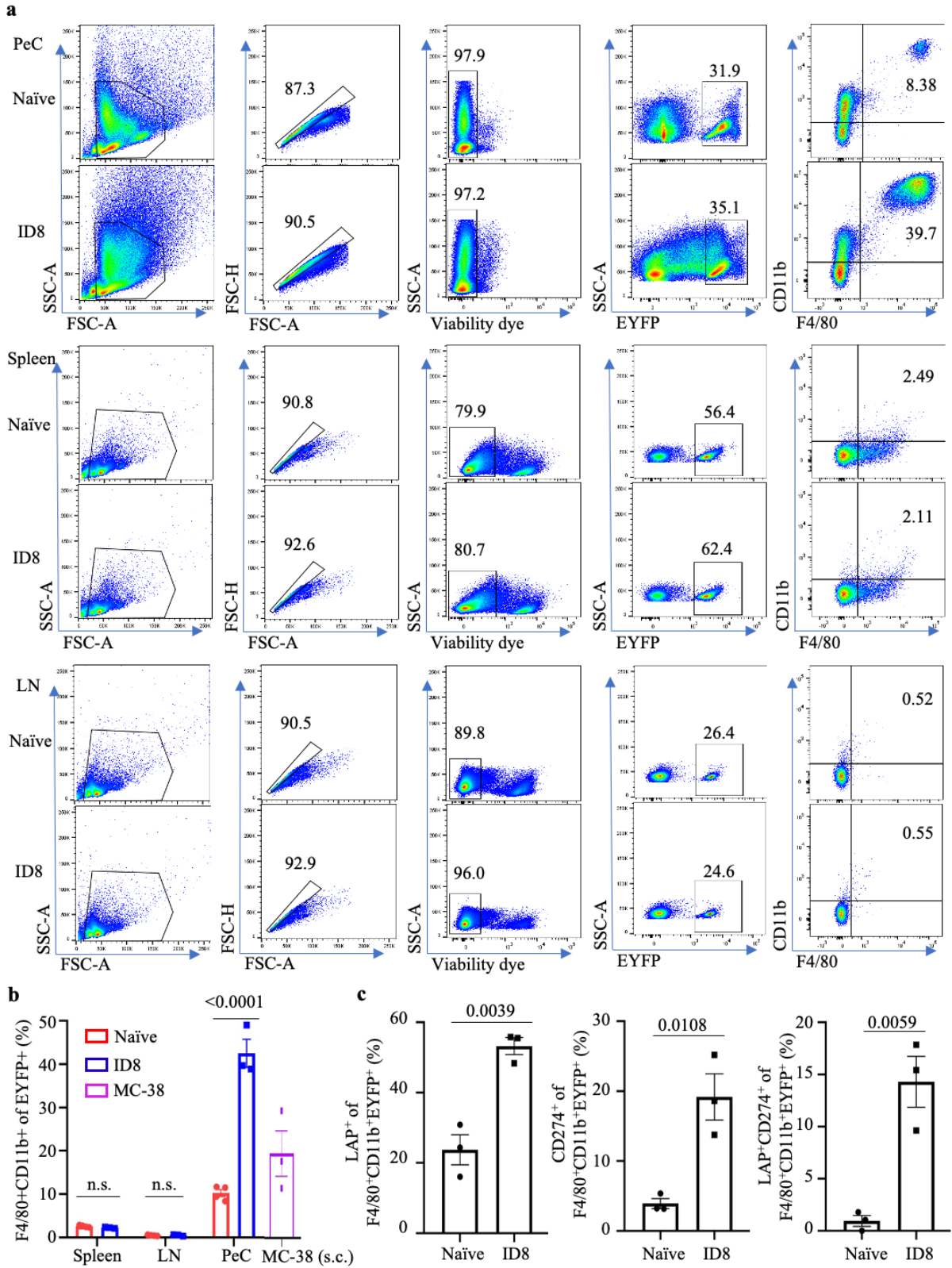


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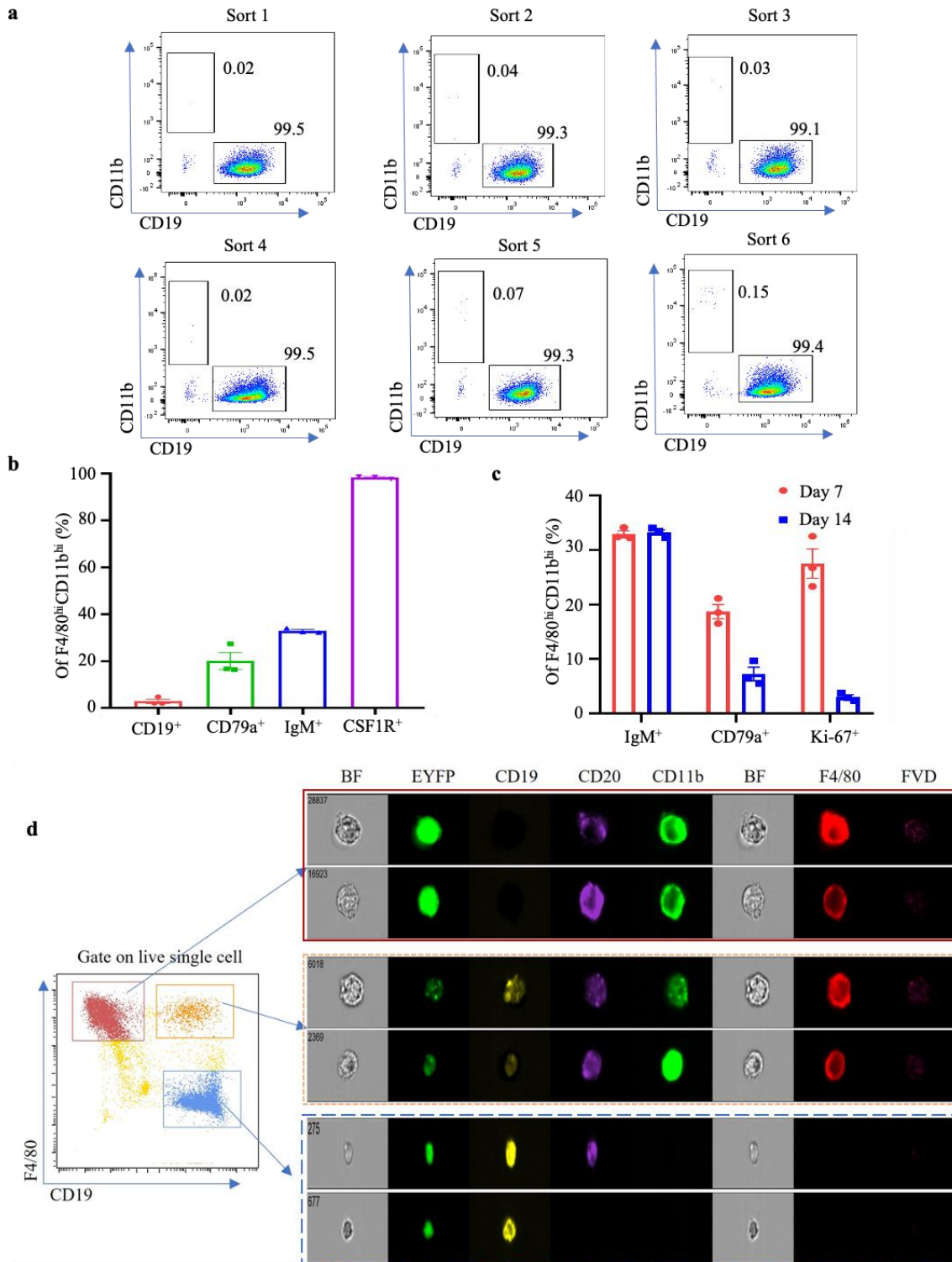
Supplementary Figure 1. Co-expression of B-cell and macrophage specific markers. a, gating strategy showing co-expression of B-cell specific markers (CD19, CD79a, CD20) and macrophage markers (F4/80 and CD11b) in TIB and TAM in the tumor from BALB/c mice with s.c. 4T1.2 cancer. **b,** Representative FACS profiling data of surface markers in CD79a⁺

TAM (presumably B-MF) as compared with CD79a⁻ TAM (“classic” TAM) in in the tumor of mice with 4T1.2 breast cancer (n=4). (CD20⁺ $P=0.0003$, IgM⁺ $P=0.0001$, IgD⁺ $P=0.0015$, F4/80 MFI $P=0.0016$, Mean SSC-A $P=0.0004$, CD206⁺ $P=0.0029$, IL4Ra⁺ $P=0.0002$, Filipin III MFI $P=0.0047$ in CD79a⁺ vs CD79a⁻). P values were calculated using two-tailed paired t test. **c**, Representative histogram showing B-cell specific makers (CD19, CD20 and IgM) within TAM (F4/80⁺CD11b⁺) in the peritoneum of B-cell sufficient (Mogp) and B-cell deficient (Mogp-J_HT) C57BL/6 mice with peritoneal Mogp ovarian cancer. **d**, Box-Whiskers plot quantification of tumor infiltrated CD19⁺ TIB frequency from B-cell sufficient (BALB/c) vs B-cell deficient (μ MT BALB/c) mice with 4T1.2 cancer (n=14 for BALB/c group, n=9 for μ MT BALB/c group, $P<0.0001$). P values were calculated using two-tailed unpaired t test. Representative FACS profiling data of surface markers in CD79a⁺ TAM as compared with CD79a⁻ TAM in Mogp mice (n=7, CD20⁺ $P=0.0004$, IgM⁺ $P=0.0005$, IgD⁺ $P=0.0073$, F4/80 MFI $P=0.0004$, Mean SSC-A $P<0.0001$, IL4Ra⁺ $P=0.0018$, CD274⁺ $P<0.0001$, Filipin III MFI $P<0.0001$ in CD79a⁺ vs CD79a⁻, **e**) and C57BL/6 mice with MC-38 cancers (n=5, CD20⁺ $P=0.0007$, IgM⁺ $P=0.0115$, IgD⁺ $P<0.0001$, F4/80 MFI $P=0.0012$, Mean SSC-A $P=0.0002$, CD206⁺ $P=0.0018$, IL4Ra⁺ $P=0.0062$, CD274⁺ $P=0.0001$, Filipin III MFI $P=0.0237$ in CD79a⁺ vs CD79a⁻, **f**). P values were calculated using two-tailed paired t test. Representative immunohistochemistry staining for B cells (CD19) and macrophages (CD68) in the tumor of BALB/c and C57BL/6 mice with s.c. 4T1.2 cancer (**g**) and MC38 colon cancer (**h**), respectively. White arrow shows CD19⁺CD68⁺ cells, and scale bars represent 50 μ m (upper panel) and 10 μ m (lower 2 panels). G and H, n=3 mice per group. B, E and F, each dot is an independent mouse. Results shown here were reproduced at least twice. From here on, Error bars are for SEM.



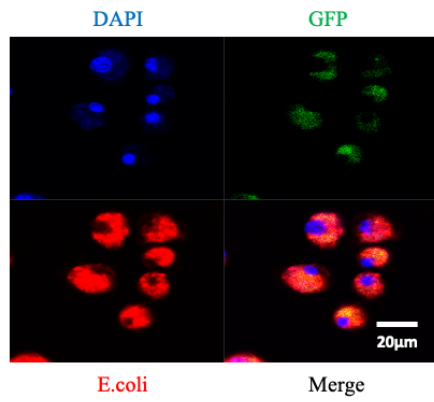
Supplementary Figure 2. EYFP⁺F4/80⁺CD11b⁺ cells in the tumor of naïve and tumor bearing Mb1-EYFP mice. Representative FACS plots (a) and quantification (b) showing

Mean \pm SEM of F4/80⁺CD11b⁺ of EYFP⁺ cells in spleen, LN and PeC ($P < 0.0001$) of Mb1-EYFP mice vs without (naïve) peritoneal ID8 cancer. “MC-38 (s.c.)” is for F4/80⁺CD11b⁺ EYFP⁺ cells (Mean \pm SEM) in the tumor of Mb1-EYFP mice with MC38 tumor. (n=4 for naïve group, n=3 for ID8 group and n=3 for MC-38 group). P values were calculated using two-tailed unpaired t test. **c**, Quantification of LAP/TGF β and PD-L1 expression in F4/80⁺CD11b⁺EYFP⁺ cells (Mean \pm SEM) in the peritoneum of naïve vs ID8 cancer-bearing mice (LAP⁺ $P=0.0039$, CD274⁺ $P=0.0108$, and LAP⁺CD274⁺ $P=0.0059$, n=3/group). P values were calculated using two-tailed unpaired t test. Each symbol is an independent mouse.

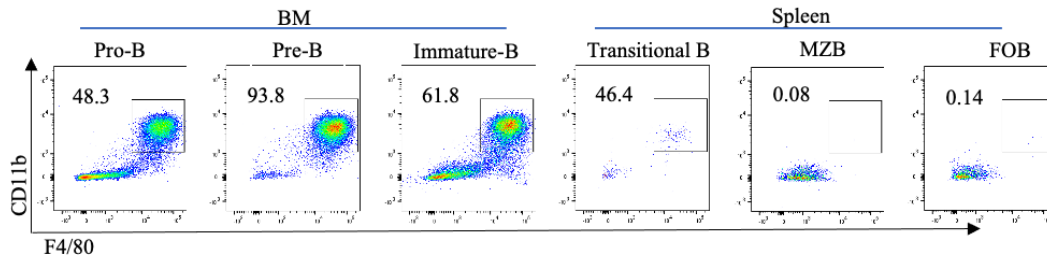


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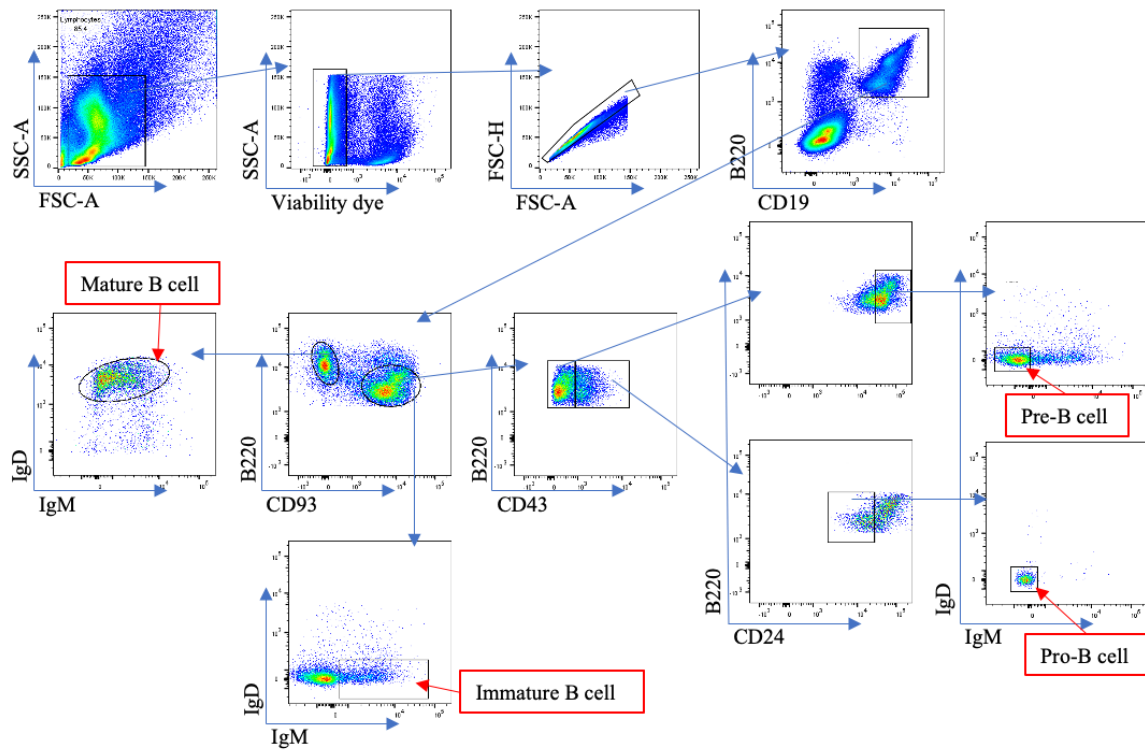
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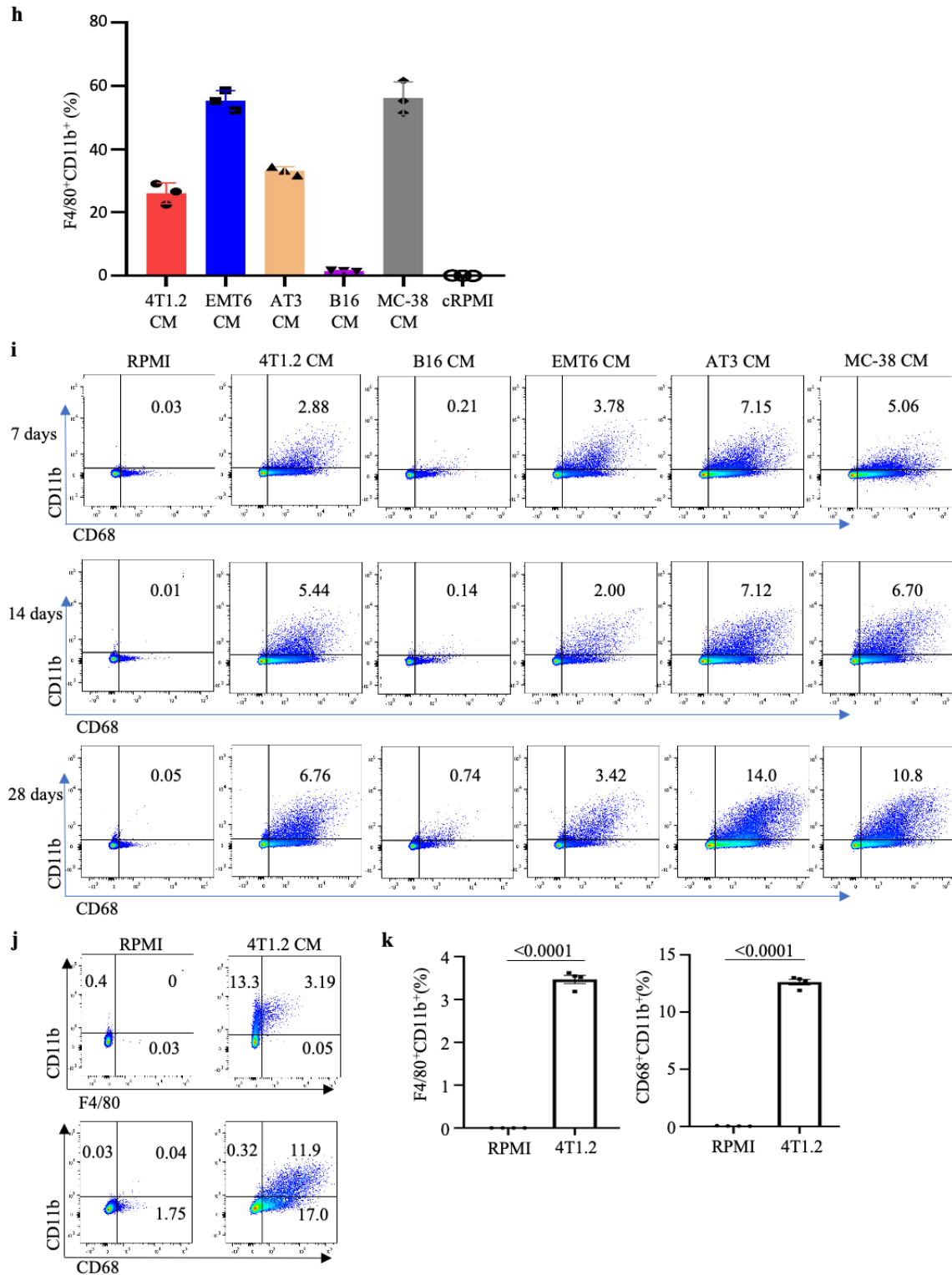
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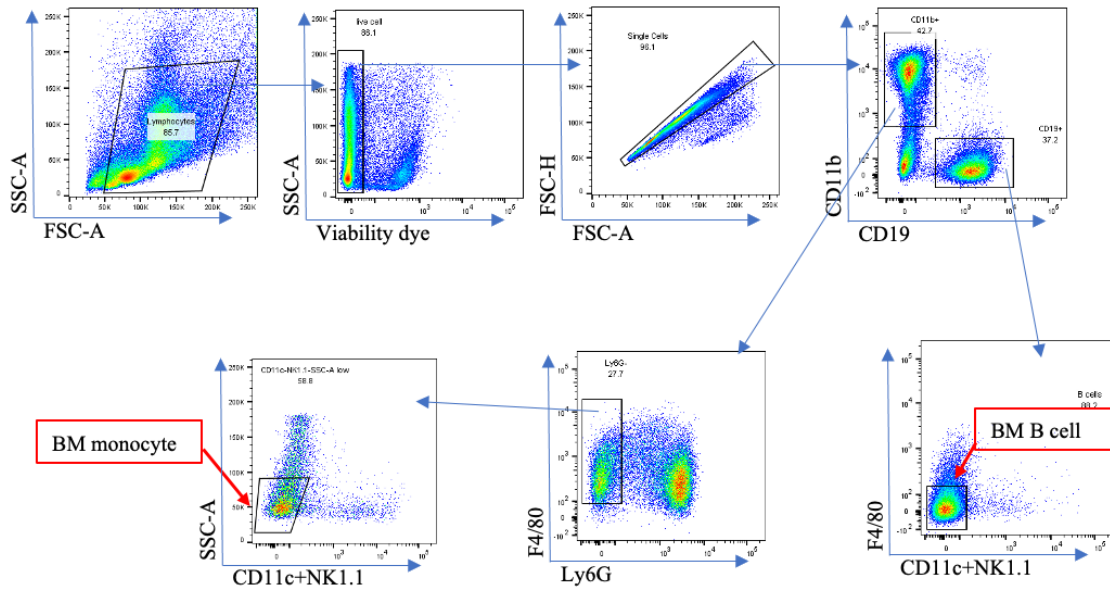


Supplementary Figure 3. Cancer CM causes differentiation of BM B cells into macrophages. **a**, representative FACS plots showing the purity (> 99%) of CD19⁺ B cells independently FACS-purified from 6 mice (sort 1-6), which we used in the *in vitro* B-MF

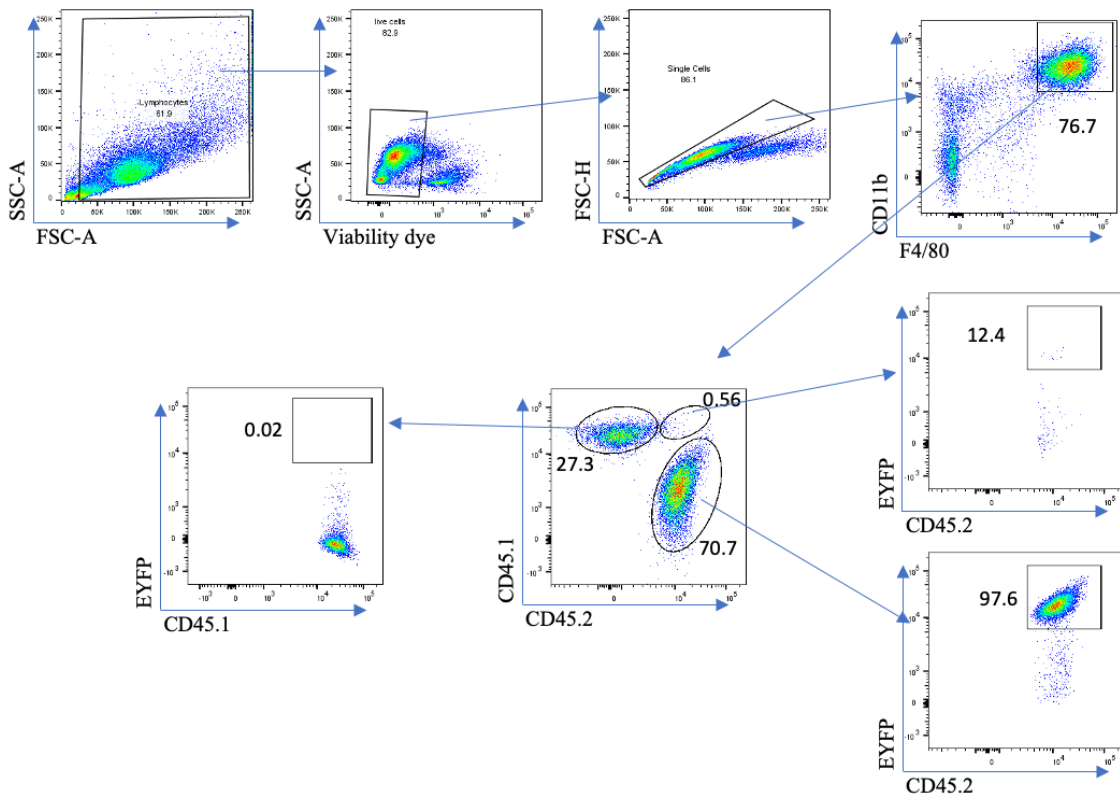
conversion assays. **b** and **c**, Quantification of FACS results (Mean \pm SEM) for indicated expression of markers (X-axis) in F4/80⁺CD11b⁺ cells generated from BM B cells cultured in 4T1.2 CM for 7 (n=3, **b**) and 14 days (red, 7 days, and blue, 14 days, n=3/group, **c**). **d**, Representative Imagestream images for expression of EYFP, F4/80, CD11b, CD19, and CD20 in single CD19⁺F4/80⁻, CD19⁺F4/80⁺ and CD19⁻F4/80⁺ cells (gated and arrows) after incubation of BM B cells of EYFP⁺ mice in 4T1.2-CM for 7 days. See that pure B cells (F4/80⁻CD19⁺ cells) are smaller in size than F4/80⁺ cells. **e**, Immunofluorescent microscopy image showing B-MF generated from RAG2-GFP⁺ BM B-cells can phagocytize pHrodo™ red-labeled *E. coli*. Reproduced twice. Scale bar represents 20 μ m. **f**, Representative FACS plots showing FACS-purified BM B-cell precursors (pro-B and pre-B cells) and immature B cells, but not splenic transitional, marginal zone and follicular B cells, can generate F4/80⁺CD11b⁺ cells upon culture with 4T1.2-CM for 7 days. **g**, Gating strategy of FACS-purified BM B-cell subsets used. **h**, Quantification of results shown in Fig2G, showing Mean \pm SEM of F4/80⁺CD11b⁺ cells after culture of BM B cells in cRPMI or CM from 4T1.2, EMT6, AT3, B16-F10, and MC38 cancer cells (n=3 per group). **i**, Representative FACS plots of pre-B cell line 70z/3 cultured for 7, 14, or 28 days with CM from indicated cancer cells. Shows surface upregulation of CD11b and CD68. **j**, Representative FACS plots and **k**, quantification (Mean \pm SEM, n=4 per group) for surface expression of macrophage markers in 70z/3 cells after 30-day culture in cRPMI vs 4T1.2-CM. *P* values were calculated using two-tailed unpaired *t* test. The results shown here were independently reproduced at least three times.

a

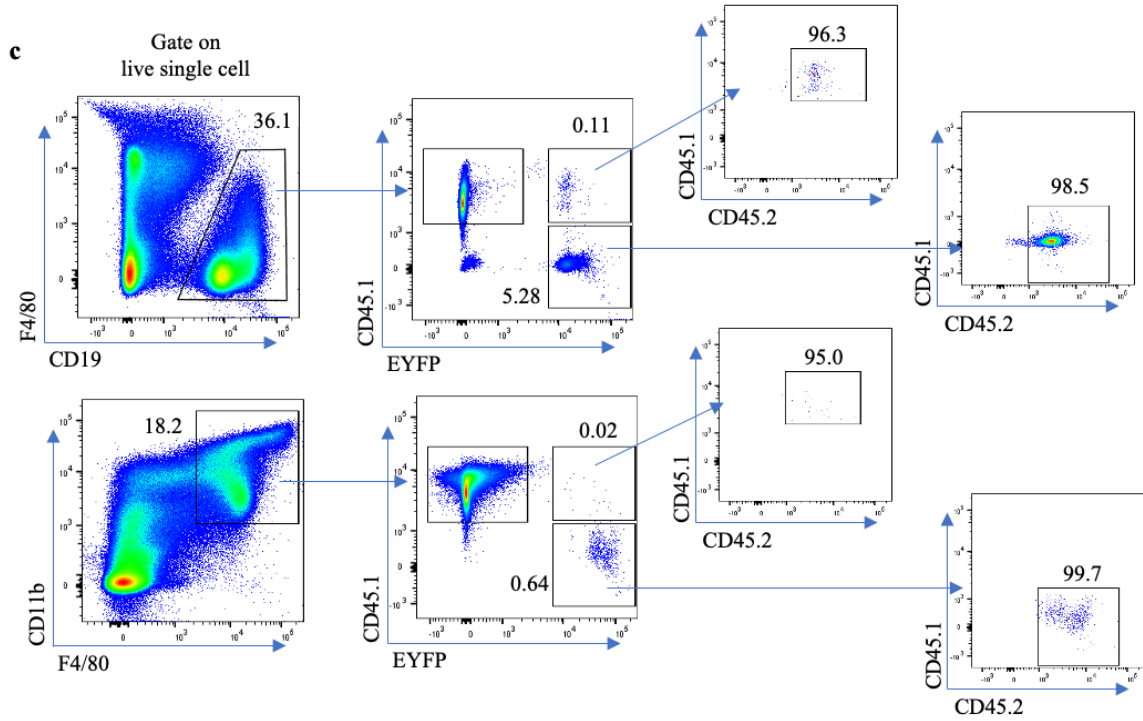
BM B cell and monocyte sort gating strategy



b

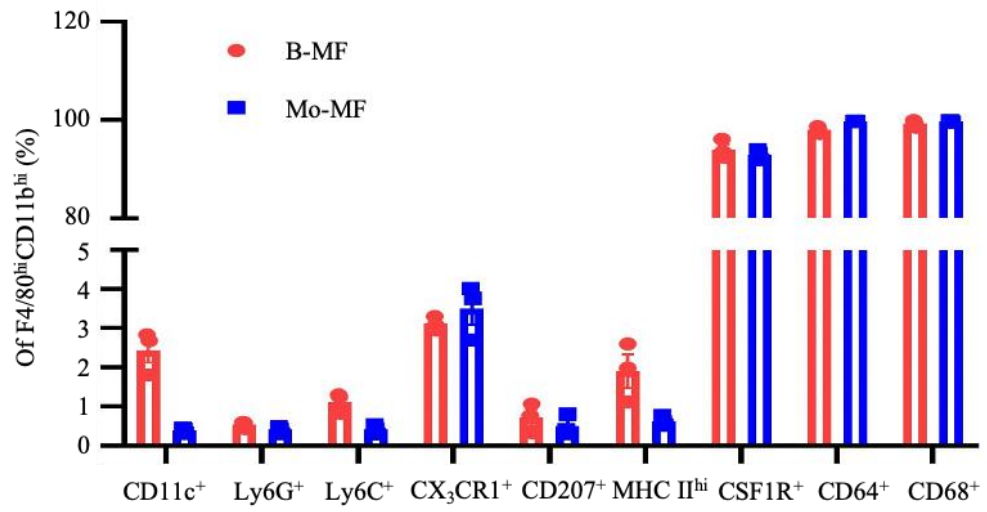


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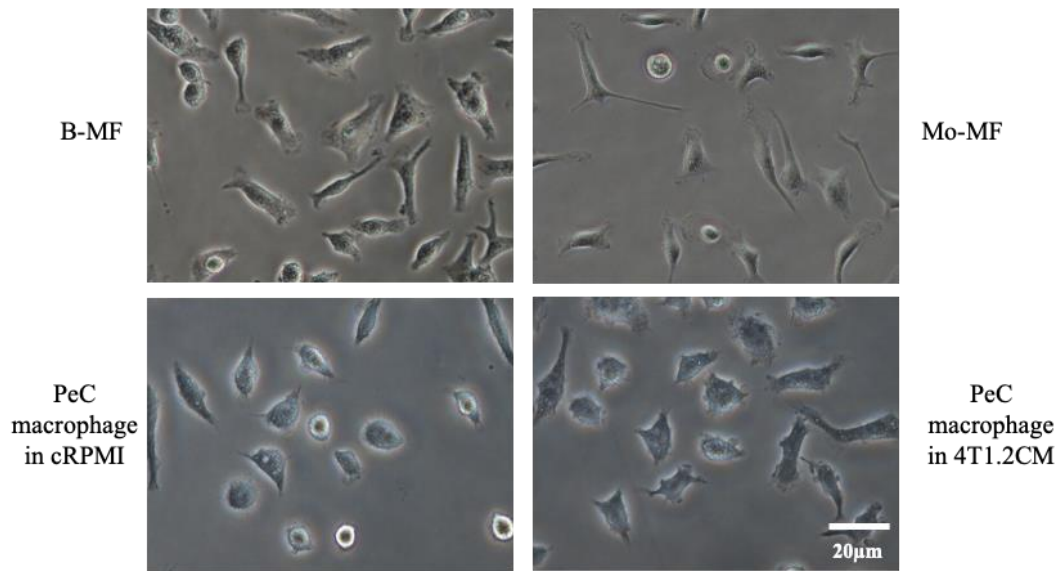


Supplementary Figure 4. Gating strategy of FACS-sorted cells and cells used in vivo in Fig.2h-j. **a**, Gating strategy of FACS-sorted BM B cells and monocytes. **b**, Gating strategy of the experiment depicted in Fig 2H. It shows CD45.1⁺ and CD45.2⁺ F4/80⁺CD11b⁺ macrophages after a mixed culture of CD45.1⁺ monocytes and EYFP⁺CD45.2⁺ BM B cells in 4T1.2-CM. **c**, Gating strategy of the experiment depicted in Fig 2I. It shows EYFP, surface CD45.1 and CD45.2 expression in peritoneal F4/80⁺CD11b⁺ (macrophages) and CD19⁺ B cells in CD45.1 mice with ID8 cancer. EYFP⁺CD45.2⁺ BM B cells were i.p. transferred into CD45.1 mice with ID8 cancer in the peritoneum.

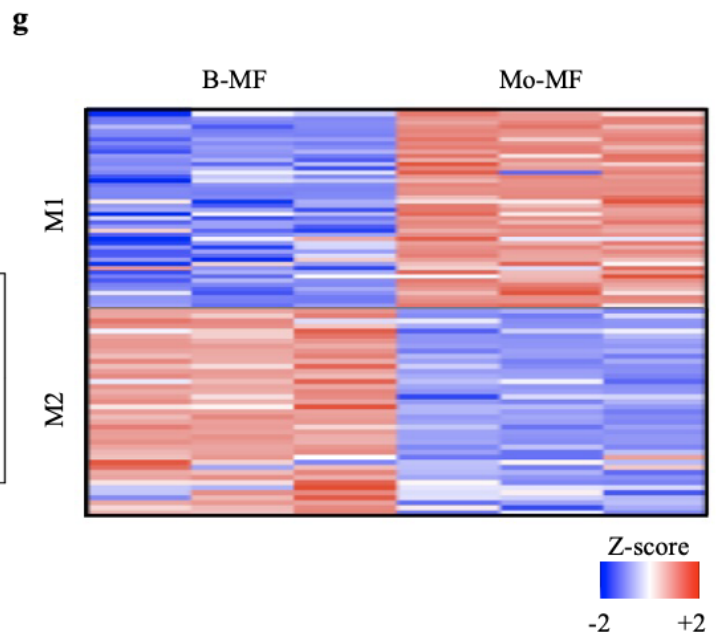
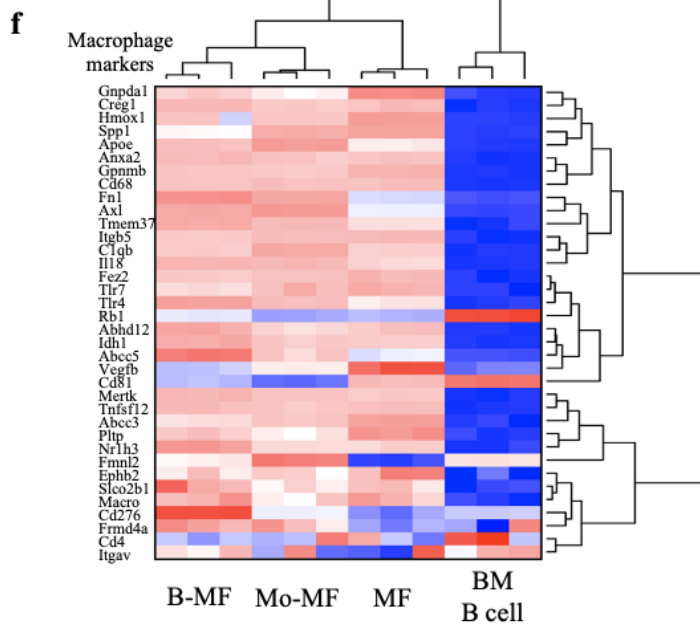
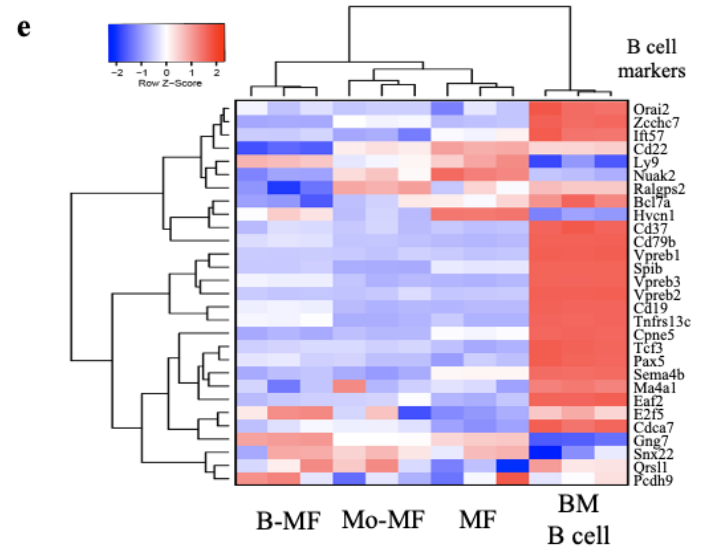
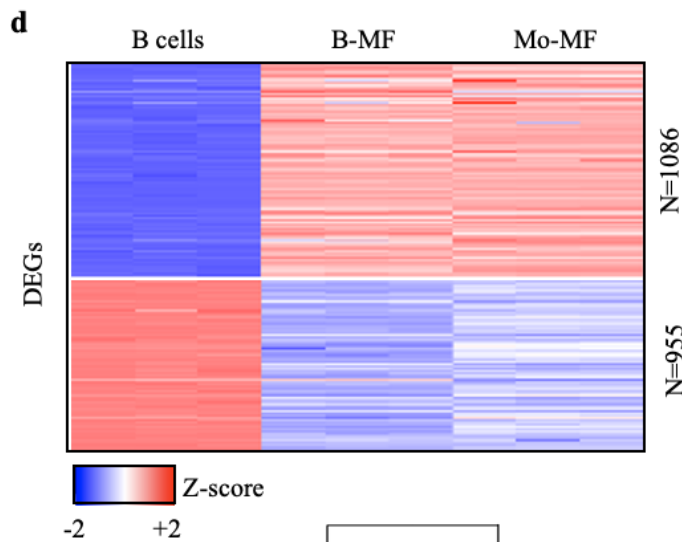
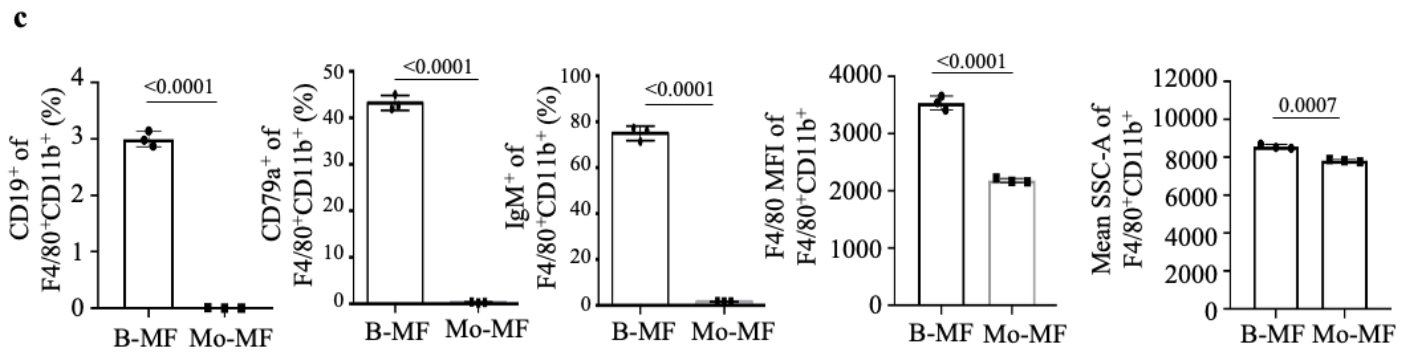
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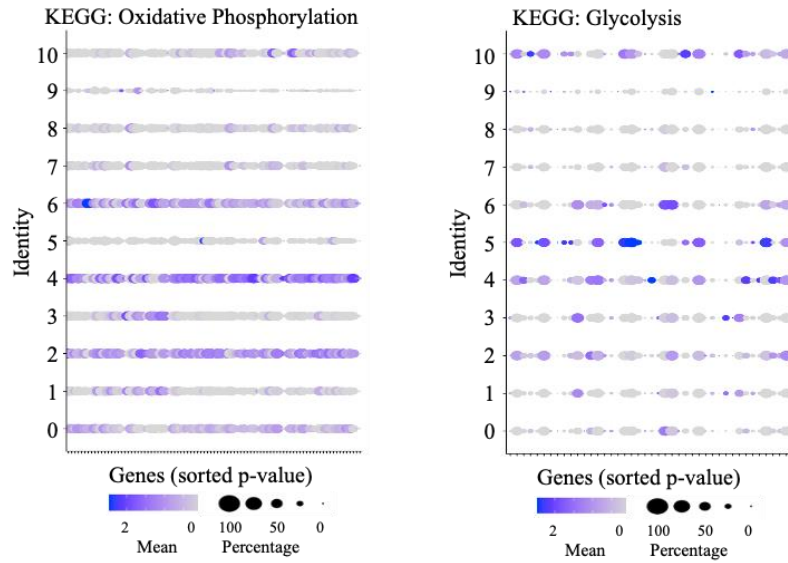
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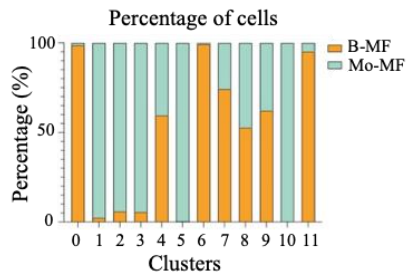
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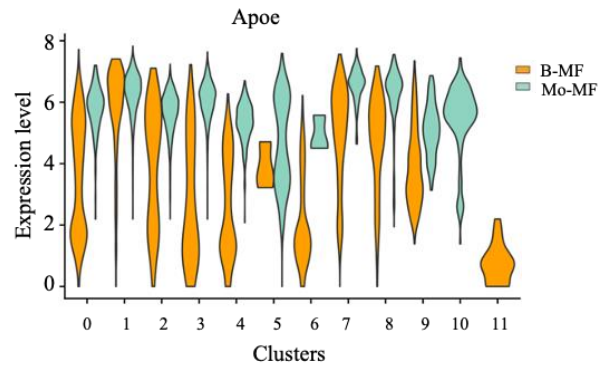
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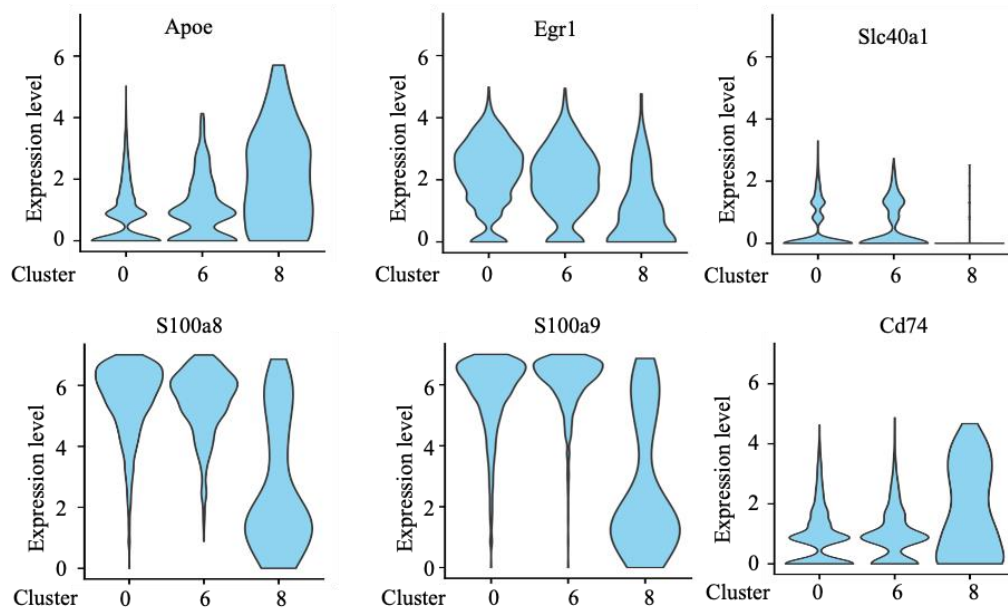
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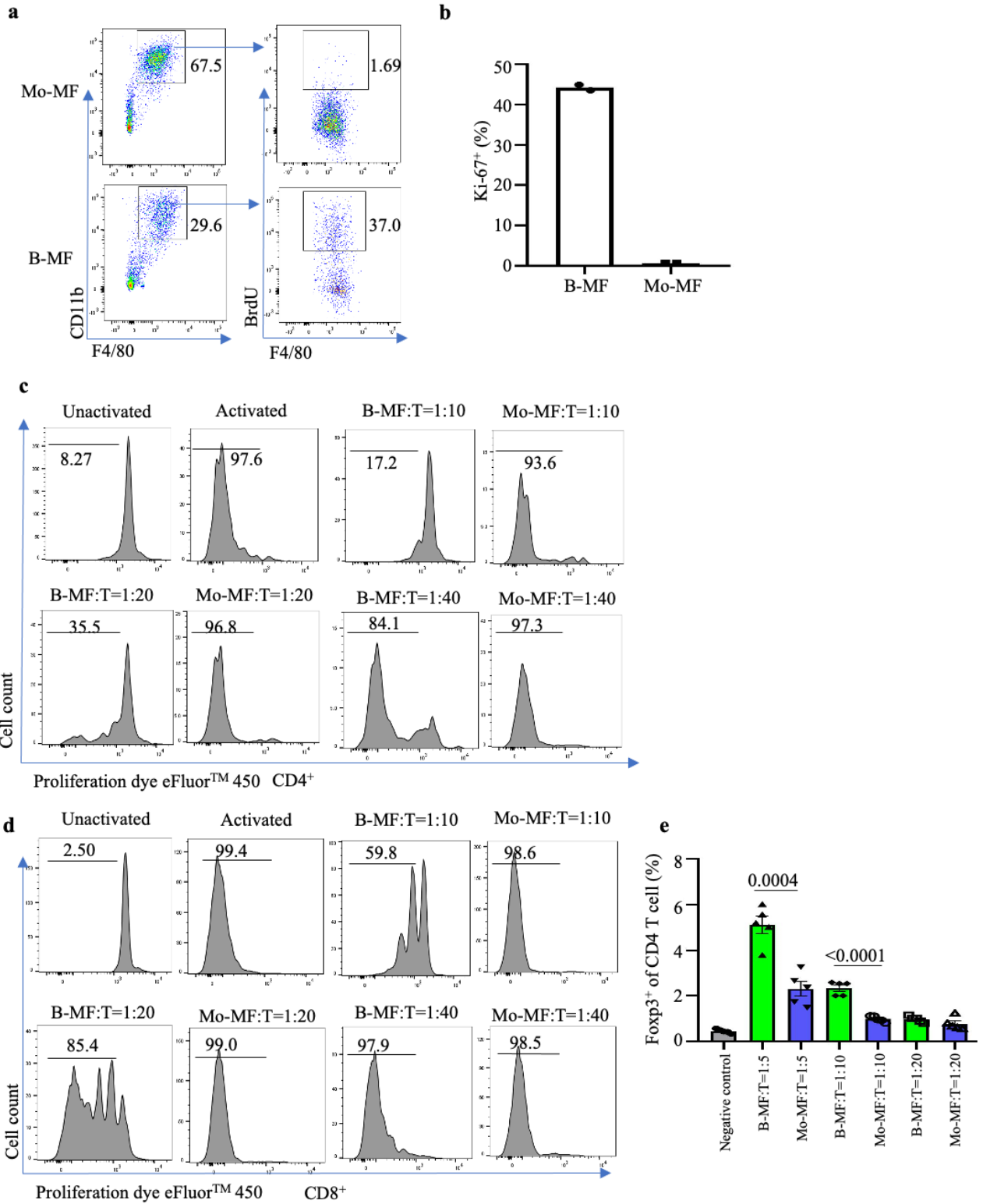


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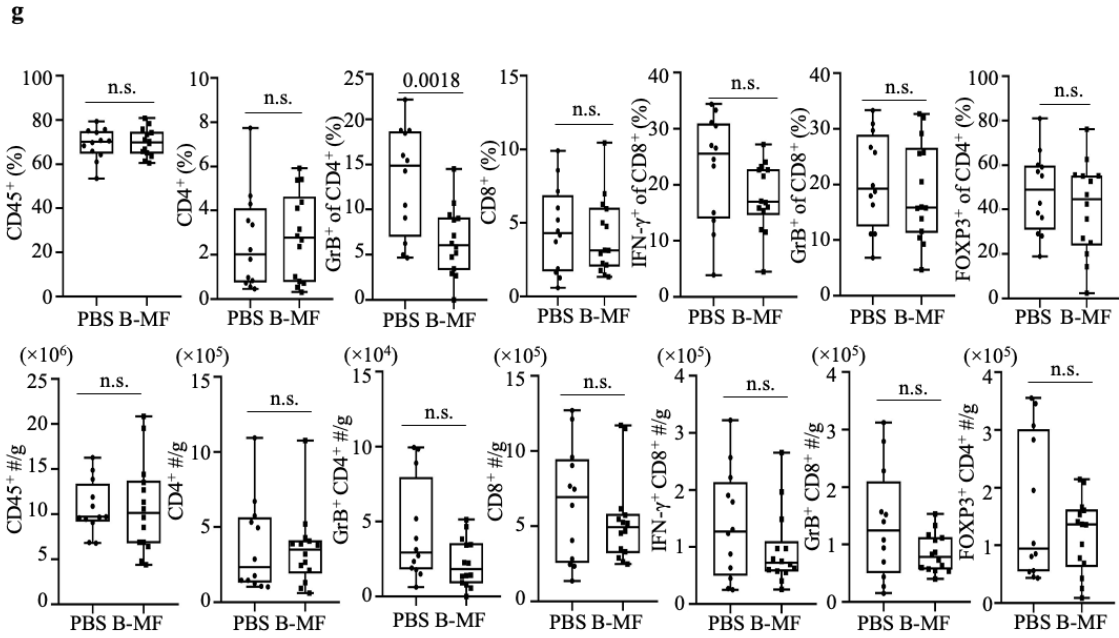
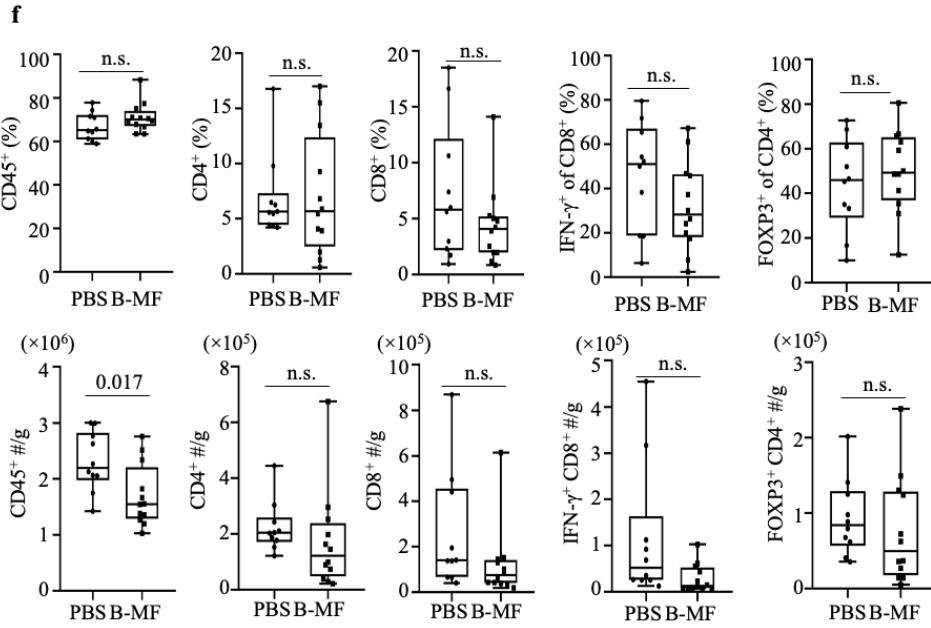


Supplementary Figure 5. Distinct gene expression profiles of B-MF and Mo-MF. a,

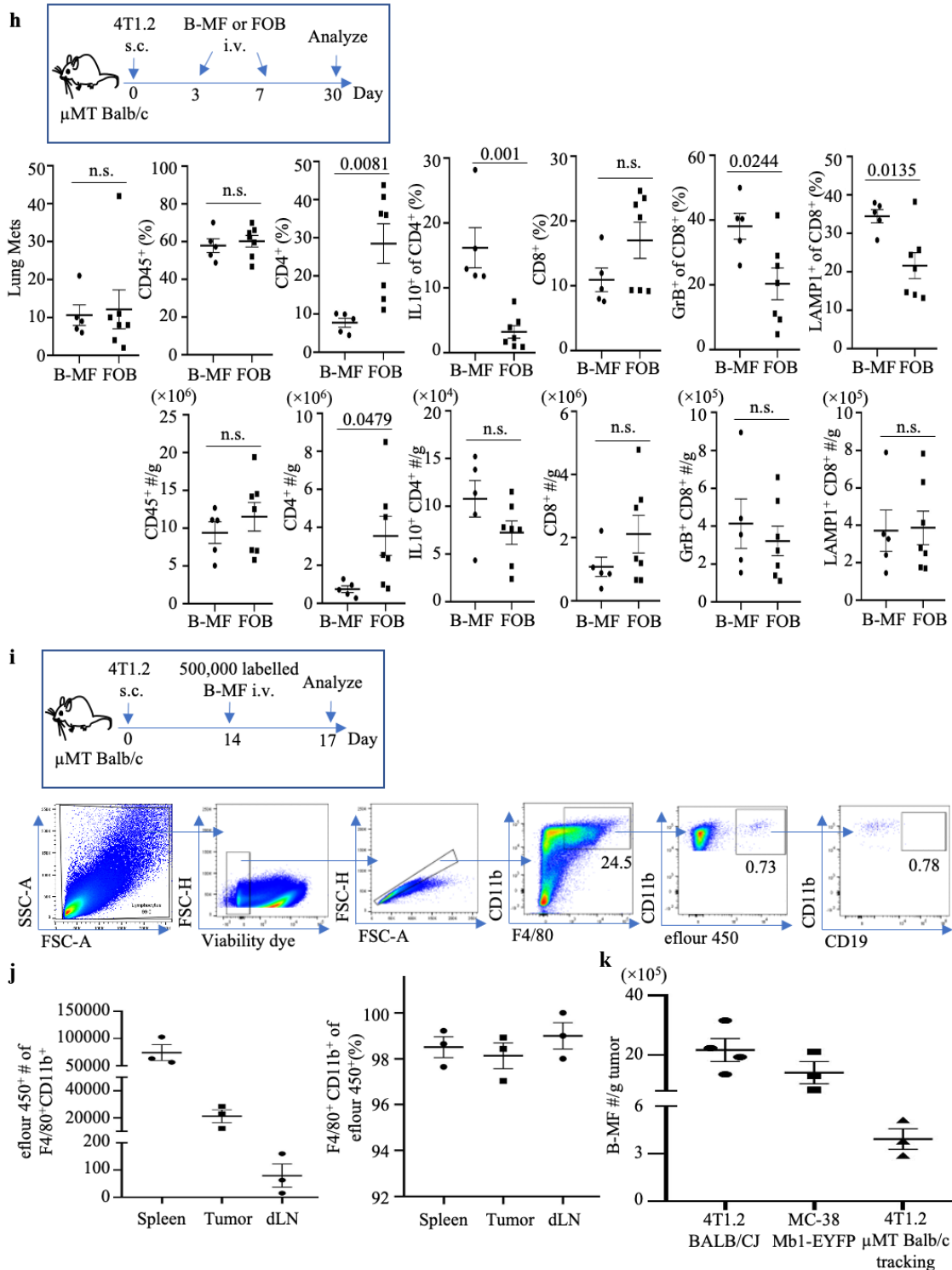
Quantification of FACS staining shows Mean \pm SEM expression of indicated markers (X-axis) on the surface of *in vitro* generated F4/80⁺CD11b⁺ B-MF vs Mo-MF (CD19⁺ P <0.0001, CD79a⁺ P <0.0001, IgM⁺ P <0.0001, F4/80 MFI P <0.0001, and Mean SSC-A P =0.0007, n=3 per group). **b**, Representative light microscopy images showing morphology of *in vitro* generated B-MF and Mo-MF as compared to macrophages from peritoneum (PeC) of naïve mice, which were cultured in cRPMI or 4T1.2-CM. Independently reproduced twice. Scale bar represented 20 μ m. **c**, *in vitro* generated B-MF differ from Mo-MF. Shown are frequency Mean \pm SEM of CD19, CD79a and IgM expression and MFI Mean \pm SEM of F4/80 within F4/80⁺ CD11b⁺ cells and Mean \pm SEM of cell sizes (n=3 per group). P values were calculated using two-tailed unpaired t test between B-MF vs Mo-MF. **d**, Heatmap showing 2041 differential expressed genes between B cells and macrophages (B-MF and Mo-MF) identified from mRNA expression profiling (microarray). Scale bar is for expression z-score. Heatmap of expression of canonical B-cell (**e**) and macrophage (**f**) genes in B-MF, Mo-MF, peritoneal macrophages (MF) and BM B cells. **g**, Heatmap showing expression of M1 and M2 signature genes (M1/M2 signature genes were from GSE5099) in B-MF and Mo-MF. **h**, KEGG oxidative phosphorylation and glycolysis pathway gene signature in *in vitro* generated B-MF and Mo-MF in scRNA-seq data. **i**, Frequency of B-MF or Mo-MF in the individual cell clusters **j**, Violin plot of *Apoe* expression in the B-MF and Mo-MF among 12 clusters. **k**, Violin plot of expression of *Apoe*, *Egr1*, *Slc40a1*, *S100a8*, *S100a9* and *CD74* in clusters 0, 6, 8 identified from scRNA-seq results of TAM from mice with 4T1.2 cancer.



Suppl.Fig.6

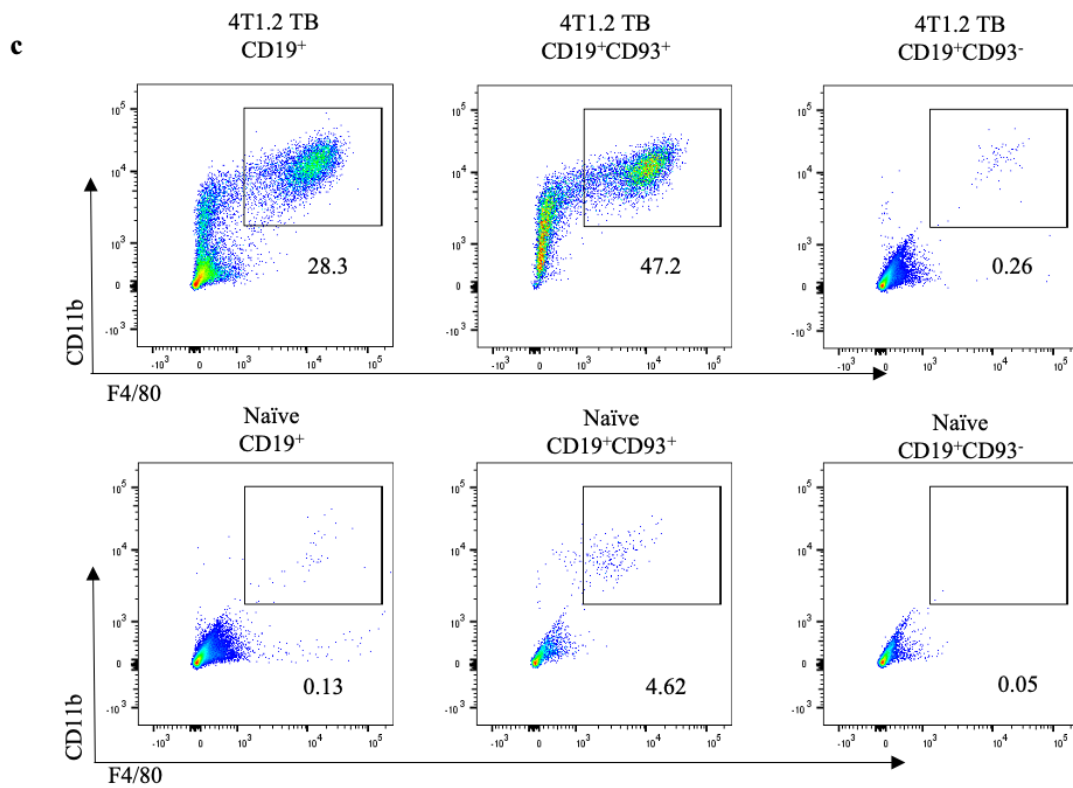
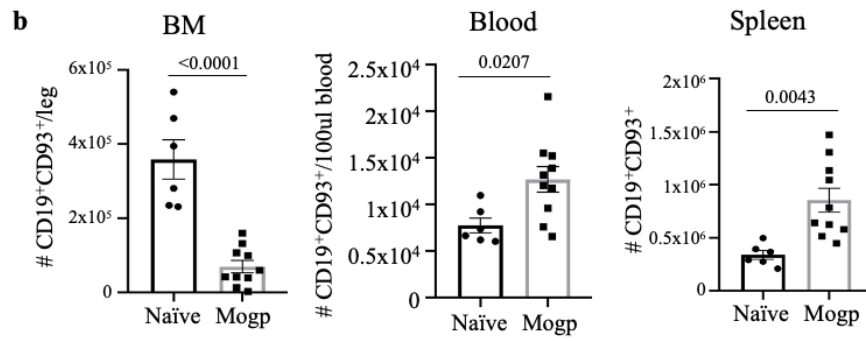
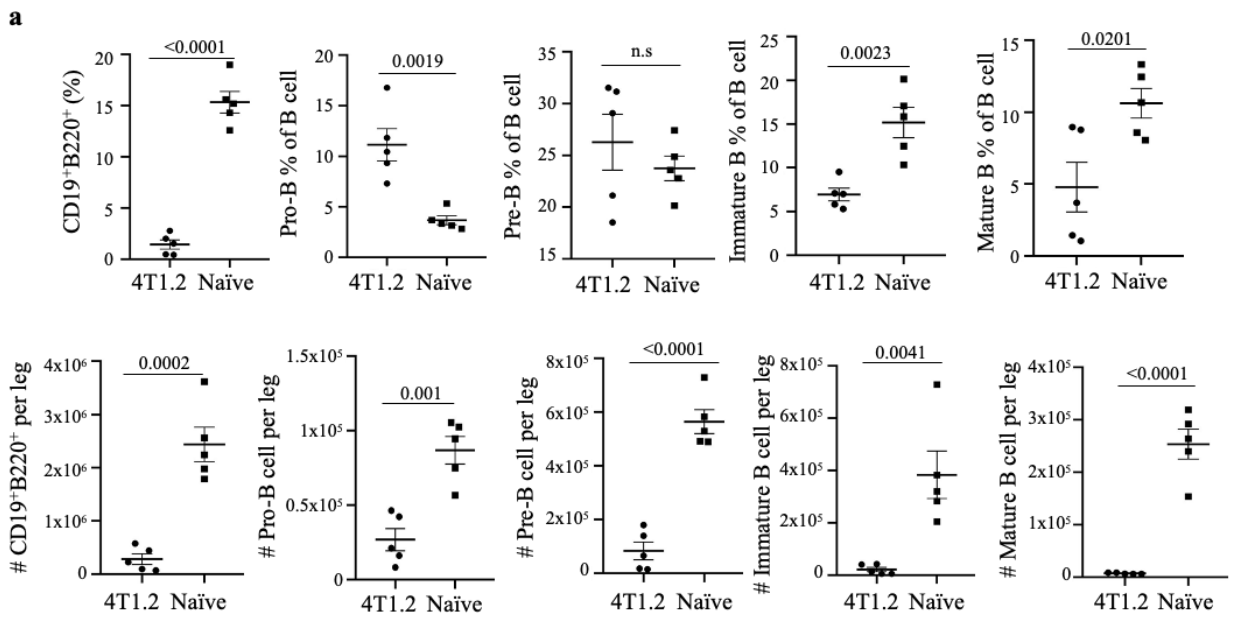


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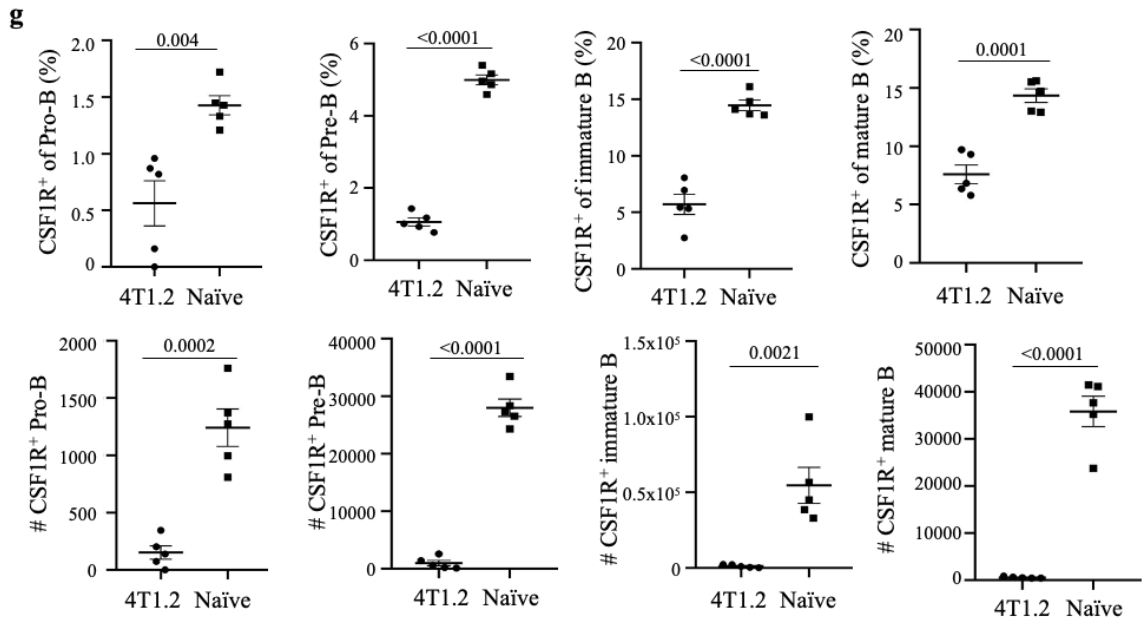
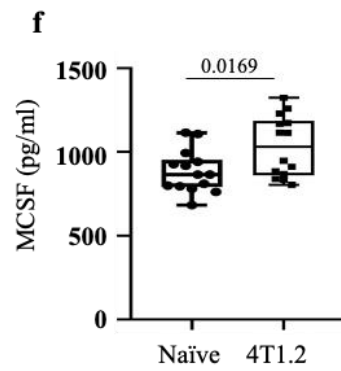
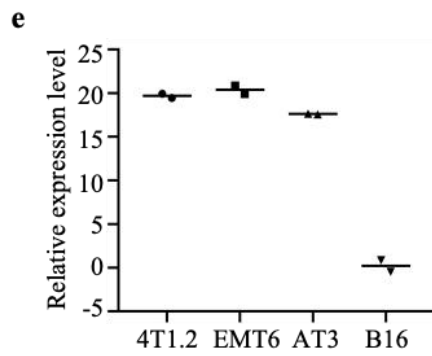
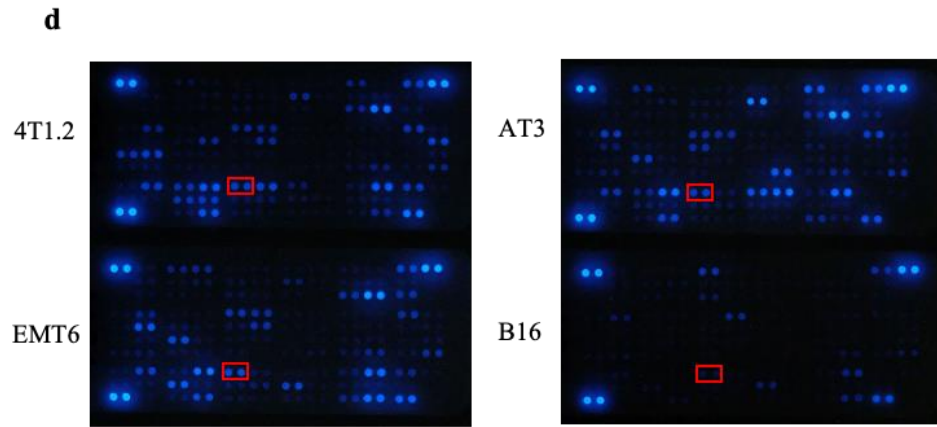


Supplementary Figure 6. B-MF and Mo-MF are functionally different cells. a, Representative FACS plots showing BrdU⁺ frequency in *in vitro* generated B-MF and Mo-MF after pulsing with BrdU for 24h. **b,** Quantification of Ki-67 expression of *in vitro* generated B-MF and Mo-MF, n=2. **c and d,** Unlike Mo-MF, B-MF suppress proliferation of T cells *in vitro*. Representative histogram showing the proliferation of CD4⁺ T cells (**c**)

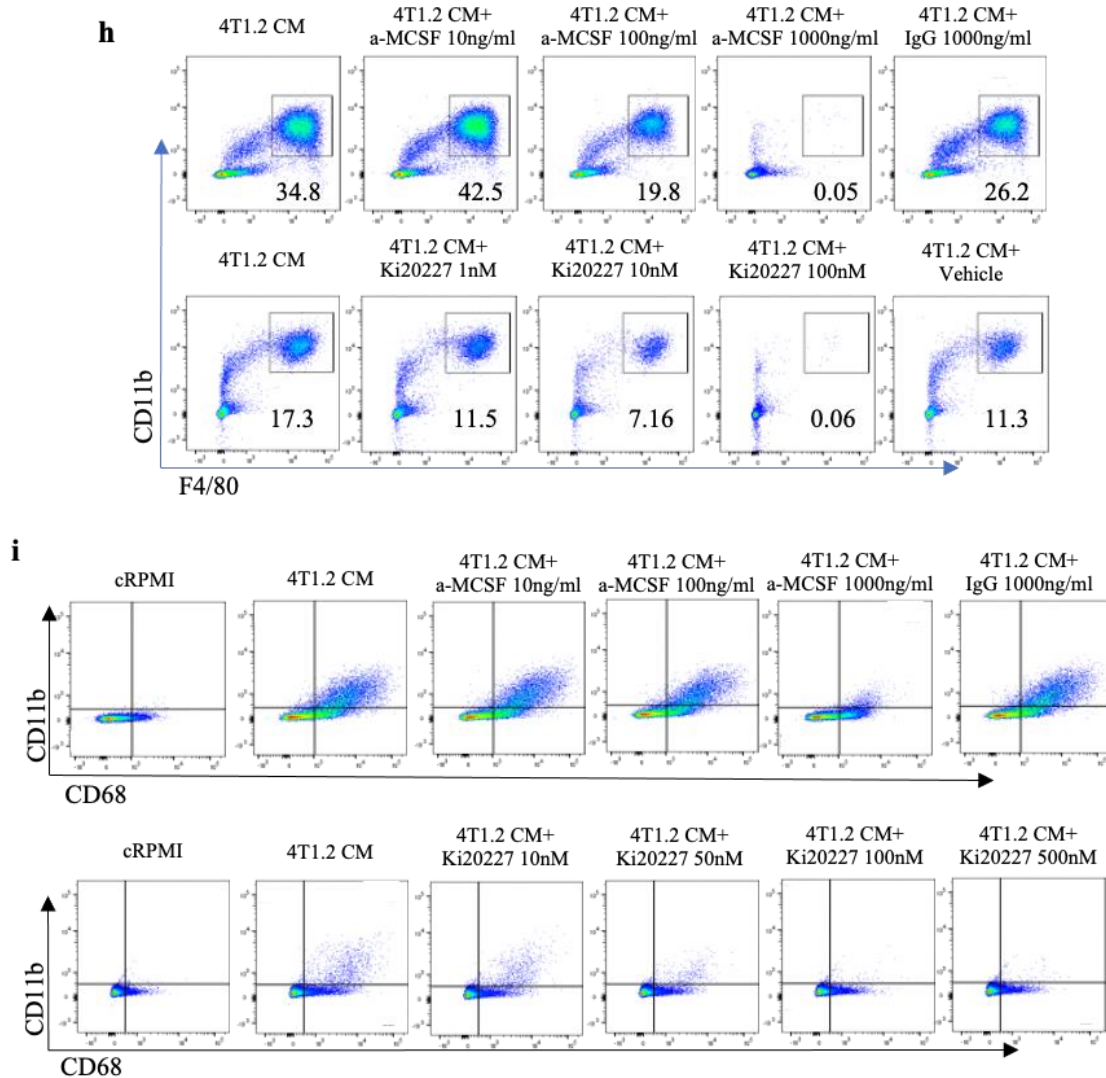
and CD8⁺ T cells (**d**) after B-MF and Mo-MF were cultured with T cells stimulated with anti-CD3/CD28 Abs at 10:1, 20:1 and 40:1 (T cell: MF) ratio for 4 days. **e**, Quantification (Mean \pm SEM, n=5 per group) of FoxP3⁺ Tregs *in vitro*-converted from CD25⁻CD4⁺ non-Tregs T cells (isolated from naïve mouse spleen) after 5-day co-culture with B-MF vs Mo-MF at 5:1 ($P=0.0004$), 10:1 ($P<0.0001$) and 20:1 (T cell: MF) ratio. P values were calculated using two-tailed unpaired t test. **f** and **g**, Box-Whiskers plot quantification (frequency and absolute numbers) of tumor-infiltrating CD4⁺ T cells and CD8⁺ T cells within CD45⁺ cells and their GrB and IFN- γ -expressing cells, and FoxP3⁺CD4⁺ T cells in s.c. tumor of μ MT C57BL/6 mice with B16-F10 melanoma (n=10 for PBS group, n=12 for B-MF group, CD45⁺#/g $P=0.017$, **f**) vs PBS and 4T1.2 μ MT BALB/c mice (n=12 for PBS group, n=14 for B-MF group, GrB⁺% of CD4⁺ $P=0.0018$, **g**) vs PBS. P values were calculated using two-tailed unpaired t test. Schema of adoptive transfer experiment in μ MT BALB/c mice (upper panel, **h**). Equal numbers of *in vitro*-generated B-MF or FACS-purified FOB (3×10^5 cells/mouse) were i.v. injected 3 and 10 days after s.c. challenge with 4T1.2. cancer (n=5 for B-MF group, n=8 for FOB group). Lung metastasis and tumor-infiltrated cells (TIL) were quantified at day 30 (lower panel, **h**). Shown are frequency within CD45⁺ cells and absolute numbers of TIL (CD4⁺% $P=0.0081$, IL10⁺% of CD4⁺ $P=0.001$, GrB⁺% of CD8⁺ $P=0.0244$, LAMP1⁺% of CD8⁺ $P=0.0135$, CD4⁺#/g $P=0.0479$, **h**) after B-MF vs FOB. P values were calculated using two-tailed unpaired t test. Schema of B-MF tracking experiment (**i**). eFluor450-labelled B-MF (5×10^5) were i.v. injected into μ MT BALB/c mice bearing a 14-day 4T1.2 tumor. After 3 days, absolute numbers (Mean \pm SEM, n=3 per group) of eFluor450⁺ in CD11b⁺F4/80⁺ were quantified (left panel, **j**) and proportion of CD11b⁺F4/80⁺ in eFluor450⁺ cells were determined by FACS (right panel, **j**) in indicated tissues and in the tumor. Absolute numbers of transferred eFluor450⁺ CD11b⁺F4/80⁺ cells per gram tumor was compared with that of B-MF in 4T1.2 tumor in BALB/c mice (CD79a⁺F4/80⁺CD11b⁺CD19⁻) and in MC38 tumor in Mb1-EYFP mice (EYFP⁺F4/80⁺CD11b⁺CD19⁻, **k**). Each symbol in **b** and **e-j** is for independent mouse.



Suppl. Fig.7

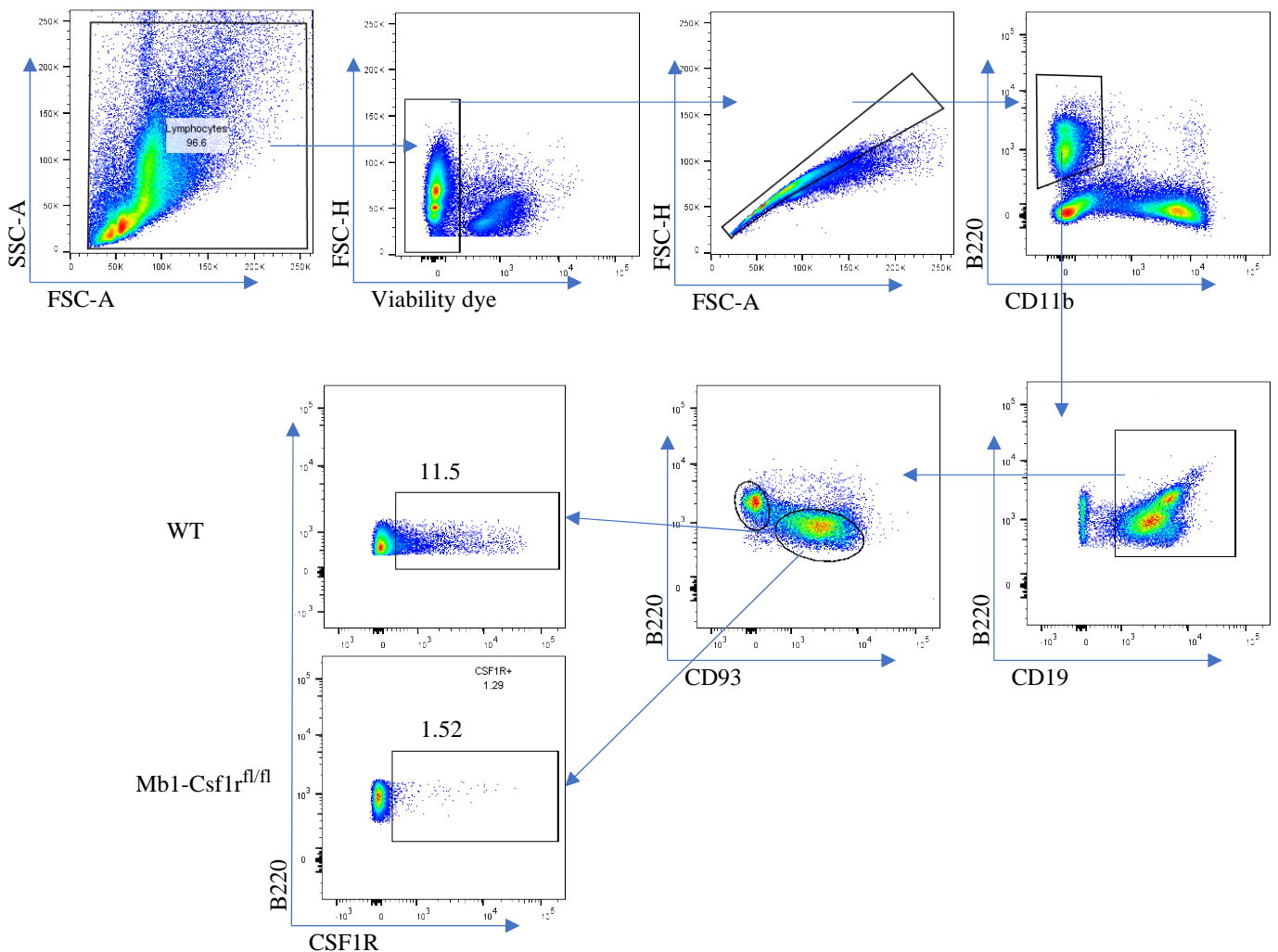


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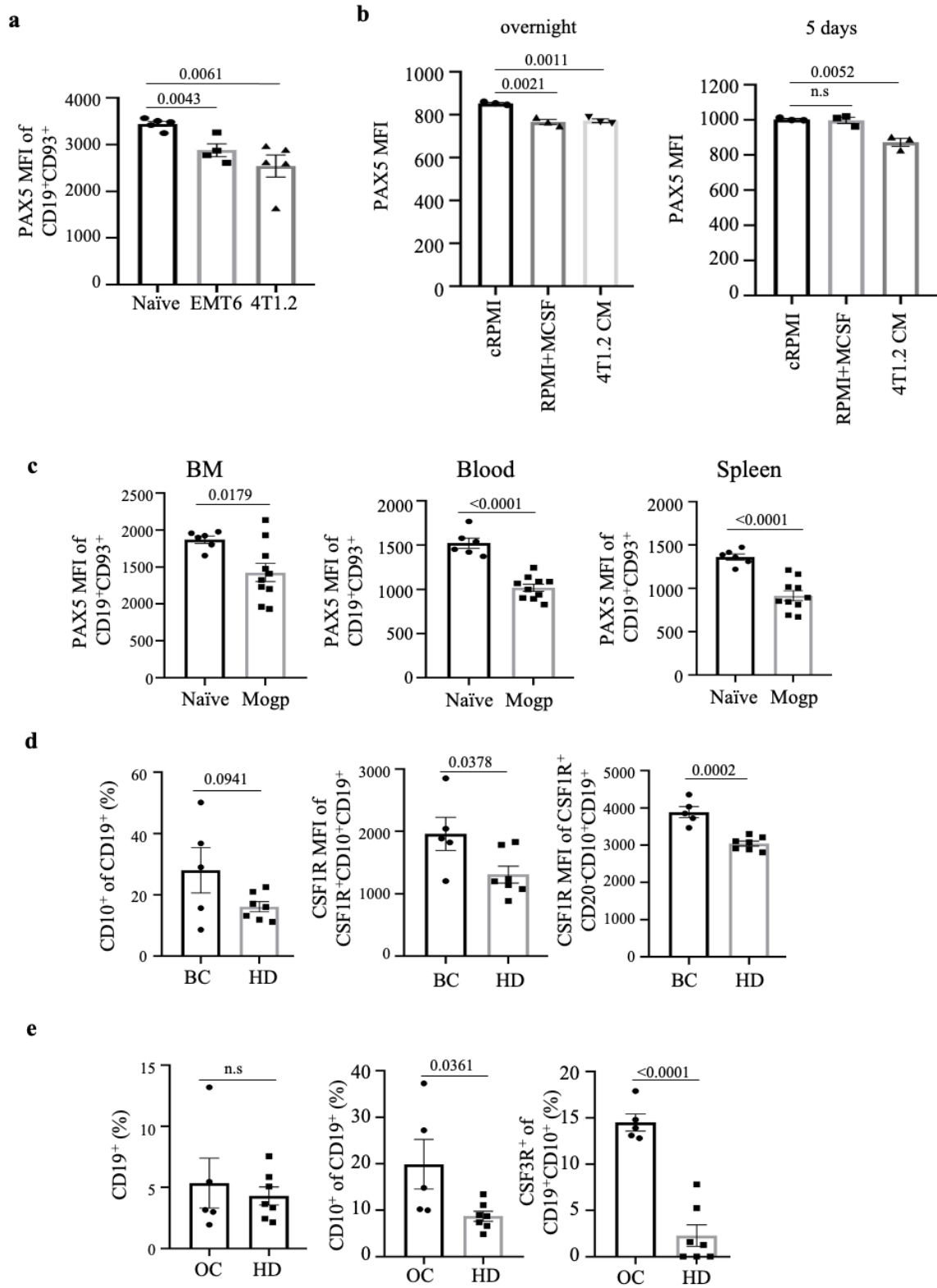


Supplementary Figure 7. CSF1/CSF1R signaling axis is needed for the generation B-MF. **a**, Mean \pm SEM frequency (top) and numbers (bottom) of CD19⁺B220⁺ B cells (frequency $P < 0.0001$, number $P = 0.0002$) and their subsets (pro-B frequency $P = 0.0019$ number $P = 0.001$, pre-B number $P < 0.0001$, immature B frequency $P = 0.0023$ number $P = 0.0041$ and mature B cells frequency $P = 0.0019$ number $P = 0.001$) in BM of mice with 4T1.2 cancer or naïve mice ($n = 5/\text{group}$). P values were calculated using two-tailed unpaired t test. **b**, Numbers of B-cell precursors (CD19⁺CD93⁺, Mean \pm SEM) in BM ($P < 0.0001$), PB ($P = 0.0207$), and spleen ($P = 0.0043$) of mice with Mogp cancer and naïve mice ($n = 6$ for naïve group, $n = 10$ for Mogp group). P values were calculated using two-tailed unpaired t test. **c**, Representative FACS plots showing F4/80⁺CD11b⁺ cells after *in vitro* culture of splenic CD19⁺, CD19⁺CD93⁺, and CD19⁺CD93⁻ B cells isolated from mice with 4T1.2 cancer (top) or naïve (bottom) mice in 4T1.2-CM for 7 days. **d** and **e**, Cytokine array image for expression of secreted factors in cancer CM (**d**) and relative levels (Mean) of M-CSF (red boxes in **d**) in CM of 4T1.2, EMT6, AT3 and B16-F10 cells ($n = 2$ mouse samples per group, **e**). **f**, Box-Whiskers plot amount of MCSF (pg/ml) in sera of naïve and 4T1.2 cancer-bearing mice ($n = 14/\text{group}$, ELISA data, $P = 0.0169$, **f**).

P values in **f** were calculated using two-tailed unpaired *t* test. **g**, Mean \pm SEM frequency (top) and numbers (bottom) of indicated CSF1R⁺ B-cell subsets in BM of mice with 4T1.2 cancer or naïve mice (pro-B frequency *P*=0.004 number *P*=0.0002, pre-B frequency *P*<0.0001 number *P*<0.0001, immature B frequency *P*<0.0001 number *P*=0.0021 and mature B cells frequency *P*=0.0001 number *P*<0.0001, n=5/group). *P* values were calculated using two-tailed unpaired *t* test. **h** and **i**, anti-MCSF Ab or Ki20227 (a specific *c-fms*/CSF1R inhibitor) inhibits ability of 4T1.2-CM to generate B-MF from of BM B-cell precursors (**h**) and 70z/3 cells (**i**) in dose-dependent manner. Numbers in dot plots are for % of CD11b⁺F/80⁺ cells. **a-c** and **g-i** were independently reproduced at least three times.



Supplementary Figure 8. Gating strategy for data presented in Fig.5F, G, showing surface CSF1R expression in BM CD93⁺CD19⁺ B-cell precursors of Mb1-CSF1R^{Flx/Flx} (bottom) and littermate Mb1-Cre (WT, top) mice. Numbers in the pots are for expression frequency of CSF1R (%) in the gated areas.



Supplementary Figure 9. Cancers induce PAX5 downregulation in B-cell precursors. **a**, FACS evaluation of Pax5 (MFI Mean \pm SEM) in PB CD19⁺CD93⁺ B cells from naïve mice and mice with EMT6 and 4T1.2 cancers (n=5 for naïve group, n=4 for

EMT6 group and $n=5$ for 4T1.2 group, $P=0.0043$ naïve vs EMT6 and $P=0.0061$ naïve vs 4T1.2). P values were calculated using two-tailed unpaired t test. **b**, Expression of Pax5 (MFI Mean \pm SEM) in 70z/3 cells after treatment with M-CSF or 4T1.2-CM ($n=3$ /group) for overnight ($P=0.0021$ cRPMI vs RPMI+MCSF and $P=0.0011$ cRPMI vs 4T1.2 CM, left) or 5 days ($P=0.0052$ cRPMI vs 4T1.2 CM, right). P values were calculated using two-tailed unpaired t test. **c**, Pax5 (MFI Mean \pm SEM) in B-cell precursors ($CD19^+CD93^+$) from BM($P=0.0179$), PB($P<0.0001$) and spleen($P<0.0001$) from mice with Mogp cancer and naïve mice ($n=6$ for naïve group, $n=10$ for Mogp group). P values were calculated using two-tailed unpaired t test. **d** and **e**, FACS evaluation of PB cells of breast cancer patients ($n=5$) and healthy human donors ($n=7$). P values were calculated using two-tailed unpaired t test. **d**, Mean \pm SEM frequency of CD10 expression in $CD19^+$ B cells($P=0.0941$) and CSF1R expression in $CD19^+CD10^+$ ($P=0.0378$) and $CD19^+CD10^+CD20^-$ B cells($P=0.0002$); and **e**, $CD19^+$ cells and CD10 expression in $CD19^+$ cells($P=0.0361$) and CSF3R expression in $CD19^+CD10^+$ cells($P<0.0001$) in PB of ovarian cancer patients and healthy donors. P values were calculated using two-tailed unpaired t test.

Supplementary Table 1. List of antibodies used in the study

Antibody	Clone	Species reactivity	Catalog #	Company
CD19-PE	1D3	Mouse	152408	BioLegend
CD19-APC	1D3	Mouse	152410	BioLegend
CD19-BUV421	HIB19	Human	302234	BioLegend
F4/80-APC	BM8	Mouse	123116	BioLegend
F4/80-BV421	BM8	Mouse	123137	BioLegend
CD11b-BV510	M1/70	Mouse/Human	101263	BioLegend
CD11b-FITC	M1/70	Mouse/Human	101206	BioLegend
CD64-PE/Dazzle 594	X54-5/7.1	Mouse	139319	BioLegend
CX3CR1-PerCP/Cyanine5.5	SA011F11	Mouse	149009	BioLegend
CD68-BV421	FA-11	Mouse	137017	BioLegend
CD93-APC	AA4.1	Mouse	136510	BioLegend
CSF1R-PE	AFS98	Mouse	135505	BioLegend
CSF1R-PE/Cyanine7	AFS98	Mouse	135524	BioLegend
IgM-PE/Cyanine7	RMM-1	Mouse	406514	BioLegend
IgD-BV510	11-26c.2a	Mouse	405723	BioLegend
CD20-BV421	SA275A11	Mouse	150405	BioLegend
CD20-APC	2H7	Human	302310	BioLegend
CD10-PE	HI10a	Human	312204	BioLegend
CD79a-PE	F11-172	Mouse	133103	BioLegend
TER119-Biotin	TER-119	Mouse	116204	BioLegend
CD11b-Biotin	M1/70	Mouse/Human	101204	BioLegend
Gr-1-Biotin	RB6-8C5	Mouse	108404	BioLegend
CD3 ϵ -Biotin	145-2C11	Mouse	100304	BioLegend
NK1.1-Biotin	PK136	Mouse	108704	BioLegend
CD49b-Biotin	HMA2	Mouse	103522	BioLegend
CD49b-Biotin	DX5	Mouse	108904	BioLegend
Ly6C-Biotin	HK1.4	Mouse	128004	BioLegend
Ly6C-FITC	HK1.5	Mouse	128005	BioLegend
Ly6G-Biotin	1A8	Mouse	127604	BioLegend
Ly6g-BV650	1A9	Mouse	127641	BioLegend
CD11c-Biotin	N418	Mouse	117304	BioLegend
CD11c-BV510	N419	Mouse	117353	BioLegend
CD117-Biotin	2B8	Mouse	105804	BioLegend

Alexa Fluor® 488					
Streptavidin	-	-	405235	BioLegend	
I-A/I-E-PerCP/Cyanine5.5	M5/114.15.2	Mouse	107626	BioLegend	
CD206-FITC	C068C2	Mouse	141704	BioLegend	
CD207-PE/Cyanine7	4C7	Mouse	144209	BioLegend	
		Mouse/Huma			
B220-FITC	RA3-6B2	n	103206	BioLegend	
IgM-PE/Dazzle™ 594	RMM-1	Mouse	406530	BioLegend	
CD93-PE	AA4.1	Mouse	136504	BioLegend	
CD43-APC	S11	Mouse	143208	BioLegend	
CD24-BV605	M1/69	Mouse	101827	BioLegend	
CD274-PE/Cyanine7	10F.9G2	Mouse	124314	BioLegend	
LAP-PerCP/Cyanine5.5	TW7-16B4	Mouse	141410	BioLegend	
CD4-PerCP/Cyanine5.5	GK1.5	Mouse	100434	BioLegend	
CD8a-FITC	53-6.7	Mouse	100706	BioLegend	
IFN-γ-APC	XMG1.2	Mouse	505810	BioLegend	
		Mouse/Huma			
PAX5-Alexa Fluor® 488	1H9	n	562816	BD	
CD71-BV605	C2	Mouse	563013	BD	
CD124-BV421	Mil4R-M1	Mouse	564086	BD	
CD366-BV421	5D12	Mouse	747626	BD	
CD19-BUV395	1D3	Mouse	563557	BD	
		Mouse/			
CD11b-BUV395	M1/70	Human	565976	BD	
		Mouse/			
CD11b-BV786	M1/70	Human	740861	BD	
CSF1R-BB515	9-4D2-1E4	Human	565346	BD	
LDLR-BUV805	C7	Human	748950	BD	
CSF3R-BUV395	LMM741	Human	743524	BD	
			12-8898-	eBioscienc	
Granzyme B-PE/Cyanine7	NGZB	Mouse	82	e	
			12-5773-	eBioscienc	
FOXP3-PE	FJK-16s	Mouse	82	e	
		Mouse/Huma	48-5698-	eBioscienc	
Ki67-eFluor 450	SolA15	n	82	e	
			61-0689-	eBioscienc	
CD68-PE-eFluor 610	Y1/82A	Human	42	e	
Anti-CD19	rabbit		ab24523		
antibody[EPR23174-145]	monoclonal	Mouse	5	abcam	
	Rat				
Anti-CD68 antibody[FA-11]	monoclonal	Mouse	ab53444	abcam	