

## Supplemental Information

### Linking the *Salmonella enterica* 1,2-propanediol utilization bacterial microcompartment shell to the enzymatic core via the shell protein PduB

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#### Section A: Details of kinetic model of the 1,2-propanediol utilization (Pdu) pathway

The kinetic model for the 1,2-propanediol utilization (Pdu) pathway here was developed based on the original work of Jakobson et al. [1], with modifications described herein. Specifically, we assume in this model that the cytosol is well-mixed, such that we are working with a compartmental model. We also incorporate the effects of cell growth over time, and explicitly model the metabolite concentrations in the external media. This required several adjustments to the original model, which are outlined in detail below. We focus our analysis here on metabolite profiles over time in a batch reactor setting instead of at steady-state to best study the conditions under which we ran our growth curves.

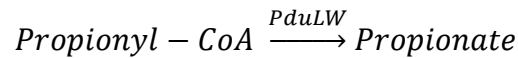
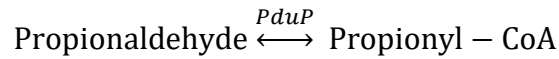
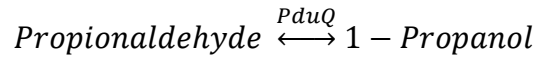
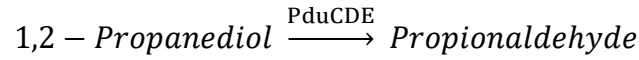
##### *A.1: Model assumptions*

We assume the following in our model:

1. At time  $t$ , there are  $N(t)$  identical, non-interacting cells in a well-mixed solution.
2. The substrates 1,2-propanediol, propionaldehyde, propionyl-CoA, propionate, and 1-propanol passively diffuse across the cell membrane at rates specified by permeability parameters.
3. The substrates 1,2-propanediol, propionaldehyde, propionyl-CoA, propionate, and 1-propanol passively diffuse across the microcompartment (MCP) shell or polar body surface at rates specified by permeability parameters.
4. There are  $n_{\text{MCP}}$  non-interacting MCPs or polar bodies in the cytosol of each cell.
5. Reactions catalyzed by PduCDE, PduP, and PduQ, forward or reverse, can only occur in the MCP or polar body.
6. Reactions catalyzed by PduL/W can only occur in the cytosol.
7. The external media, cytosol of the cell, and internal MCP volume are well-mixed such that the concentration in each compartment is assumed uniform at any given point in time.
8. The volume of the external media up to the leading order is the volume of the entire culture.
9. The volume of the cytosol up to the leading order is the volume of the cell.
10. All enzymes behave according to Michaelis-Menten kinetics.

## A.2: Chemical reactions

The following reactions are considered in our model, with the assumptions described above:



## A.3: Equations used in mathematical model

The differential equations described below were integrated forward in time from a starting condition of 55 mM 1,2-propanediol in the external media, the starting condition used in our growth curve experiments.

### A.3.1: Variable definition

Concentrations of all substrates are defined by the following variables:

- $P_i$ : 1,2-propanediol concentration in volume  $i$  (MCP/polar body, cytosol, or external media)
- $A_i$ : Propionaldehyde concentration in volume  $i$  (MCP/polar body, cytosol, or external media)
- $Pol_i$ : 1-Propanol concentration in volume  $i$  (MCP/polar body, cytosol, or external media)
- $PCoA_i$ : Propionyl-CoA concentration in volume  $i$  (MCP/polar body, cytosol, or external media)
- $Pate_i$ : Propionate concentration in volume  $i$  (MCP/polar body, cytosol, or external media)

Other constraints and variables used in the model are defined as follows:

- $V_{\text{compartment}}$ : Volume of the compartment (MCP or polar body)
- $SA_{\text{compartment}}$ : Surface area of the compartment (MCP or polar body)
- $V_{\text{cell}}$ : Volume of the cell
- $SA_{\text{cell}}$ : Surface area of the cell
- $Perm_i^j$ : Permeability of substrate  $i$  at interface  $j$  (cell surface or MCP/polar body surface)
- $R_i(X_j)$ : Reaction rate of enzyme  $i$  as a function of concentrations  $X$  in a given volume  $j$  (MCP/polar body, cytosol, or external media)
- $K_M^{i,j}$ : Michaelis constant of enzyme  $i$  for substrate  $j$

- $V_{\max}^{i,j}$ : Maximum reaction velocity of enzyme  $i$  for substrate  $j$
- $N(t)$ : Number of cells at time  $t$ , calculated from the experimental growth profile
- PB: Refers to polar body
- MCP: Refers to MCP
- $n_{\text{MCP/PB}}$ : Number of MCPs/polar bodies per cell

### A.3.2: Differential equations: MCP or polar body

The differential equations for the MCP or polar body volume were as follows:

$$\frac{dP_{\text{MCP/PB}}}{dt} = -R_{\text{PduCDE}}(X_{\text{MCP/PB}}) + \frac{\text{Perm}_{\text{MCP/PB}}^{\text{P}} SA_{\text{MCP/PB}}}{\text{Vol}_{\text{MCP/PB}}} (P_{\text{cytosol}} - P_{\text{MCP/PB}})$$

$$\begin{aligned} \frac{dA_{\text{MCP/PB}}}{dt} = & R_{\text{PduCDE}}(X_{\text{MCP/PB}}) - R_{\text{PduP},f}(X_{\text{MCP/PB}}) + R_{\text{PduP},r}(X_{\text{MCP/PB}}) - R_{\text{PduQ},f}(X_{\text{MCP/PB}}) \\ & + R_{\text{PduQ},r}(X_{\text{MCP/PB}}) + \frac{\text{Perm}_{\text{MCP/PB}}^{\text{A}} SA_{\text{MCP/PB}}}{\text{Vol}_{\text{MCP/PB}}} (A_{\text{cytosol}} - A_{\text{MCP/PB}}) \end{aligned}$$

$$\begin{aligned} \frac{dPol_{\text{MCP/PB}}}{dt} = & R_{\text{PduQ},f}(X_{\text{MCP/PB}}) - R_{\text{PduQ},r}(X_{\text{MCP/PB}}) \\ & + \frac{\text{Perm}_{\text{MCP/PB}}^{\text{Pol}} SA_{\text{MCP/PB}}}{\text{Vol}_{\text{MCP/PB}}} (Pol_{\text{cytosol}} - Pol_{\text{MCP/PB}}) \end{aligned}$$

$$\begin{aligned} \frac{dPCoA_{\text{MCP/PB}}}{dt} = & R_{\text{PduP},f}(X_{\text{MCP/PB}}) - R_{\text{PduP},r}(X_{\text{MCP/PB}}) \\ & + \frac{\text{Perm}_{\text{MCP/PB}}^{\text{PCoA}} SA_{\text{MCP/PB}}}{\text{Vol}_{\text{MCP/PB}}} (PCoA_{\text{cytosol}} - PCoA_{\text{MCP/PB}}) \end{aligned}$$

$$\frac{dPate_{\text{MCP/PB}}}{dt} = \frac{\text{Perm}_{\text{MCP/PB}}^{\text{Pate}} SA_{\text{MCP/PB}}}{\text{Vol}_{\text{MCP/PB}}} (Pate_{\text{cytosol}} - Pate_{\text{MCP/PB}})$$

### A.3.3: Differential equations: cytosol

The differential equations for the cytosol of the cell were as follows:

$$\begin{aligned} \frac{dP_{\text{cytosol}}}{dt} = & \frac{n_{\text{MCP}} \text{Perm}_{\text{MCP/PB}}^{\text{P}} SA_{\text{MCP/PB}}}{\text{Vol}_{\text{cell}}} (P_{\text{MCP/PB}} - P_{\text{cytosol}}) \\ & + \frac{\text{Perm}_{\text{cell}}^{\text{P}} SA_{\text{cell}}}{\text{Vol}_{\text{cell}}} (P_{\text{external}} - P_{\text{cytosol}}) \end{aligned}$$

$$\frac{dA_{\text{cytosol}}}{dt} = \frac{n_{\text{MCP}} \text{Perm}_{\text{MCP/PB}}^{\text{A}} \text{SA}_{\text{MCP/PB}}}{\text{Vol}_{\text{cell}}} (A_{\text{MCP/PB}} - A_{\text{cytosol}}) + \frac{\text{Perm}_{\text{cell}}^{\text{A}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{cell}}} (A_{\text{external}} - A_{\text{cytosol}})$$

$$\frac{dPol_{\text{cytosol}}}{dt} = \frac{n_{\text{MCP}} \text{Perm}_{\text{MCP/PB}}^{\text{Pol}} \text{SA}_{\text{MCP/PB}}}{\text{Vol}_{\text{cell}}} (Pol_{\text{MCP/PB}} - Pol_{\text{cytosol}}) + \frac{\text{Perm}_{\text{cell}}^{\text{Pol}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{cell}}} (Pol_{\text{external}} - Pol_{\text{cytosol}})$$

$$\frac{dPCoA_{\text{cytosol}}}{dt} = -R_{\text{PduLW}}(X_{\text{cytosol}}) + \frac{n_{\text{MCP}} \text{Perm}_{\text{MCP/PB}}^{\text{PCoA}} \text{SA}_{\text{MCP/PB}}}{\text{Vol}_{\text{cell}}} (PCoA_{\text{MCP/PB}} - PCoA_{\text{cytosol}}) + \frac{\text{Perm}_{\text{cell}}^{\text{PCoA}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{cell}}} (PCoA_{\text{external}} - PCoA_{\text{cytosol}})$$

$$\frac{dPate_{\text{cytosol}}}{dt} = R_{\text{PduLW}}(X_{\text{cytosol}}) + \frac{n_{\text{MCP}} \text{Perm}_{\text{MCP/PB}}^{\text{Pate}} \text{SA}_{\text{MCP/PB}}}{\text{Vol}_{\text{cell}}} (Pate_{\text{MCP/PB}} - Pate_{\text{cytosol}}) + \frac{\text{Perm}_{\text{cell}}^{\text{Pate}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{cell}}} (Pate_{\text{external}} - Pate_{\text{cytosol}})$$

#### A.3.4: Differential equations: external media

The differential equations for the external media of the cell were as follows:

$$\frac{dP_{\text{external}}}{dt} = N(t) \frac{\text{Perm}_{\text{cell}}^{\text{P}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{external}}} (P_{\text{cytosol}} - P_{\text{external}})$$

$$\frac{dA_{\text{external}}}{dt} = N(t) \frac{\text{Perm}_{\text{cell}}^{\text{A}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{external}}} (A_{\text{cytosol}} - A_{\text{external}})$$

$$\frac{dPol_{\text{external}}}{dt} = N(t) \frac{\text{Perm}_{\text{cell}}^{\text{Pol}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{external}}} (Pol_{\text{cytosol}} - Pol_{\text{external}})$$

$$\frac{dPCoA_{\text{external}}}{dt} = N(t) \frac{\text{Perm}_{\text{cell}}^{\text{PCoA}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{external}}} (PCoA_{\text{cytosol}} - PCoA_{\text{external}})$$

$$\frac{dPate_{\text{external}}}{dt} = N(t) \frac{\text{Perm}_{\text{cell}}^{\text{Pate}} \text{SA}_{\text{cell}}}{\text{Vol}_{\text{external}}} (Pate_{\text{cytosol}} - Pate_{\text{external}})$$

### A.3.5: Definition of reaction rate

Reaction rates were assumed to follow Michaelis-Menten kinetics.

$$R_{PduCDE} = V_{\max}^{PduCDE} \frac{P_{MCP/PB}}{K_M^{PduCDE} + P_{MCP/PB}}$$

$$R_{PduP,f} = V_{\max}^{PduP,f} \frac{A_{MCP/PB}}{K_M^{PduP,f} + A_{MCP/PB}}$$

$$R_{PduP,r} = V_{\max}^{PduP,r} \frac{PCoA_{MCP/PB}}{K_M^{PduP,r} + PCoA_{MCP/PB}}$$

$$R_{PduQ,f} = V_{\max}^{PduQ,f} \frac{A_{MCP/PB}}{K_M^{PduQ,f} + A_{MCP/PB}}$$

$$R_{PduQ,r} = V_{\max}^{PduQ,r} \frac{Pol_{MCP/PB}}{K_M^{PduQ,r} + Pol_{MCP/PB}}$$

$$R_{PduLW} = V_{\max}^{PduLW} \frac{PCoA_{\text{cytosol}}}{K_M^{PduLW} + PCoA_{\text{cytosol}}}$$

## Section B: Bioinformatic analysis of PduB homologs

The shell protein PduB from *Salmonella enterica* serovar Typhimurium LT2 is a circularly permuted trimeric shell protein that is critical for linking the enzymatic core of the 1,2-propanediol utilization (Pdu) microcompartment (MCP) to its shell. As described in detail in Lehman *et al.*, this is likely mediated by an N-terminal extension that is hypothesized to interact with the enzymatic core [2]. To determine if this putative mechanism for enzyme encapsulation is broadly distributed across MCP types, we performed bioinformatic analyses on PduB homologs. These analyses are based on the in-depth studies published by Sutter *et al.* and Axen *et al.* [3, 4].

A list of 45 diverse PduB homologs was curated based on the comprehensive list available in Sutter *et al.* A representative from each of the subclades (designated based on the color palette available at <https://xkcd.com/color/rgb/>) of PduB homologs (including all BMC-T<sup>P</sup> variants) was chosen from the full data set. The full set of sequences used for this analysis is available in **Table S5**. Of the 46 BMC-T<sup>P</sup> homolog sequences analyzed, 20 contained N-terminal extensions (43%) (**Fig S4, Supplemental Alignment 1**). Within close PduB homologs (based on the distinction made in Sutter *et al.*), the N-terminal extension of PduB is not well conserved relative to other highly conserved motifs within PduB. The most highly conserved positions within the N-terminal extension are a leucine at position 6 (73% identical) and a valine at position 14 (77% identical). After position 19 conservation scores drop off rapidly, with no positions between 19 and 37 having greater than 30% conservation. By comparison, the conserved lysine residues at position 102 and 207 are 100% conserved among the PduB homologs analyzed. This implies that while this mechanism may be important for the close PduB homologs present in Pdu and GRM MCPs, other encapsulation mechanisms are likely present in Eut and SPU MCPs.

## Section C: Supplemental Tables

**Table S1.** Strains used in this study.

| Strain         | Organism                                   | Genotype  |
|----------------|--|---|
| DTE003         | <i>S. enterica</i> serovar Typhimurium LT2 | Wild type   |
| CMJS256        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pocR   |
| TUC01 [5], [6] | <i>E. coli</i> W3110                       | gal490 pgl $\Delta$ 8 $\lambda$ cl857<br>$\Delta$ (cro-bioA)<br>int<>cat/sacB |
| NWks083        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduA $\Delta$ pduJ   |
| CEMS179        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduB   |
| NWks365        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduB::pduB-K102A-K207A   |
| NWks337        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduA::pduA-mCherry   |
| NWks338        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduA::pduA-mCherry $\Delta$ pduB                                     |
| NWks344        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduA::pduA-mCherry $\Delta$ pocR                                     |
| NWks364        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduA::pduA-mCherry<br>$\Delta$ pduB::pduB-K102A-K207A                |
| TMDS025        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduD::ssD-GFPmut2  |
| CEMS299        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pocR $\Delta$ pduD::ssD-GFPmut2                                      |
| CEMS298        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduA $\Delta$ pduD::ssD-GFPmut2 $\Delta$ pduJ                        |
| CEMS218        | <i>S. enterica</i> serovar Typhimurium LT2 | $\Delta$ pduB $\Delta$ pduD::ssD-GFPmut2                                      |

CEMS296

*S. enterica* serovar Typhimurium LT2

$\Delta pduB::pduB-K102A-$   
 $K207A \Delta pduD::ssD-$   
 $GFPmut2$

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**Table S2.** Plasmids used in this study.

| <b>Name</b> | <b>Plasmid</b>                          | <b>Origin</b>                   | <b>Resistance</b> |
|-------------|---|---------------------------------|-------------------|
| CMJ069      | pBAD33t-ssD-GFPmut2                     | p15A                            | Chloramphenicol   |
| EYK208      | pBAD33t-ssP-GFPmut2                     | p15A                            | Chloramphenicol   |
| EYK193      | pBAD33t-PduG-GFPmut2                    | p15A                            | Chloramphenicol   |
| NWKp041     | pBAD33t-PduO-GFPmut2                    | p15A                            | Chloramphenicol   |
| NWKp042     | pBAD33t-PduW-GFPmut2                    | p15A                            | Chloramphenicol   |
| NWKp043     | pBAD33t-ssL-GFPmut2                     | p15A                            | Chloramphenicol   |
| CMJ140      | pBAD33t-PduB-FLAG                       | p15A                            | Chloramphenicol   |
| NWKp044     | pBAD33t-PduB-K102A-K207A                | p15A                            | Chloramphenicol   |
| pSIM6 [6]   | $\lambda$ Red system repressed by cl857 | pSC101 <i>repA<sup>ts</sup></i> | Ampicillin        |

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**Table S3.** Primers used in this study.

| <b>Name</b> | <b>Purpose</b>         | <b>Description</b>  | <b>Sequence*</b>  |
|-------------|------------------------|---|---|
| NWK<br>o493 | Recombineering         | Amplify <i>cat/sacB</i> with<br>homology upstream<br>of <i>pduB</i> For                   | gccctcacaccgatgtagaaaaatcttaccg<br>aaggaattagccaatgaTGTGACGGA<br>AGATCACTTCG                                    |
| NWK<br>o494 | Recombineering         | Amplify <i>cat/sacB</i> with<br>homology<br>downstream of <i>pduB</i><br>Rev              | ttcgccagtgcttcaaacttttcgatctcatgaa<br>tcagcctcgtgggtaATCAAAGGGAAA<br>ACTGTCCATAT                                |
| NWK<br>o517 | Recombineering         | Amplify <i>pduB-K102A-<br/>K207A</i> with<br>homology upstream<br>of <i>pduB</i> For      | ccacgccctcacaccgatgtagaaaaatctt<br>accgaaggaattagccaATGAGCAG<br>CAATGAGCTGG                                     |
| NWK<br>o518 | Recombineering         | Amplify <i>pduB-K102A-<br/>K207A</i> with<br>homology<br>downstream of <i>pduB</i><br>Rev | gccagtgcttcaaacttttcgatctcatgaatc<br>agcctcgtgggatcaGATGTAGGACG<br>GACGATCG                                     |
| NWK<br>o499 | Recombineering         | Knockout <i>pduB</i>  | cgcgtccgcacagcgatgttgaggccatttac<br>cgaaatcagcctaatcgggcaagcgagggtg<br>aagcgtaatggataaagagctctgcaatcaa<br>cggtc |
| NWK<br>o466 | Sequencing             | Amplify from<br>upstream of <i>pduA</i>   | CTGCGAACCTGTCTCC  |
| NWK<br>o467 | Sequencing             | Amplify from<br>downstream of <i>pduA</i>   | CGTCTCTCGTATAGGTTGG   |
| NWK<br>o501 | Golden Gate<br>cloning | Amplify <i>pduO</i> with<br>GG overhang For   | AttGGTCTCACATGGCGATTTAT<br>ACCCGAAC   |
| NWK<br>o504 | Golden Gate<br>cloning | Amplify <i>pduO</i> with<br>GG overhang and<br>GS linker Rev                              | ATTGGTCTCAGCTGCCttgatgagtt<br>cccacgtaatag  |
| NWK<br>o502 | Golden Gate<br>cloning | Amplify <i>pduW</i> with<br>GG overhang For   | AttGGTCTCACATGTCTTACAAAA<br>TAATGGCCATTA  |

|             |                               |   |  |
|-------------|-------------------------------|---|--|
| NWK<br>o505 | Golden Gate<br>cloning        | Amplify <i>pduW</i> with<br>GG overhang and<br>GS linker Rev    | ATTGGTCTCAGCTGCCggctggtac<br>acaaagcc                          |
| EYKP<br>263 | Sacl cloning                  | Amplify <i>pduG</i> with<br>Sacl recognition For                | TATTGAGCTCTTAAAGAGGAGA<br>AAGGTCatgcatatatagctggcattgac<br>atc |
| EYKP<br>264 | Xbal cloning                  | Amplify <i>pduG</i> with<br>Xbal recognition Rev                | TATTTCTAGActgtccatgcgcaaactcc<br>ttatg                         |
| NWK<br>o506 | Golden Gate<br>cloning        | Amplify <i>GFPmut2</i><br>with GG overhang<br>and GS linker For | AttGGTCTCACAGCAgtaaaggagaa<br>gaacttttactgg                    |
| NWK<br>o507 | Golden Gate<br>cloning        | Amplify <i>GFPmut2</i><br>with GG overhang<br>Rev               | ATTGGTCTCATTTAttgtatagttcatc<br>catgccatgtg                    |
| TMD<br>P058 | Golden Gate<br>cloning        | Amplify <i>ssL-<br/>GFPmut2</i> with GG<br>overhang For         | ATTGGTCTCACATGgataaagagctt<br>ctgcaatcaac                      |
| NWK<br>o514 | QuikChange                    | <i>PduB-K102A</i> For   | ATGGCGGCGGACGAAGCGGTG<br>GCGGCCACCAACACC                       |
| NWK<br>o515 | QuikChange                    | <i>PduB-K102A</i> Rev   | CTGCTTCGCCACCGCCGGTGG<br>TTGTGGCTTCACCAG                       |
| NWK<br>o516 | QuikChange                    | <i>PduB-K207A</i> For   | ATACCGCGCTGGCGTCAGCCA<br>ACGTTGAAGTCG                          |
| NWK<br>o517 | QuikChange                    | <i>PduB-K207A</i> Rev   | ACTACCGGCTATGGCGCGACC<br>GCAGTCGGTTGC                          |
| MPV0<br>31  | Sequencing                    | Sequence<br>confirmation of <i>pduB</i><br>locus For            | gccctcacaccgatgtagaaaaaatc                                     |
| MPV0<br>30  | Sequencing                    | Sequence<br>confirmation of <i>pduB</i><br>locus Rev            | tccatcgccatcacttcttcg  |
| TMD<br>P021 | Sequencing,<br>recombineering | Amplify from<br>upstream of <i>pduD</i><br>locus For            | ggcaacaggttatcgctctgc  |

|             |                               |  |  |
|-------------|-------------------------------|--|--|
| TMD<br>P022 | Sequencing,<br>recombineering | Amplify from<br>downstream of <i>pduD</i><br>locus Rev                       | ccctgcaggctgttcatgc  |
| TMD<br>P072 | Sequencing                    | Amplify from<br>upstream of <i>pduD</i><br>locus For                         | catccagaaagccaagctaacc   |
| TMD<br>P073 | Sequencing                    | Amplify from<br>downstream of <i>pduD</i><br>locus Rev                       | cgtccagcgttttattggtgg  |
| TMD<br>P016 | Recombineering                | Amplify ssD-<br>GFPmut2 with<br>homology upstream<br>of <i>pduD</i> For      | TTGATCCCAACGAGATTGATTA<br>AGGGGTGAGAAatggaaattaatgaa<br>aaattgctgcgc |
| TMD<br>P019 | Recombineering                | Amplify ssD-<br>GFPmut2 with<br>homology<br>downstream of <i>pduD</i><br>Rev | ATTGCGTCGGTATTCATGGAGT<br>TATCCTTTAttattgtatagttcatccatgc<br>catgtg  |

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**Table S4.** Parameter values used in kinetic model of pathway performance in MCPs and polar bodies.

| Parameter Name           | MCP         | Polar Body  | Unit                        |
|--------------------------|-------------|-------------|-----------------------------|
| CDE_con                  | 0.462467024 | 0.462467024 | mM                          |
| CDE_tot [7]              | 6000        | 6000        | enzymes/cell                |
| cell_length              | 2.47E-06    | 2.47E-06    | m                           |
| cell_radius              | 3.75E-07    | 3.75E-07    | m                           |
| cell_surface_area        | 5.82E-12    | 5.82E-12    | m <sup>2</sup>              |
| cell_volume              | 9.81E-19    | 9.81E-19    | m <sup>3</sup>              |
| external_volume          | 5.00E-05    | 5.00E-05    | m <sup>3</sup>              |
| kcatCDE [8]              | 300         | 300         | 1/s                         |
| kcatL                    | 100         | 100         | 1/s                         |
| kcatPf [9]               | 55          | 55          | 1/s                         |
| kcatPr [9]               | 6           | 6           | 1/s                         |
| kcatQf [10]              | 55          | 55          | 1/s                         |
| kcatQr [10]              | 6           | 6           | 1/s                         |
| L_con [11]               | 0.1         | 0.1         | mM                          |
| mcp_surface_area         | 6.16E-14    | 3.75E-13    | m <sup>2</sup> /compartment |
| mcp_volume               | 1.44E-21    | 2.16E-20    | m <sup>3</sup> /compartment |
| Navogadro                | 6.02E+23    | 6.02E+23    | molecules/mole              |
| nmcp                     | 15          | 1           | MCPs/polar bodies per cell  |
| P_con                    | 0.693700536 | 0.693700536 | mM                          |
| P_tot [7]                | 9000        | 9000        | enzymes/cell                |
| PermMCPNonPolar          | 3.98E-08    | 1.00E+01    | m/s                         |
| PermMCPPolar             | 3.98E-08    | 1.00E+01    | m/s                         |
| Q_con                    | 0.520275402 | 0.520275402 | mM                          |
| Q_tot [7]                | 6750        | 6750        | enzymes/cell                |
| radius_mcp [12]          | 7.00E-08    | 1.73E-07    | m                           |
| VmaxCDEf                 | 138.7401071 | 138.7401071 | mM/s                        |
| VmaxLf                   | 10          | 10          | mM/s                        |
| VmaxPf                   | 38.15352946 | 38.15352946 | mM/s                        |
| VmaxPr                   | 4.162203213 | 4.162203213 | mM/s                        |
| VmaxQf                   | 28.61514709 | 28.61514709 | mM/s                        |
| VmaxQr                   | 3.12165241  | 3.12165241  | mM/s                        |
| KmCDEPropanediol [8]     | 0.5         | 0.5         | mM                          |
| KmPfPropionaldehyde [9]  | 15          | 15          | mM                          |
| KmPrPropionyl [9]        | 95          | 95          | mM                          |
| KmQfPropionaldehyde [10] | 15          | 15          | mM                          |
| KmQrPropanol [10]        | 95          | 95          | mM                          |
| KmLPropionyl             | 20          | 20          | mM                          |

|  |          |          |     |
|--|----------|----------|-----|
| PermCellPropanediol [13] [14] [15]     | 1.00E-04 | 1.00E-04 | m/s |
| PermCellPropionaldehyde [13] [14] [15] | 1.00E-02 | 1.00E-02 | m/s |
| PermCellPropanol [13] [14] [15]        | 1.00E-04 | 1.00E-04 | m/s |
| PermCellPropionyl [13] [14] [15]       | 1.00E-05 | 1.00E-05 | m/s |
| PermCellPropionate [13] [14] [15]      | 1.00E-07 | 1.00E-07 | m/s |

**Table S5.** Sequences used for PduB phylogenetic tree and multiple sequence alignment (information obtained from Sutter et al. [3]).

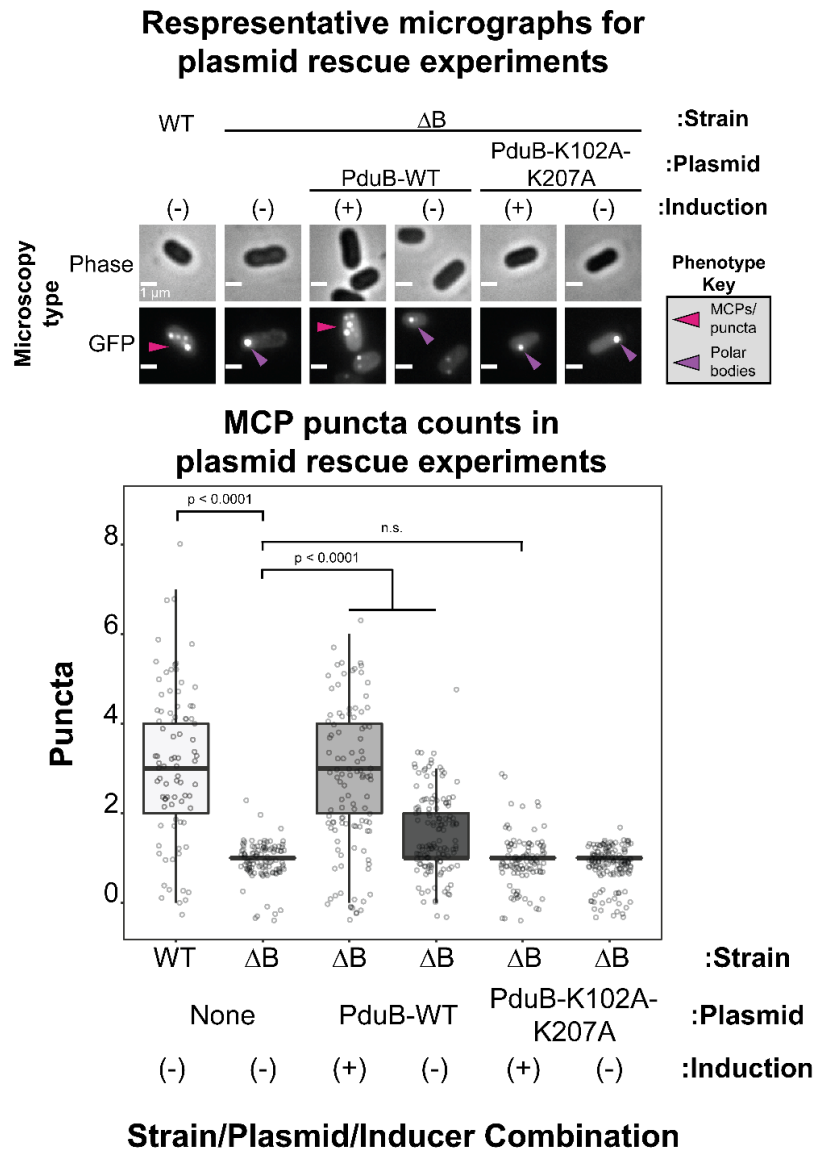
| Bacteria  | BMC Type                 | Length | Accession Number (Uniprot) | Subclade Color Designation |
|---|--------------------------|--------|----------------------------|----------------------------|
| Escherichia 9.0111 (Proteobacteria)               | bmcType=PDU1AB (main)    | 277aa  | [I2WHE2] pduB              | greenishBeige0             |
| Klebsiella PO552 (Proteobacteria)                 | bmcType=PDU1AB (main)    | 270aa  | [A0A206YP55] pduB          | greenishBeige1             |
| Citrobacter MGH105 (Proteobacteria)               | bmcType=GRM4 (main)      | 271aa  | [A0A0J1LFA2] pduB          | greenishBeige2             |
| Listeria ATCC 33091 (Firmicutes)                  | bmcType=PDU1E (main)     | 267aa  | [H1GCR5] pduB              | sunYellow1                 |
| Enterococcus ATCC 43197 (Firmicutes)              | bmcType=PDU1E (main)     | 269aa  | [R2PCN6] pduB              | sunYellow2                 |
| Lactobacillus DSM 20253 (Firmicutes)              | bmcType=PDU1C (main)     | 263aa  | [A0A0R2D124] pduB          | sunYellow3                 |
| Clostridium ATCC 25763 (ADT22) (Firmicutes)       | bmcType=MUF1 (main)      | 274aa  | [A0A0L9YAN3] pduB          | chartreuse1                |
| unclassified_Eubacteriaceae CHKC1004 (Firmicutes) | bmcType=PDU1D (sat)      | 274aa  | [A0A143XJ34] pduB          | chartreuse2                |
| Blautia MGS:257 (Firmicutes)                      | bmcType=PDU1D_GRM5 (sat) | 274aa  | [R5WST7] pduB              | chartreuse3                |
| Ruminococcus AM49-8 (Firmicutes)                  | bmcType=GRM5 (main)      | 242aa  | [A0A373QR46] pduB          | chartreuse4                |
| Propionibacterium F0372 (Actinobacteria)          | bmcType=PDU1D (main)     | 262aa  | [S3X7K1] pduB              | turquoiseBlue1             |
| Romboutsia FRIFI (Firmicutes)                     | bmcType=PDU1D (main)     | 270aa  | [A0A2P2BR70] pduB          | turquoiseBlue2             |
| Buttiauxella ATCC 51602 (Proteobacteria)          | bmcType=GRM3B (main)     | 261aa  | [A0A1B7IB09] pduB          | turquoiseBlue3a            |
| Azospirillum TSO35-2 (Proteobacteria)             | bmcType=GRM3C (main)     | 275aa  | [A0A2U1VFS1] pduB          | turquoiseBlue3b            |
| Quasibacillus SGZ-8 (QY95) (Firmicutes)           | bmcType=PDU1D (main)     | 260aa  | [A0A0F5HND3] pduB          | lichen1                    |
| Fusobacterium 1_1_41FAA (Fusobacteria)            | bmcType=PDU1F (main)     | 271aa  | [D6LIA1] pduB              | lichen2                    |
| Desulfofundulus DSM 16057 (Firmicutes)            | bmcType=PDU1D (main)     | 274aa  | [A0A1M6DW06] pduB          | darkGrassGreen             |
| Streptococcus NCTC13765 (Firmicutes)              | bmcType=GRM3A (main)     | 261aa  | [A0A380L0X7] pduB          | aqua1                      |
| Megasphaera 28L (Firmicutes)                      | bmcType=GRM3A (main)     | 270aa  | [D3LW76] pduB              | aqua2                      |
| Pelosinus JBW45 (Firmicutes)                      | bmcType=PDU1D (main)     | 265aa  | [I8TWY2] pduB              | pistachio1                 |
| Paenibacillus KCTC 13049 (Firmicutes)             | bmcType=GRM6 (main)      | 280aa  | [A0A2Z2KG56] pduB          | pistachio2                 |
| Morganella GCSL-Mp20 (AYY16) (Proteobacteria)     | bmcType=EUT1 (main)      | 218aa  | [A0A1B8H666] eutL          | rosa                       |
| Photobacterium Gung47 (Proteobacteria)            | bmcType=EUT1 (main)      | 218aa  | [A0A0C5WBE8] eutL          | darkLavender1              |
| Pseudoflavonifractor ATCC 29799 (Firmicutes)      | bmcType=EUT2G (main)     | 217aa  | [A6NPH4] eutL              | darkLavender2              |
| Cellulosilyticum ATCC 49066 (Firmicutes)          | bmcType=EUT2I (main)     | 217aa  | [F2JLY6] eutL              | darkPeach1                 |
| Brevibacillus LAK 1210 (EX87) (Firmicutes)        | bmcType=EUT2I (main)     | 217aa  | [A0A0F6Y0T9] eutL          | darkPeach2                 |
| Bacillus 2046851 (Firmicutes)                     | bmcType=EUT2I (main)     | 217aa  | [A0A268IB25] eutL          | darkPeach3                 |
| Lachnoclostridium An196 (Firmicutes)              | bmcType=EUT2B (main)     | 217aa  | [A0A1Y4IQA1] eutL          | mutedPink1                 |
| Dethiosulfovibrio peptidovorans (Synergistetes)   | bmcType=EUT2B (main)     | 217aa  | [A0A2G6M5H7] eutL          | mutedPink2                 |
| Hathewayia DSM 3090 (Firmicutes)                  | bmcType=EUT2I (main)     | 218aa  | [A0A1M6KC11] eutL          | scarlet1                   |
| Fusobacterium ATCC 23726 (Fusobacteria)           | bmcType=EUT2F (main)     | 221aa  | [D5RF36] eutL              | scarlet2                   |
| Halanaerobium sapolanicus (Firmicutes)            | bmcType=PDU_EUT (sat)    | 217aa  | [E4RJS6] eutL              | scarlet3                   |

|  |                      |       |                   |             |
|--|----------------------|-------|-------------------|-------------|
| Natronincola DSM 18979 (Firmicutes)        | bmcType=EUT2B (main) | 217aa | [A0A1I0EUU4] eutL | amethyst1   |
| Fusicatenibacter (ERS852537) (Firmicutes)  | bmcType=EUT2B (main) | 218aa | [A0A174T4Q4] eutL | amethyst2   |
| Vagococcus bH819 (Firmicutes)              | bmcType=EUT2A (main) | 218aa | [A0A1X6WPL8] eutL | amethyst3   |
| Tepidanaerobacter JL (Firmicutes)          | bmcType=EUT2B (main) | 218aa | [A0A0U9HIK8] eutL | ceriseX     |
| Sporomusa DSM 5090 (Firmicutes)            | bmcType=UA (sat)     | 215aa | [A0A1W2F4B5] eutL | cerise1     |
| Minicystis DSM 24000 (Proteobacteria)      | bmcType=SPU3 (sat)   | 226aa | [A0A1L6L3A8] eutL | cerise2     |
| Levilinea KIBI-1 (Chloroflexi)             | bmcType=SPU6 (sat)   | 216aa | [A0A0P6XV62] eutL | cerise3     |
| Alkalispirochaeta ASpG1 (Spirochaetes)     | bmcType=EUT3 (main)  | 218aa | [A0A1N6TZP1] eutL | cerise4     |
| Melghirimyces DSM 45787 (Firmicutes)       | bmcType=EUT2C (main) | 217aa | [A0A2T6C8Q0] eutL | orangePinkX |
| Brevibacillus AK1 (Firmicutes)             | bmcType=EUT2E (main) | 216aa | [M8D890] eutL     | orangePink1 |
| Fictibacillus ZFHKF-1 (Firmicutes)         | bmcType=EUT2C (main) | 217aa | [I8AM68] eutL     | orangePink2 |
| Salimicrobium WN024 (Firmicutes)           | bmcType=EUT2C (main) | 216aa | [A0A2A3A540] eutL | orangePink3 |
| Domibacillus DSM 4337 (AWH48) (Firmicutes) | bmcType=EUT2C (main) | 217aa | [A0A177KYD5] eutL | orangePink4 |

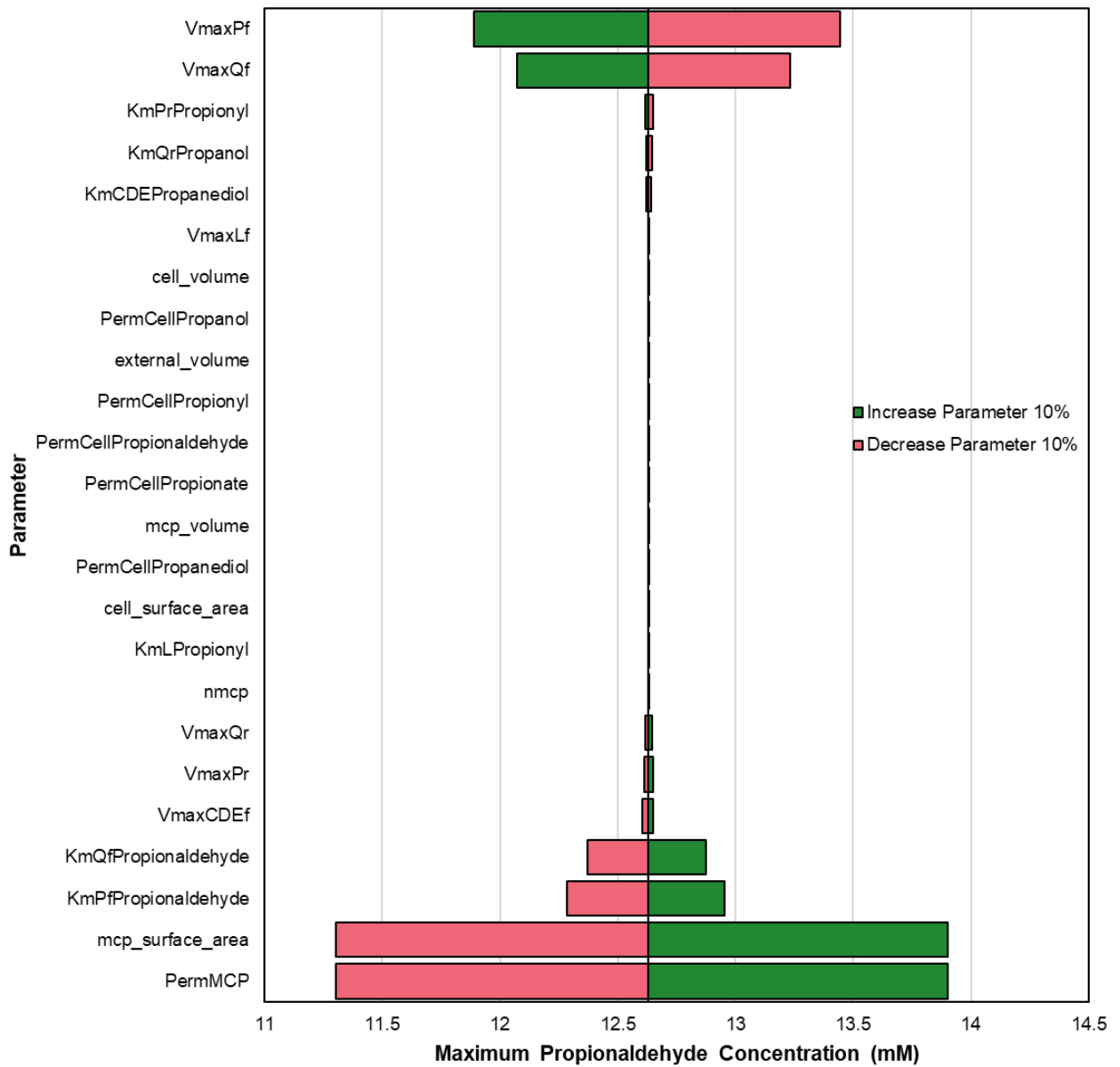


## Section D: Supplemental Figures

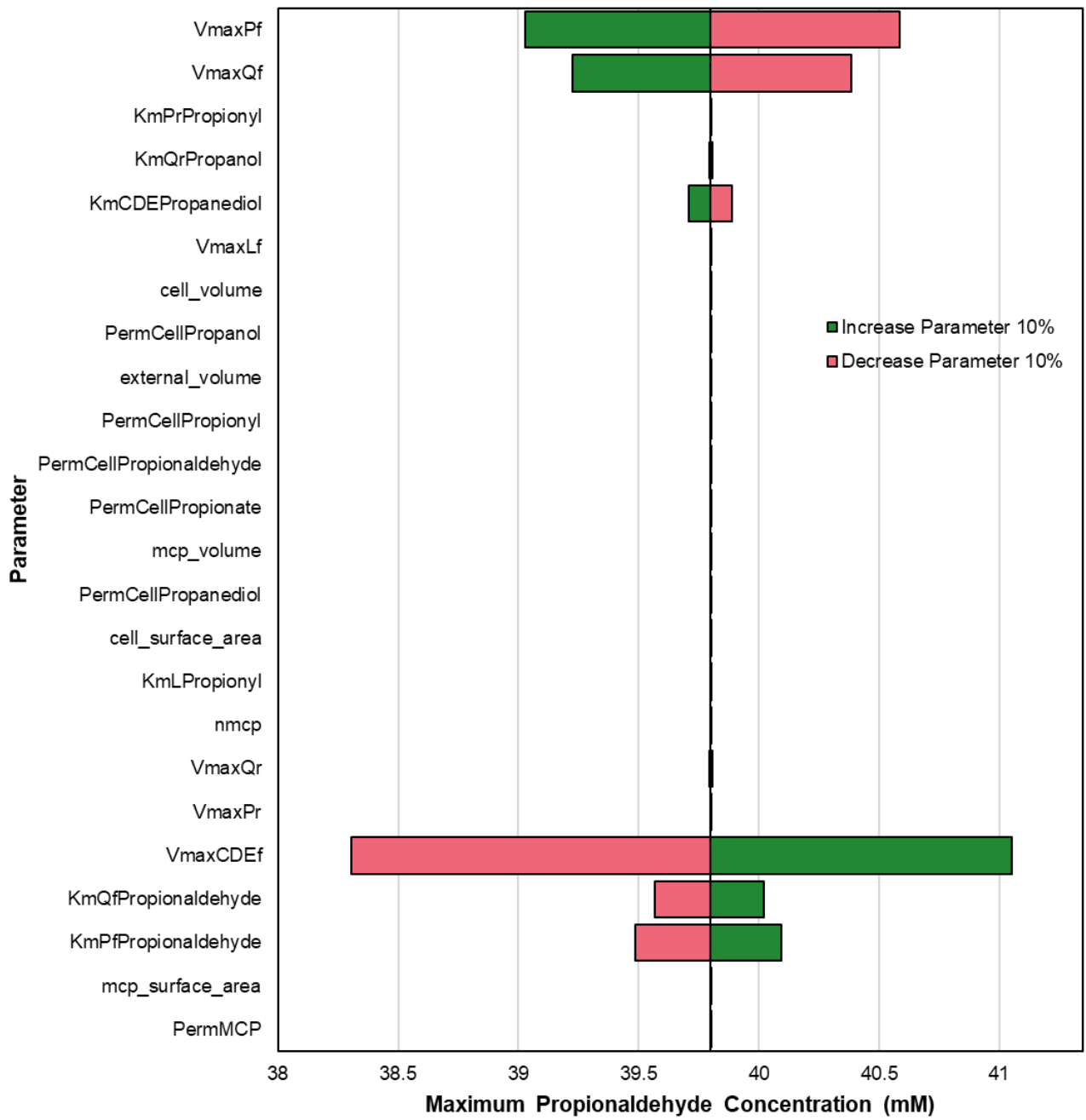
**Figure S1.** Representative micrographs (top) and puncta quantification (bottom) for plasmid rescue experiments. PduB-WT and PduB-K102A-K207A were expressed from a pBAD33t plasmid in the PduB knockout strain ( $\Delta B$ ) to rescue MCP formation, visualized by the formation of fluorescent puncta. The  $\Delta B$  strain contained significantly fewer puncta than the WT strain ( $p < 0.0001$ , one-tailed t-test).  $\Delta B$  with the PduB-WT plasmid had significantly more puncta than the  $\Delta B$  strain without a plasmid ( $p < 0.0001$ , one-tailed t-test).  $\Delta B$  with PduB-K102A-K207A overexpressed did not have significantly different puncta counts (one-tailed t-test). Note that all strains contain ssD-GFP at the *pduD* locus to allow visualization of cargo location within the cell. Puncta were counted from three biological replicates and at least 85 puncta were counted in each condition. Scale bar = 1  $\mu\text{m}$ .



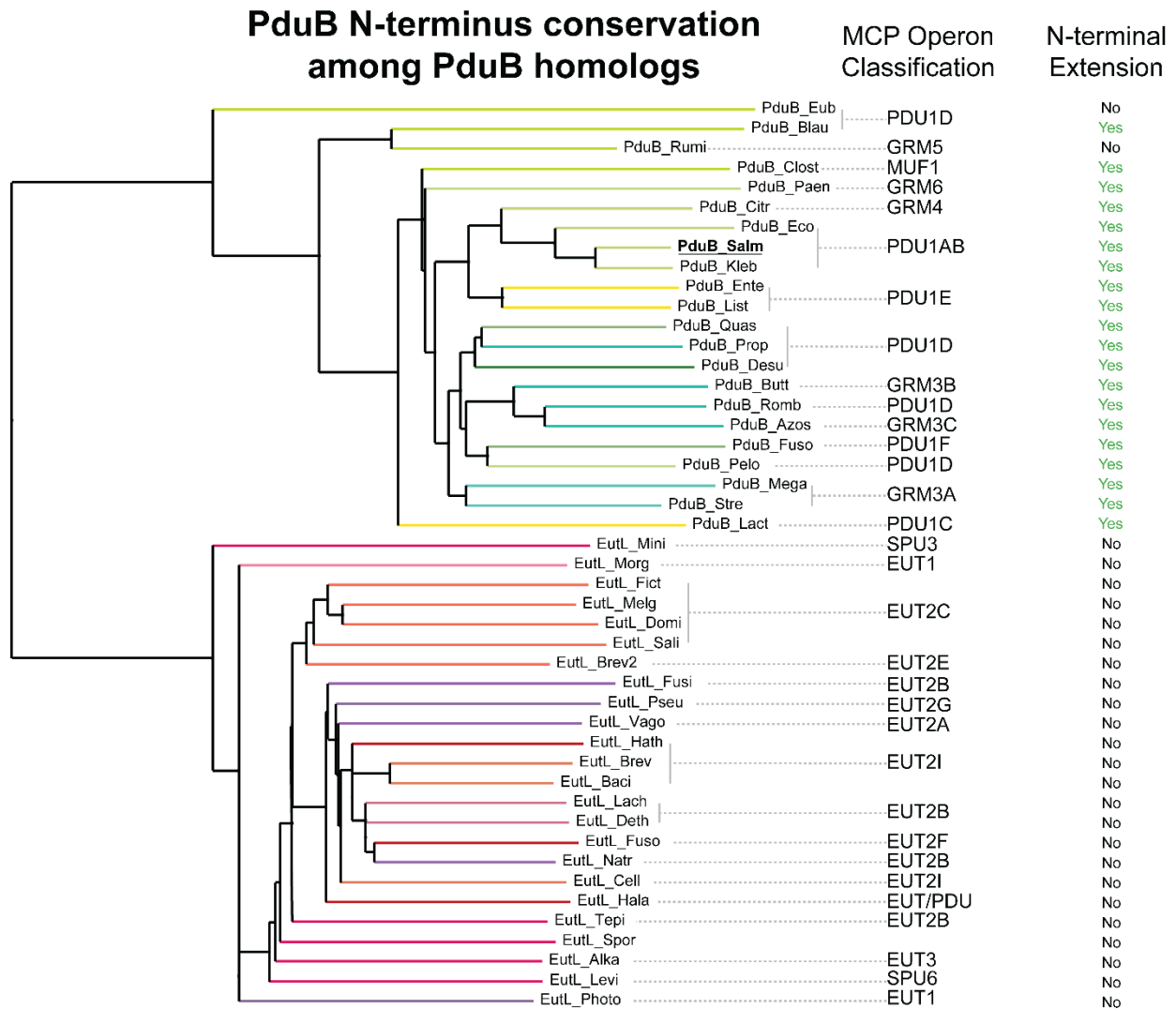
**Figure S2.** Tornado plot showing changes in maximum propionaldehyde level observed in MCP model with a 10% increase and decrease in the noted parameter.



**Figure S3.** Tornado plot showing changes in maximum propionaldehyde level observed in polar body model with a 10% increase and decrease in the noted parameter.



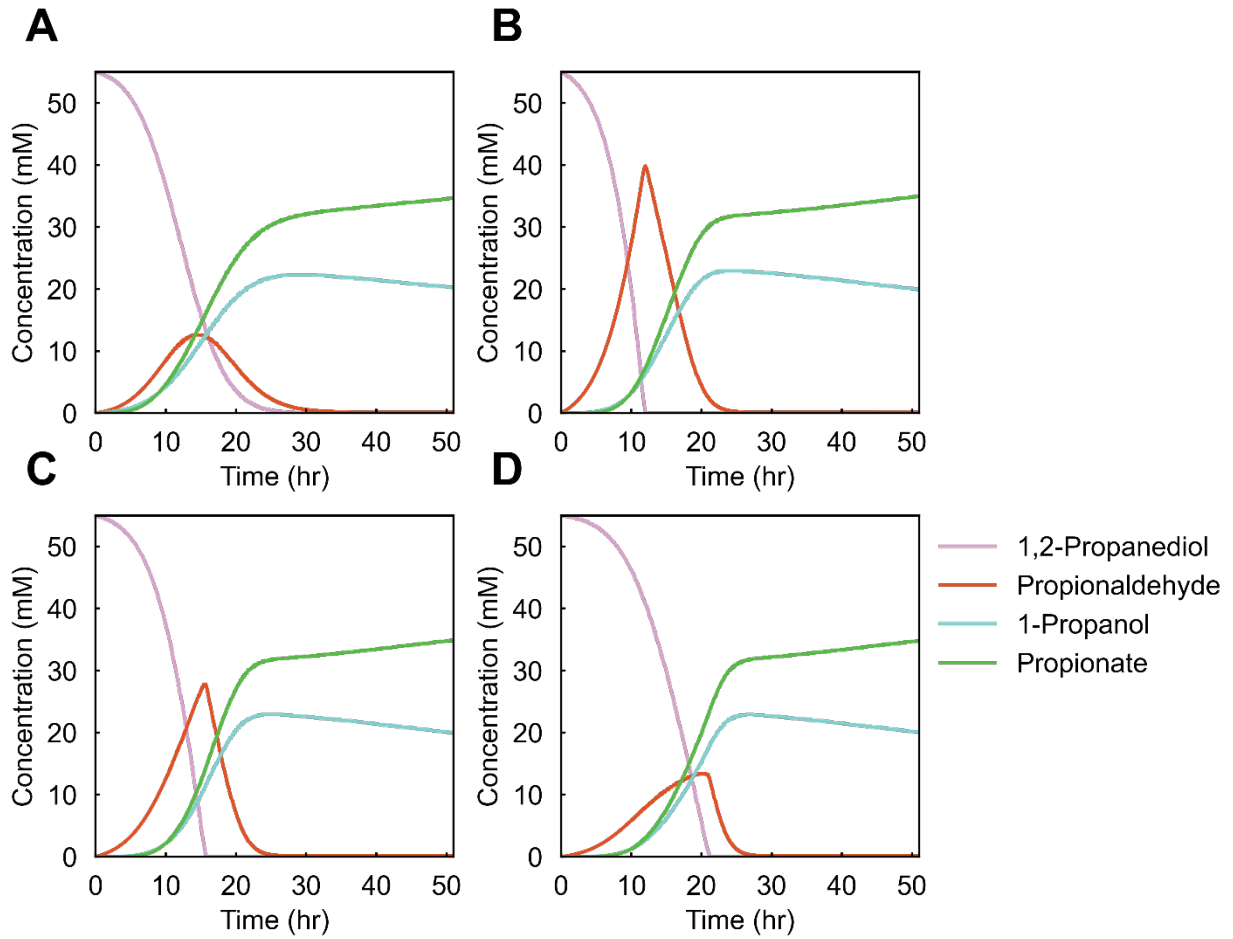
**Figure S4.** Phylogenetic tree of PduB homologs and their associated MCP types (information adapted from Sutter et al. [2]). The PduB sequence from *S. enterica* investigated in this work is underlined and bolded (PduB\_Salm).



**Acronym definitions**

- PDU Propanediol utilization microcompartment
- GRM Glycyl radical enzyme containing microcompartment
- MUF Metabolosome microcompartment of unknown function
- SPU Sugar phosphate utilization micorcompartment
- EUT Ethanolamine utilization microcompartment

**Figure S5.** Full profiles of all metabolites for the kinetic model results shown in Figure 7. Profiles are shown for the (a) MCP model with 0.46 mM PduCDE and the polar body model with (b) 0.46 mM PduCDE, (c) 0.23 mM PduCDE, and (d) 0.12 mM PduCDE.



## Section E: Supplemental Alignments

### Supplemental Alignment S1. Multiple sequence alignment of PduB homologs.

CLUSTAL O(1.2.4) multiple sequence alignment

```
EutL_Mini      -----MSVLEPIIPKV 11
EutL_Morg      -----MPVLDLIRPNI 11
EutL_Fict      -----MSLRRIQTDI 10
EutL_Melg      -----MDLQPVKAEI 10
EutL_Domi      -----MTSQKIFAEI 10
EutL_Levi      -----MLDPIPANA 9
EutL_Sali      -----MIRPVYAEM 9
EutL_Photo     -----MAILDKIQPAV 11
EutL_Alki      -----MATLDPIKPTI 11
EutL_Spor      -----MIEPIKPKV 9
EutL_Brev2     -----MEKPIRATP 9
EutL_Fusi      -----MTGDLIKAEI 10
EutL_Pseu      -----MIDELIPVKV 10
EutL_Tepi      -----MYGFKPIKAQV 11
EutL_Vago      -----MINDSLGANV 10
EutL_Hath      -----MIGEQUIPVNI 10
EutL_Cell      -----MKNDKVRANV 10
EutL_Hala      -----MRDDPLRAEV 10
EutL_Brev      -----MRNEPIYAKV 10
EutL_Baci      -----MKNERLQANI 10
EutL_Lach      -----MKRDPVRASV 10
EutL_Deth      -----MKGDPKTTL 10
EutL_Fuso      -----ME-VKMINDPLRASV 14
EutL_Natr      -----MRNDPIRAQV 10
PduB_Eub      -----MGQV-----IQMADYFTPDAEQF 18
PduB_Blau     MFFLLK-----YVKTLNNTGKSIEIPQLLSYIKVTIPKGDFFMNVKDI STHCTRTEF 52
PduB_Rumi      -----MIDVHHIEQISQDCSLTEF 19
PduB_Clost    --MQEQLIRNII SEVLKNVENTIVANPNENKEE---LKN---IQNFQNNKISLENDIPEF 52
PduB_Paen     --MDEAILKQVIEAAVRAIPGGVPSSPAADAAAGLPLNNQYTEKREQDVTDYKAITEY 58
PduB_Lact     MKNENEIVDRVMSRILANKPTE-----AESGSNALKAKGQALTEF 40
PduB_Citr     -MSSDNLVENIIAEVLGKVGDLKTPSAASQP-----GISQQEPAIPRKLKGLTEF 49
PduB_Ente     MANTDQKIEEIMKRVLAKMETGNEL----K-----TTNGGDHKMIEKQCALTEF 45
PduB_List     MSEQNKLIDEILKQVME TVNDAPEKIV-----EDGGSRQMSEKAVQLTEF 45
PduB_Eco      -MSSNELVEQIMAQVIARVATPEHEVLP-ET-----THQTRETAMAEEKSCTLLEF 48
PduB_Salm     -MSSNELVEQIMAQVIARVATPEQQAI P-GQ-----PQPIRETAMAEEKSCLTEF 48
PduB_Kleb     -MSSNELIDQIMTQVIARMAKPE SNVIP-AP-----SHSTRETAMAENCSLLEF 48
PduB_Quas     --MREQLIEKIMEEVIKKVDAPATEEAP-----QAEKKHSSLTEF 38
PduB_Desu     --MDEQLVQKVLEQVMKKVGAASSSAAASSVPAS-----SSPVAEQVAGRGLTSLPTEF 52
PduB_Prop     --MSNELVDQVMAEVLKKGGSAPQAAPAAV-----TPGVRPQSTEF 40
PduB_Butt     --MRDNLVEQII SEVMNKQTDVSA-----TGKNTANIPACGLTEF 38
PduB_Romb     --MKEDLMSKIMDEVMMKMGETPKLEVKEETA-----EEAKYEEPNSCSLLEF 46
PduB_Azo      --MKDELVEKIMGEVLRKMGPDLLPAIPDSPRA-----APPPSSPSRSLGGCGLTEF 52
PduB_Mega     --MDQQLLEKVMKQVQHELESQVQVNTNGES-----IFPKEKEAGIKTMAGCTEF 48
PduB_Fuso     MSMQENLVQKMVAEVVEKLDKKGICETPKKV-----ESDCACASKND--CRLTEF 48
PduB_Stre     --MDQELLNKVVQGV RDELNKTT----PTT-----SSDC----PINCATSITEF 39
PduB_Pelo     --MQEQLIDKVMDEIKKR MENHA----PTA-----SQGVQEKPKASVNPGIT EF 43
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EutL\_Mini LS-----VRRLPAADAALLRAYGADPERHTSLGLVTVDQDD-STYVALDEATKHAHV 62  
 EutL\_Morg TA-----MRLIANVQQDFCRELNLPK-HIRSLGLITVDSDD-VAYIAADEATKQAMV 61  
 EutL\_Fict LA-----VRLIPNADAALCEALHVPS-SFKSLGILTTTSDD-VGFTALDEATKQANV 60  
 EutL\_Melg LA-----AKMIPNV DAGLAESLALKP-HQRSIGILT TNIDD-VGYTALDAATKKADV 60  
 EutL\_Domi LA-----MKVIPNV DAGLAEKLLKLP-YQRSIGLFTSTIDD-VGYTAADEATKKADV 60  
 EutL\_Levi LA-----VRLIPSVAP EYARQLGLRP-DQRSLGLLTADNDD-ATYVSI DEATKMAV 59  
 EutL\_Sali LA-----TRII PNVDKQLARQFGLGP-SHRSLGMFTVTLDD-VGV TALDEATKRADV 59  
 EutL\_Photo LA-----TRVIASVDPVYKEKMGLEP-HHNSIGLITADCDD-VTYCALDEATKAANV 61  
 EutL\_Alki LA-----VRLIPNVHTDFAKRLELRP-DQRSIAIITCDMDD-PLYVALDEATKMAV 61  
 EutL\_Spor LA-----VRLIPNVAPDLAAQLGLKD-YQRSIGMITSNMDD-PTYTALDEATKKAEV 59  
 EutL\_Brev2 LA-----VRMIPNVDPRLAEKLNVP-HIRSLGLLTSTIDD-VGYTAIDEATKKAHV 59  
 EutL\_Fusi LA-----AKIIPNVHPELAGKLLKGG-GSCSLGLLTCTSDDD-VAYTALDEATKKADV 60  
 EutL\_Pseu LA-----THTIANVSEGLAKRLSLPD-GHRSIGILTDCDD-MTYCALDEATKAAV 60  
 EutL\_Tepi LA-----VKLIPNVNEYMKKELNLKP-EHRSIGMLT VTIDD-VGYTAVDEATKKADV 61  
 EutL\_Vago LS-----SKII SNIDPGLAKALNLD PVKHRSLGLVTGDCDD-VTYVALDEATKKAMV 61  
 EutL\_Hath LS-----VKMIPNVNPD MVKQFNLDP-NHKSIGMITADCDD-VTYTALDEATKASDV 60  
 EutL\_Cell LG-----VRVISNVSP ELAAKLELGP-KHKSIGIITADSDD-VTYTALDEATKDADV 60  
 EutL\_Hala LS-----VQLIPNVNDALKEKLNLEA-GHRSIGLITCDIDD-VGYTALDEATKKAHV 60  
 EutL\_Brev LG-----TKLISNVSPDMAEKLELKP-HHKSIGIITVDIDD-VTYTALDEATKAAEV 60  
 EutL\_Baci LS-----VKIISNISPDMAKELELKP-YHKSGLITADTDD-VTYTALDEATKAADV 60  
 EutL\_Lach LA-----TKIIPNVSPDMAKEFNLDP-SMKSLALITADSDD-VTYTALDEATKKADV 60  
 EutL\_Deth LS-----IKIIPNVNPDLAASLGLRT-DQRSLGLVTADCDD-VTYTALDEATKKADV 60  
 EutL\_Fuso LS-----VKLIPNVDAKMAEELNLNP-GYRSIGIITADSDD-VTYTALDEATKMAEV 64  
 EutL\_Natr LS-----IKLIPNINPDMA TELKLEK-QHRSIGIITDIDD-VSYTALDEATKKADV 60  
 PduB\_Eub VGT-AAAHTIGLMIGNINGMVRILDIPE-KYNALGISSRTGTVAQAI AIDDAVKACNA 76  
 PduB\_Blau VGT-TVLDTIGLVI SGIDHTLLQKMNIGT-KYYALGLFSSRTGAAGQLTAIDDAVKATNT 110  
 PduB\_Rumi VGT-TVLDTIGLVI PGIDPSLLEKMNLKP-PYKSLGLISSRTGAAGQINA ADEAVKSTNT 77  
 PduB\_Clost VGS-AIGDTVGLVIANVNE LLLNKMNVSK-KYRSIGIIGARTGASAQI ISADEAVKGTNS 110  
 PduB\_Paen VGS-AMGDTVGLVIANLDRHLHESMGLDP-RYRSIGI ISSRTGAGPHIMAADEAVKATNT 116  
 PduB\_Lact VGTAPIGDTIGLVIANVDANLAEKMNLDK-KYRSIGVISARTGASPQVMATDVAVKATNT 99  
 PduB\_Citr VGT-ATGDSIGLVIANVDSALLDAMHLEK-KYRSIGILGARTGAGPQIMAADEAVKATNT 107  
 PduB\_Ente VGATEFGDTIGLVIANVDS SLEETMKLEK-KYRSIGIVGARTGAGPHIMAADEAVKATNT 104  
 PduB\_List VGT-AIGDTIGLVIANVDG LLEAMKLEK-SYRSIGILGARTGAGPHIMAADEAVKATNT 103  
 PduB\_Eco VGT-AIGDTVGLVIANVDS ALLDAMKLEK-RYRSIGILGARTGAGPHIMAADEAVKATNT 106  
 PduB\_Salm VGT-AIGDTLGLVIANVD TALLDAMKLEK-RYRSIGILGARTGAGPHIMAADEAVKATNT 106  
 PduB\_Kleb VGT-AIGDTVGLVIANVD GALLEAMKLEK-RYRSIGILGARTGAGPHIMAADEAVKATNT 106  
 PduB\_Quas VGT-GLGDSIGLVIANVD HSIHEAMNLDK-KYRSIGILGARTGAGPQIMAADEAVKATNT 96  
 PduB\_Desu VGT-AMGDTIGLVIANVD P LLEHEKMG LDS-RYRSIGILGGRTGAGPQIMAADEAVKATNT 110  
 PduB\_Prop VGT-GLGDTIGLVIANVD PALTEKMNIDP-KYRSIGIIGGRTGAGPQLFA ADEAVKATNT 98  
 PduB\_Butt VGT-AMGHTIGLVIANVD NQLHEVMKIDK-KYRSIGILGARTGAGPHI FA ADEAIKATNS 96  
 PduB\_Romb VGT-AIGNTIGLVIANVD YNLHELMNIDK-KYRSIGIVGGRTGAGPHI FA ADEAVKATNS 104  
 PduB\_Azos VGT-AIGHTIGLVIANVD P LLEHEKMKISP-KYRSIGIVGARTGAGPHI FA ADEAVKATNS 110  
 PduB\_Mega VGT-AIGDTIGLVIAGVD P LKDTMKLG--KFRSIGIIGRIGAGPQIMAVDDAVKATNT 105  
 PduB\_Fuso VGLTTHGHGIGLVIANVDS ALHEAMGLDK-KYRSIGIIGARTGAGPFI MA ADEAVKATNT 107  
 PduB\_Stre VGT-AIGDTIGLVIANVD HELHEKMGIDP-KYRSIGIIGARV GAGPHIMAVDEAVKATNT 97  
 PduB\_Pelo VGT-AIGDTIGLVIANVD PMLHEKMKIDP-KYRSIGILGARTGAGPHIMAADEAVKATNT 101

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EutL\_Mini DVV---FARISFYAGSAHASGRLSSEIILGVLAASDPDEIEHGLTALLRCLAHDA-CFYAAD 118  
EutL\_Morg EVV---FGRSFLYAGASNSPSPSTSGEVMIMLGGPNPAEVRAGLDSMMATIESGP-AFRWAN 117  
EutL\_Fict EVA---YAKSFYAGASHASGPLSGEFIFIGIVAGPTPDEVRSGIDAAIHTIEHEA-AFEALD 116  
EutL\_Melg EVV---YGSSFYAGAAHASGPLSGEFIFIGILAGPTPAEVKSGMEVFCYTAENEA-WFYALD 116  
EutL\_Domi EVV---YAKSFYAGAEHASGPLSGEFIMAILAGPTSEEVKSSALDAIQIKVESDA-YFEMIS 116  
EutL\_Levi EVV---YAHSFYAGARHASGPLSGEIIALLAGPNPAEVRAGLDAALRYLSTRA-LWYSAN 115  
EutL\_Sali DGV---YAASFYGGSDYPSGPLSGEFLGVLAGPDPEEVRSGMDAVKQMVESGA-HFEALD 115  
EutL\_Photo DVV---YARISFYAGAAHSSGPTSGEVIGILAGACPADVIAGLERAKAFIEDEA-AFQCAD 117  
EutL\_Alki EVV---YQSFYAGSAHASGKLSGEVIGILAGNDPAEVRSGVNACVAYCEESA-WFYSAN 117  
EutL\_Spor EVV---YAKSFYAGAAHASGPRSSEIIGILAAPTPAEISAGIDACINHLENEA-FFY-TC 114  
EutL\_Brev2 EVV---YARISFYAGSAHASGPLSGEFIFIGVIGGATPSEVESGLDAAIAFMANGA-CFYSLN 115  
EutL\_Fusi KVV---YAKSFLYAGSGNASTRFAGEVIGI IAGPDPEEVRSGLEACAHIQNSA-FFHSAN 116  
EutL\_Pseu EVV---YGRSFLYAGAGNASTKLAGEVIGI IAGPNPAEVQSGLRAAVAFLEGG-GFRSAN 116  
EutL\_Tepi EVV---YARISFYAGAAHASGPLSGEFIFIGILAGPNPAEVKSGLNAAIDCIENDA-CFWAAD 117  
EutL\_Vago DVV---YARISFLYAGAGSASTKLAGEVIGI IAGPNPAEVKSGLDVVVYEIENSP-GFVSAN 117  
EutL\_Hath KVV---YARISFYGGAGNASTKLAGEVIGI ISGSPAEVRSGLNAIVDFIESGVATFRMAN 117  
EutL\_Cell EVV---YARISMYAGAGNASTKLAGEVIGI IGGPSPAEVRSGLKAALDFFGSDA-CFISAN 116  
EutL\_Hala DIV---YAKSFYAGADNATTKLAGEFIFIGIMAGPSPAENVSGLDAAVDFIENGA-CFYSAN 116  
EutL\_Brev DVV---YAKSMYAGSANASTKLAGEVIGI IAGPSPAEVKSGLSAAISYIEDSA-HFVGAN 116  
EutL\_Baci EVV---YAKSFLYAGSANASTKLAGEIIGILAGPSPAEVRSGLNAAVDFIENGA-HFISAN 116  
EutL\_Lach KVV---YAKSFYAGAAANANTKLAGEIIGILAGPNPAEVRSGLEAAVDMIENQA-YFISAN 116  
EutL\_Deth KVV---YAKSFYGGAAANANTKLAGEIIGILAGPNPAEVRSGLNALADFIENEA-WFISAN 116  
EutL\_Fuso VIV---YAKSFYGGAAANANTKLAGEVIGI IAGPNPAEVKSGLNAAIDFIENGG-CFYSAN 120  
EutL\_Natr QVA---YAKSFYGGAAANASTKLAGEFIFIGILAGPNPAEVRSGLNAAIEFIENGA-CFYSAN 116  
PduB\_Eub KLLRFEMAI DAGRQCQGC-----LFLIGAETISDARKVVEESLHNID-YWSQCIYIN 128  
PduB\_Blau EVLSIELPRDTKGGGGHGC-----YIVLGGNDVSDVRHAVQLALELLE-KYAGELYIS 162  
PduB\_Rumi EIVSIELPRDTKGGGGHGC-----FVIGGDVSDVRRRAVDITLEYIG-KYAGELYIS 129  
PduB\_Clost EIIKVELARDTKGNAGHGV-----LIVFGAMDVSDVRQAVEITLNDLN-RTFGDLYIN 162  
PduB\_Paen EVVSIEMARDTKGGAGHGC-----LIIIFASEDVSDRRRAVEVVLKELP-RTFGDVYPC 168  
PduB\_Lact AIISVEMPRDTKTGAGHGS-----LITIGGEDVSDVRRRAVEVALAQT-P-KMFGDIYTN 151  
PduB\_Citr EVVAIELARDTKGGAGHGS-----LVLFGGQDVSDVKRAVEVALGELE-RTFGDIYAN 159  
PduB\_Ente EVLILIEPRDTKGGAGHGS-----LIVIFGGDDVSDVKRAVEVTLKEVDTRTFGDVYGN 157  
PduB\_List EVVKIELPRDTKGGAGHGS-----LIIIFGGDDVSDVKRAVEVALNELD-KTFGDVYGN 155  
PduB\_Eco EVISIELPRDTKGGAGHGS-----LILLGGNDVSDVVRGIEVALKELD-RTFGDVYAN 158  
PduB\_Salm EVVSIELPRDTKGGAGHGS-----LIIIFGGNDVSDVVRGIEVALKELD-RTFGDVYGN 158  
PduB\_Kleb EVVSIELPRDTKGGPGHGS-----LIIIFGGNDVSDVVRGIEVALKELD-RTFGDVYAN 158  
PduB\_Quas EILAVELARDTKGGAGHGS-----LIIIFGAEDVADARRAVEVALKELE-RTFGDVHAN 148  
PduB\_Desu EVVAVELPRDTRGGAGHGS-----LIIIFGAEEVSDARRAVEVALQELQ-RTFGDVYAN 162  
PduB\_Prop EIVLAEPRDTKGGAGHGS-----LIVFAEDVSDARRAVEVTLKELE-RTFGDVYGT 150  
PduB\_Butt EILSIELARDTEGGGGHGC-----LIIIFGATDVSDVRRRAIEVALAEID-RTMGDVYGS 148  
PduB\_Romb EILLIELPRDTEGGAGHGS-----LIIIFGAEDVSDARRAVEVTLNELE-RTFGDVYGS 156  
PduB\_Azos EVVLIELPRDTEGGAGHGC-----LIIIFGAEDVSDARRAVEVTLKEVE-RTMGDVYGT 162  
PduB\_Mega EIISVELPRDTKGGAGHGS-----LILIFGAEDVSDARRAVETALQVLP-KYFGDVYGN 157  
PduB\_Fuso EVISIELPRDTEGGAGHGS-----LILIFGAEDVSDVKRAVEVAINNVT-EKFGDVYAN 159  
PduB\_Stre EVLSIELPRDTKGGAGHGS-----LIIIFGSDVSDARRGIEVALATLD-QTFGDVYSN 149  
PduB\_Pelo EIISIELPRDTKGGAGHGS-----LIIIFGAEEVSDARRAIEVALKELD-RTFGDVYGN 153

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|            |  |               |       |
|------------|--|---------------|-------|
| EutL_Mini  | FPPPT-ETNFAAAW---LTGSLPACQAAAEAYAEAVVRIA----       | AE-PYE-----   | 218   |
| EutL_Morg  | VAPPS-ETNYSAAF---LTGSQAACQAACNAFTEAVLDIA----       | RN-PIQ-----   | 216   |
| EutL_Fict  | YGPPS-ETNYGGGL---VTGTESACRAAAEAFREAIISIA----       | KS-PRS-----   | 216   |
| EutL_Melg  | YKPPS-ETNFGGGL---LTGSQSSCKSATQAFMDTVIRIA----       | EN-PKV-----   | 216   |
| EutL_Domi  | FAPPS-ETNFGGGL---LTGSQSSCQAAADAFRQTILILIT----      | AN-PKS-----   | 216   |
| EutL_Levi  | TPPPS-ETNFMGAV---LTGTQSACQAAALAFQDAVLEVA----       | RR-PIE-----   | 215   |
| EutL_Sali  | YGPPPT-ETNFGGAL---LTGSQSACQAAADAFRDAVRSIA----      | DS-PLS-----   | 215   |
| EutL_Photo | FAPPS-ETNFGGGH---LAGSQAACRAACEAFQAAVLEVA----       | RN-PIA-----   | 217   |
| EutL_Alki  | FKPPS-ETNFGGAL---MAGSQSACKAACQAFSDAVVEVA----       | AH-PLK-----   | 217   |
| EutL_Spor  | YGPPPT-ETNFGGAL---LTGSQSACRAACDAFQRAVIDVT----      | LN-ALK-----   | 214   |
| EutL_Brev2 | FGPPT-ETNFGGAL---LTGSQSACSAAAEAFAEAVRSVA----       | AT-PVK-----   | 215   |
| EutL_Fusi  | YGPPPT-ETNFAGGL---LTGSQESCRAACDAFARAVEQTA----      | SE-PRD-----   | 216   |
| EutL_Pseu  | YEPPS-ETNFGGGL---LTGTQSACNAACAAFAEAVKAI A----      | AR-PRE-----   | 216   |
| EutL_Tepi  | FGPPS-ETNFAGGH---LTGSQSACKAACDAFAAAVMEVA----       | KY-PKR-----   | 217   |
| EutL_Vago  | YGPPS-ETNFAGGL---LTGSQSACQAACDAFAQAVQSVA----       | AN-PKS-----   | 217   |
| EutL_Hath  | FGPPS-ETNFGGAL---LTGSQSACKAACDAFADAVKFVA----       | AN-PTD-----   | 217   |
| EutL_Cell  | FGPPS-ETNFGGGL---LTGTQSACHAACAAFAEAIKAVA----       | DN-PRD-----   | 216   |
| EutL_Hala  | YGPPS-ETNFGGAL---LTGSQSACQAACNAFANAVKSVA----       | DK-PCN-----   | 216   |
| EutL_Brev  | FGPPS-ETNFGGAL---LTGSQSACRAACDAFAAAVQFVA----       | DN-PTS-----   | 216   |
| EutL_Baci  | YGPPS-ETNFGGGL---LTGSQSACKAACDAFAEAVQFVA----       | EN-PTN-----   | 216   |
| EutL_Lach  | YAPPS-ETNFGGAL---LTGSQSACKAACDAFAAAVEFVA----       | DN-PKE-----   | 216   |
| EutL_Deth  | FAPPS-ETNFAGGL---LTGSQSACKAACDAFAQAVIFVA----       | EN-PVL-----   | 216   |
| EutL_Fuso  | FGPPS-ETNFGGGL---LTGSQSACKSACDAFAEAVKFVA----       | QN-PKK-----   | 220   |
| EutL_Natr  | YGPPS-ETNFGGGL---LTGSQSACKAACDAFAAAVEYVA----       | DN-PTK-----   | 216   |
| PduB_Eub   | GSPSH-NLRMMNEFVAGVTGDIAAVQKAMDVGKETGCQLLTALGIT-    | PISIQDVAEECGE | 244   |
| PduB_Blau  | MTPDM-GTAHSNEVILALSGDAGAVKEAVLNARQIGLELLIAMGSY-    | PEIPGTP-----  | 272   |
| PduB_Rumi  | MTPDQ-GTSHSNEVIVAVTGDADAVKNAVLTARDLGLQLLVSMGSY-    | PESPATP-----  | 239   |
| PduB_Clost | WSPAY-KTSYSNECVTTIYGDSEAVKQALITAREVGINVLSKIGST-    | PTSQSTP-----  | 272   |
| PduB_Paen  | ASPDK-GTARSNEIIVTITGDSGAVRQSVISARDVGVSVLRSMGHN-    | PVSMTQSN----- | 279   |
| PduB_Lact  | ASPGK-GTSLTNEAMIHISGDSGAVRQAVVTARDVGIKLLSTLGSE-    | PKN-DYP-----  | 260   |
| PduB_Citr  | NSPAK-GTSFTNEVILIVSGDSGAVRQAVRSARDVGLTLETMGGK-     | APSATTP-----  | 269   |
| PduB_Ente  | ATPQN-GQSFNEVTMTITGDSGAVRQAIISARDIGCQLLGTMGST-     | PKN-DQP-----  | 266   |
| PduB_List  | SSPAD-GTSFSNEVILCISGDSGAVRQAVISAREIGKLLGALGDE-     | PKN-DRP-----  | 264   |
| PduB_Eco   | SSPAQ-GTSYSNEVILIIISGDSGAVARRSSPHAKSVKPCFRLSAIH-   | RKTIALPT----- | S 270 |
| PduB_Salm  | SSPAH-GTSFSNEAILVLSGDSGAVRQAVTSAREIGKTVLATLGSE-    | PKN-DRP-----  | 267   |
| PduB_Kleb  | SSPAH-GTSFSNEAILVLSGDSGAVRQAVISAREIGKTI LSTLGCE-   | PKN-DRP-----  | 267   |
| PduB_Quas  | SSPAK-GTSFTNEVILTFSGDSGAVRQALITAREVGLDLLGTLGGK-    | PASTTVP-----  | 258   |
| PduB_Desu  | ASPAK-GTSFTNEAILFISGDSGAVRQALIAAREVGIALLSTLGDK-    | PQSLTRP-----  | 272   |
| PduB_Prop  | ATPQH-GQSFNEVTMTFSGDSGAVRQAVIAAREVGKTL LGAFGGD-    | VVSTTTP-----  | 260   |
| PduB_Butt  | ASPAN-GTSFSNEVIFTFAGDSGAVRQAVIAAREVGLQLLATLDPAPI-  | QSATKP-----   | 259   |
| PduB_Romb  | STPGNGGTSYSNEVILTFSGDSGAVRQSI IAAREVGKQLLSTLSNEEL- | KSSSTIP-----  | 268   |
| PduB_Azos  | ASPGS-GTSYSNEVIFLFTGDSGAVRQAVIAAREVGKQVLATLDPAEI-  | KSVTRP-----   | 273   |
| PduB_Mega  | ASPGNGKTSYSNEVILCFSGDSGAVRQAVKAAIVVGKLLGALGDE-     | PKSTTTP-----  | 268   |
| PduB_Fuso  | SSPSKG-TSYSNEAILAICGDSGAVKQAVLAAKEVGVKLLLEAMGGE-   | APSASHP-----  | 269   |
| PduB_Stre  | SSPSNG-TSYSNEVILTGTGDSGAVRQAVSAAAREVGLNLLGSLGEQ-   | AVSTTKP-----  | 259   |
| PduB_Pelo  | GSPASG-TSYSNEVILQVTGDSGAVRQAVLAAKEVGVKLLLEAMAGP-   | APSSTKP-----  | 263   |

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|            |                               |     |
|------------|-------------------------------|-----|
| EutL_Mini  | -RHFTAFSR-----                | 226 |
| EutL_Morg  | -RA-----                      | 218 |
| EutL_Fict  | -Y-----                       | 217 |
| EutL_Melg  | -Y-----                       | 217 |
| EutL_Domi  | -Y-----                       | 217 |
| EutL_Levi  | -Y-----                       | 216 |
| EutL_Sali  | -K-----                       | 216 |
| EutL_Photo | -N-----                       | 218 |
| EutL_Alki  | -Y-----                       | 218 |
| EutL_Spor  | -F-----                       | 215 |
| EutL_Brev2 | -R-----                       | 216 |
| EutL_Fusi  | -II-----                      | 218 |
| EutL_Pseu  | -I-----                       | 217 |
| EutL_Tepi  | -Y-----                       | 218 |
| EutL_Vago  | -Y-----                       | 218 |
| EutL_Hath  | -F-----                       | 218 |
| EutL_Cell  | -Y-----                       | 217 |
| EutL_Hala  | -Y-----                       | 217 |
| EutL_Brev  | -Y-----                       | 217 |
| EutL_Baci  | -Y-----                       | 217 |
| EutL_Lach  | -E-----                       | 217 |
| EutL_Deth  | -Y-----                       | 217 |
| EutL_Fuso  | -I-----                       | 221 |
| EutL_Natr  | -F-----                       | 217 |
| PduB_Eub   | DYLRNMSPAVCQEEKAASIHINKKCDEER | 274 |
| PduB_Blau  | -YL-----                      | 274 |
| PduB_Rumi  | -YLQ-----                     | 242 |
| PduB_Clost | -YI-----                      | 274 |
| PduB_Paen  | --L-----                      | 280 |
| PduB_Lact  | SYI-----                      | 263 |
| PduB_Citr  | -YI-----                      | 271 |
| PduB_Ente  | SYI-----                      | 269 |
| PduB_List  | SYI-----                      | 267 |
| PduB_Eco   | EFSEADL-----                  | 277 |
| PduB_Salm  | SYI-----                      | 270 |
| PduB_Kleb  | SYI-----                      | 270 |
| PduB_Quas  | -YI-----                      | 260 |
| PduB_Desu  | -YI-----                      | 274 |
| PduB_Prop  | -YI-----                      | 262 |
| PduB_Butt  | -YI-----                      | 261 |
| PduB_Romb  | -YI-----                      | 270 |
| PduB_Azos  | -YI-----                      | 275 |
| PduB_Mega  | -YI-----                      | 270 |
| PduB_Fuso  | -YI-----                      | 271 |
| PduB_Stre  | -YI-----                      | 261 |
| PduB_Pelo  | -YI-----                      | 265 |

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