

Figure S1. (A) $Ccn1^{WT/WT}$ liver macrophages isolated 48 hrs after injection of vehicle or CCl_4 were cultured overnight and stained with anti-CD68 and anti-F4/80 antibodies and counterstained with DAPI. (B) $Ccn1^{WT/WT}$ liver macrophages isolated 48 hrs after injection of vehicle were co-stained with anti-CD206 antibody and anti-Galectin-3 antibody or anti-CD206 antibody and anti-CD11b antibody. The percentages of positive cells were quantified (*n=4*). Data represents means ± SD. Scale bars: 100 µm.



Figure S2. $Ccn1^{WT/WT}$ and $Ccn1^{D125A/D125A}$ mice were i.p. injected with $CCl_{4.}$ and Ccn1 (A) or Ccn2 (B) expression was analyzed by qRT-PCR in liver RNA isolated at the indicated times (*n*=6). (C) Liver sections from $Ccn1^{WT/WT}$ and $Ccn1^{D125A/D125A}$ mice after injection with CCl_4 for indicated times were stained with anti-CCN1 antibody. (D) Immunoblots of protein extracts from the indicated livers were probed with antibodies against CCN1 and β -actin. Scale bars: 100 µm.



Figure S3. (A) Liver sections from *Ccn1^{WT/WT}* and *Ccn1^{D125A/D125A}* mice fed DDC diet for 4 weeks or subjected to BDL 7 days were immunostained for α -smooth muscle actin (α SMA, green) and counterstained with DAPI (*n*=6) or stained with Sirius Red to reveal collagen deposition. The fibrotic areas were measured by Image J. The percentages of fibrotic areas were assessed by Image J analysis of microphotographs of six randomly selected fields. (B) Expression of *Tgfb1* in *Ccn1^{WT/WT}* and *Ccn1^{D125A/D125A}* mice livers which were subjected to BDL for 7 days was assessed by qRT-PCR (*n*=4). Data represent means ± SD. **P*<0.033, ***P*<0.002. Scale bars: 100 µm.



Figure S4. (A) Serum ALT levels were measured 48 hrs after CCI_4 injection (*n*=6). (B) Macrophages were isolated from injured livers and expression of indicated genes was assessed by qRT-PCR (*n*=6). Data represents means ± SD.**P*<0.033.



Figure S5. (A) $Ccn1^{D125A/D125A}$ mice were injected with CCl₄, followed by *i.p.* injections of either vehicle or CCN1 protein (1 mg/kg) at 4, 24, and 40 hrs thereafter. Livers were collected at 48 hrs after CCl₄ injection, and liver sections were stained with anti-Ly6G or anti-MPO antibody as indicated and counterstained with hematoxylin (blue). (B) $Ccn1^{WT/WT}$ and $Ccn1^{D125A/D125A}$ hepatic macrophages isolated 48 hrs after injection of CCl₄ were co-stained with CD206 and Galectin-3 or CD206 and CD11b. The percentages of positive cells were quantified (*n*=4). **P*<0.033, ***P*<0.002, ****P*<0.001. Data represents means ± SD. Scale bars: 100 µm.



Figure S6. Bone marrow derived macrophages (BMMs) from $Ccn1^{WTWT}$ and $Ccn1^{D125A/D125A}$ mice were incubated for 4 hrs with their own apoptotic neutrophils, which were pre-treated with their own type of CCN1. (A) The expression of cd206 and Tgfb1 were assessed by qRT-PCR (n=4). (B) HSCs isolated from uninjured $Ccn1^{WT/WT}$ and $Ccn1^{D125A/D125A}$ mice were co-cultured for 5 days with BMMs from $Ccn1^{WT/WT}$ and $Ccn1^{D125A/D125A}$ mice were co-cultured for 5 days with BMMs from $Ccn1^{WT/WT}$ and $Ccn1^{D125A/D125A}$ mice with or without apoptotic PMN and stained with anti- α SMA antibody. (C) The expression of α SMA was assessed by qRT-PCR (n=4). Data represents means ± SD. *P<0.033, **P<0.002. Scale bars: 100 µm.



Figure S7. HSCs isolated from $Ccn1^{WT/WT}$ mice 48 hrs after injection with CCI_4 were grown in culture. (A) Integrin α_6 mRNAs level and (B) Integrin α_v mRNAs level were measured by qRT-PCR (*n*=4). Data represents means ± SD.

Gene	Orientation	Sequence (5' to 3')
αSMA	sense	ctgacagaggcaccactgaa
	antisense	gaaggaatagccacgctcag
Tgf β 1	sense	gttttgaccccgaggagc
	antisense	gagaagagagcgcagaatcca
Ccn1	sense	ggaactggcatctccaca
	antisense	tttgggccggtatttctt
Ccn2	sense	taggccctcagcctcact
	antisense	cttgacaggcttggggat
Cyclophilin	sense	ttcacaaaccacaatggcacaggg
	antisense	tgccgtccagccaatctgtcttat
	sense	ccaagggtaacagcggtgaa
Col1a1	antisense	cctcgttttccttcttctccg
	sense	taacctggatgccgtcgt
Mmp2	antisense	ttcaggtaataagcacccttgaa
	sense	cgtcgtgatccccacttact
Mmp9	antisense	aacacacagggtttgccttc
Mmp13	sense	agttgacaggctccgagaaa
	antisense	ggcactccacatcctggttt
Pdgfa	sense	gtgcgacctccaacctga
	antisense	ggctcatctcacctcacatct
	sense	cggcctgtgactagaagtcc
Pdgf β	antisense	gagcttgaggcgtcttgg
Egf	sense	agaaacaccaagaccccaag
	antisense	tgtgcccattccatctatgtg
lgf	sense	accgaggggcttttacttca
	antisense	tggctcacctttccttctcc

Table S1. Primer sequences used in this study.

Elisa kit	Company and catalog number		
TGF-β1	BioLegend (436707)		
Free active TGF-β1	BioLegend (437707)		

Table S2. Elisa kits used in this study.

Antibody	dilution	Company and catalog number
α-SMA	1:200	abcam (ab7817)
Ly6G	1:300	BD pharmigen (551460)
Cleaved-Caspase3	1:300	Cell signaling (9661S)
CD206	1:500	Cell signaling (24595T)
Galectin-3	1:300	Cell signaling (89572S)
CD11b	1:300	Cell signaling (17800S)
F4/80	1:300	Bio-Rad (MCA497RT)
МРО	1:150	invitrogen (PA5-16672)
CD68	1:200	Abcam (ab53444)

Table S3. Antibodies used in staining.

Antibody	dilution	Company and catalog number
Anti-Phospho-Smad2	1:1000	Cell signaling (18338)
anti-Smad2	1:1000	Cell signaling (5339)
anti-CCN1	1:800	R&D Systems (AF4055)
anti-β-actin	1:20000	Abcam (ab8226)

Table S4. Antibodies used in Western blots