Supporting information of

## In situ bioorthogonal conjugation of delivered bacteria with gut inhabitants for enhancing probiotics colonization

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Figure S1 to S11



**Figure S1.** *In vitro* bacterial biological properties after azido and DBCO modification. The OD600 changes of GFP, azido-modified GFP and DBCO modified GFP during 24 hours. Significant differences were assessed in this figure using one-way ANOVA (Turkey; n. s., not significant)



Figure S2. Zeta potential of bacteria after bacterial adhesion (n = 3). Significant differences were assessed in this figure using one-way ANOVA (Turkey; *n. s.*, not significant)



Figure S3. *In vitro* bacterial adhesion between bioorthogonal groups modified *E. coli* MG1655 and *P. anaerobius*. (A), Confocal fluorescence images of azide modified *E. coli* MG1655 (Cy5 labeled) after incubation with DBCO-decorated *P. anaerobius* (FITC labeled) for 2 h (n = 3). (B), Confocal fluorescence images of *E. coli* MG1655 (Cy5 labeled) after incubation with *P. anaerobius* (FITC labeled) for 2 h (n = 3).



**Figure S4. TEM images of bioorthogonal groups modified** *E. coli* MG1655 and *P. anaerobius.* TEM image of bacterial adhesion of azide decorated MG1655 (N3-MG1655) and DBCO decorated *P. anaerobius* (DBCO-*P. a*). Scale bar, 1 μm



Figure S5. *In vitro* bacterial adhesion between bioorthogonal groups modified *E. coli* MG1655 and *S. aureus*. (A), Fluorescence colonization assay of azide-modified *S. aureus* (FITC labeled) and DBCO-functionalized *E. coli* MG1655 (Cy5 labeled). (B), Representative confocal fluorescence images of *E. coli* MG1655 (Cy5 labeled) after incubation with DBCO decorated *S. aureus* (FITC labeled) for 2 h (n = 3).



Figure S6. *In vitro* characterization of bacterial aggregation at different bacterial densities. (A), Representative confocal fluorescence images of 1:1 mixed cocultures at a bacterial density of OD = 0.06. (B), Aggregation ratio of bacterial aggregation in N3-DBCO and PBS groups (n = 3). Significant differences were assessed in (B) using t test (\*P  $\leq$  0.05). The mean values and SEM are presented.



Figure S7. *In vivo* long-term colonization of bacteria in GI tract of mice at different time points. Representative IVIS images for evaluating bacterial retention in the GI tract of mice mediated by biorthogonal reactions (n = 3).



Figure S8. *In vitro* colonization of bacteria in GI tract of mice at different time points. Representative IVIS images of mice colons in each group for evaluating bacterial retention (n = 3).



Figure S9. DSS induced colitis of mice with 3 % DSS in drinking water for 7 consecutive days. (A), Representative photographs of mice colon in DSS and PBS groups, respectively (n = 3). (B), Body weight changes over 7 days (n = 3). (C), Colon length of mice in each group (n = 3). Significant differences were assessed in (B) and (C) using t test (\*\*P  $\leq 0.01$ ; \*\*\*\*P  $\leq 0.0001$ ). The mean values and SEM are presented.



Figure S10. In vivo bioorthogonal-mediated therapy in DSS-induced colitis mice model with E. coli Nissle1917. (A), Schematic of administration schedule. C57BL/6 mice were provided with PBS, ETEC (108 CFU per mouse) or N3-ETEC (108 CFU per mouse) for 3 days after antibiotic cocktail treatment. On days 4, 5 and 6, mice were orally administered with PBS or E. coli Nissle1917 (108 CFU per mouse), DBCO modified *E. coli* Nissle1917 (10<sup>8</sup> CFU per mouse). Mice were euthanized on day 10. (B), Representative photographs of mice colon with different treatments on day 10 (n = 5). (C), Colon length of mice in each group (n = 5). (D), Body weight changes (n = 5). 5). (E) and (F), IL-10 and IL-1 $\beta$  proteins in colon tissue homogenates by ELISA (n = 3). Significant differences were assessed in (C) using one-way ANOVA and using t test in (D) (\* $P \le 0.05$ ; \*\*\*\* $P \le 0.0001$ ; n.s., not significant). The mean values and SEM are presented.



Figure S11. Bacterial counting by bacterial colony-forming units of ETEC in chromogenic *E. coli* agar plate (n = 3).