## **Supplementary information**



Fig. S1. Immunostaining of CD4<sup>+</sup> and CD8<sup>+</sup> cells in human pancreatic tumor tissues. Tumor tissues were processed for immunostaining of CD4 and CD8 staining. Representative images are shown of three tumors from untreated PDAC patients or treated with gemcitabine, as shown in Fig. 3I. Scale bars: 50  $\mu$ m. Quantitative analyses are shown in Fig. 3H.



Fig. S2. Immunostaining of FoxP3<sup>+</sup> T cells in mouse pancreatic tumor tissues. PDAC tumor-bearing C57BL/6 mice were treated as described in Fig. 1E. Tumor tissues were processed for immunostaining of FoxP3 staining. Representative images are shown. Scale bars: 50  $\mu$ m. Quantitative analyses are shown in Fig. 4G.





Fig. S3. Immunostaining of CD8<sup>+</sup> T cells in mouse colon CT26 tumor tissues. CT26 tumor-bearing BALB/c mice were treated as described in Fig. 2. Tumors were harvested at end-points and were processed for immunostaining of CD8<sup>+</sup> T cells. Representative images are shown. Scale bars: 50  $\mu$ m. Quantitative analyses are shown in Fig. 4H.

## Mouse CT26 tumors: CD8 positive cells



Fig. S4. Cisplatin induces the expression of CXCL11 in a cGAS-dependent manner. (A) Mouse pancreatic cancer cells were incubated with gemcitabine (Gem) or cisplatin (Cis) for 48 h (as shown in Fig. 7A-C). The expression of CXCL11 was measured by qRT-PCR. (B) Mouse pancreatic wild-type (WT) or the sgRNA-mediated cGAS-disrupted cells (cGAS-sgRNA) were transfected with dsDNA fragments for 48 h (as shown in Fig. 7E-G). The expression of CXCL11 was measured by qRT-PCR. (C) Mouse pancreatic wild-type (WT) or cGAS-disrupted cells were incubated with cisplatin for 48 h (as shown in Fig. 7H-J). The expression of CXCL11 was measured by qRT-PCR. (D) Wild-type mouse pancreatic cancer cells were incubated with gemcitabine. Eight hours post-treatment, cells were transfected with dsDNA fragments for 40 h (as shown in Fig. 7K-M). The expression of CXCL11 was measured by qRT-PCR. Statistical analysis: data are means  $\pm$  SEM of three separate experiments; one-way ANOVA followed by Tukey *post hoc* test for A-D. \*\*, P < 0.01; \*\*\*, P < 0.001.

Quantitative RT-PCR					
Genes	Forward primer	Reverse primer			
Mouse cxcl9	CTTTTCCTTTTGGGCATCATCT	GCAGGAGCATCGTGCATTC			
Mouse cxcl10	GAATCCGGAATCTAAGACCATCAA	GTGCGTGGCTTCACTCCAGT			
Mouse cxcl11	TGCTCAAGGCTTCCTTATGTT	CTTTGTCGCAGCCGTTACTC			
Mouse ifnb1	GGAGCAGCTGAATGGAAAGA	CATCTCTTGGATGGCAAAGG			
Mouse ifng	GAACTGGCAAAAGGATGGTG	GCTGATGGCCTGATTGTCTT			
Mouse prf1b	TCTTCGGGAACCAAGCTACA	CTTCGGGTTCTGTTCTTCCA			
Mouse gzmb	GTGTGCTATGTGGCTGGTTG	CACACTCCCGATCCTTCTGT			
Mouse tbx21	AGCAAGGACGGCGAATGTT	GGGTGGACATATAAGCGGTTC			
Mouse ifngr1	CTGGCAGGATGATTCTGCTGG	GCATACGACAGGGTTCAAGTTAT			
Mouse gata3	CTCGGCCATTCGTACATGGAA	GGATACCTCTGCACCGTAGC			
Mouse il4	GGTCTCAACCCCCAGCTAGT	GCCGATGATCTCTCTCAAGTGAT			
Mouse foxp3	CCCATCCCCAGGAGTCTTG	ACCATGACTAGGGGGCACTGTA			
Mouse roryt	GACCCACACCTCACAAATTGA	AGTAGGCCACATTACACTGCT			
Mouse il9	ATCACGTGTCCGTCCTTTTC	GCTTTTCTGCCTTTGCATCT			
Mouse il22	CAGACAGGTTCCAGCCCTAC	CGCCTTGATCTCTCCACTCT			
Mouse il17a	TTTAACTCCCTTGGCGCAAAA	CTTTCCCTCCGCATTGACAC			
Mouse il23a	CATGCTAGCCTGGAACGCACAT	ACTGGCTGTTGTCCTTGAGTCC			
Mouse rpl19	GAAGGTCAAAGGGAATGTGTTCA	CCTTGTCTGCCTTCAGCTTGT			
(reference)					
Human	GTGGCATTCAAGGAGTACCTC	TGATGGCCTTCGATTCTGGATT			
CXCL10					
Human	GGCAGATATTGAGAAAGCCTCC	GCCTTGCTTGCTTCGATTTG			
CXCL11					
Human EF1	GCTTCACTGCTCAGGTGAT	GCCGTGTGGCAATCCAAT			
(reference)					

Table S1.	Oligonucleotide see	nuences of primer	s used in aRT-P(	CR analyses and	transfection.
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## Transfection

	Forward primer	Reverse primer
dsDNA (60-bp	CCATCAGAAAGAGGTTTAATATTT	AAGTTTTATCTCTTTTCTCTCTCGAT
fragment, HSV-	TGTGAGACCATCGAAGAGAGAAAAG	GGTCTCACAAAAATATTAAACCTCT
1 sequence)	AGATAAAACTT	TTCTGATGG