

Supplementary Material

Table S1 Gene editing efficiency of different RNP delivery systems

Delivery system	Cell Type	Efficiency	Refs
CRISPR/Cas9_{-3NLS}/sgHMGA2@PDA	MKN-45 cells	82%	This work
Metal organic frameworks	MCF-7 cells	~60%	[1]
Graphene oxide	AGS cells	~39%	[2]
Black phosphorus nanosheets	MCF-7 cells	32.1%	[3]
Calcium phosphate	Protoplast cells	20%	[4]
Gold nanocluster	A375 cells	26.2%	[5]
DNA nanoclews	3T3-L1 cells	75%	[6]
Chitosan nanoparticles	U2OS cells	55.8%	[7]
Poly(β -amino ester) nanoparticles	HEK293T cells	77%	[8]
Lipid nanoparticles	HEK cells	70%	[9]
Lipid nanoparticles	Jurkat T cells	8%	[10]

Amino acid sequences of the CRISPR/Cas9:

10	20	30	40	50
MNFKILPIAI DLGVKNTGVF SAFYQKGTSL ERLDNKNGKV YELSKDSYTL				
60	70	80	90	100
LMNNRTARRH QRRGIDRKQL VKRFLKLIWT EQLNLEWDKD TQQAISFLFN				
110	120	130	140	150
RRGFSFITDG YSPEYLNIVP EQVKAILMDI FDDYNGEDDL DSYLKLATEQ				
160	170	180	190	200
ESKISEIYNK LMQKILEFKL MKLCTDIKDD KVSTKTL KEI TSYEFELLAD				
210	220	230	240	250
YLANYSESLK TQKFSYTDKQ GNL KE LSYYH HDKYNIQEFL KRHATINDRI				
260	270	280	290	300
LDTLLTDDLD IWNFNFEKFD FDKNEEKLN QEDKDHIQAH LHHFVFAVNK				
310	320	330	340	350
IKSEMASGGR HRSQYFQEIT NVLDENNHQE GYLKNFCENL HNKKYSNLSV				
360	370	380	390	400
KNLVNLIGNL SNLELKPLRK YFNDKIHAKA DHWDEQKfte TYCHWILGEW				
410	420	430	440	450
RVGVKDQDKK DGAKYSYKDL CNELKQKVTk AGLVDFLLEL DPCRTIPPYL				
460	470	480	490	500
DNNNRKPPKC QSLILNPKFL DNQYPNWQQY LQELKKLQSI QNYLDSFETD				
510	520	530	540	550
LKVLKSSKDQ PYFVEYKSSN QQIASGQRDY KDL DARILQF IFDRVKASDE				
560	570	580	590	600
LLLNEIYFQA KKLKQKASSE LEKLESSKKL DEVIANSQLS QILKSQHTNG				
610	620	630	640	650
IFEQGTFLHL VCKYYKQRQR ARDSRLYIMP EYRYDKKLHK YNNTGRFDDDD				
660	670	680	690	700
NQLLYC�HK PRQKRYQLLN DLAGVLQVSP NFLKDKIGSD DDLFISKWLV				
710	720	730	740	750
EHIRGFKKAC EDSLKIQKDN RGLLNHKINI ARNTKGKCE KE IFNLICKIE				
760	770	780	790	800
GSEDKKGNyK HGLAYELGVL LFGEPNEASK PEFDRKIKKF NSIYSFAQIQ				
810	820	830	840	850
QIAFAERKGN ANTCAVCSAD NAHRMQQIKI TEPVEDNKDK IILSAKAQRL				
860	870	880	890	900
PAIPTRIVDG AVKKMATILA KNIVDDNWQN IKQVLSAKHQ LHIPIITESN				
910	920	930	940	950
AFEFEPALAD VKGKSLKDRR KKALERISPE NIFKDKNNRI KE FAKGISAY				
960	970	980	990	1000

SGANLTDGDF DGA**KE**EELDHI IPRSHKKYGT LNDEANLICV TRGDNKNKGN
 1010 1020 1030 1040 1050
 RIFCLRDLAD NYKLLKQFETT DDLEIEKKIA DTIWDANKKD FKFGNYRSFI
 1060 1070 1080 1090 1100
 NLTPQEQKAF RHALFLADEN PIKQAVIRAI NNRNRTFVNG TQRYFAEVLA
 1110 1120 1130 1140 1150
 NNIYLRAK**KE**NLNTDKISFD YFGIPTIGNG RGIAEIRQLY EKVDSDIQAY
 1160 1170 1180 1190 1200
 AKGDKPQASY SHLIDAMLAF CIAADEHRND GSIGLEIDKN YSLYPLDKNT
 1210 1220 1230 1240 1250
 GEVFTKDIFS QIKITDNEFS DKKLVRKKAI EGFNTHRQMT RDGIYAENYL
 1260 1270 1280 1290 1300
 PILH**KE**LNE VRKGYTWKNS EEIKIFKGKK YDIQQLNNLV YCLKFVDKPI
 1310 1320 1330 1340 1350
 SIDIQISTLE ELRNILTTNN IAATAEYYYI NLKTQKLHEY YIENYNTALG
 1360 1370 1380 1390 1400
 YKKYS**KE**MEF LRSLAYRSER VKIKSIDDVK QVLDKDSNFI IGTKITLPFK**K**
 1410 1420 1430 1440 1450
EWQRLYREWQ NTTIKDDYEF LKSFFNVKSI TKLHKKVRKD FSLPISTNEG
 1460 1470 1480 1490 1500
 KFLVKRKTWD NNFIYQILND SDSRADGTKP FIPAFDISKN EIVEAIIDSF
 1510 1520 1530 1540 1550
 TSKNIFWLPK NIELQKVDNK NIFAITSKW FEVETPSDLR DIGIATIQYK
 1560 1570 1580 1590 1600
 IDNNSRPKVR VKLDYVIDDD SKINYFMNHS LLKSRYPDKV LEILKQSTII
 1610 1620 1621
 EFESSGFNKT I**KE**MLGMKLA GIYNETSNN

Fig. S1. Amino acid sequences of the CRISPR/Cas9. The KE sequence has been highlighted in yellow.

The sequences are extracted from UniprotKB-A0Q5Y3 (CAS9_FRATN).

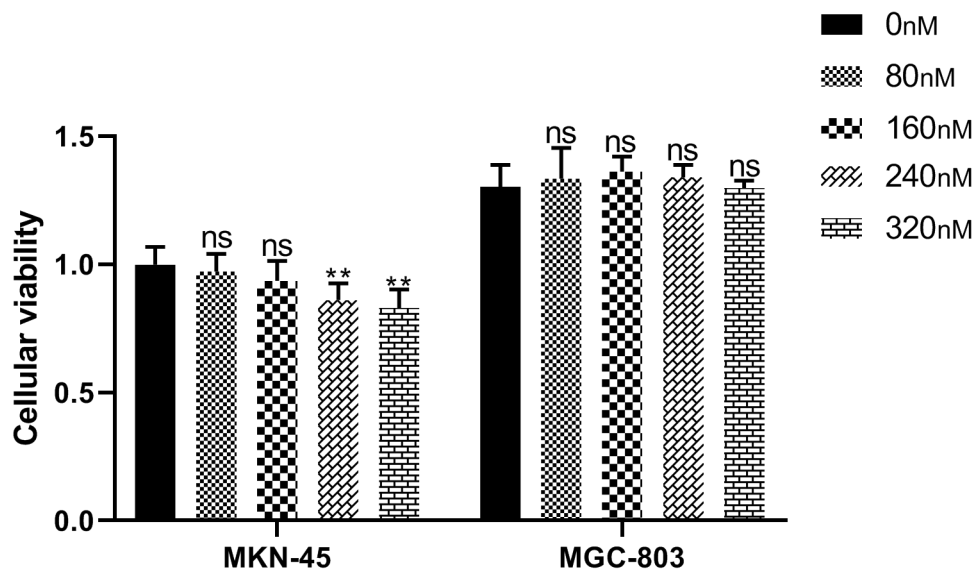


Fig. S2. Cell viability of MKN-45 and MGC-803 cells after incubating with CRISPR/Cas9-3NLS@PDA (n=3), ns: no significant differences; ** $P < 0.01$; **** $P < 0.001$, Student's t test. Error bars represent SD.

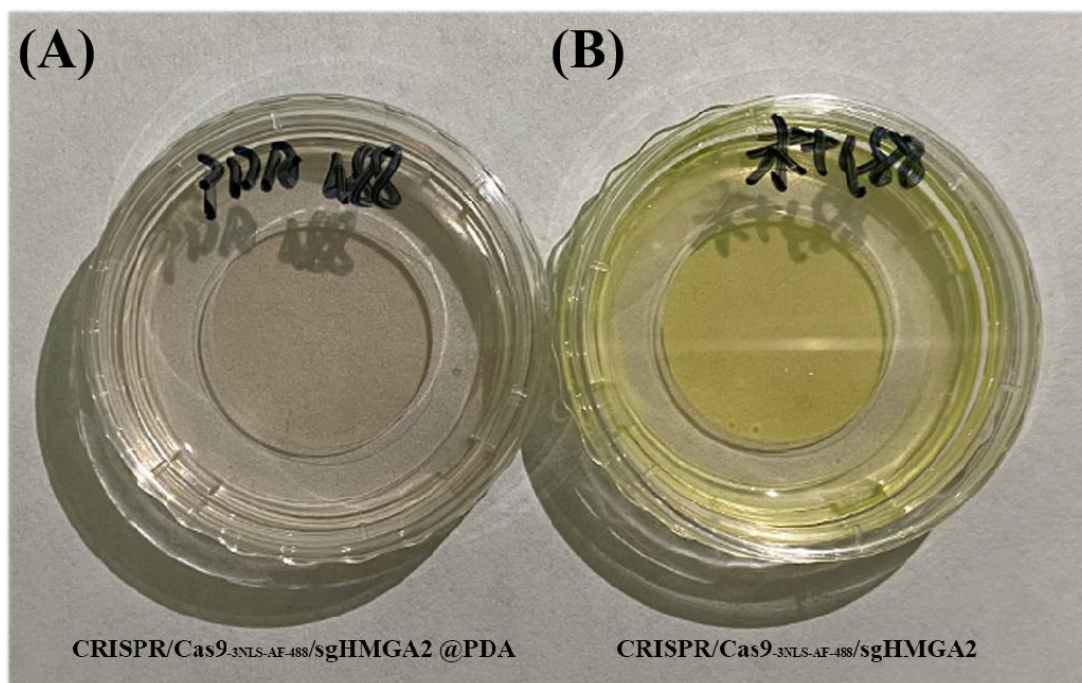


Fig. S3. (A) The color of CRISPR/Cas9-3NLS-AF-488/sgHMGA2@PDA solution. (B) The color of CRISPR/Cas9-3NLS-AF-488/sgHMGA2 solution

References

- [1] M.Z. Alyami, S.K. Alsaiani, Y. Li, S.S. Qutub, F.A. Aleisa, R. Sougrat, J.S. Merzaban, N.M. khashab, Cell-Type-Specific CRISPR/Cas9 Delivery by Biomimetic Metal Organic Frameworks, *J Am Chem Soc* 142 (2020) 1715-1720.
- [2] H. Yue, X. Zhou, M. Cheng, D. Xing, Graphene oxide-mediated Cas9/sgRNA delivery for efficient genome editing, *Nanoscale* 10 (2018) 1063-1071.
- [3] W. Zhou, H. Cui, L. Ying, Enhanced Cytosolic Delivery and Release of CRISPR/Cas9 by Black Phosphorus Nanosheets for Genome Editing, *Angew Chem Int Edit* 57 (2018) 10268-10272.
- [4] S. Li, Z. Song, Biomimetic Mineralization-Based CRISPR/Cas9 Ribonucleoprotein Nanoparticles for Gene Editing, *ACS Appl Mater Interfaces* 11 (2019) 47762-47770.
- [5] E. Ju, T. Li, S. Ramos da Silva, S.J. Gao, Gold Nanocluster-Mediated Efficient Delivery of Cas9 Protein through pH-Induced Assembly-Disassembly for Inactivation of Virus Oncogenes, *ACS Appl Mater Interfaces* 11 (2019) 34717-34724.
- [6] W. Sun, J. Wang, CRISPR-Cas12a delivery by DNA-mediated bioresponsive editing for cholesterol regulation, *Sci Adv* 6 (2020) eaba2983.
- [7] J. Qiao, W. Sun, S. Lin, R. Jin, L. Ma, Y. Liu, Cytosolic delivery of CRISPR/Cas9 ribonucleoproteins for genome editing using chitosan-coated red fluorescent protein, *Chem Commun* 55 (2019) 4707-4710.
- [8] Y. Rui, D.R. Wilson, Carboxylated branched poly(β -amino ester) nanoparticles enable robust cytosolic protein delivery and CRISPR-Cas9 gene editing, *Sci Adv* 5 (2019) eaay3255.
- [9] M. Wang, J.A. Zuris, F. Meng, H. Rees, S. Sun, P. Deng, Y. Han, X. Gao, D. pouli, Q. Wu, I. Georgakoudi, D.R. Liu, Q. Xu, Efficient delivery of genome-editing proteins using bioreducible lipid nanoparticles, *Proc Natl Acad Sci U S A* 113 (2016) 2868-2873.

- [10] P. Gee, M.S.Y. Lung, Extracellular nanovesicles for packaging of CRISPR-Cas9 protein and sgRNA to induce therapeutic exon skipping, *Nat Commun* 11 (2020) 1334.