

Supplemental Tables

Supplemental Table 1. Mastocytosis Subtypes (Variants)

Cutaneous mastocytosis

- Urticaria pigmentosa / maculopapular cutaneous mastocytosis
- Diffuse cutaneous mastocytosis
- Mastocytoma of skin

Systemic mastocytosis

- Indolent systemic mastocytosis (includes bone marrow mastocytosis)^a
- Smoldering systemic mastocytosis ^a
- Aggressive systemic mastocytosis ^a
- Mast cell leukemia
- Systemic mastocytosis with an associated myeloid neoplasm

Mast cell sarcoma

^a The diagnosis of these variants of systemic mastocytosis require correlation with B and C findings.

Supplemental Table 2. Systemic Mastocytosis: B-findings

1. High mast cell burden, >30% of BM cellularity by mast cell aggregates (assessed on BM biopsy) and serum tryptase >200 ng/mL
2. Cytopenia (not meeting criteria for C findings) or -cytosis. Reactive causes are excluded, and criteria for other myeloid neoplasms are not met.
3. Hepatomegaly without impairment of liver function, or splenomegaly without features of hypersplenism including thrombocytopenia, and/ or lymphadenopathy (>1 cm size) on palpation or imaging

Supplemental Table 3. Mast Cell Leukemia

1. Meets diagnostic criteria for SM
2. BM aspirate shows $\geq 20\%$ atypical immature mast cells^{a/b}

^a Atypical immature mast cells include promastocytes, metachromatic blast-like forms, multinucleated or highly pleomorphic mast cells.

^b In the presence of a suboptimal aspirate (dry tap), a BM biopsy showing a dense, diffuse mast cell infiltration of atypical immature mast cells is sufficient to support the diagnosis of mast cell leukemia.

Supplemental Table 4. Features of the various clonal cytopenias

Type		Dysplasia ^a	Other features
Clonal cytopenia of undetermined significance (CCUS)		Absent	No comorbid condition that explains the cytopenia; no MDS-defining genetic abnormality ^b
VEXAS syndrome		Absent ^c	Autoimmune manifestations <i>UBA1</i> mutation; no MDS-defining genetic abnormality ^b
Myelodysplastic syndrome		Present, or an MDS-defining genetic abnormality ^b	Classified according to Table 20

^aDefined as dysplastic cytologic changes in $\geq 10\%$ of erythroid cells, granulocytic cells, and/or megakaryocytes; absence of dysplasia requires less than these thresholds in all lineages in an adequate bone marrow sample.

^bMDS-defining genetic abnormalities are *SF3B1* mutation (VAF $>10\%$), multi-hit *TP53* mutation (VAF $>10\%$), or del(5q), del(7q), -7 or complex karyotype detected by karyotype. Additional features may be required in the setting of a germline predisposition condition (see “Diagnosis of Myelodysplastic Syndrome in the Setting of Germline Predisposition” section).

^cVacuolation of erythroid and myeloid precursors are not considered to be MDS-defining dysplastic features in the setting of VEXAS.

Supplemental Table 5. Symptoms and signs at diagnosis of JMML.¹

Pallor	64%	Splenomegaly	97%
Fever	54%	Hepatomegaly	97%
Infection	45%	Lymphadenopathy	76%
Bleeding	46%	Skin rash	36%
Cough	40%	Café au lait spots	12%
Abdominal pain	7%	Xanthoma	7%
Bone pain	< 4%	CNS infiltration*	< 3%
Diarrhea	< 4%		

*myeloid sarcoma, diabetes insipidus, facial palsy.

Supplemental Table 6. Acute myeloid leukemia (AML) with other rare recurring translocations

- AML with t(1;3)(p36.3;q21.3)/*PRDM16::RPN1*
- AML with t(3;5)(q25.3;q35.1)/*NPM1::MLF1*
- AML with t(8;16)(p11.2;p13.3)/*KAT6A::CREBBP*
- AML (megakaryoblastic) with t(1;22)(p13.3;q13.1)/*RBM15::MRTF1**
- AML with t(5;11)(q35.2;p15.4)/ *NUP98::NSD1**
- AML with t(11;12)(p15.4;p13.3)/*NUP98::KMD5A**
- AML with *NUP98* and other partners*
- AML with t(7;12)(q36.3;p13.2)/*ETV6::MNX1**
- AML with t(10;11)(p12.3;q14.2)/*PICALM::MLLT10*
- AML with t(16;21)(p11.2;q22.2)/*FUS::ERG*
- AML with t(16;21)(q24.3;q22.1)/*RUNX1::CBFA2T3*
- AML with inv(16)(p13.3q24.3)/*CBFA2T3::GLIS2**

* Occurs predominantly in infants and children

Supplemental Table 7. Provisional entities in acute lymphoblastic leukemia/lymphoma (ALL) (See references for reviews and additional references therein) ²⁻¹⁴

B-ALL					
Subtype	Frequency	Gene expression pattern	Genomics	Immunophenotype	Comment
<i>ETV6::RUNX1</i> -like	<5%, mostly children	GEP Clusters with <i>ETV6::RUNX1</i>	Fusions or CNAs in ETS family genes including <i>ETV6</i> , <i>FUS</i> and also <i>IKZF1</i> ; some cases harbor germline LOF <i>ETV6</i> mutations	CD27+CD44- or low, like <i>ETV6::RUNX1</i>	Good prognosis in children, not adults
<i>PAX5</i> alteration** (<i>PAX5alt</i>)	10% children and adults	Broad GEP cluster near Ph like	Various mutations (esp. compound heterozygosity for R38, R140), intragenic amplifications and non-kinase fusions (<i>ETV6</i> most common)		Intermediate prognosis
<i>ZEB2</i> (p.H1038R) or <i>IGH::CEBPE</i>	< 1%	GEP cluster of <i>ZEB2</i> H1038R and <i>IGH::CEBPE</i> fusion	<i>ZEB2</i> (H1038R) mutation, mostly coexisting with <i>IGH::CEBPE</i> or other known gene fusions, Frequent <i>NRAS</i> mutations (50%), LMO1 upregulation and downregulation of <i>SMAD1</i> and <i>BMP2</i>		
<i>KMT2A</i> -like	<1%	GEP Clusters with <i>KMT2A</i>	Some with <i>HOXA</i> fusions		
<i>ZNF384</i> -like	<1%	GEP clusters with <i>ZNF384</i> or <i>ZNF362</i>	unknown	Like <i>ZNF384</i>	
T-ALL/LL					
Subtype	Frequency	Partner genes/other rearrangements	Immunophenotype	Comment	
<i>HOXA</i> dysregulated	15-25%	<i>HOXA::TRB/TRG</i> ; <i>KMT2A</i> -Rearranged; <i>PICALM::MLLT10</i> ; <i>SET::NUP214</i>		Immature, some ETP	

<i>SPI1</i> rearrangement	<5%, children	<i>STMN1; TCF7; BCL11B</i>	CD4-, CD8+/-, DR+	Very poor prognosis
<i>TLX1</i> rearrangement	5-10% children; near 30% adult	TCR	CD4+, CD8+/-, CD1a+, cortical thymocyte	Good prognosis
<i>TLX3</i> rearrangement	20-25% children <5% adult	TCR; <i>BCL11B; CDK6</i>	CD4+, CD8+/-, CD1a+, cortical thymocyte, some ETP or near ETP	Good prognosis; <i>BCL11B</i> overexpression different from ETP group
<i>NKX2</i> rearrangement	<5% children	<i>NKX2.1/NKX2.2/NKX2.5::TCR; BCL11B; CDK6</i>	CD4+, CD8+	Similar GEP to <i>TLX1-R</i>
<i>TAL1-2</i> rearrangement	30-40% (TAL2 rare)	TRA/D; TRB (<i>TAL2</i>); 1p32 deletion (<i>STIL</i>); intergenic SNV (super enhancer)	CD3+, late cortical	Poor prognosis
<i>LMO1-2</i> rearrangement	<i>LMO1-R</i> -5% <i>LMO2-R</i> 10%	TCR; cryptic deletion; enhancer/promoter mutations	Immature but not-ETP	Form LMO complex with bHLH factors. Extremely high LMO expression.
<i>BHLH</i> , other	<2%	<i>LYL1::TRB</i> <i>OLIG2/BHLHB1::TCR</i>	Immature but not ETP	Extremely high LMO expression <i>LYL1-R</i> shows stem cell-like signature

^a Not all examples of these entities can be defined by conventional FISH or WTS alone because of intragenic alterations that dysregulate these genes and produce leukemias with an identical gene expression pattern.

WTS; whole transcriptome sequencing; GEP; gene expression profile, LMO; LIM-domain-only, BHLH; basic helix-loop-helix, CNA: copy number alteration

Supplemental Table 8. ICC Myeloid Neoplasm/Acute Leukemia Disease Co-chairs and Work Group Participants.

Co-Chairs

Daniel Arber
Robert Hasserjian
Attilio Orazi
Mario Cazzola
Hartmut Döhner
Ayalew Tefferi

Chronic Myeloid Leukemia

Hans-Michael Kvasnicka (lead)
Ayalew Tefferi (lead)
Susan Bradford
Carlos Bueso-Ramos
Jorge Cortes
Attilio Orazi
Annette Schmitt-Graeff

Myeloproliferative Neoplasms

Hans-Michael Kvasnicka (lead)
Ayalew Tefferi (lead)
Naseema Gangat
Umberto Gianelli
Attilio Orazi
Jürgen Thiele
Alessandro M. Vannucchi

Eosinophilic and Mast Cell Disorders

Sa A. Wang (lead)
Ayalew Tefferi (lead)
Daniel Arber
Jason Gotlib
Robert Hasserjian
Attilio Orazi
Andreas Reiter
Alexander Tzankov

Myelodysplastic/Myeloproliferative Neoplasms

Attilio Orazi (lead)
Luca Malcovati (lead)
Daniel Arber
Carlos Bueso-Ramos
Mario Cazzola
Robert Hasserjian
Michael Savona

Pre-malignant Clonal Cytopenias and Myelodysplastic Syndromes

Robert Hasserjian (lead)

Mario Cazzola (lead)

Benjamin Ebert

Eva Hellström-Linberg

Attilio Orazi

Alberto Orfao

Maria Rozman

Sa A. Wang

Pediatric and/or Germline Mutation-associated Disorders

Katherine Calvo (lead)

Irith Baumann

Charlotte Niemeyer (lead)

Martina Rudelius

Akiko Shimamura

Olga Weinberg

Acute Myeloid Leukemias

Daniel Arber (lead)

Harmut Döhner (lead)

Hervé Dombret

Eric Duncavage

Kathryn Foucar

Robert Hasserjian

Bob Löwenberg

Attilio Orazi

Olga Weinberg

Acute Lymphoblastic Leukemia

Michael Borowitz (lead)

Harmut Döhner (lead)

Daniel Arber

Kathryn Foucar

Nicola Gökbuget

Elias Jabbour

Mignon Loh

Charles Mullighan

Ching-Hon Pui

Cytogenetics

Paola Dal Cin

Harmut Döhner

Michelle Le Beau

1. Niemeyer CM, Aric M, Basso G, et al. Chronic myelomonocytic leukemia in childhood: A retrospective analysis of 110 cases. *Blood*. 1997;89(10):3534-3543.
2. Iacobucci I, Kimura S, Mullighan CG. Biologic and Therapeutic Implications of Genomic Alterations in Acute Lymphoblastic Leukemia. *J Clin Med*. 2021;10(17).
3. Zaliova M, Stuchly J, Winkowska L, et al. Genomic landscape of pediatric B-other acute lymphoblastic leukemia in a consecutive European cohort. *Haematologica*. 2019;104(7):1396-1406.
4. Li JF, Dai YT, Lilljebjorn H, et al. Transcriptional landscape of B cell precursor acute lymphoblastic leukemia based on an international study of 1,223 cases. *Proc Natl Acad Sci U S A*. 2018;115(50):E11711-E11720.
5. Paietta E, Roberts KG, Wang V, et al. Molecular classification improves risk assessment in adult BCR-ABL1-negative B-ALL. *Blood*. 2021;138(11):948-958.
6. Lilljebjorn H, Fioretos T. New oncogenic subtypes in pediatric B-cell precursor acute lymphoblastic leukemia. *Blood*. 2017.
7. Liu Y, Easton J, Shao Y, et al. The genomic landscape of pediatric and young adult T-lineage acute lymphoblastic leukemia. *Nat Genet*. 2017;49(8):1211-1218.
8. Chang YH, Yu CH, Jou ST, et al. Targeted sequencing to identify genetic alterations and prognostic markers in pediatric T-cell acute lymphoblastic leukemia. *Sci Rep*. 2021;11(1):769.
9. Chen B, Jiang L, Zhong ML, et al. Identification of fusion genes and characterization of transcriptome features in T-cell acute lymphoblastic leukemia. *Proc Natl Acad Sci U S A*. 2018;115(2):373-378.
10. Cordo V, van der Zwet JCG, Cante-Barrett K, Pieters R, Meijerink JPP. T-cell Acute Lymphoblastic Leukemia: A Roadmap to Targeted Therapies. *Blood Cancer Discov*. 2021;2(1):19-31.
11. Girardi T, Vicente C, Cools J, De Keersmaecker K. The genetics and molecular biology of T-ALL. *Blood*. 2017;129(9):1113-1123.
12. van der Zwet JCG, Cordo V, Cante-Barrett K, Meijerink JPP. Multi-omic approaches to improve outcome for T-cell acute lymphoblastic leukemia patients. *Adv Biol Regul*. 2019;74:100647.
13. Bardelli V, Arniani S, Pierini V, et al. T-Cell Acute Lymphoblastic Leukemia: Biomarkers and Their Clinical Usefulness. *Genes (Basel)*. 2021;12(8).
14. Tran TH, Langlois S, Meloche C, et al. Whole-transcriptome analysis in acute lymphoblastic leukemia: a report from the DFCI ALL Consortium Protocol 16-001. *Blood Adv*. 2022;6(4):1329-1341.