

Supplemental information

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NUDT7 stimulates *de novo*
lipogenesis in hepatocytes**

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Deficiency of peroxisomal NUDT7 stimulates de novo lipogenesis in hepatocytes

Jinsoo Song^{1,2}, In-Jeoung Baek³, Sujeong Park¹, Jinjoo Oh¹, Deokha Kim¹, Kyung Song⁴, Mi Kyung Kim⁵, Hye Won Lee⁶, Byoung Kuk Jang⁵, Eun-Jung Jin^{1, 2, 7*}

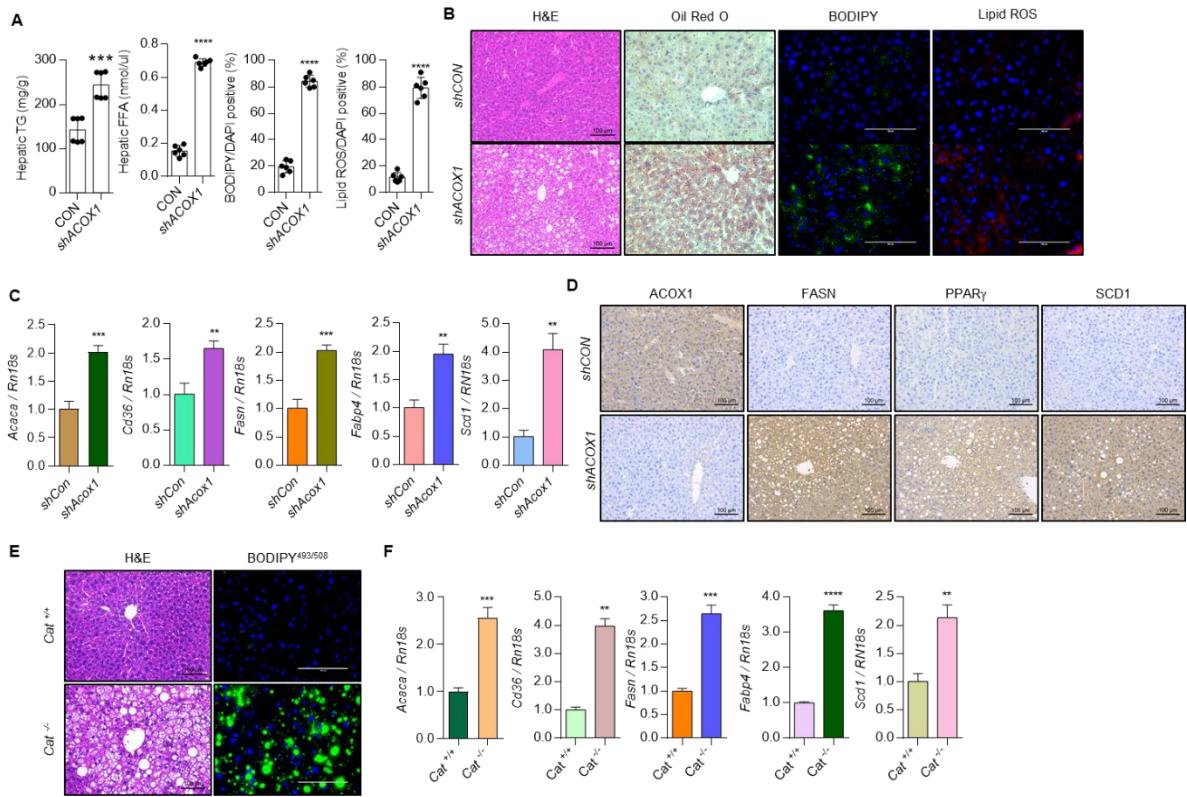


Figure S1. Peroxisome dysfunction induces hepatic lipid accumulation, related to Figure 1.

(A) Positive percentage of TG, FFA, BODIPY and Lipid ROS in *shCON* and *shACOX1* mouse liver (n = 6). (B) Representative images of H&E, Oil Red , BODIPY, and Lipid ROS staining in *shCON* and *shACOX1* mouse liver (n = 5; Scale bars, 100 μ m). (C) Expression levels of *Acaca*, *Cd36*, *Fasn*, *Fabp4*, and *Scd1* of *shCon* and *shACOX1* mouse liver (n = 3). (D) Representative images of ACOX1, FASN, PPAR γ , and SCD1 staining (n = 3; Sale bars, 100 μ m). (E) Representative images of H&E and BODIPY staining in *Cat*^{+/+} and *Cat*^{-/-} mouse liver (n = 3; Scale bars, 100 μ m). (F) Expression levels of *Acaca*, *CD36*, *Fasn*, *Fabp4* and *Scd1* in *Cat*^{+/+} and *Cat*^{-/-} mouse liver (n = 3). Values were expressed as means + s.d. An unpaired *t*-test was used for statistical analysis. **P < 0.01, ***P < 0.001, ****P < 0.0001.

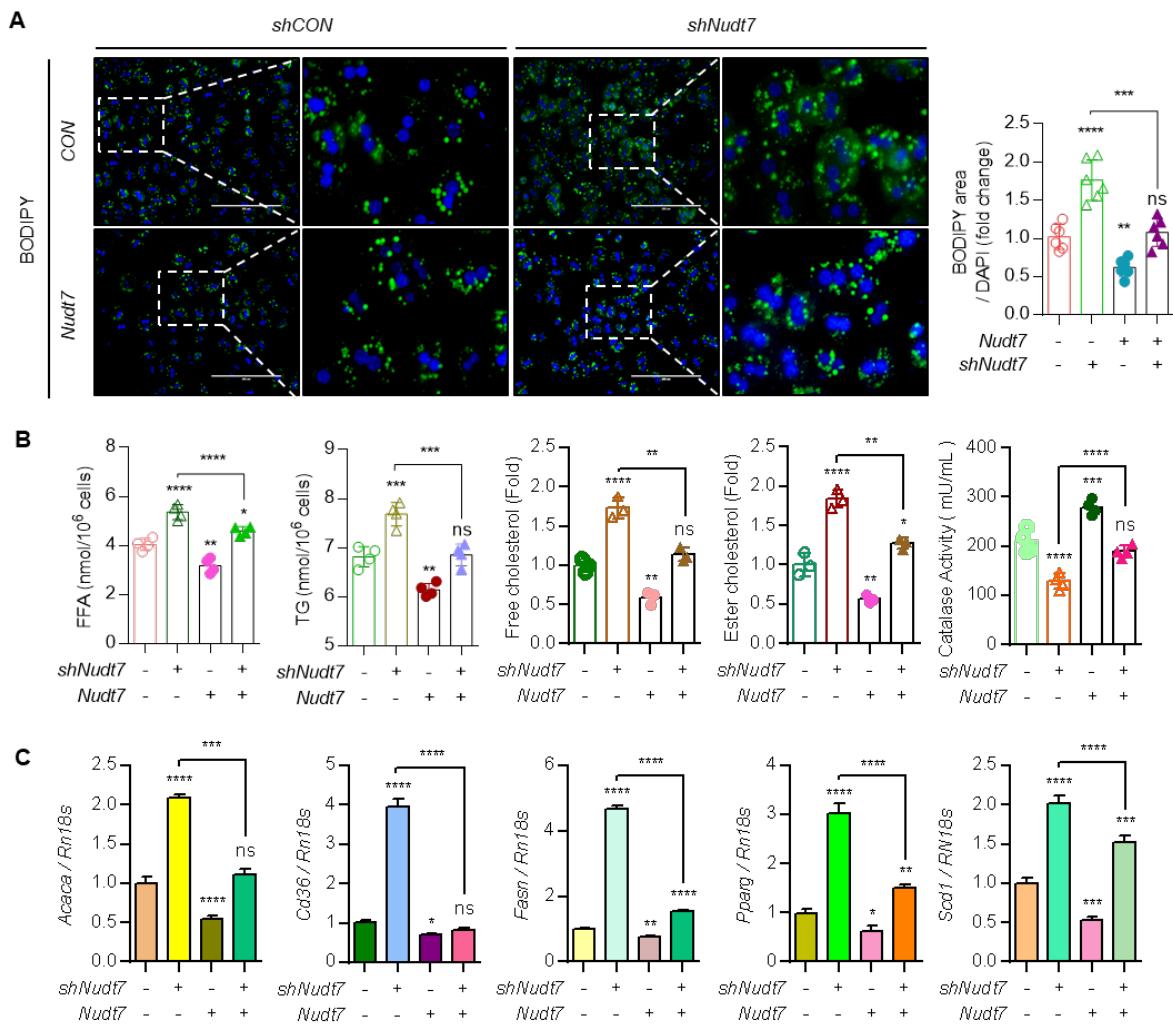


Figure S2. In-vitro modulation of Nudt7 alters lipid accumulation, related to Figure 2.

(A) Representative image of BODIPY staining in *shNudt7* or *Nudt7* hepatocytes and bar graph of BODIPY positive percentage was represented as the fold of CON (n=5). (B) Analysis of FFA (n = 4), TG (n = 4), free cholesterol (n = 3), ester cholesterol (n = 3) levels, and catalase activity (n = 4). (C) Expression levels of *Acaca*, *Cd36*, *Fasn*, *Pparg*, and *Scd1* were analyzed using qRT-PCR (n = 3). Values were expressed as means + s.d. An unpaired Student's t-test and one-way ANOVA multiple test were used for statistical analysis. ns = non-significant, *P ≤ 0.05 , **P < 0.01, ***P < 0.001, ****P < 0.0001.

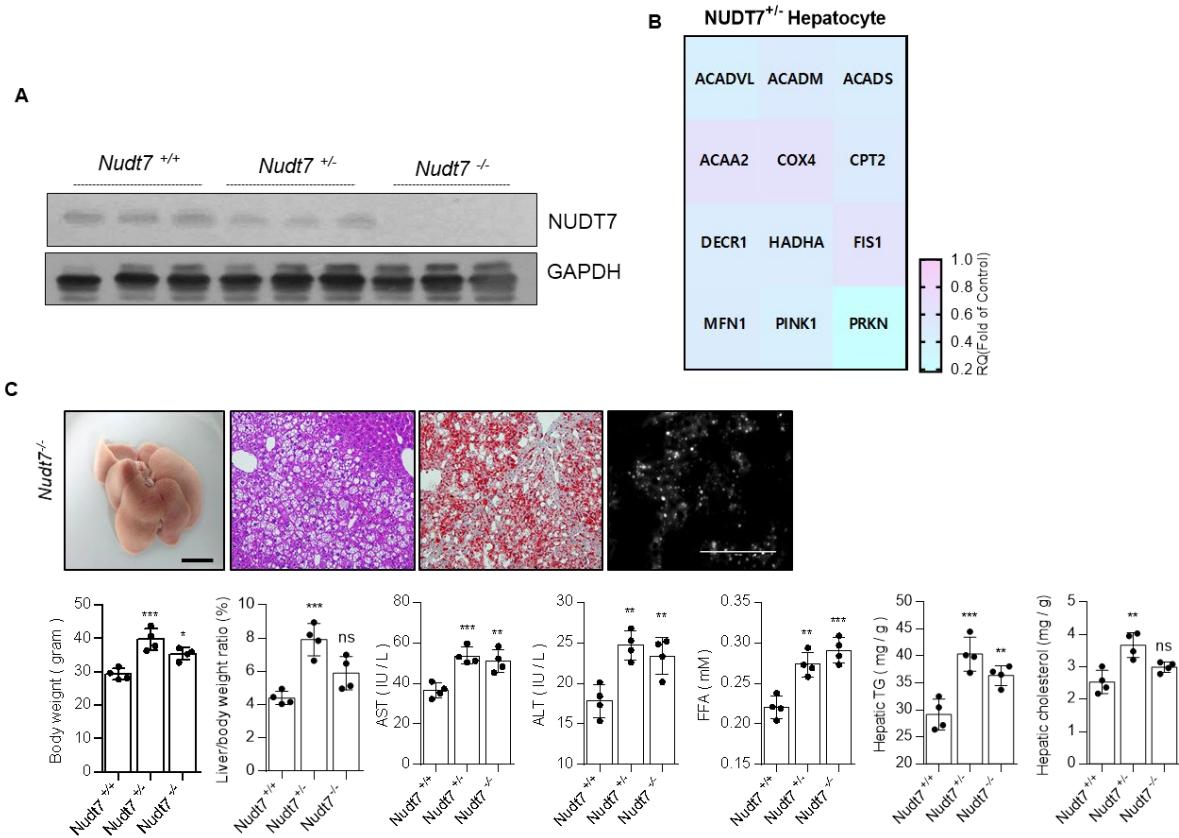


Figure S3. *Nudt7*^{-/-} mice induces hepatic lipid accumulation, related to Figure 3.

(A) The expression level of NUDT7 was analyzed by immunoblotting in *Nudt7*^{+/+}, *Nudt7*^{+/-}, and *Nudt7*^{-/-} liver (n = 3; GAPDH was used for loading control). (B) The expression level of mitochondria related genes was analyzed by qRT-PCR (n = 3). (C) Representative images of liver, H&E, Oil Red O, and Filipin staining of *Nudt7*^{-/-} NCD liver (n = 4). The weight of body and liver, and the level of AST, ALT, FFA, TG, and cholesterol were analyzed in *Nudt7*^{+/+}, *Nudt7*^{+/-}, and *Nudt7*^{-/-} mouse (n = 4). Values were expressed as means + s.d. An unpaired Student's t-test and one-way ANOVA multiple test were used for statistical analysis. ns = non-significant, *P ≤ 0.05, **P < 0.01, ***P < 0.001.

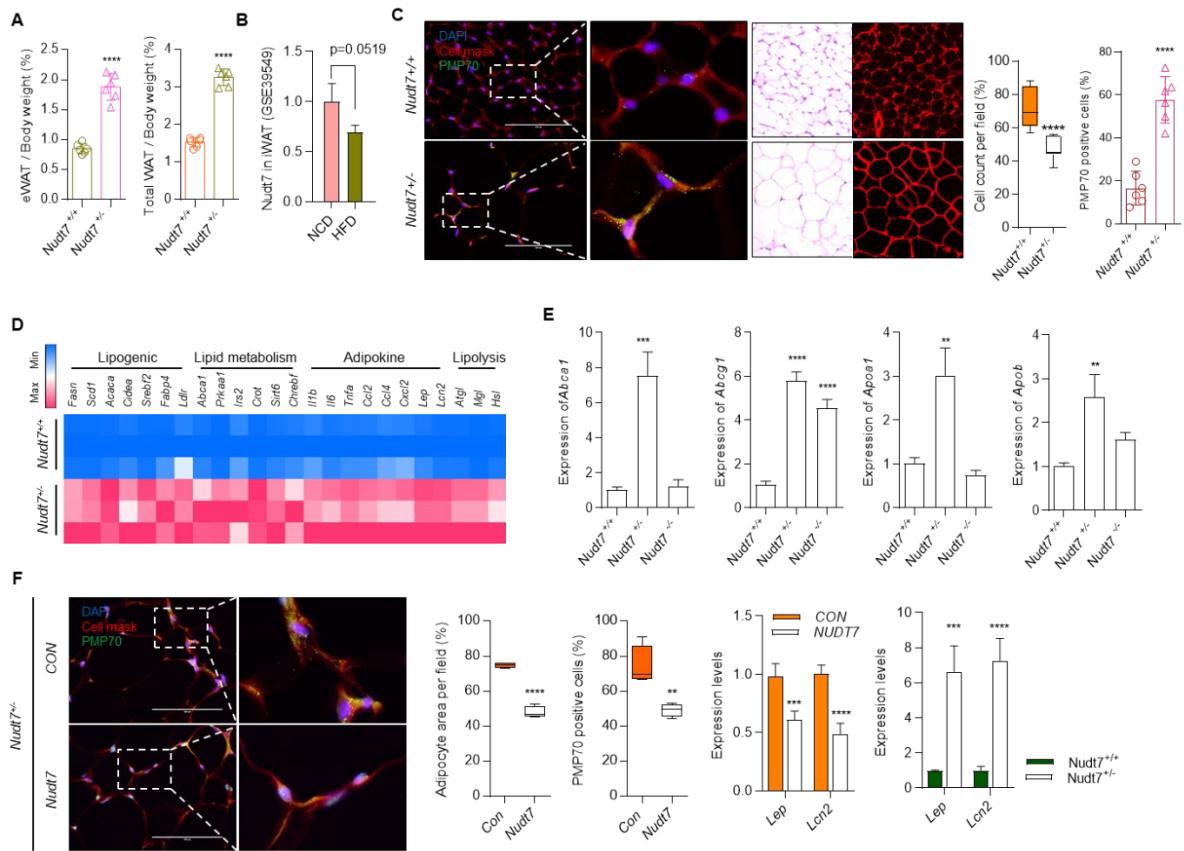


Figure S4. *Nudt7*⁺⁻ mice displays adipocyte enlargement and adipose expansion, related to Figure 3.

(A) The weight of eWAT and total WAT to body weight ($n = 4$). (B) The expression level of Nudt7 in GSE39549. (C) Representative images of cell mask orange and PMP70 staining and graphs with adipocyte area per filed and percentage of PMP70-positive cell counting ($n = 3$; Scale bars, 100 μ m). (D) The expression levels of genes in lipid metabolism were analyzed by qRT-PCR ($n = 3$). (E) The expression levels of Aba1, Abcg1, Apoal, and Apob in *Nudt7*^{+/+}, *Nudt7*⁺⁻, and *Nudt7*^{-/-} adipocytes ($n = 3$). (F) Representative images of cell mask orange and PMP70 staining and graphs with adipocyte area per filed and percentage of PMP70-positive cell counting ($n = 4$; Scale bars, 100 μ m), and expression levels of *Lep* and *Lcn2* in *Nudt7*⁺⁻, adipocytes with NUDT7 restoration. Values were expressed as means + s.d. An unpaired

Student's t-test and one-way ANOVA multiple test were used for statistical analysis.

P < 0.01, *P < 0.001, ****P < 0.0001.

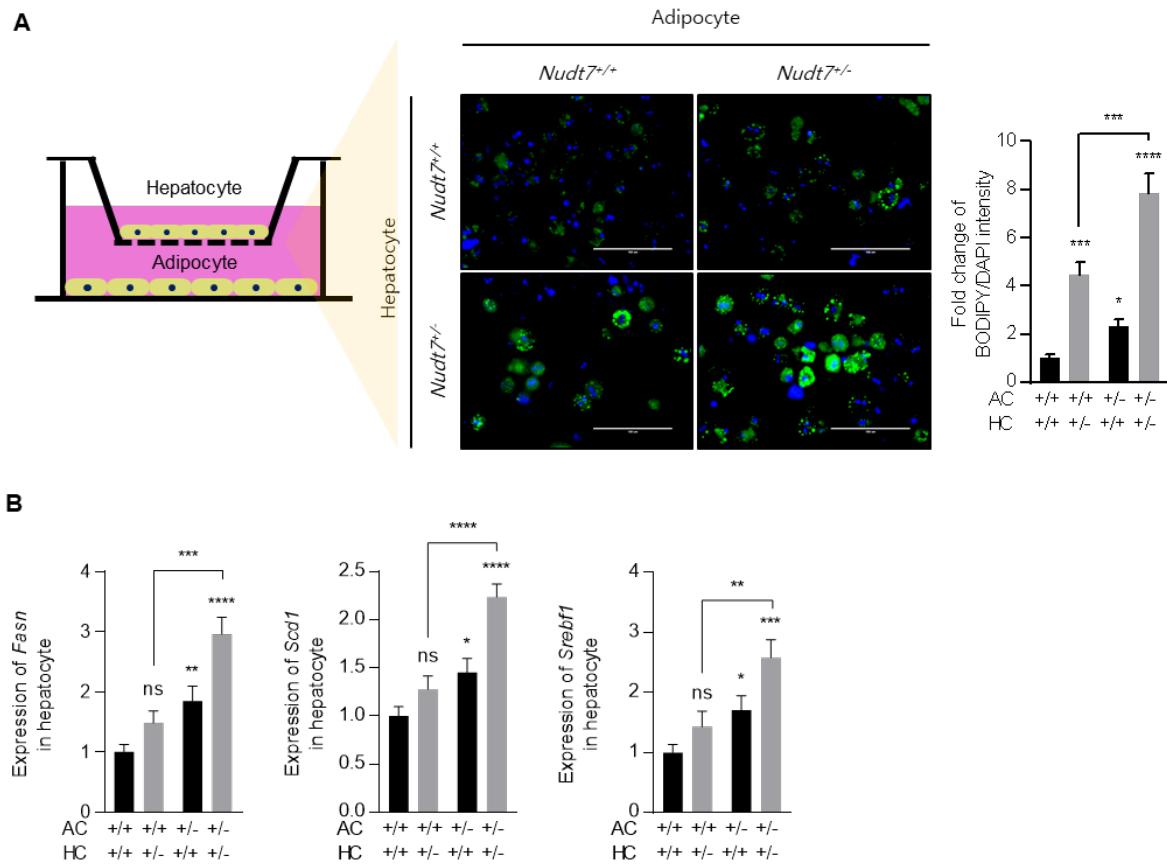


Figure S5. Increased lipolysis of adipose tissue affects hepatic lipid accumulation in *Nudt7*^{-/-} mice, related to Figure 3.

(A) Co-culture of hepatocytes and adipocytes isolate from *Nudt7*^{+/+} or *Nudt7*^{-/-} mice. Representative image of BODIPY staining and bar graph of BODIPY intensity in hepatocytes was represented as fold of control. (B) The expression levels of *Fasn*, *Scd1*, and *Srebf1* in hepatocyte were analyzed by qRT-PCR (n = 3). Values were expressed as means + s.d. An unpaired t-test was used for statistical analysis. ns = non-significant, *P ≤ 0.05, **P < 0.01, ***P < 0.001, ****P < 0.0001.

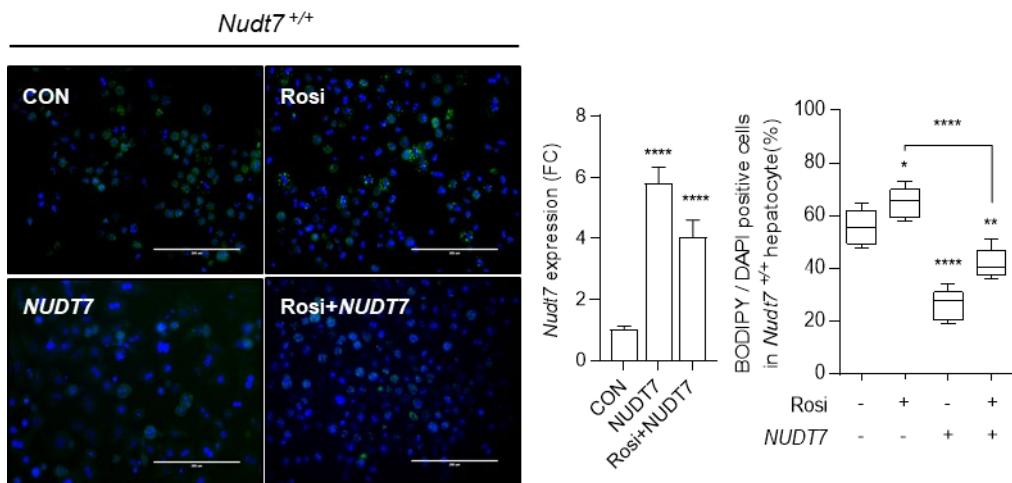


Figure S6. Exposure of PPAR γ agonist induces lipid accumulation in *Nudt7*^{+/+} hepatocytes, related to Figure 4.

Representative image of BODIPY staining, positive cell counting ($n=3$), and *Nudt7* expression level in *Nudt7*^{+/−} hepatocytes in the presence of 10 μ M rosiglitazone (Rosi) with/without introduction of pcDNA-*Nudt7*. Scale bars, 200 μ m. Values were expressed as means + s.d. An unpaired t-test or one-way ANOVA were used for statistical analysis. * $P \leq 0.05$, ** $P < 0.01$, *** $P < 0.001$, **** $P < 0.0001$.

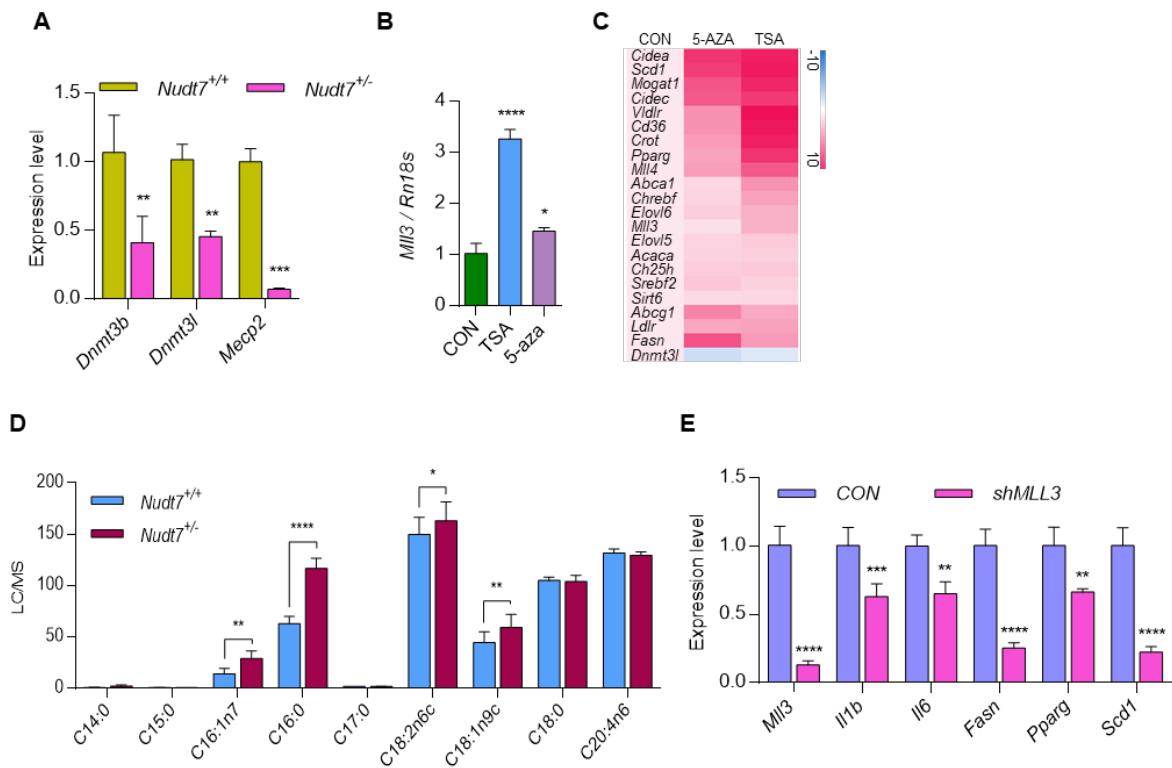


Figure S7. The possible involvement of *Mll3* in hepatic lipogenesis, related to Figure 5.

(A) The expression levels of Dnmt3b, Dnmt3l, and MeCP2 were analyzed by qRT-PCR ($n = 3$). (B) Primary cultured hepatocyte was treated with TSA or 5-aza and the expression level of *Mll3* was analyzed by qRT-PCR ($n = 3$). (C) The expression level of lipogenic genes ($n = 3$) was analyzed by qRT-PCR. (D) Lipidomic analysis of *Nudt7^{+/+}* and *Nudt7^{-/-}* liver ($n = 3$). (E) The expression levels of *Mll3*, lipogenic genes, and inflammatory cytokine ($n = 3$). Values were expressed as means \pm s.d. An unpaired *t*-test or one-way ANOVA multiple comparisons were used for statistical analysis. * $P \leq 0.05$, ** $P < 0.01$, *** $P < 0.01$, **** $P < 0.0001$, **** $P < 0.0001$.

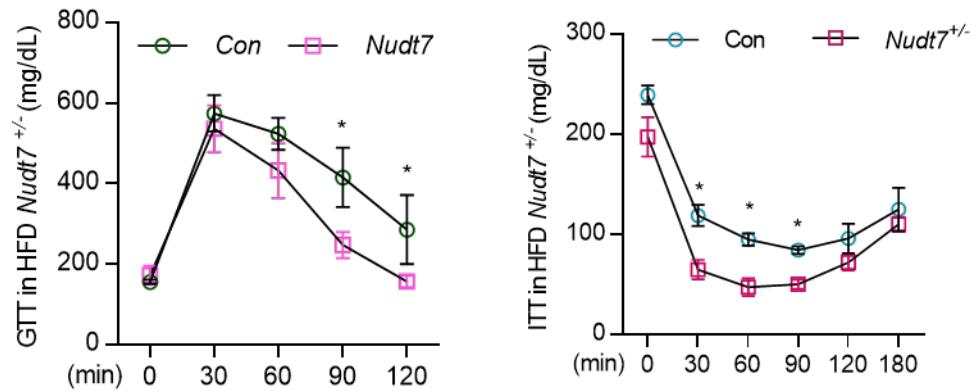


Figure S8. Impaired glucose and insulin tolerance by *Nudt7* deficiency could rescue by the restoration of *Nudt7*, related to Figure 6. Analysis of glucose and insulin tolerance levels in HFD *Nudt7*^{+/+} and *Nudt7*^{+/−} mice. An unpaired Student's *t*-test was used for statistical analysis. *P ≤ 0.05.

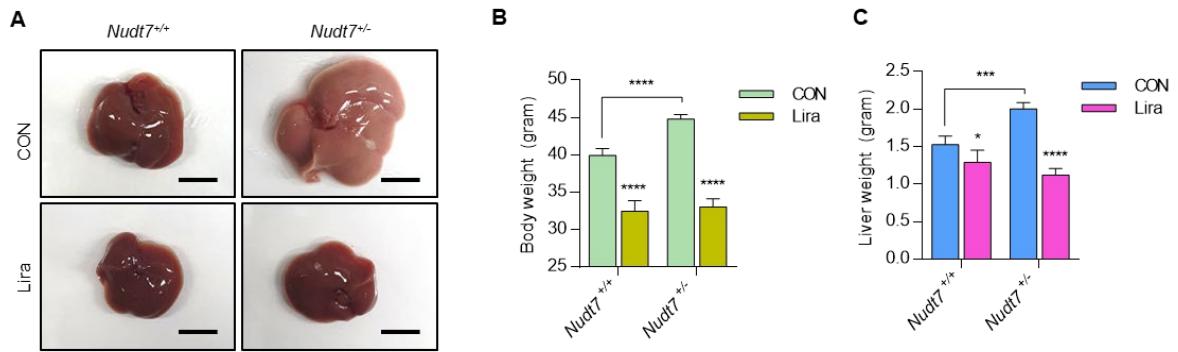


Figure S9. The effect of liraglutide in *Nudt7*^{+/−} liver, related to Figure 7.

(A) Representative images of mouse liver of HFD fed *Nudt7*^{+/−} mice with liraglutide or equal volume of sterile saline i.p. injected mouse. (B) Body weight and (C) liver weight were analyzed (n = 4).

Table S1. Clinical characteristics of enrolled patients, related to Figure 2.

Variables	Median (IQR) or N (%)
Male : Female (%)	10: 12 (4.5 :54.5)
Age (year)	39.00 (33.00-47.75)
Hypertension, N (%)	10 (45.5)
Type 2 Diabetes, N (%)	10 (45.5)
Dyslipidemia, N (%)	10 (45.5)
Body mass index, kg/m ²	37.67 (33.95-44.02)
Total body fat (%)	47.15 (42.45-51.68)
Systolic BP, mmHg	140.00 (12.6.25-150.00)
Diastolic BP mmHg	80.00 (70.00-90.00)
eGFR mL/min/1.73 m ²	113.60 (79.45-121.25)
ALP, U/L	64.00 (50.75-79.00)
AST, IU/L	34.00 (20.75-43.00)
ALT, IU/L	33.50 (19.50-44.50)
γ-GT, U/L	31.00 (18.50-47.00)
Total cholesterol (mg/dL)	178.00 (147.50-201.00)
LDL cholesterol (mg/dL)	124.00 (92.25-142.00)
HDL- cholesterol (mg/dL)	45.50 (37.00-51.25)
Triglyceride (mg/dL)	124.50 (79.25-203.75)
HbA1C (%)	5.80 (5.60-6.30)
Fasting glucose (mg/dL)	105.00 (89.75-114.25)
HOMA_IR	4.56 (2.49-7.44)
Fibrotic Stage	
F1	4 (18.2)
F2	7 (77.3)
F3	1 (4.5)
NAS	
1	0 (0.0)
2	6 (27.3)
3	11 (50.0)
4	3 (13.6)
5	2 (9.1)

BP, Blood pressure; γ-GT, γ-glutamyltransferase; ALT, alanine aminotransferase; AST, aspartate aminotransferase; HDL, high-density lipoprotein; HbA1c, glycated hemoglobin A1c; HOMA-IR, Homeostasis Model Assessment of Insulin Resistance; LDL, low-density lipoprotein, NAS; Nonalcholic fatty liver disease Activity Score.

Table S2. Primers used for qRT-PCR analysis, related to STAR Methods.

Description	Symbol	Forward (5' to 3')	Reverse (5' to 3')
Peroxisome profiling	<i>Abcd2</i>	GACCATGCCTATGAGACCTATT	GTAAGAGACTGGTCAGGGTTG
	<i>Acad11</i>	CCCAAAGAGTGTGGGTTATAG	GTGTCCTGGGCCAATAGTAAA
	<i>Acot1</i>	CGTCATGGCTCTGGCTTATTA	CAGGTAGTTCACGGCTTCTT
	<i>Acot2</i>	CGTGATGGCTCTGGCTTATTA	CAGGTAGTTCACGGCTTCTT
	<i>Acot8</i>	ACTGGAGCCCAAACAGATG	AGGAAGGCGTAGTCAGAGATA
	<i>Acox1</i>	CGCACATCTGGATGGTAGT	GGCTCGAGTGAGGAAGTTATAG
	<i>Acox3</i>	ACCCACGGATAAGGAAGAGA	GAGAAGTGGCCAAGGCATAG
	<i>Acsf3</i>	GAGTGGAAGTACGCATCATCTC	GAACCCCTGGAGTCACCTTG
	<i>Acs1</i>	GCTTGTTGGATGTGGAAGAAATG	TCTTGCTGGGTCTTCAGTAG
	<i>Acs3</i>	GGCTGTGCACTGGAGATATT	GTACTCTCCTGCCTGTAGTTTC
	<i>Aldh3a2</i>	GGCTTCTCCTGACTATGAAAGG	CATCCATCTCTCCACCGAAAG
	<i>Crat</i>	GGAGGCCATATCAACTTCTC	ATGTCCAGCAGAGCTTCTC
	<i>Crot</i>	TGCTGTGATCATGCTCCTTAC	CTGAACCCTTCCATCTCCTTC
	<i>Ephx2</i>	GTAAAGGGTGGGACGAAAGA	GACATTTCAGGACGGAGTACAA
	<i>Far1</i>	CAAGAGCGAGTGGAGGAAAT	AGCCAGTTAGGTTGGGTAAG
	<i>Hacl1</i>	TTTGGACTGCCTCCAAGATAC	TCACAGAAGGCTGACATTATT
	<i>Idh1</i>	GTGGAGATGCAAGGAGATGAA	ATGCAGATCCAGTCCACATAG
	<i>Idi1</i>	ACAGCAGAGATCAGATGCTAA	GCTCGCCTGGGTTACTTAAT
	<i>Idi2</i>	CCCAGGACCAGATTCCATAAA	ACCAACAGAACGGTAGCCAATC
	<i>Mlycd</i>	CTGCCATCTTCTACTCCATCAG	GCTCCTTGACCACTCTTTAT
	<i>Mpv17</i>	GCCCTCATACCAACTACTATC	CAATAGCAACACACTGGACAAC
	<i>Mvk</i>	GGAGCAACTGGAGAACGCTAAA	TGCCAGGTACAGGTAGAGAA
	<i>Nudt12</i>	GGAAGAGGAAAGTGGAGTCAAA	GTAGACACTGCCACAGCTAAA
	<i>Nudt7</i>	CAGTTGCTAGATGATGCTAAGG	AAAGGACGGAGTATTGTTATG
	<i>Peci</i>	CCTCTGGTTGCGGTAGTAAAT	GAGTTGGCTGAATGGAGTATGA
	<i>Pecr</i>	GCGAAATGGGACAAACCTTATT	ACAGCAGGAAACATACCAAGAG
	<i>Pex1</i>	GGAGATGGAGAATGTGGCTTAG	CCAAAGTAGAGCCGGTACATATT
	<i>Pex11a</i>	TCCTTCCTCCTCCTCTTATT	GCCGAGATTGGACTTGAGAT
	<i>Pex11b</i>	ATCTGAGCCGTGATGCTTATG	CCTGAGACTCCTACTCCAGAAA
	<i>Pex11g</i>	AGTATGGCCTGGGACAAAG	GCGATATGCTACAAGGATAGT
	<i>Pex12</i>	CAGAAGCGTTGGTGAGAAGATA	CAGGAAGAACATACCCACAGAG
	<i>Pex19</i>	GCTAGATCTCATGCAGCAGTTA	TTGAGAGCATCCAGGTCAAAG
	<i>pex26</i>	CTGGAGCTGTGCATCCTT	AGGAAGGCCCTGGTTATCT
	<i>Pex6</i>	CTGACTGGTGCAGACTCTTATT	CAGCTCTAGCCCTCCTCTA
	<i>pex7</i>	CATGTCCTGTCACCTGTAGTG	CTCCTGCGTGTGCTCTTAT
	<i>Pipox</i>	CAAGGGATCGACCATGAGTATC	ATAGAGGACACCTCCAGTCTT
	<i>Prdx1</i>	CCGCTCTGTGGATGAGATTATAC	GTACTCACTGCCAGGTTCCA
	<i>Rhoc</i>	TCCGAAAGAAGCTGGTGATTG	AAAGACGGTAGGCACGTAGA
	<i>Scp2</i>	CGGACCACCTCAACTGATAAA	TCCCAGCATACCCAAACATC
	<i>Slc22a5</i>	ATAATCCTGTGGCTGACCATATC	CCAGTAGGAAGCAGTCACATAG
	<i>Sod1</i>	CTCAGGAGAGCATTCCATCATT	CTCCCAGCATTCCAGTCTT
	<i>Xdh</i>	CCACATGGACAACGCCATAAA	GCTGTGTTAGAGGGCAGATTAG
	<i>Rn18s</i>	CCAGTAAGTGCAGGGTCATAAG	GGCCTCACTAACCATCCAA
Lipogenic genes	<i>Acaca</i>	AGCCAGAAGGGACAGTAGAA	CTCAGCCAAGCGGATGAAA
	<i>Cd36</i>	CTGGGACCATTGGTGATGAAA	CACCACTCCAATCCCAAGTAAG
	<i>Cidea</i>	GCAACCAAAGAAATCGGAATAG	CTCGTACATCGTGGCTTGA
	<i>Elov15</i>	GGTGTGTGGGAAGGCAAATAC	TGGAGAAGTAGTACCAACAGAG
	<i>Fabp4</i>	GGATGGAAAGTCGACCAATA	TGGCTCATGCCCTTCTATAA
	<i>Fasn</i>	AGACCCGAACCTCAAGTTATT	GCAGCTCCTGTATACTCTCC

	<i>Ldlr</i>	ATCCACCGAACATCTACTG	GGAACAGTGTCCCTCCTCTTAC
	<i>Scd1</i>	CAACTTCACCACGTTCTTCATC	CCCGTCTCCAGTTCTCTTAATC
	<i>Srebf1</i>	ACTTCCCTGGCTATTGACC	GGCATGGACGGGTACATCTT
Lipid metabolic genes	<i>Abca1</i>	GTTCCGGGAAGTGTCTAAA	CTGGGAGAGGATGCTGAATATC
	<i>Cat</i>	GATGGTAACTGGGATCTTGTGG	GTGGGTTCTCTTCTGGCTATG
	<i>Chreb1</i>	CAGCTGCCGGATGAAATAGA	CAAAGCGCTGATGTGTGATG
	<i>Crot</i>	TGCTGTGATCATGCTCCT	CTGAACCCCTCCATCTCCTT
	<i>Irs2</i>	ATGGGTACATGAGCATGGATAG	CAGGCGTGGTTAGGGAATAA
	<i>Prkaa1</i>	GATCCTTCCGGTGTGGATTAT	GAAAGACCAAAGTCGGCTATCT
	<i>Ppara</i>	CGGTGTGTATGAAGCCATCT	TAAGGAACTCGCCTGTGATAAA
	<i>Pparg</i>	CTGGCCTCCCTGATGAATAAG	AGGCTCCATAAGTCACCAAAG
	<i>Sirt6</i>	CGTCTGGTATTGTCAACCT	GAGTCTGCACATCACCTCATC
Adipokine genes	<i>Ccl2</i>	CTCACCTGCTGCTACTCATTC	ACTACAGCTTCTTGGGACAC
	<i>Ccl4</i>	CCACTTCCTGCTGTTCTCTTA	CAAAGACTGCTGGTCTCATAGT
	<i>Ccl5</i>	GCCCACGTCAAGGAGTATT	CTTGAACCCACTTCTCTGG
	<i>Cxcl2</i>	GACAGAAGTCATAGCCACTCTC	GCCTTGCCCTTGTTCAGTATC
	<i>Il1b</i>	CCACCTCAATGGACAGAAATATCA	CCCAAGGCCACAGGTATT
	<i>Il6</i>	CCAGAGTCCTTCAGAGAGATACA	CCTCTGTGACTCCAGCTTATC
	<i>Lep</i>	GACTTCATTCCCTGGGCTTCA	ATTCTCCAGGTCTTGGCTATC
	<i>Lcn2</i>	TGGCCCTGAGTGTCTATGTG	CTCTTAGCTCATAGATGGTGC
	<i>Tnfa</i>	TTGTCTACTCCCAGGTTCTCT	GAGGTTGACTTCTCCTGGTATG

Lipolysis genes	<i>Atgl</i>	CATCCGTGGCTGTACTAAAG	GACGTTCTCCGTCTGAAAC
	<i>Hsl</i>	CATCAACCACTGTGAGGGTAAG	AAGGGAGGTGAGATGGTAACT
	<i>Mgl</i>	AAGAGTGGAGCGAGCAATG	GATGATTCCATGAGCAGGTAGG
Histone modification genes	<i>Dnmt3b</i>	GGTCTCCAGCCTCTGAATTAC	CAGAGCCATTCCCATCATCTAC
	<i>Dnmt3l</i>	GCAGAGACTACCAAGAACATG	TGACTTGGGCTTGCAGATAC
	<i>Mecp2</i>	AGGCAGGCAAAGCAGAAACATCAG	TCATACTTCCAGCAGATCGGCCA
	<i>Mll1</i>	CGAGGAAAGAGATCAGCAGAAAG	AGTCCGCGTGAAGTTGTAATAG
	<i>Mll2</i>	AGTACCTGAAAGGCGAAGAAC	CGCCCGTAGCGGAATAAATA
	<i>Mll3</i>	CGACTCTCTGTTGACCCCTTATG	ATTGCTGGGTGTGGAGTTAG
	<i>Mll4</i>	GTTCGGATAGGAAGGGAGCTTATG	CATCCTCAAGCCCTGGTAAAT