Supporting Information

A MALDI-TOF assay identifies nilotinib as an inhibitor of inflammation in acute myeloid leukemia

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Table S1. List of compounds used in MALDI-TOF MS screen. The table indicates the drug name,CHEMBL ID, and molecular formula.

Table S2. TMT labels. The table indicates the sample name, TMT label reagent, and HCD monoisotopic report mass (Da).

Table S3. Proteins used to normalize TPP data. Document indicates description and data.

Table S4. TPP analysis. Document indicates description and data.

Table S5. Proteomics ANOVA analysis. Document indicates description and data.

 Table S6. Proteomics Limma analysis and Gene ontology data. Document indicates description,

 data, downregulated and upregulated gene ontology and KEGG enrichment analyses.

Table S7. MALDI-TOF MS, RTqPCR and ELISA prices for 96 drugs discovery assay and per sample. The table indicates the techniques (MALDI-TOF MS, RTqPCR and ELISA), company and reference number, price and units, price for 96 samples and price per sample.







1.0 5654.7 0.6 0.4 0.2 0.0 5657.7 5657.7 5657.7 5657.7 5657.7 5657.7 5657.7 5657.7 5657.7 5654.7 5657.7 5677.7 5777.



11313.0

DMSO





Figure S1. MALDI-TOF MS spectra. A) Representative MALDI-TOF MS spectra of untreated (resting) THP-1 cells and treated with 100 ng/mL LPS, 100 ng/mL Pam₂CSK₄, 100 ng/mL Pam₃CSK₄, 100 U/mL IFN-γ, 1 μg/mL poly(I:C) or 1 μg/mL poly(A:U) for 24 h. B) Representative MALDI-TOF MS spectra of THP-1 cells pre-treated with vehicle control (DMSO), 5 µM NG-25, 5 µM BI2536 or 1 µM MRT68601 for one hour before 100 ng/mL LPS-treatment for up to 24 h. C-D) Representative MALDI-TOF MS spectra of THP-1 cells pre-treated with DMSO, 5 µM nilotinib, 5 µM imatinib, 1 µM dasatinib, 1 µM bosutinib or 1 µM ponatinib C) for one hour before 100 ng/mL LPS-treatment for up to 24 h; D) for one hour before stimulation with live E. coli for up to 24 h. E-F) Representative MALDI-TOF MS spectra of THP-1 cells pre-treated with DMSO, 5 µM nilotinib, 5 µM imatinib, 1 µM losmapimod or 1 µM MRT68601 E) for one hour before stimulation with live E. coli for up to 24 h; F) for one hour before 100 ng/mL LPS-treatment for up to 24 h. G-H) Representative MALDI-TOF MS spectra of G) OCI-AML2 and H) NCI-H929 cells pre-treated with DMSO, 5 µM nilotinib or imatinib for one hour before stimulation with live E. coli for up to 24 h. I-K) Representative MALDI-TOF MS spectra of I) primary monocytes and J-K) primary AML cells pre-treated with DMSO, 5 µM nilotinib, 5 μM imatinib, 1 μM dasatinib, 1 μM bosutinib, 1 μM ponatinib or 1 μM losmapimod for one hour before 100 ng/mL LPS-treatment for up to 24 h. L and M) Represent XTT cell viability assays. Significant differences between two groups were determined by Mann-Whitney U-test. The statistical significance of the comparisons with resting is indicated as follows: **, $P \le 0.01$; *, $P \le 0.05$. Error bars represent the standard deviation of four biological replicates.



Figure S2. Nilotinib reduces IL-6 and IL-1ß secretion. A) IL-6 and **B)** IL-1ß secretion of THP-1 cells treated in either vehicle control-treated (DMSO), 100 ng/mL LPS-treated, or pre-treated with 5 μ M NG-25, BI2536 or 1 μ M MRT68601 for one hour before 100 ng/mL LPS-treatment for up to 24 h measured by ELISA. **C)** IL-6 and **D)** IL-1ß secretion of in THP-1 cells in vehicle control-treated (DMSO), 100 ng/mL LPS-treated, or pre-treated with 5 μ M nilotinib, imatinib, 1 μ M dasatinib, bosutinib or ponatinib for one hour before 100 ng/mL LPS-treatment for up to 24 h measured by ELISA. Significant differences between two groups were determined by a Mann-Whitney U-test. The statistical significance of the comparisons with LPS is indicated as follows: ns, not significant; **, $P \le 0.01$; ***, $P \le 0.001$. Error bars represent the standard deviation of four biological replicates.



Figure S3. MALDI-TOF MS screening quality control. A) *Z*' scores of biological replicates 1, 2 and 3 from the 96 compounds screened across the 8 staggered sets. **B**) Correlation plots of biological replicates 1, 2 and 3 showing correlation $R^2 > 0.80$ and $P \le 0.001$. **C**) MALDI-TOF MS IC50 curve of nilotinib- and imatinib-pre-treated cells one hour before 100 ng/mL LPS-treatment for up to 24 h. **D**) TNF- α secretion IC50 curve of nilotinib- and imatinib-treated THP-1 cells. Table below shows IC50 concentration (nM) and R². Error bars represent the SEM of six (**C**) or four (**D**) biological replicates.



Figure S4. TPP ratio heatmap. Heat map of protein thermal stabilities in THP-1 cells with 5 μ M nilotinib and imatinib for one hour before 100 ng/mL LPS-treatment for up to 15 min. The median relative abundance across replicates (n = 4) at the indicated temperature is shown for each protein as fold change relative to the lowest measured temperature (40 °C).

Figure S5



Figure S5. TPP analysis. A) Determination by TPP analysis of the thermostability of p38y (MAPK12), B) SH2 domain-containing protein 4A (SH2D4A), C) protein TSSC4, D) protein SET, E) putative phospholipase B-like 2 (PLBD2), F) Actin-related protein 2/3 complex subunit 5 (ARPC5), G) Mitogen-activated protein kinase kinase 4 (MKK4), H) Mitogen-activated protein kinase kinase 6 (MKK6) protein at the indicated temperatures with 5 µM nilotinib or imatinib for one hour before 100 ng/mL LPS-treatment for up to 15 min. Table insert shows melting temperature (Tm °C) and R². I-J) Determination of the thermostability of MKK3 at the indicated temperatures in THP-1 cells pre-treated with 5 µM nilotinib or imatinib for one hour before 100 ng/mL LPS-treatment for up to 15 min. J) Representative western blots of the thermostability of MKK3. GAPDH served as a loading control. Table insert shows melting temperature (Tm °C) and R². K-L) Determination of the thermostability of p38a (MAPK14) at 56°C in DMSO, 100 ng/mL LPS and pre-treated with the indicated concentrations of nilotinib or imatinib for one hour before 100 ng/mL LPS-treatment for up to 15 min. Table insert below shows effective concentration (EC50) (μ M) and R². M-N) Determination of the thermostability of p38a (MAPK14) at 56°C in 5 µM nilotinib or imatinib for one hour. Table insert shows melting temperature (Tm °C) and R². Error bars represent the SEM of four biological replicates. GAPDH served as a loading control. A representative image of four replicates is shown. Relative mobilities of reference proteins (masses in kilo Daltons) are shown on the left of each blot.



Figure S6. Nilotinib reduces the pro-inflammatory phenotype. A) MALDI-TOF MS relative quantitation of the ratio m/z 4964 / 4632 in THP-1 cells pre-treated with DMSO, 5 μ M nilotinib, imatinib, 1 μ M dasatinib, bosutinib or ponatinib for one hour before stimulation with live *E. coli* for up to 24 h. **B)** MALDI-TOF MS relative quantitation of the ratio m/z 4964 / 4632 in THP-1 cells pre-treated with DMSO, 5 μ M nilotinib, imatinib, 1 μ M losmapimod or MRT68601 for one hour before stimulation with LPS for up to 24 h. **C)** Relative quantification of four replicates (Figure 4B) is shown

on the right for p38 α and MK2 phosphorylation levels. **D**) Western blot analysis of the p38 α MAPK pathway in THP-1 cells pre-treated with DMSO, 5 μ M nilotinib, 5 μ M nilotinib, imatinib, 1 μ M dasatinib, bosutinib or ponatinib for one hour before stimulation with live *E. coli* for up to 15 min. α -tubulin serves as a loading control. A representative image with two biological replicates of four replicates is shown. Relative mobilities of reference proteins (masses in kilo Daltons) are shown on the left of each blot. Relative quantification is shown on the right for p38 α and MK2 phosphorylation levels. Error bars represent the standard deviation of four biological replicates. Significant differences between two groups were determined by a Mann-Whitney U-test. The statistical significance of the comparisons with *E. coli* is indicated as follows: ns, not significant; nd, not detected; *, $P \le 0.05$; **, $P \le 0.01$; ***, $P \le 0.001$.



Figure S7. Nilotinib inhibits the pro-inflammatory phenotype in *E. coli*-activated monocytes. A-C) ELISA IC50 curves of A) TNF-α, B) IL-6, and C) IL-1β were measured by ELISA at 24 h. Error bars represent the standard deviation of four biological replicates. Significant differences between two groups were determined by a Students t-test. The statistical significance of the comparisons with E. coli is indicated as follows: ns, not significant; **, $P \le 0.01$; ***, $P \le 0.001$. Table insert shows IC50 (nM) and R^2 . D) *TNF-a*, and E) *IL-6* expression were determined by RT-qPCR in THP-1 cells pre-treated with DMSO, 5 µM nilotinib or imatinib for one hour before stimulation with live *E. coli* for up to 24 h. The results were analyzed using the $2^{-\Delta\Delta Ct}$ method and normalized using GAPDH and TBP as the reference genes; and DMSO as the reference sample. Error bars represent the standard deviation of four biological replicates. The statistical significance of the comparisons with E. coli is indicated as follows: ns, not significant; **, $P \le 0.01$; ***, $P \le 0.001$. F) IL-6 expression was determined by RT-qPCR in non-transfected or transfected HEK293 cells with p38a WT or p38a M107T for 48 h and pre-treated with DMSO, 5 µM nilotinib or imatinib for one hour before stimulation with LPS for up to 24 h. The results were analyzed using the $2^{-\Delta\Delta Ct}$ method and normalized using *GAPDH* and *TBP* as the reference genes; and DMSO as the reference sample. The statistical significance of the comparisons with: LPS non-transfected is indicated as ‡; LPS transfected with p38a WT is indicated as †; LPS transfected with p38a T106M is indicated as \$. The statistical significance of the comparisons between p38a WT and p38 α M107T is indicated as follows: ns, not significant; ***, $P \leq 0.001$. Error bars represent the standard deviation of four biological replicates. Significant differences between two groups were determined by Mann-Whitney U-test.



Figure S8. Assessment of TKIs against p38 α using surface plasmon resonance (SPR). (A-G) Representative multicycle SPR sensograms for A) nilotinib, B) ponatinib, C) dasatinib, D) bosutinib, E) imatinib, and F) MRT68601showing a dose-dependent concentration series against immobilized p38 α . K_D , equilibrium dissociation rate constant; K_{on} (M⁻¹ s⁻¹), on-rate constant or association reaction; K_{off} (s⁻¹), off-rate constant or dissociation reaction; Kd (affinity fit), dissociation rate constant. Experiments performed in triplicate.

0

-8

-4 -2 0 2 4 Log₂ Fold change



⊲α-tubulin

25 55

8

Figure S9. Differences in the proteome of nilotinib- and imatinib-treated monocytes. A) The matrix of correlation plots revealed very high correlations between \log_2 transformed LFQ intensities (Pearson correlation coefficient between 0.982 and 0.998). The color code indicates the values of correlation coefficients. **B)** Volcano plot of THP-1 cells treated with 5 µM nilotinib *vs. E. coli*, cut-off of FDR <0.05 and 1.5-fold change between conditions. **C)** MMP9 and p38 α levels in THP-1 pre-treated with 5 µM nilotinib or imatinib for one hour before stimulation with live *E. coli* for up to 24 h. α -tubulin serves as a loading control. A representative image with two biological replicates of four replicates is shown. Relative mobilities of reference proteins (masses in kilo Daltons) are shown on the left of each blot.



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Figure S10. Nilotinib and imatinib show different sensitivity in blood malignancies. Nilotinib reduces the inflammation phenotype in OCI-AML2 and NCI-H929 cell line. A) IC50 values of nilotinib and imatinib in various types of cancer, and B) in hematological malignancies according to TCGA classification from the Genomics of Drug Sensitivity in Cancer (GDSC) database. C) MALDI-TOF MS relative quantitation of the ratio m/z 4964/4632 in NCI-H929 cells pre-treated with DMSO, 5 μ M nilotinib or imatinib for one hour before stimulation with live E. coli for up to 24 h. D) p38 α MAPK pathway status in NCI-H929 cells. α -tubulin was detected as a loading control. A representative image of four replicates is shown. Relative mobilities of reference proteins (masses in kilo Daltons) are shown on the left of each blot. E) TNF- α , IL-6, and IL-1 β secretion was measured by ELISA at 24 h. Error bars represent the SEM of four biological replicates. Significant differences between two groups were determined by a Mann-Whitney U-test. The statistical significance of the comparisons with *E. coli* is indicated as follows: ns, not significant; na, not applicable; *, $P \le 0.05$; **, $P \le 0.01$; ***, $P \le 0.001$.