Biophysical Journal, Volume 121

Supplemental information

Comparative evaluation of spin-label modeling methods for protein

structural studies

Maxx H. Tessmer, Elizabeth R. Canarie, and Stefan Stoll

Comparative Evaluation of Spin Label Modeling Methods for Protein Structural Studies

Maxx Tessmer, Elizabeth R. Canarie, Stefan Stoll

Department of Chemistry, University of Washington, Seattle, Washington 98195, USA

Supporting Information

Figure S1: Spin label skeletal diagrams.

Figure S2. X-band CW EPR of all nine site pairs (rows) and all five labels (columns) in the presence (red) and absence (blue) of maltose.

Figure S3. X-band CW EPR of WT MBP spin labeled compared to the CW spectra of the double mutants with the lowest overall signal (double integral) for each label.

Figure S4. DEER time domain signals and fits. Raw data is shown as gray dots. Apo fits are shown in blue and holo fits are shown in red with 95% confidence intervals shown as transparent bands of the same. Background fits are shown as gray lines with 95 % confidence intervals shown as gray transparent bands

Figure S5. Effect of glycerol concentration on MBP conformation. Top. DEER Traces of MBP S238R1 L275R1 5 mM Maltose, the construct with the largest dynamic range, in 20% and 50% glycerol. Bottom. Comparison of the distance distributions of the same, showing nearly identical distributions.

Figure S6. DEER distance distribution predictions of spin label modeling methods for R1 labeled site pairs (left). Simulated time domain signals of distance distributions predictions overlayed on experimental data (right). Time domain signal modulation depth and background were fit to optimize the root mean squared deviation of the simulated and experimental data.

Figure S7. DEER distance distribution predictions of spin label modeling methods for R7 labeled site pairs (left). Simulated time domain signals of distance distributions predictions overlayed on experimental data (right). Time domain signal modulation depth and background were fit to optimize the root mean squared deviation of the simulated and experimental data.

Figure S8. DEER distance distribution predictions of spin label modeling methods for V1 labeled site pairs (left). Simulated time domain signals of distance distributions predictions overlayed on experimental data (right). Time domain signal modulation depth and background were fit to optimize the root mean squared deviation of the simulated and experimental data.

Figure S9. DEER distance distribution predictions of spin label modeling methods for M1 labeled site pairs (left). Simulated time domain signals of distance distributions predictions overlayed on experimental data (right). Time domain signal modulation depth and background were fit to optimize the root mean squared deviation of the simulated and experimental data.

Figure S10. DEER distance distribution predictions of spin label modeling methods for I1 labeled site pairs (left). Simulated time domain signals of distance distributions predictions overlayed on experimental data (right). Time domain signal modulation depth and background were fit to optimize the root mean squared deviation of the simulated and experimental data.

Figure S12. Extended comparison of prediction Kullback**–**Leibler divergence (KLD, Simulated||Experimental) . A) Violin plots comparing all methods and all spin labels. Dots indicate individual KLDs, and horizontal lines indicate the mean KLD of the group. B) Scatterplot shows difference between off-rotamer sampling and other methods. Shaded bands are a density estimate of all data points in the group, indicating the characteristics of the distribution of the data. C) Histograms of the change in log(KLD) between modeling methods for each site pair. Colors indicate which method performed better for the bin. Numbers indicate the number of site pairs that showed improvement over the compared method.

Figure S13. Extended comparison of prediction earth-movers distance (EMD) . A) Violin plots comparing all methods and all spin labels. Dots indicate individual EMDs, and horizontal lines indicate the mean EMD of the group. Shaded bands are a density estimate of all data points in the group, indicating the characteristics of the distribution of the data. B) Scatterplot shows difference between off-rotamer sampling and other methods. C) Histograms of the change in EMDs between modeling methods for each site pair. Colors indicate which method performed better for the bin. Numbers indicate the number of site pairs that showed improvement over the compared method.

Figure S14. Extended comparison of prediction mean absolute deviation (MAD). A) Violin plots comparing all methods and all spin labels. Dots indicate individual MADs, and horizontal lines indicate the mean MAD of the group. Shaded bands are a density estimate of all data points in the group, indicating the characteristics of the distribution of the data B) Scatterplot shows difference between off-rotamer sampling and other methods. C) Histograms of the change in MAD between modeling methods for each site pair. Colors indicate which method performed better for the bin. Numbers indicate the number of site pairs that showed improvement over the compared method.

Figure S15. Comparison of clash evaluation potentials

Figure S16. Polar histograms of spin label model dihedral angles for R1. Rows group the spin label modeling method and columns group the dihedral angels. Dihedral definitions for R1 are as follows (χ₁: N-CA-CB-SG, χ2: CA-CB-SG-SD, χ3:CB-SG-SD-CE, χ4: SG-SD-CE-C3, χ5: SD-CE-C3-C4) using the MMM atom names.

Figure S17. Polar histograms of spin label model dihedral angles for R7. Rows group the spin label modeling method and columns group the dihedral angels. Dihedral definitions for R7 are as follows $(\chi_1: N$ -CA-CB-SG, χ_2 : CA-CB-SG-S2, χ_3 :CB-SG-S2-C4, χ_4 : SG-S2-C4-C5, χ_5 : S2-C4-C5-C6) using the MMM atom names.

Figure S18. Polar histograms of spin label model dihedral angles for V1. Rows group the spin label modeling method and columns group the dihedral angels. Dihedral definitions for V1 are as follows (χ₁: N-CA-CB-SG, χ₂: CA-CB-SG-S2, χ₃:CB-SG-S2-C4, χ₄: SG-S2-C4-N2) using the MMM atom names.

Figure S19. Polar histograms of spin label model dihedral angles for M1. Rows group the spin label modeling method and columns group the dihedral angels. Dihedral definitions for M1 are as follows $(\chi_1:$ N-CA-CB-SG, χ₂: CA-CB-SG-C4, χ₃:CB-SG-C4-C5, χ₄: C5-N2-C8-C9) using the MMM atom names.

Figure S20. Polar histograms of spin label model dihedral angles for I1. Rows group the spin label modeling method and columns group the dihedral angels. Dihedral definitions for I1 are as follows (χ₁: N-CA-CB-SG, χ_2 : CA-CB-SG-CD, χ_3 :CB-SG-CD-CE, χ_4 : SG-CD-CE-NZ, χ_5 : CD-CE-NZ-C3, χ_6 : CE-NZ-C3-C2) using the MMM atom names.

Description of metrics

For all metrics described below, *p(r)* and *q(r)* are the probability distribution functions being compared, *P(r)* and *Q(r)* are their respective cumulative distribution functions and *ε* is the machine epsilon. All metrics used for comparing distributions were computed as follows

Overlap

$$
\\nverlap = \sum_{r} \min(p(r_i), q(r_i))
$$

Earth Mover's Distance (Wasserstein distance)

$$
EMD = \sum_{r} |P(r_i) - Q(r_i)|
$$

Kullback–Leibler divergence

$$
KLD = \sum_{r} \max(p(r_i), \varepsilon) \log \left(\frac{\max(p(r_i), \varepsilon)}{\max(q(r_i), \varepsilon)} \right)
$$

Mean Absolute Deviation

$$
MAD = \left| \sum_{r} r_i p(r_i) - \sum_{r} r_i q(r_i) \right|
$$

Table S1. List of DEER parameters.

^a Fitted modulation depth of the experimental DEER trace.

^b Semiquantitative labeling efficiency on a with * being the lowest efficiency and ***** being the highest.

c Signal to noise ratio.

^d four pulsed DEER pump pulse time increment.

