



- 962 **Supplementary Fig. 1.** Protein-sharing network between viral contigs from Challenger
- 963 Deep ( $n = 1$  628), other hadal and non-hadal deep-sea ( $n = 2$  885) and wetland sediments
- 964 (n = 1 212), Global Ocean Virome 2.0 (n = 195 728), thawed permafrost soils (n=1 907),
- 965 and vContact2 RefSeq prokaryotic viral genomes  $(n = 2 010)$ . Nodes (circles) represent
- 966 viral contigs. Shared edges (lines) indicate shared protein content. Well-supported viral
- 967 clusters containing CD vOTUs were shown in this figure.



 **Supplementary Fig. 2.** Normalized coverages of viral populations (species level) in CD and reference deep-sea metagenomes. Each row represents an individual CD metagenome, while each column represents an individual CD viral population. The normalized relative abundance values (log2-transformed) are shown in color scale. Red, blue and black lines represent the hierarchical clustering of slope, bottom-axis and other deep-sea sediments, respectively.



slope and bottom-axis samples. The bubbles indicate the normalized coverage (coverage

per giga base), and the predicted lifestyle of viruses for each sample.



 **Supplementary Fig. 4.** Sequencing depth of CD viromes in number of total paired reads per metagenome. (a) The number of paired-end reads mapped to CD vOTUs. (b) Percentage of paired-end reads mapped to CD vOTUs in metagenomes. *p*-value was estimated using the two-sided Wilcoxon test. For boxplots, centre line indicates median, bounds of box indicate 25th and 75th percentiles, and whiskers indicate minimum and maximum.



 **Supplementary Fig. 5** Overview of putative AMGs in CD viral communities. (**a**) Number of putative AMGs involved ranked by AMGs prevalence in CD virome (top fifteen shown). (**b**) Abundance of viruses that have genes belonging to the fifteen most prevalent AMGs identified in CD sediment viromes. The genes identified in various KEGG-described metabolisms are listed in Supplementary Data 9. Relative abundances were log2 transformed and normalized with a z-score method based on normalized vOTUs coverage.



 **Supplementary Fig. 6.** Bray-Curtis dissimilarity principal coordinate analyses (PCoA) of the viral AMGs. The pink and blue areas covered slope and bottom-axis samples, 1000 respectively.





 **Supplementary Fig. 7.** Similarity of each Cys protein (CysC/CysH) with the five closest homologues recruited from public viromes and the eggNOG database, respectively. Three CysC (**a**) and six CysH proteins (**b**) from CD viral contigs. Boxplots represent the median with 25th and 75th percentiles, and whiskers show the 1.5 interquartile range.



1010<br>1011 **Supplementary Fig. 8.** Maximum-likelihood phylogenetic tree between the identified CD CysH proteins (this study) and the CysH proteins found in eggNOG database (V5.0). The tree is rooted in the mid-point. Two CysH homologs from Uniport database are in blue, and indicate protein function based on experimental evidence. Bootstrap values (1 000 1015 replicates)  $\geq$  70% are indicated at nodes.



1019 **Supplementary Fig. 9.** Total arsenic and mercury concentrations ( $\mu$ g g<sup>-1</sup> of dry sediment) in 13 CD sites (5 400-10 911 m depth). *p* values were estimated using two-sided Wilcoxon

test for pairwise comparisons. CD sediments were analyzed for total arsenic and mercury

using Atomic Fluorescence Spectrometry. Bar graphs identify mean and standard error bars.



**Supplementary Fig. 10.** Genome maps of the viral contigs that contain LPS and dTDP-

L-rhamnose synthetic genes. The maps indicate the positions of these genes in the

viromes, and the upstream and/or downstream phage genes. Viral genes were annotated

by CheckV and VIBRANT. AMGs, non-phage-like and uncharacterized genes are shown

in blue, pink and gray, respectively. Detailed functional annotation of the viral scaffolds

can be found in Supplementary Data 8.

Cruise	Sample station	Sampling method	Latitude [North]	Longitude [East]	Water Depth [m]	Position in trench	Layers for metagenome [cmbsf]	Layers for metatranscriptome [cmbsf]
DY37II	MC <sub>02</sub>	push core (submersible)	11.764	141.976	5 4 0 0	slope	$8-10; 28-30$	
DY37II	$D1T1^*$	push core (submersible)	11.801	142.117	5 5 3 3	slope	$0-2; 4-6$	
DY37II	$D1T2^*$	push core (submersible)	11.801	142.117	5 5 3 3	slope	$4-6$ ; 24 $-26$	
<b>TS01</b>	T <sub>1</sub> B <sub>5</sub>	box core (hydrographic cable)	10.924	141.799	7 0 6 1	slope	$0-2$ ; 8-10; 28-30; 38-40	
<b>TS01</b>	T <sub>1</sub> B <sub>3</sub>	box core (hydrographic cable)	11.553	141.873	7082	slope	$0 - 3$	
<b>TS01</b>	T1B8	box core (hydrographic cable)	11.602	142.228	7 1 4 3	slope	$2-4$ ; 16-18	
<b>TS01</b>	T <sub>1</sub> L <sub>6</sub>	push core (lander)	11.091	142.073	7850	slope	$0-2$ (two push cores)	
<b>TS01</b>	T1B10	box core (hydrographic cable)	11.195	141.812	8638	slope	$0-2$ ; 36-38; 44-46	
<b>TS01</b>	T1B11	box core (hydrographic cable)	11.228	141.689	9 1 5 0	slope	$0 - 3$	
<b>TS03</b>	T3L11	push core (lander)	11.325	142.191	10 908	bottom-axis	$0-3$ ; 6-9; 12-15; 18-21	6-9; 12-15; 18-21
<b>TS03</b>	T <sub>3</sub> L <sub>8</sub>	push core (lander)	11.327	142.194	10 909	bottom-axis	$0-3$ ; 6-9; 12-15; 18-21	
<b>TS03</b>	T3L14	push core (lander)	11.325	142.189	10 911	bottom-axis	$0-2$ ; 4-6; 6-8; 12-14; 18-20	
<b>TS01</b>	T1L10	push core (lander)	11.328	142.202	10 900	bottom-axis	$0-3$ ; 6-9; 12-15; 18-21	

1032 **Supplementary Table 1.** Sampling sites and sampling methods used for collecting the sediment samples in CD

1033 #: sediment samples from two slope sites (~ 100m apart) collected in a single dive of the submersible "Jiaolong".



## 1035 **Supplementary Table 2. List of marine viruses related datasets**



1036 \* Estimated by multiplying the number of reads by the average read length

1037 # Shows datasets used for comparisons in this study

## **Supplementary references**

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