

Original Papers

Medicinal Plants for the Treatment of Mental Diseases in Pregnancy: An *In Vitro* Safety Assessment

Deborah Spiess^{1,2,*}, Moritz Winker^{3,4,*}, Antoine Chauveau², Vanessa Fabienne Abegg², Olivier Potterat², Matthias Hamburger², Carsten Gründemann^{3,4,**}, Ana Paula Simões-Wüst^{1,**}

Affiliations

¹ Department of Obstetrics, University Hospital Zurich, University of Zurich, Zurich, Switzerland

² Division of Pharmaceutical Biology, Department of Pharmaceutical Sciences, University of Basel, Basel, Switzerland

³ Center for Complementary Medicine, Institute for Infection Prevention and Hospital Epidemiology, University of Freiburg, Faculty of Medicine, Freiburg, Germany

⁴ Translational Complementary Medicine, Department of Pharmaceutical Sciences, University of Basel, Basel, Switzerland

* These authors contributed equally to the work and should be considered as joint first authors.

** These authors contributed equally to the work and should be considered as joint last authors.

Correspondence

PD Dr. Ana Paula Simões-Wüst

University Hospital of Zurich

Department of Obstetrics

Schmelzbergstrasse 12/PF 125, Path G51a

8091 Zurich

Switzerland

Phone: +41 44 255 51 31

Fax: +41 44 255 44 30

anapaula.simoes-wuest@usz.ch

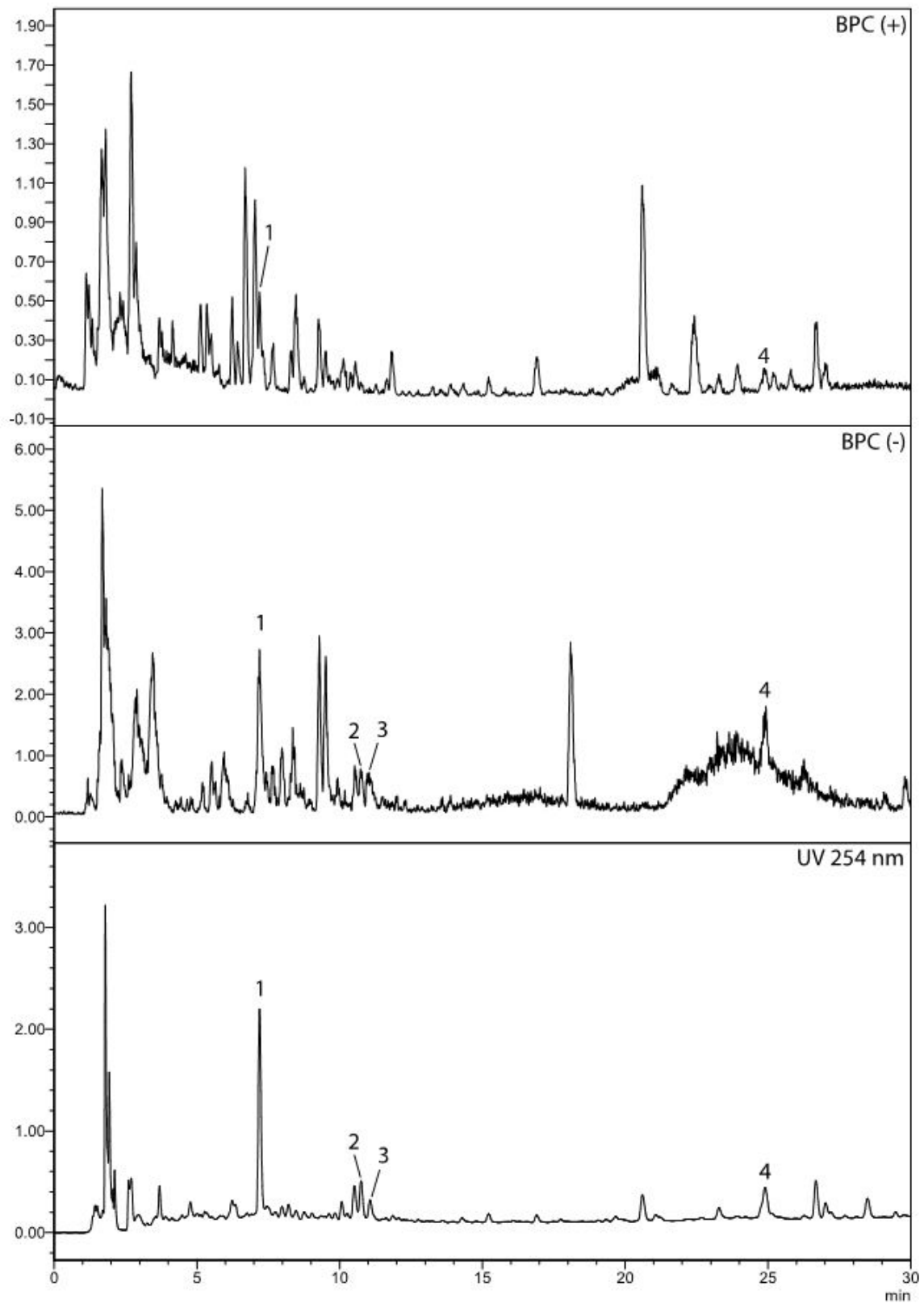


Fig. 1S. HPLC-UV-MS analysis of *Valeriana officinalis* 70% EtOH extract. BPC: base peak chromatogram. **1:** chlorogenic acid, **2** and **3:** dicaffeoylquinic acid isomers (tentative assignment), **4:** valerenic acid.

Table 1S. Retention times and spectroscopic data of annotated peaks in *Valeriana officinalis* 70% EtOH extract.

Peak number	Retention time (min)	UV maxima (nm)	ESI MS (+) (m/z)	ESI MS (-) (m/z)	Identification
1	7.2	206, 310, 327	354.9 [M+H] ⁺	353.1 [M-H] ⁻	Chlorogenic acid
2	10.8	209, 310, 327	516.9 [M+H] ⁺ 499.0	515.1 [M-H] ⁻	Dicaffeoylquinic acid
3	11.1	217, 310, 327	516.9 [M+H] ⁺ 499.0	515.1 [M-H] ⁻	Dicaffeoylquinic acid
4	24.9	222	235.1 [M+H] ⁺	233.1 [M-H] ⁻	Valerenic acid



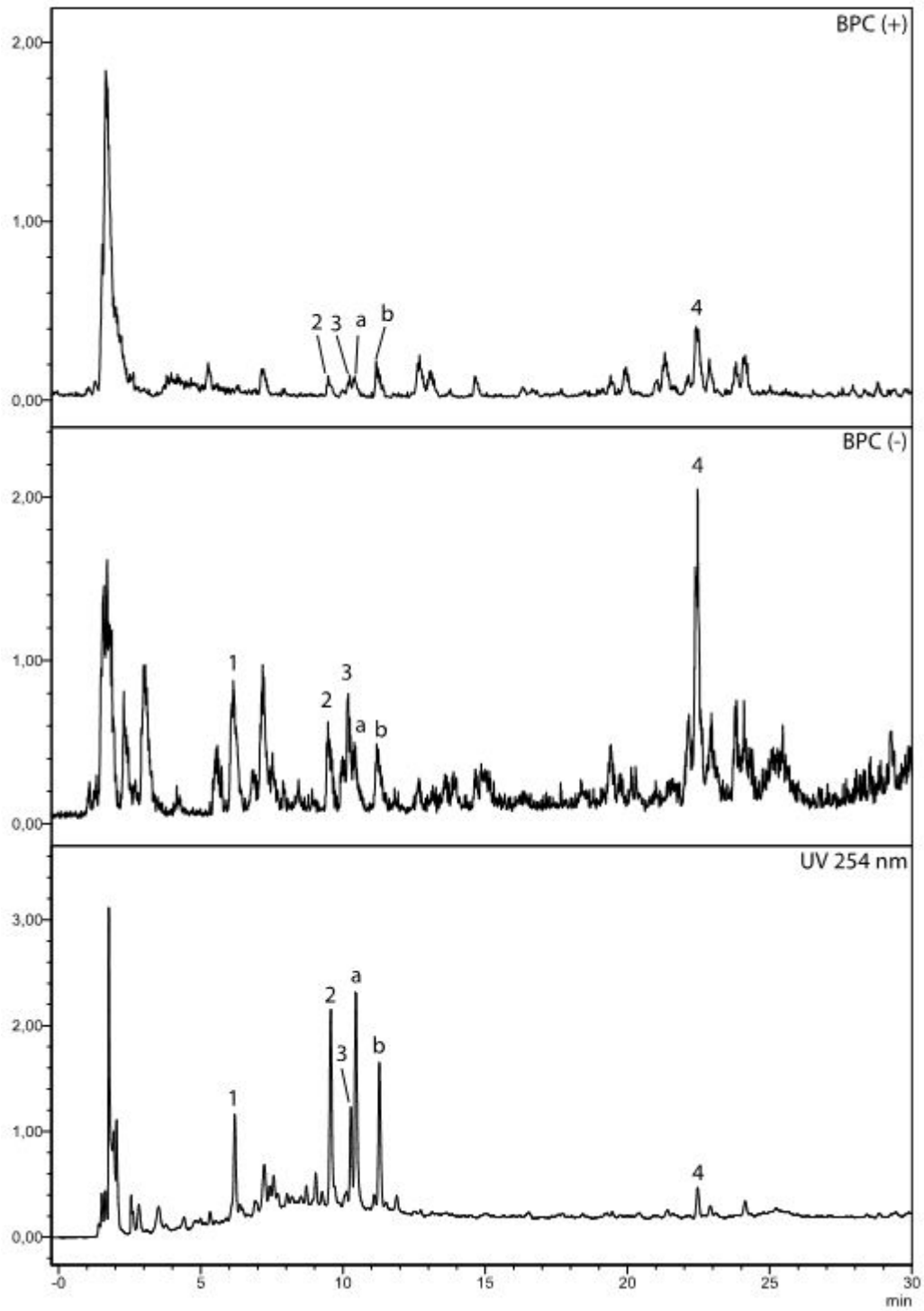


Fig. 2S. HPLC-UV-MS analysis of *Humulus lupulus* 70% EtOH extract. BPC: base peak chromatogram. **1:** chlorogenic acid, **2:** isoquercitrin, **3:** kaempferol hexoside (tentative assignment), **4:** xanthohumol, **a:** unidentified flavonoid, MW 550 amu, **b:** unidentified flavonoid, MW 534 amu.

Table 2S. Retention time and spectroscopic data of annotated peaks in *Humulus lupulus* 70% EtOH extract.

Peak number	Retention time (min)	UV maxima (nm)	ESI MS (+) (m/z)	ESI MS (-) (m/z)	Identification
1	6.2	203, 300, 325	354.9 [M+H] ⁺	353.1 [M-H] ⁻	Chlorogenic acid
2	9.6	203, 255, 353	465.1 [M+H] ⁺	463.0 [M-H] ⁻	Isoquercitrin
3	10.3	201, 265, 343	448.9 [M+H] ⁺ 286.7	447.0 [M-H] ⁻	Kaempferol hexoside
4	22.5	222, 369	354.9 [M+H] ⁺	353.0 [M-H] ⁻	Xanthohumol
a	10.5	204, 255, 353	550.8 [M+H] ⁺	549.2 [M-H] ⁻	Unidentified flavonoid
b	11.3	205, 265, 346	534.9 [M+H] ⁺	533.0 [M-H] ⁻	Unidentified flavonoid

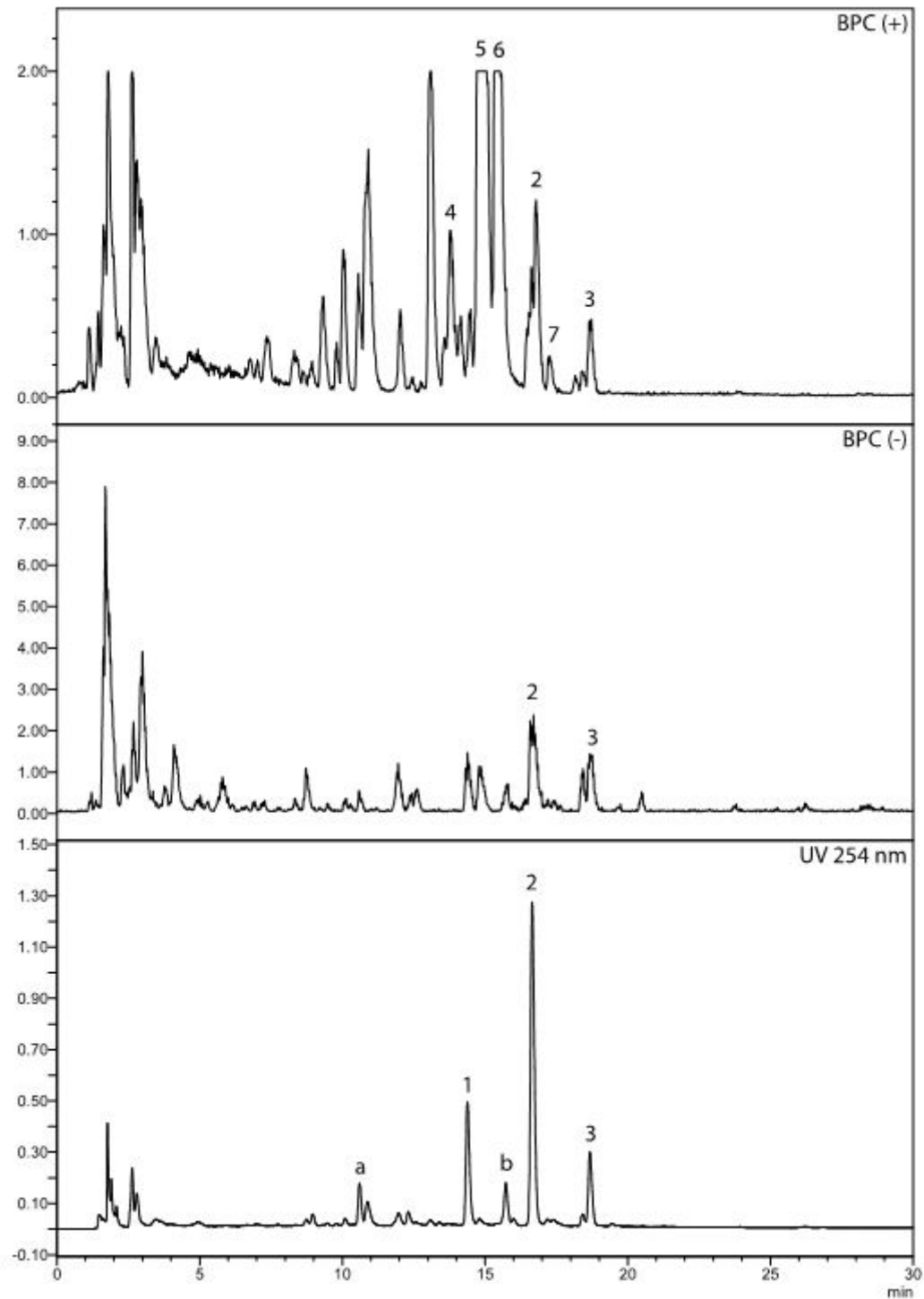


Fig. 3S. HPLC-UV-MS analysis of *Eschscholzia californica* 70% EtOH extract. BPC: base peak chromatogram. **1:** rutin rhamnoside (tentative assignment), **2:** rutin, **3:** rhamnoside of *O*-methyl quercetin (tentative assignment), **4:** protopine, **5:** californidine, **6:** escholzine, **7:** neocaryachine-7-*O*-methyl ether *N*-metho salt (CAS-Nr. 1310405-84-5, tentative assignment), **a:** unidentified flavonoid, MW 772 amu, **b:** unidentified flavonoid, MW 624 amu.

Table 3S. Retention time and spectroscopic data of annotated peaks in *Eschscholzia californica* 70% EtOH extract.

Peak number	Retention time (min)	UV maxima (nm)	ESI MS (+) (m/z)	ESI MS (-) (m/z)	Identification
1	14.4	219, 255, 353	757.1 [M+H] ⁺	755.1 [M-H] ⁻	Rutin rhamnoside
2	16.7	216, 255, 353	611.0 [M+H] ⁺	609.1 [M-H] ⁻	Rutin
3	18.7	204, 254, 353	625.1 [M+H] ⁺	623.1 [M-H] ⁻	Rhamnoside of O-methyl quercetin
4	13.8	* OV	354.0 [M+H] ⁺	-	Protopine
5	14.9	* OV	337.9 [M+H] ⁺	-	Californidine
6	15.5	* OV	324.0 [M+H] ⁺	-	Escholzine
7	17.3	* OV	340.0 [M+H] ⁺	-	Neocaryachine-7-O-methyl ether N-metho salt
a	10.6	202, 255, 353	773.0 [M+H] ⁺	771.1 [M-H] ⁻	Unidentified flavonoid
b	18.7	204, 254, 353	625.1 [M+H] ⁺	623.1 [M-H] ⁻	Unidentified flavonoid

* UV spectrum overlapped by coeluting flavonoids.

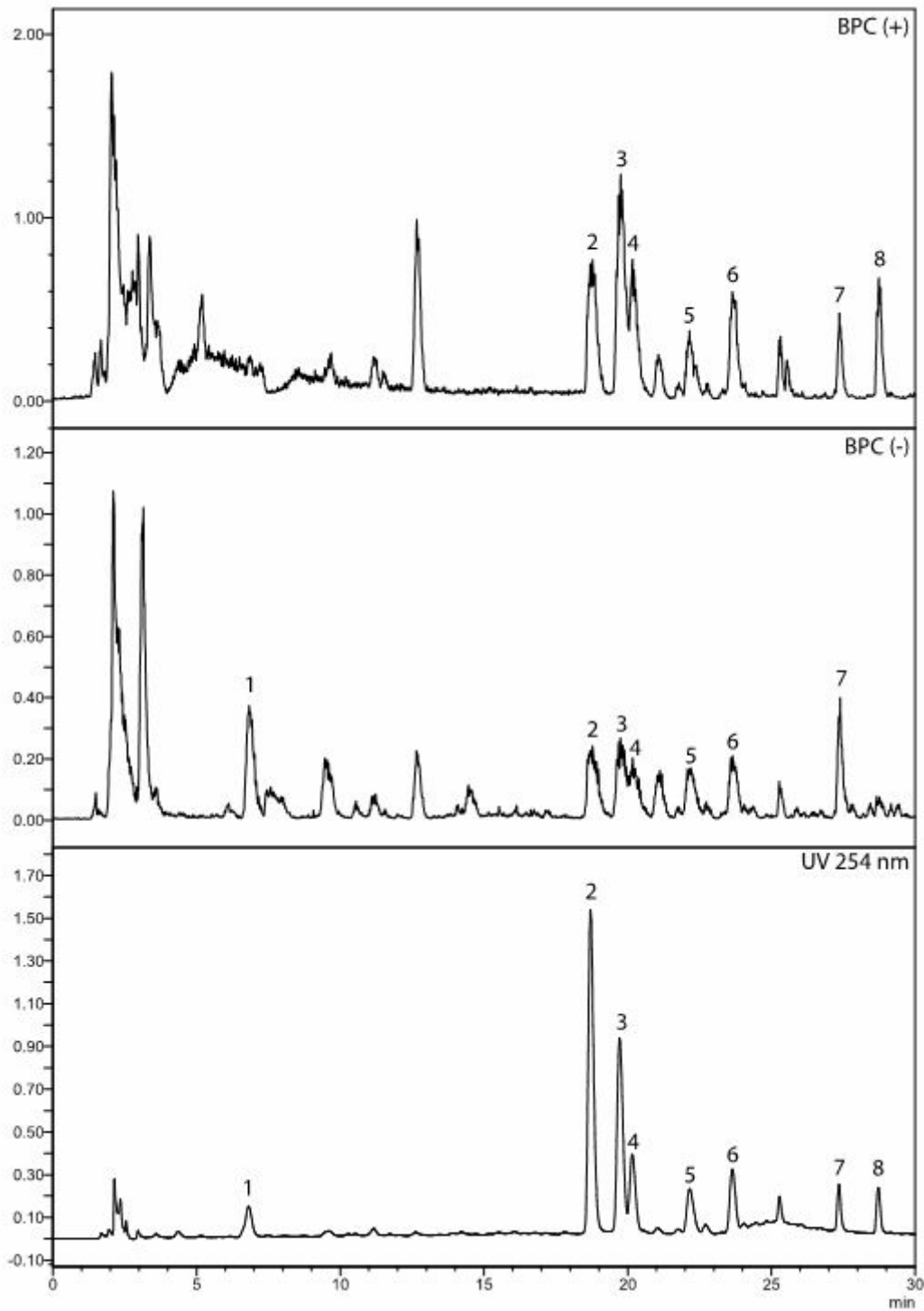
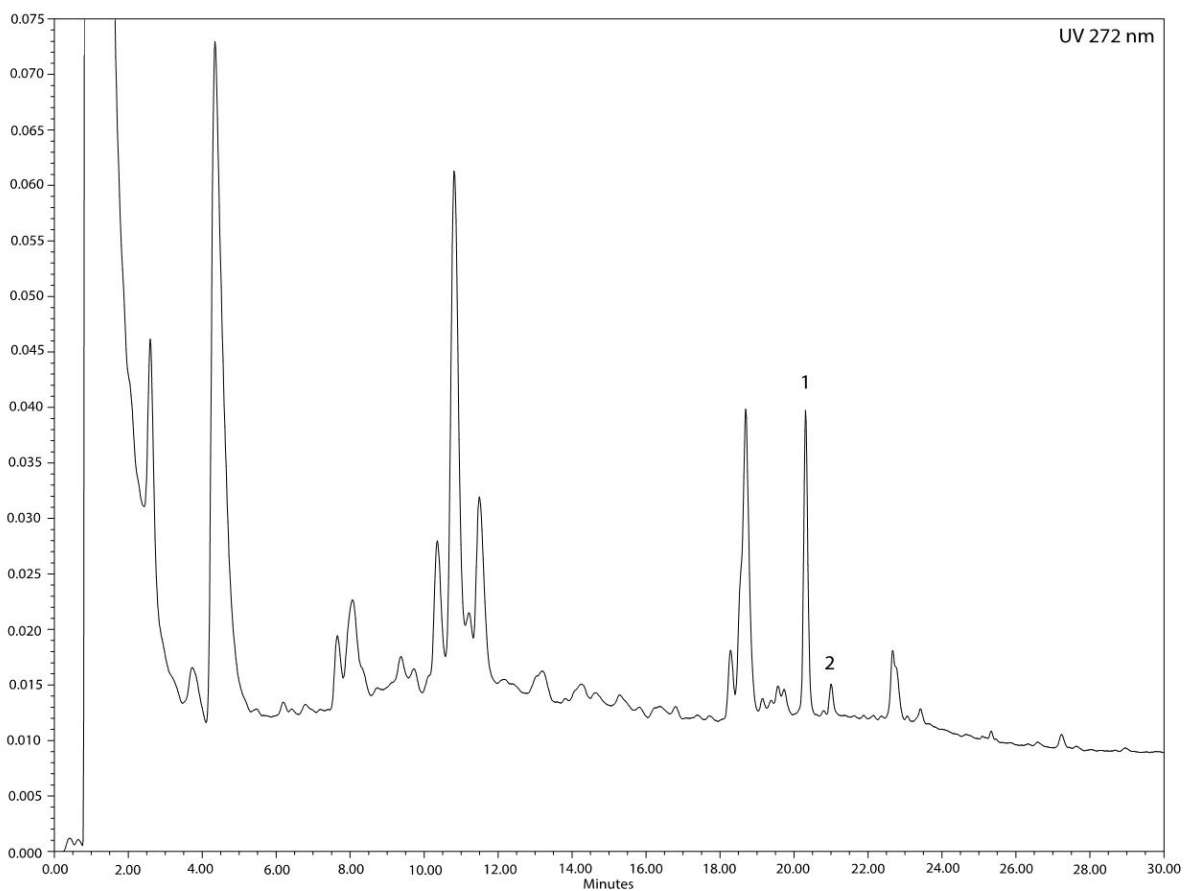


Fig. 4S. HPLC-UV-MS analysis of *Hypericum perforatum* 70% EtOH extract. BPC: base peak chromatogram. **1:** chlorogenic acid isomer (tentative assignment), **2:** rutin, **3:** hyperosid, **4:** isoquercitrin, **5:** miquelianin (quercetin-3-*O*-glucuronide), **6:** quercitrin, **7:** quercetin, **8:** biapigenin.

Table 4S. Retention time and spectroscopic data of annotated peaks in *Hypericum perforatum* 70% EtOH extract.

Peak number	Retention time (min)	UV maxima (nm)	ESI MS (+) (m/z)	ESI MS (-) (m/z)	Identification
1	6.8	210, 300, 325	355.0 [M+H] ⁺	353.1 [M-H] ⁻	Chlorogenic acid isomer
2	18.7	221, 255, 351	609.0 [M+H] ⁺	611.0 [M-H] ⁻	Rutin
3	19.7	215, 255, 353	464.9 [M+H] ⁺	463.1 [M-H] ⁻	Hyperosid
4	20.2	201, 255, 353	465.0 [M+H] ⁺	463.1 [M-H] ⁻	Isoquercitrin
5	22.2	201, 255, 353	478.9 [M+H] ⁺	477.0 [M-H] ⁻	Miquelianin
6	23.6	201, 254, 349	449.0 [M+H] ⁺	447.0 [M-H] ⁻	Quercitrin
7	27.4	204, 254, 370	302.9 [M+H] ⁺	301.0 [M-H] ⁻	Quercetin
8	28.7	208, 268, 330	538.9 [M+H] ⁺	537.0 [M-H] ⁻	Biapigenin

**Fig. 5S.** HPLC-UV analysis of hyperforin and adhyperforin in *Hypericum perforatum* extract. **1:** hyperforin, **2:** adhyperforin.

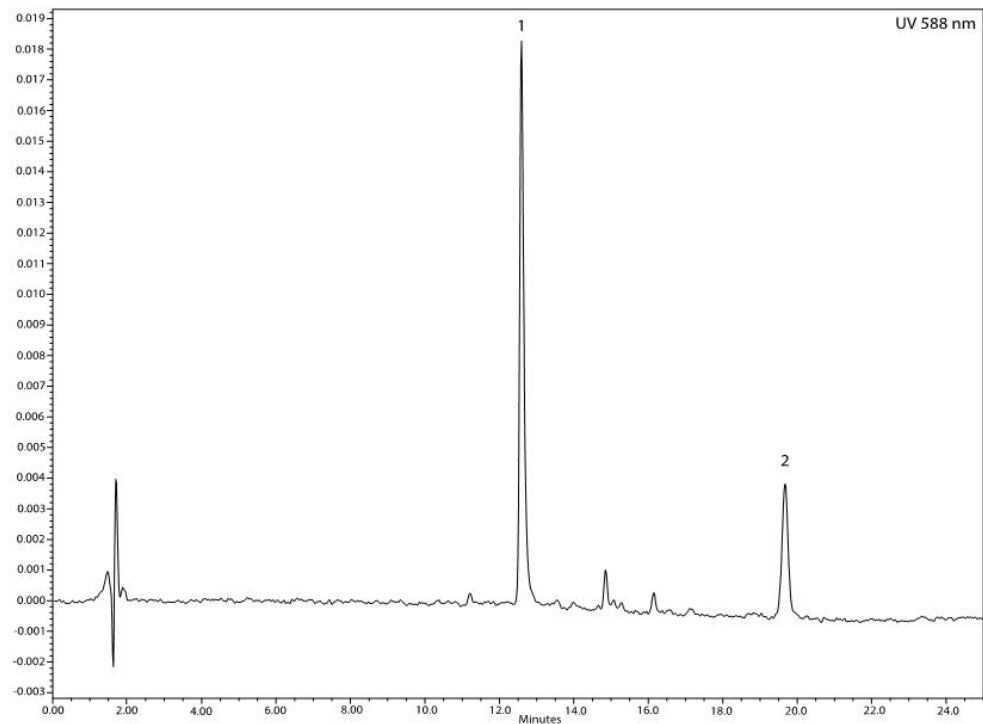


Fig. 6S. HPLC-UV analysis of hypericin and pseudohypericin in *Hypericum perforatum* extract. **1:** pseudohypericin, **2:** hypericin.

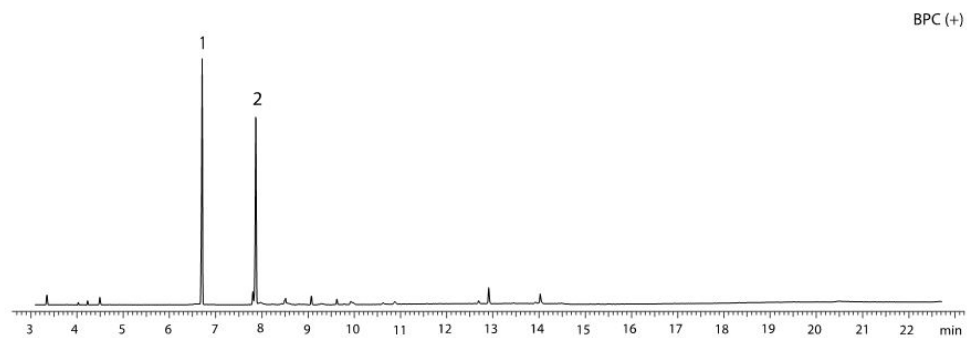


Fig. 7S. GC-MS analysis of *Lavandula angustifolia* essential oil. **1:** linalool, **2:** linalyl acetate.

Table 5S. Retention time, peak area, and identification of constituents of *Lavandula angustifolia* essential oil.

Peak number	Retention time (min)	Area (%)	Identification	Spectrum identity (% NIST)
1	6.71	43.4	Linalool	97
2	7.87	38.6	Linalyl acetate	90

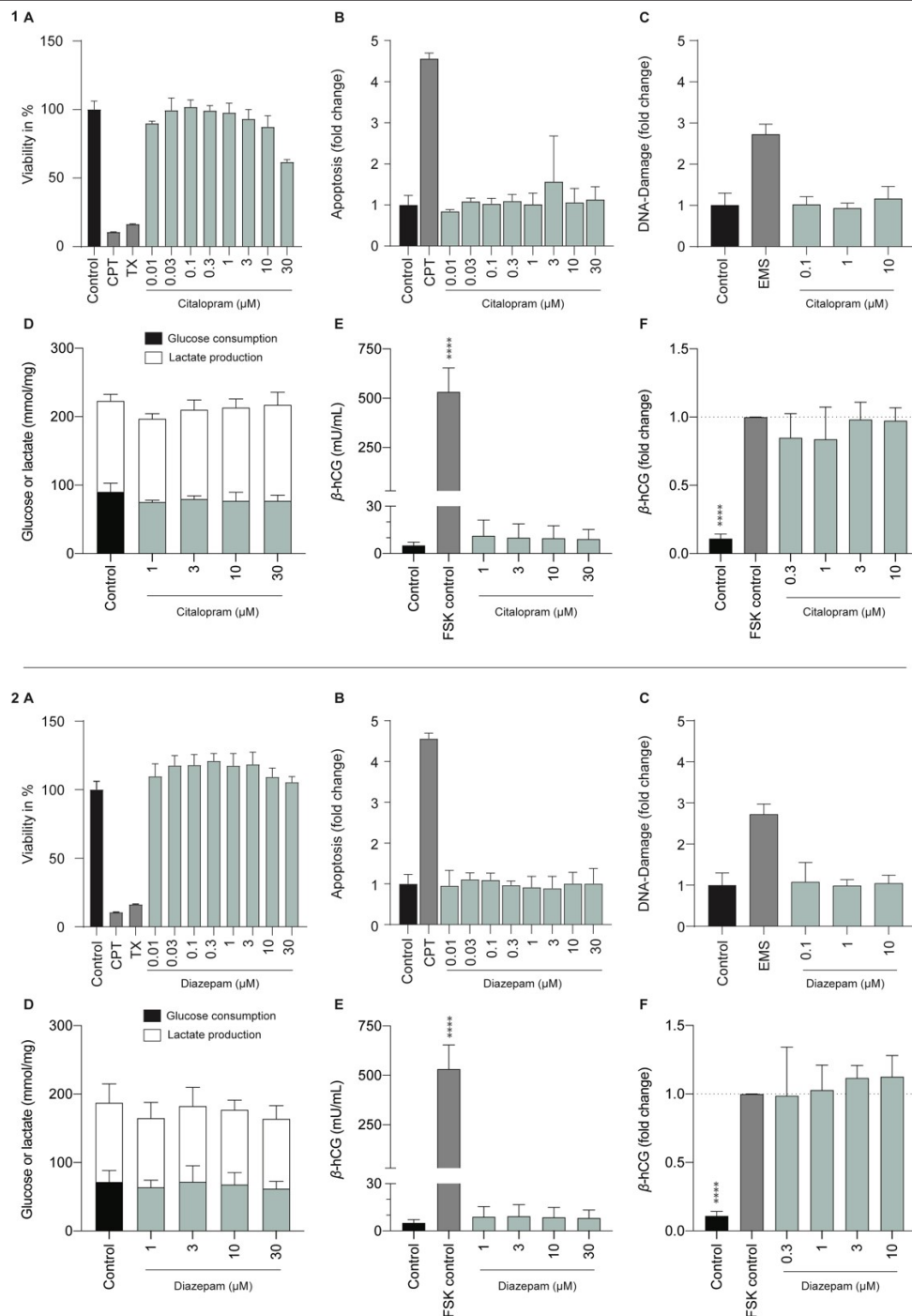


Fig. 8S. Effects of different concentrations of citalopram (**1**) and diazepam (**2**) on *in vitro* safety using undifferentiated BeWo b30 cells. All controls consisted of cell culture media containing 0.2% of DMSO. All data were obtained from at least 3 independent experiments (D–F in triplicate) and are shown as mean \pm SD: * $p < 0.05$. (**A**) Effects on cell viability after treatment for 72 h as percent compared to the untreated control; treatments with camptothecin (CPT) and Triton-X-100 (TX) serve as toxicity controls. Results were normalized to untreated control signal = 100%. (**B**) Effects on cell death after treatment for 72 h as fold changes compared to the untreated controls with camptothecin (CPT) for apoptosis. (**C**) Effects on tail DNA after treatment for 3 h as fold changes compared to the untreated control with ethyl methanesulfonate (EMS) as a positive control. (**D**) Effects on glucose consumption and lactate production after treatment for 48 h expressed in mmol and normalized per amount of protein (mg). (**E**) Effects on β -hCG secretion (mU/mL) by BeWo b30 cells upon 48 h treatment with increasing concentrations of citalopram (**1**) or diazepam (**2**) and 50 μ M FSK as a positive control. (**F**) Effects on FSK-induced differentiation of BeWo b30 cells as fold changes compared to the FSK control. Cells were pre-treated with test compound or cell culture medium for 24 h, before adding 5 μ M FSK for a further 24 h.

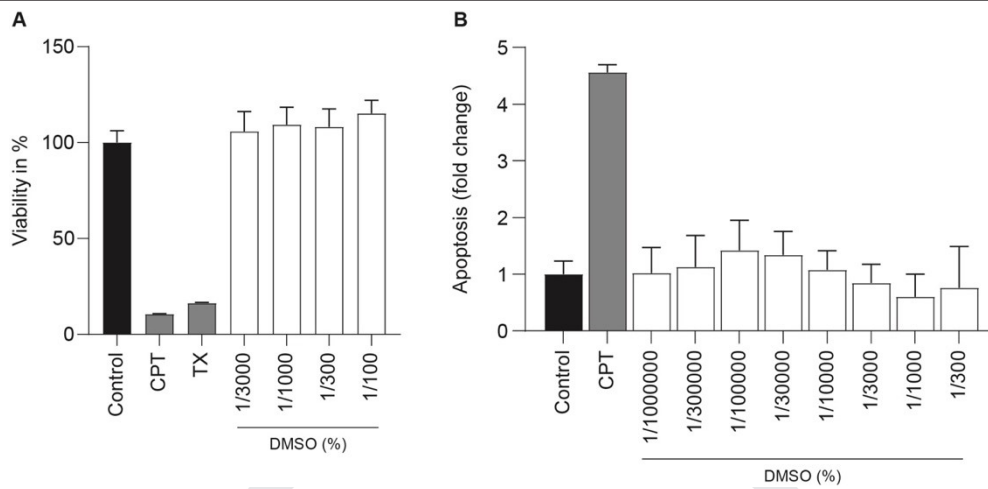


Fig. 9S. (A) Cytotoxic effects of DMSO (0.03–1%) on BeWo b30 cells. Viability assays were performed after a 72 h incubation period. Camptothecin (CPT) and Triton-X-100 (TX) were used as toxicity controls. **(B)** Effects of DMSO (0.0001–0.3%) on cell death after treatment for 72 h as fold changes compared to the untreated controls with camptothecin (CPT) for apoptosis. All data are represented as mean \pm SD of three independent experiments ($n = 3$; $p^* < 0.05$).