Figure S1



Fig. S1: ECP effects on T cell activation. (A) Intragraft abundance for indicated effector T cells. Data shown is mean percent abundance  $\pm$  S.D.; 2-sided Mann Whitney U test; \*p< 0.05, \*\*p < 0.01. (B) IL-4 expression following stimulation of saline (open circles) or ECP-treated (closed circles) intragraft CD4<sup>+</sup> T cells with B6 splenocytes (syngeneic antigens), FVB splenocytes (donor antigens) or B6 splenocytes pulsed with lung self-antigens Col V and K $\alpha$ 1T (N=5/group). Data shown are mean IL-4 culture supernatant concentration  $\pm$  S.D.; 2-sided Mann Whitney U test; where n.s. means non-significant.



Fig. S2: ECP-treated leukocytes are rapidly engulfed by AM. Cell Trace<sup>633</sup>-labeled ECP-treated leukocytes and control EGFP<sup>+</sup> non-ECP-treated leukocytes were co-transferred into 3T-FVB lung recipients and AM engulfment was analyzed two hours later. FACS plots are a representative result from 4 experiments. Histograms show pooled data of 4 experiments with mean AM percent engulfment  $\pm$  S.D.



Fig. S3: ECP-treated leukocytes are engulfed by AM, which in turn induces resistance to TGF- $\beta$  mediated  $\alpha\nu\beta5$  upregulation. Saline and ECP-treated AM were pre-treated with 10 mM SB43152 or vehicle (DMSO) and then stimulated with or without TGF- $\beta1$ . FACS plots shown are a representative result from 4 experiments. Fold MFI changes in  $\alpha\nu\beta5$  expression is normalized to respective non-TGF- $\beta$ -treated DMSO-pretreated controls. Data in bar graphs is pooled data from 4 experiments; Mean Fold MFI  $\pm$  S.D.; 2-sided Mann Whitney U test; where \*\*\*p < 0.001.



**Fig. S4**: **ECP stimulates DCM expression in lung allograft CD64+ macrophages following induction of BOS pathogenesis**. POD 16 3T-FVB allograft tissue treated with saline vehicle (N=3) or ECP (N=4) and analyzed for CD64 and DCN expression. Arrows show CD64<sup>+</sup> DCN<sup>+</sup> macrophages in alveolar spaces. Data shown are a representative result for each group.



Fig. S5: ECP-treated AM inhibits  $T_h17$  development in DCN expression-dependent manner. A representative FACS plot result from 5 experiments where plate bound CD3 $\epsilon$  and CD28 Abs were used to activate naïve B6 CD4<sup>+</sup> T cells in the presence or absence of indicated AM conditioned supernatants added in a 1:1 v/v ratio to  $T_h17$  polarization medium containing 10 ng/ml TGF- $\beta$ 1. Intracellular IL-17A expression was assessed 4 days later.



Fig. S6: Reconstitution with recipient-derived AM does not spontaneously induce OB lesions and validation of DCN antibody specificity. (A) AM depletion of donor lungs one day after clodronate depletion. Data shown is a representative FACS plot result from N=2/group (B) Intragraft neutrophilia in donor lungs one day after receiving clodronate liposomes. Data shown are representative FACS plots from N=4/group where histograms depict mean neutrophil numbers  $\pm$  S.D; 2-sided Mann Whitney U test; n.s. is non-significant. (C) Mo-AM generation in clodronate liposome-treated POD 7 DCN<sup>Δ/Δ</sup> and DCN<sup>fl/fl</sup> lung recipients of FVB lungs. Data shown are representative FACS plots from N=2/group. (D) POD 7 histology of a 3T-FVB allograft from a DCN<sup>Δ/Δ</sup> recipient that received clodronate liposome treatment. Histology shown is a representative result from 2 transplants. (E) POD 16 allograft tissue from ECP-treated DCN<sup>Δ/Δ</sup> and DCN<sup>fl/fl</sup> recipients of 3T FVB lungs evaluated for DCN expression within the CD64 compartment. Red arrows show CD64+ macrophages that express DCN. White arrows show CD64+ macrophages that lack DCN expression. Data shown are representative allograft stains from N=3/group.

## Figure S7

Α.



Β.



Fig. S7. Validation of monocyte depletion within lung recipients. (A) FACS plot gating strategy to measure allograft CCR2<sup>+</sup> monocyte abundance of CCR2<sup>DTR</sup> recipients of 3T FVB allografts 3 days after receiving PBS or 10 ng/g body weight of i.v. diptheria toxin (DTx). Data shown is a representative result of 2 independent experiments where Lin represents a cocktail of CD90.2, B220, NK1.1 and Ly6G-specific Abs. (B) B6 recipients of 3T FVB allografts received 200  $\mu$ g i.v. of CCL2 neutralizing Abs on POD 6, 9 and 12 and assessed for blood peripheral CCR2<sup>+</sup> accumulation on POD 16. FACS plots shown is representative result from 2 experiments.



Fig. S8. HA accumulation in 3T-FVB allografts and CCR2 activity inhibition prevents intragraft Mo-AM accumulation. (A) HA concentration in the BAL fluid from POD 16 3T-FVB and 2T-FVB allograft. Data shown is a representative result from 2 determinations for N=5/group with a mean HA concentration  $\pm$  S.D.; 2-sided Mann Whitney U test; \*p<0.05. Intragraft 3T FVB Mo-AM numbers transplanted into POD16 CCR2<sup>DTR</sup> recipients that received indicated treatments or transplanted into B6 recipients that received CCL2 Ab. Data shown is for N=5/group with mean Mo-AM numbers  $\pm$  S.D; 1-way ANOVA with Dunnett's multiple comparisons test; \*\*\*p<0.001.



**Fig. S9**. **Validation of TGF-**β**R2 deletion in CCR2<sup>+</sup> monocytes**. TGF-βR2<sup>fl/fl</sup> and TGF-βR2<sup>Δ/Δ</sup> mice were treated tamoxifen i.p. every other day for 10 days, rested for 5 days, FACS sorted for peripheral CCR2<sup>+</sup> blood monocytes and assessed for (**A**) *Tgfbr2* transcript levels or (**B**) stimulated overnight with 5 ng/ml of TGF-β1 and measured for *Pparg* and *Car4* transcript levels. For (**A**) data shown is normalized to the macrophage housekeeping gene *Stx5a* and (**B**) shown normalized to TGF-βR2<sup>Δ/Δ</sup> levels where bars represent means ± S.D. Data shown for **A** and **B** are representative results from 2 determinations.

Figure S10



**Fig. S10**. **T**<sub>RM</sub> **phenotype and abundance in lung transplants.** (A) Intragraft numbers and percent abundance of T<sub>RM</sub> for indicated transplant and treatment conditions. Bars represent mean levels  $\pm$  S.D.;N=4/group; 1-way ANOVA with Dunnett's's multiple comparisons test \*\*p < 0.01. (B) A representative FACS histogram result of T<sub>RM</sub> LAG-3 and TIM-3 expression of N=4/group. (C) 2T-FVB allograft T<sub>RM</sub> marker expression patterns. Data shown are a representative result from N=4/group.

## Figure S11



**Fig. S11**. **Clusters of Gzmb<sup>+</sup>CD49a<sup>+</sup>CD8<sup>+</sup>T cells in BOS subjects.** Explant tissue from patients with BOS or core biopsies from patients without evidence of CLAD or antibody-mediated rejection (Stable) were stained with indicated antibodies. Images shown are representative stains from BOS (N=4) and stable recipients (N=6). The white box represents a cell cluster and arrows show a single cell.



**Fig. S12**. **CD8**<sup>+</sup> **T** cell-specific Blimp-1 deficiency reduces intragraft IFN-γ<sup>+</sup> **CD8**<sup>+</sup> **T** cell accumulation. Flow cytometric analysis of indicated intragraft effector T cells in Blimp-1<sup>ftfl</sup> and Blimp-1<sup>Δ/Δ</sup> recipients (N=4/group). Data shown is mean percent abundance ± S.D.; 2-sided Mann-Whitney U test; \*\*p < 0.01.