

Supporting Information

Bitter peptides YFYPEL, VAPFPEVF and YQEPVLGPVRGPFPIIV, released during gastric digestion of casein, stimulate mechanisms of gastric acid secretion via bitter taste receptors TAS2R16 and TAS2R38

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Tables

Table-SI 1. Changes in gene expression of the bitter receptors after incubation for 15/30/60/120 min with peptides VAPFPEVF (17.5 µM), YFYPEL (0.03 µM) and YQEPVLGPVRGPFPIIV (0.4 µM). Normalized to the expression of PPIA and GAPDH and expression of the receptors in HGT-1 cells without treatment. Data shown as mean ± SEM, n = 3 – 5, t. r. = 3, Statistics: t test Holm-Šidák method; significant differences are expressed with * = p ≤ 0.05.

receptor	treatment	15 min	30 min	60 min	120 min
<i>TAS2R1</i>	VAPFPEVF	0.79 ± 0.12	1.36 ± 0.20	0.44 ± 0.22	1.62 ± 0.61
	YFYPEL	0.36 ± 0.25	1.03 ± 0.38	0.55 ± 0.33	0.73 ± 0.29
	YQEPVLGPVRGPFPIIV	0.33 ± 0.43	0.43 ± 0.23	1.00 ± 0.63	0.84 ± 0.26
<i>TAS2R3</i>	VAPFPEVF	0.70 ± 0.05*	1.56 ± 0.22*	1.06 ± 0.04	1.72 ± 0.12*
	YFYPEL	1.54 ± 0.07*	1.08 ± 0.16	1.19 ± 0.12	1.34 ± 0.08*
	YQEPVLGPVRGPFPIIV	1.03 ± 0.11	1.89 ± 0.19*	1.61 ± 0.22*	1.40 ± 0.02*
<i>TAS2R4</i>	VAPFPEVF	0.50 ± 0.06*	1.14 ± 0.19	0.95 ± 0.07	1.42 ± 0.14*
	YFYPEL	1.24 ± 0.07*	0.81 ± 0.12	0.92 ± 0.07	1.13 ± 0.11
	YQEPVLGPVRGPFPIIV	0.79 ± 0.08	1.71 ± 0.19*	1.60 ± 0.22*	1.21 ± 0.03*
<i>TAS2R5</i>	VAPFPEVF	0.91 ± 0.04	1.05 ± 0.07	0.93 ± 0.06	1.23 ± 0.10
	YFYPEL	1.34 ± 0.03*	1.03 ± 0.10	0.90 ± 0.04	1.02 ± 0.04
	YQEPVLGPVRGPFPIIV	1.25 ± 0.10	1.45 ± 0.10*	1.39 ± 0.13*	1.22 ± 0.03*
<i>TAS2R7</i>	VAPFPEVF	1.20 ± 0.32	0.71 ± 0.34	1.05 ± 1.05	0.62 ± 0.21
	YFYPEL	1.23 ± 0.83	2.50 ± 1.32	6.71 ± 1.56*	1.18 ± 0.41
	YQEPVLGPVRGPFPIIV	0.49 ± 0.24	0.74 ± 0.35	0.70 ± 0.94	1.82 ± 1.25
<i>TAS2R8</i>	VAPFPEVF	1.24 ± 0.13	2.25 ± 0.32*	1.34 ± 0.16	2.04 ± 0.23*
	YFYPEL	1.62 ± 0.22*	1.90 ± 0.31*	1.87 ± 0.39*	1.69 ± 0.21*
	YQEPVLGPVRGPFPIIV	1.39 ± 0.23	2.45 ± 0.28*	1.87 ± 0.32*	1.93 ± 0.23*
<i>TAS2R9</i>	VAPFPEVF	1.73 ± 0.08	1.08 ± 0.47	2.31 ± 0.75	0.99 ± 0.69
	YFYPEL	0.66 ± 0.42	2.64 ± 0.57	3.01 ± 0.75	1.41 ± 0.22
	YQEPVLGPVRGPFPIIV	0.43 ± 0.68	0.44 ± 0.80	1.91 ± 1.12	0.73 ± 0.32
<i>TAS2R10</i>	VAPFPEVF	0.34 ± 0.05*	0.76 ± 0.14	0.91 ± 0.15	1.15 ± 0.10
	YFYPEL	0.64 ± 0.08*	0.41 ± 0.05*	0.72 ± 0.20	0.90 ± 0.11
	YQEPVLGPVRGPFPIIV	0.52 ± 0.06*	0.78 ± 0.07	0.90 ± 0.12	1.04 ± 0.08
<i>TAS2R13</i>	VAPFPEVF	0.44 ± 0.09*	1.28 ± 0.33	1.01 ± 0.05	1.92 ± 0.36*
	YFYPEL	1.14 ± 0.07	0.53 ± 0.11*	0.78 ± 0.06	0.88 ± 0.36
	YQEPVLGPVRGPFPIIV	0.70 ± 0.11	1.65 ± 0.32*	1.16 ± 0.15	1.31 ± 0.08
<i>TAS2R14</i>	VAPFPEVF	0.67 ± 0.02*	0.97 ± 0.05	0.86 ± 0.03*	1.08 ± 0.03
	YFYPEL	1.11 ± 0.01*	0.92 ± 0.05	0.99 ± 0.02	1.21 ± 0.09*
	YQEPVLGPVRGPFPIIV	0.85 ± 0.04*	1.08 ± 0.06	1.03 ± 0.04	1.08 ± 0.02*
<i>TAS2R16</i>	VAPFPEVF	1.50 ± 0.08*	3.70 ± 0.97*	1.65 ± 0.24*	2.88 ± 0.28*
	YFYPEL	2.58 ± 0.43*	3.19 ± 0.50*	2.44 ± 0.58*	2.51 ± 0.83*
	YQEPVLGPVRGPFPIIV	2.61 ± 0.58*	3.46 ± 0.58*	2.50 ± 0.41*	3.00 ± 0.31*
<i>TAS2R19</i>	VAPFPEVF	0.66 ± 0.07*	1.22 ± 0.17	0.69 ± 0.02*	1.25 ± 0.09*
	YFYPEL	1.60 ± 0.02*	0.82 ± 0.09	0.79 ± 0.07	1.05 ± 0.14
	YQEPVLGPVRGPFPIIV	0.85 ± 0.14	1.58 ± 0.09*	1.27 ± 0.17	1.05 ± 0.03
<i>TAS2R20</i>	VAPFPEVF	0.70 ± 0.07*	1.33 ± 0.13*	0.87 ± 0.03*	1.78 ± 0.20*
	YFYPEL	1.37 ± 0.05*	0.93 ± 0.07	0.94 ± 0.09	1.16 ± 0.13
	YQEPVLGPVRGPFPIIV	0.95 ± 0.08	1.42 ± 0.06*	1.38 ± 0.08*	1.31 ± 0.06*
<i>TAS2R30</i>	VAPFPEVF	0.56 ± 0.09*	1.53 ± 0.32*	0.75 ± 0.04*	1.62 ± 0.19*
	YFYPEL	1.79 ± 0.06*	0.62 ± 0.09*	0.89 ± 0.08	1.15 ± 0.31
	YQEPVLGPVRGPFPIIV	0.74 ± 0.13	1.88 ± 0.20*	1.20 ± 0.17	0.89 ± 0.03
<i>TAS2R31</i>	VAPFPEVF	0.66 ± 0.05*	1.14 ± 0.14	0.70 ± 0.04	1.22 ± 0.07*
	YFYPEL	1.65 ± 0.05*	0.91 ± 0.06	0.96 ± 0.05	1.09 ± 0.11
	YQEPVLGPVRGPFPIIV	0.86 ± 0.12	1.43 ± 0.11*	1.33 ± 0.12*	0.99 ± 0.04

receptor	treatment	15 min		30 min		60 min		120 min	
<i>TAS2R38</i>	VAPFPEVF	1.31	± 0.26	2.42	± 0.55*	1.15	± 0.23	2.45	± 0.26*
	YFYPEL	2.96	± 0.64*	3.40	± 0.80*	2.39	± 0.62*	2.91	± 0.67*
	YQEPVLGPVRGPFIIV	2.01	± 0.41*	3.71	± 0.93*	2.05	± 0.49*	2.38	± 0.34*
<i>TAS2R39</i>	VAPFPEVF	0.39	± 0.13*	1.37	± 0.34	0.55	± 0.06*	1.16	± 0.25
	YFYPEL	1.21	± 0.16	0.42	± 0.09*	0.62	± 0.12*	0.45	± 0.31
	YQEPVLGPVRGPFIIV	0.53	± 0.12*	1.16	± 0.14	0.73	± 0.13	0.53	± 0.09*
<i>TAS2R40</i>	VAPFPEVF	0.81	± 0.15	1.54	± 0.37	0.88	± 0.06	1.42	± 0.28
	YFYPEL	1.90	± 0.12*	0.82	± 0.12	0.59	± 0.06*	0.75	± 0.14
	YQEPVLGPVRGPFIIV	0.92	± 0.14	1.78	± 0.29*	0.96	± 0.26	0.86	± 0.07
<i>TAS2R41</i>	VAPFPEVF	0.95	± 0.27	2.32	± 1.06	0.98	± 0.20	1.90	± 0.68
	YFYPEL	2.82	± 0.64*	1.56	± 0.32	4.58	± 2.91*	3.00	± 2.14
	YQEPVLGPVRGPFIIV	1.31	± 0.67	1.71	± 2.35	2.95	± 1.80	4.61	± 2.03*
<i>TAS2R42</i>	VAPFPEVF	0.52	± 0.09*	0.88	± 0.10	0.71	± 0.03*	1.20	± 0.14
	YFYPEL	1.17	± 0.09	0.55	± 0.12*	0.69	± 0.11*	0.93	± 0.09
	YQEPVLGPVRGPFIIV	0.62	± 0.19	1.38	± 0.15*	1.18	± 0.13	0.94	± 0.06
<i>TAS2R43</i>	VAPFPEVF	0.57	± 0.07*	1.14	± 0.25	0.72	± 0.03*	1.49	± 0.15*
	YFYPEL	1.49	± 0.04*	0.64	± 0.06*	0.91	± 0.04	1.02	± 0.21
	YQEPVLGPVRGPFIIV	0.69	± 0.09*	1.44	± 0.12*	1.15	± 0.14	0.88	± 0.04
<i>TAS2R45</i>	VAPFPEVF	0.52	± 0.05*	1.08	± 0.17	0.68	± 0.03*	1.17	± 0.11
	YFYPEL	1.15	± 0.05*	0.54	± 0.06*	0.70	± 0.03*	0.94	± 0.21
	YQEPVLGPVRGPFIIV	0.63	± 0.09*	1.25	± 0.12*	1.06	± 0.12	0.84	± 0.04*
<i>TAS2R46</i>	VAPFPEVF	0.62	± 0.07*	1.47	± 0.29*	0.85	± 0.05	1.95	± 0.21*
	YFYPEL	1.70	± 0.08*	0.73	± 0.12	1.01	± 0.11	1.30	± 0.30
	YQEPVLGPVRGPFIIV	0.69	± 0.13	1.73	± 0.18*	1.30	± 0.17	1.06	± 0.04
<i>TAS2R50</i>	VAPFPEVF	0.56	± 0.06*	1.39	± 0.35	0.83	± 0.05	1.56	± 0.18*
	YFYPEL	1.50	± 0.06*	0.70	± 0.11	0.92	± 0.09	0.91	± 0.25
	YQEPVLGPVRGPFIIV	0.67	± 0.08*	1.66	± 0.19*	1.21	± 0.13	1.07	± 0.08
<i>TAS2R60</i>	VAPFPEVF				not detected				
	YFYPEL				not detected				
	YQEPVLGPVRGPFIIV				not detected				

Table-SI 2. Optimized parameters declustering potential (DP), collision energy (CE), and collision cell exit potential (CXP) for each peptide and transition.

peptide sequence	Q1 mass (Da)	DP (volts)	Q3 mass (Da)	CE (volts)	CXP (volts)
YFYPEL	416.356	36	700.2	9	28
			571.2	13	24
			474.1	13	28
			358.1	11	22
PVVVPPFLQPEVM	726.897	180	350.7	11	12
			1058.5	15	26
			978.5	23	28
			977.5	21	26
YYVPLGTQ	470.592	21	652.2	17	20
			475.2	21	30
			794.3	11	32
			636.4	11	18
VAPFPEVF	453.135	20	614.2	15	20
			596.3	19	38
			515.2	13	16
			740.4	11	20
YQEPVLGPVRGFPFIIV	940.920	80	735.3	11	16
			641.2	13	14
			570.2	15	30
			491.3	11	18
			1151.6	45	54
			1094.5	45	54
			882.4	33	24
			730.6	39	38
			441.3	41	24

Table-SI 3. Sequences of the primer pairs used for RT-qPCR.^{13,37,38}

Gene	Direction	Sequence (5' to 3')	Amplicon length, bp
TAS2R1	Forward	AAATGGCTCCGCTGGATCTC	172
	Reverse	GTGGCAAGCAAAGTCCAA	
TAS2R3	Forward	GGGACTCACCGAGGGGGTGT	160
	Reverse	CCTCAAGAGTGCCAGGGTGGTG	
TAS2R4	Forward	GCAGTGTCTGGTTGTGACC	168
	Reverse	GCGTGATGTACAGGCAAGTG	
TAS2R5	Forward	ACACTCATGGCAGCCTATCC	107
	Reverse	CGAGCACACACTGTCTTCCA	
TAS2R7	Forward	GCAGGTGTGGATGTCAAACTC	167
	Reverse	TCTTGACCCAGTCCATGCAG	
TAS2R8	Forward	ATGTGGATTACCACCTGCCT	135
	Reverse	GGAAATGGCAAAGCATCCCAG	
TAS2R9	Forward	GCAGATTGCACTGCATGCTAC	70
	Reverse	TGCCTTTATGCCCTCATGT	
TAS2R10	Forward	GCTACGTGTAGTGGAAAGGCA	73
	Reverse	TCCATTCCCCAAAACCCCAA	
TAS2R13	Forward	GAAAGTGCCCTGCCGAGTAT	177
	Reverse	CCAGATCAGCCCAATTCTGGA	
TAS2R14	Forward	CCAGGTGATGGGAATGGCTTA	128
	Reverse	AGGGCTCCCCATCTTGAAC	
TAS2R16	Forward	ATGGCATCACTGACCAAGCA	255
	Reverse	TTTCAACGTAGGGCTGCTCA	
TAS2R19	Forward	TCTTAGGACACAGCAGAGCA	146
	Reverse	AGCGTGTCATCTGCCACAAAA	
TAS2R20	Forward	ATTGGGGAAACAAGACGCT	183
	Reverse	ACTACGGAAAAACTTGTGGAA	
TAS2R30	Forward	GGCTGGAAAAGCAACCTGTC	191
	Reverse	ACACAATGCCCTCTTGTGA	
TAS2R31	Forward	TTGAGGAGTGCAGTGTACCTTTC	218
	Reverse	ACGGCACATAACAAGAGGAAAA	
TAS2R38	Forward	CCCAGCCTGGAGGCCACATT	216
	Reverse	TCACAGCTCCTCAACTTGGCA	
TAS2R39	Forward	TTCTGTGGCTGTCCGTGTTA	207
	Reverse	GGGTGGCTGTCAGGATGAAC	
TAS2R40	Forward	CGGTGAACACAGATGCCACAGATA	150
	Reverse	GTGTTTGCCCCCTGGCCCACT	
TAS2R41	Forward	GCAGCGAACATGGCTTCATTGT	223
	Reverse	TGGCTGAGTTCAAGGAAGTGC	
TAS2R42	Forward	TCCTCACCTGCTTGGCTATC	161
	Reverse	GGCAAGCCAGGTGTCAAGT	
TAS2R43	Forward	ATATCTGGCAGTGATCAACC	148
	Reverse	CCCAACAAACATCACCGAGAATGAC	
TAS2R45	Forward	AGTACCCCTTACTGTAACCC	170
	Reverse	AGTAAATGGCACGTAACAAG	

Gene	Direction	Sequence (5' to 3')	Amplicon length, bp
TAS2R46	Forward	ACATGACTTGGAAAGATCAAAGTGAG	200
	Reverse	AGCTTTATGTGGACCTTCATGC	
TAS2R50	Forward	CGCAAGATCTCAGCACCAAGGTC	151
	Reverse	GCCTTGCTAACCATGACAACCGGG	
TAS2R60	Forward	GGTGTTCAGTGCTGCAGGTA	156
	Reverse	CACCTTGAGGAACGACGACT	
GAPDH	Forward	AGGTGGAGTCAACGGATTG	94
	Reverse	GGGGTCATTGATGGCAACAATA	
PPIA	Forward	CCACCAGATCATTCCCTCTGTAGC	144
	Reverse	CTGCAATCCAGCTAGGCATGG	

Figures

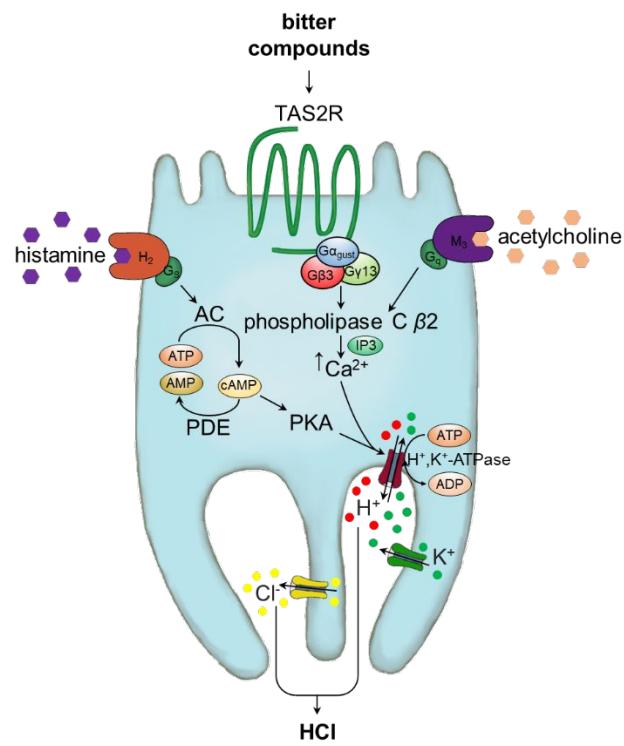


Figure-SI 1. Illustration of the proposed mechanism of proton secretion induced by bitter compounds in HGT-1 cells.^{5,56}

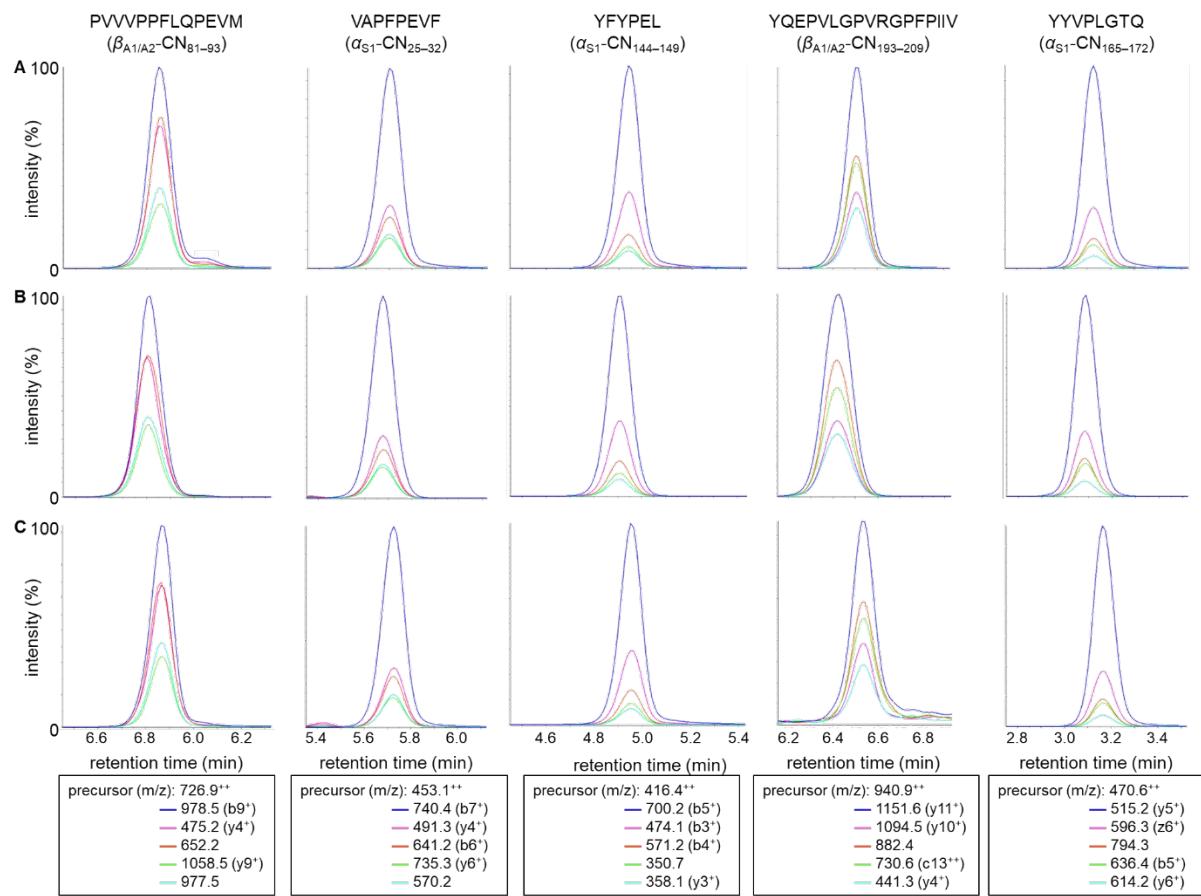


Figure-SI 2. Comparison of retention times (UPLC) and SRM mass transitions (MS/MS; 5 transitions per peptide; transitions identified with skyline 21.1.0.146) of PVVVPPFLQPEVM, VAPFPEVF, YFYPEL, YQEPVLGPVRGPFIIV and YYVPLGTQ as (A) synthesized standard, (B) formed in *in vitro* digestion, and (C) formed in *in vivo* digestion.

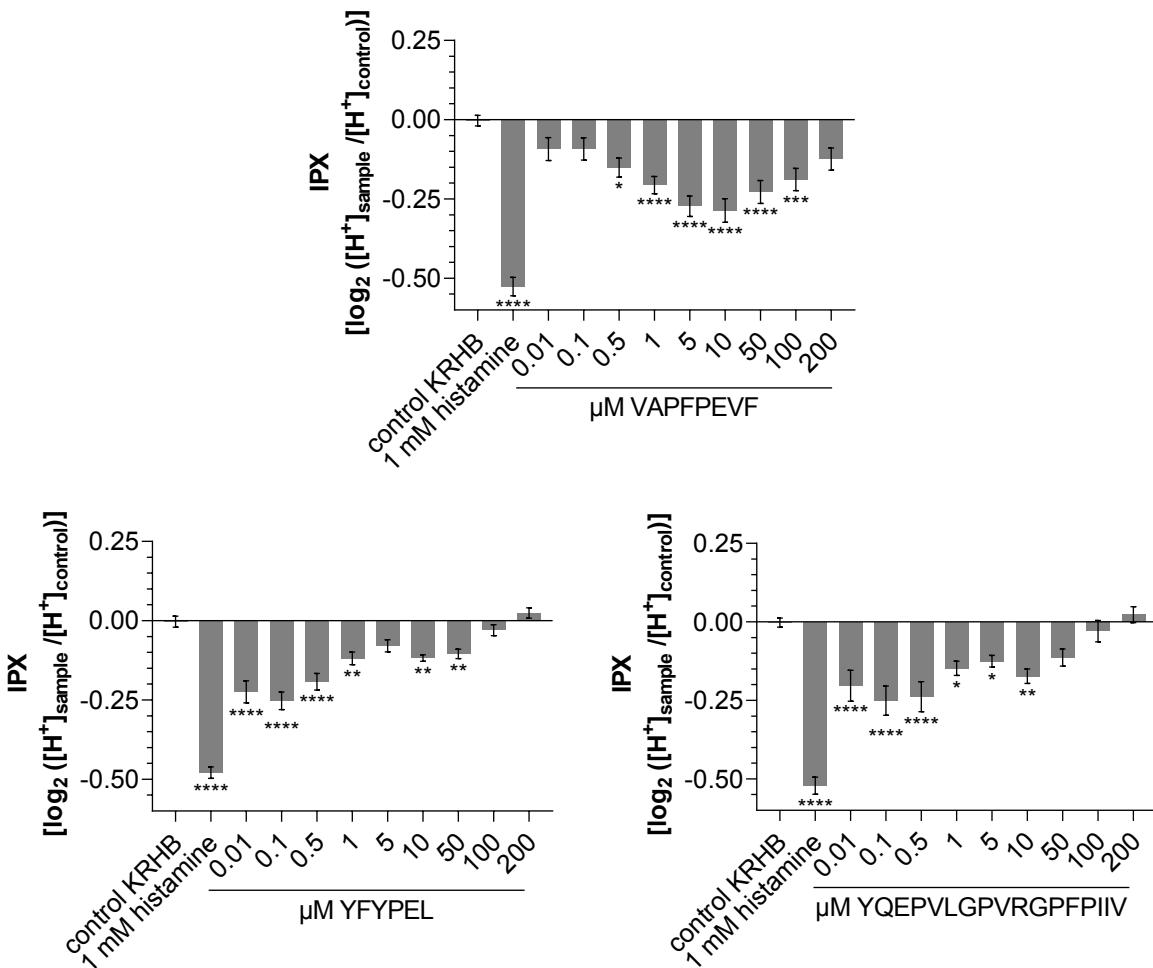


Figure-SI 3. Effect on proton secretion of HGT-1 cells incubated with VAPFPEVF (top) and the peptides YFYPEL and YQEPVLGPVRGPFPIIV with similar IPX profile (bottom). Data shown as mean \pm SEM after incubation for 10 minutes, $n = 4 - 8$, t. r. = 4 – 6, control: KRHB, Statistics: one-way ANOVA Holm-Šidák post hoc test; significant differences are expressed with * = $p \leq 0.05$, ** = $p \leq 0.01$, *** = $p \leq 0.001$, **** = $p \leq 0.0001$.

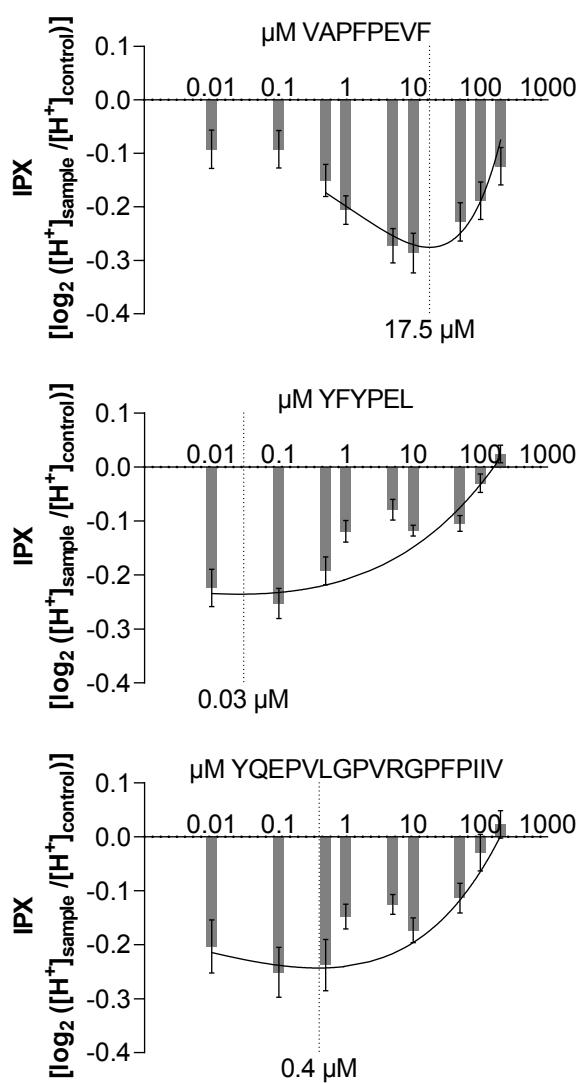


Figure-SI 4. Curve fit calculations of the proton secretion profile of the three peptides to determine which concentration has the greatest effect on proton secretion activity in HGT-1 cells in each case.

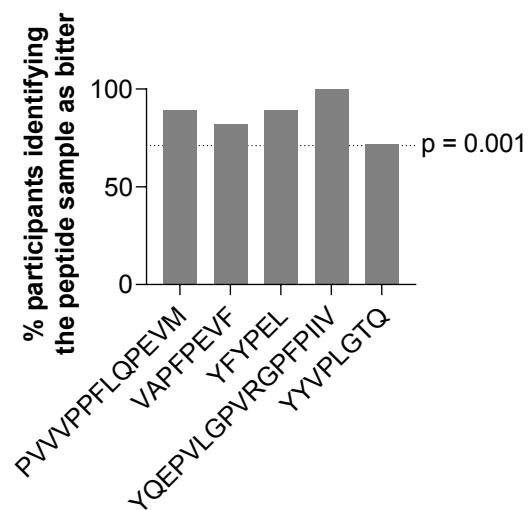


Figure-SI 5. Sensory experiments revealed significant bitterness ($p \leq 0.001$) toward water for all five selected peptides.

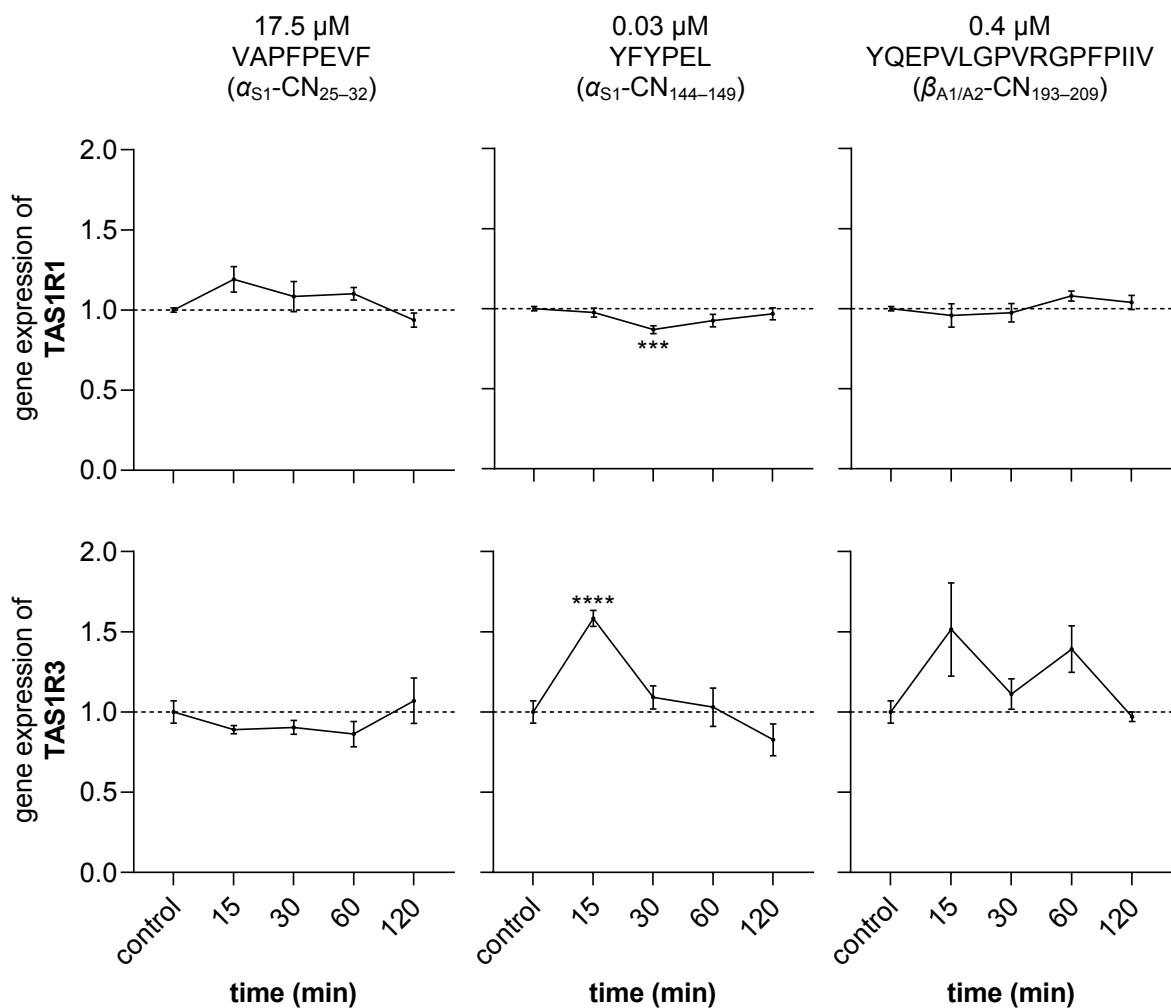


Figure-SI 6. Changes in gene expression (fold change) of taste receptors TAS1R1 (top) and TAS1R3 (bottom) as a function of incubation time with peptides VAPFPEVF (17.5 μM ; left), YFYPEL (0.03 μM ; center) and YQEPVLGPVRGPFPPIV (0.4 μM ; right). Normalized to the expression of PPIA and GAPDH (reference genes). Data shown as mean \pm SEM, n = 3, t. r. = 3, Statistics: t test Holm-Šidák method; significant differences are expressed with *** = p \leq 0.001, **** = p \leq 0.0001.

YQEPVLGPVRGPFP I I V

Figure-SI 7. Identified degradation products of YQEPVLGPVRGPFP I I V ($\beta_{A1/A2}$ -CN₁₉₃₋₂₀₉) found in the further course of digestion explaining the degradation of the peptide.

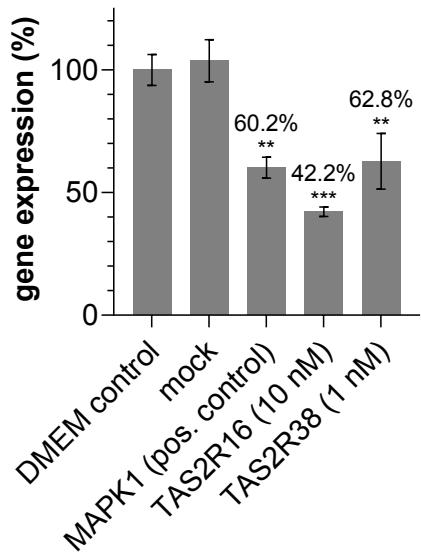


Figure-SI 8. Gene expression was reduced by transient transfection (knock-down) after 72 h.

Mock transfactions were performed with siRNA without target. Reduction in expression was best achieved with 10 nM siRNA targeting TAS2R16 (HSS121396) and 1 nM siRNA targeting TAS2R38 (HSS108754) to 42.2 and 62.8%, respectively.

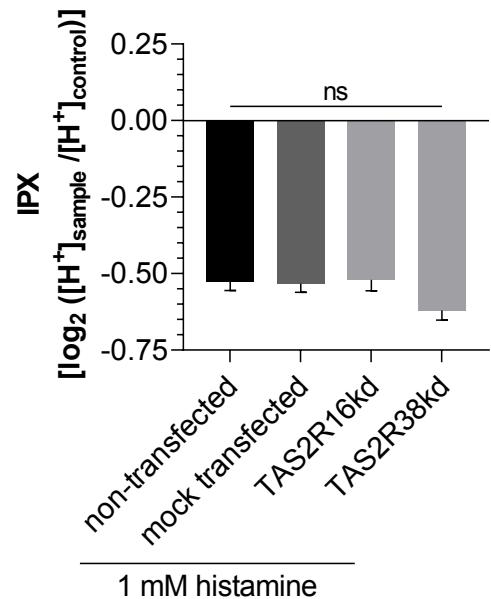


Figure-SI 9. TAS2R-independent histamine-induced stimulation of proton secretion does not differ from non-transfected cells in either mock-transfected or TAS2R16 or TAS2R38 knockdown HGT-1 cells.