

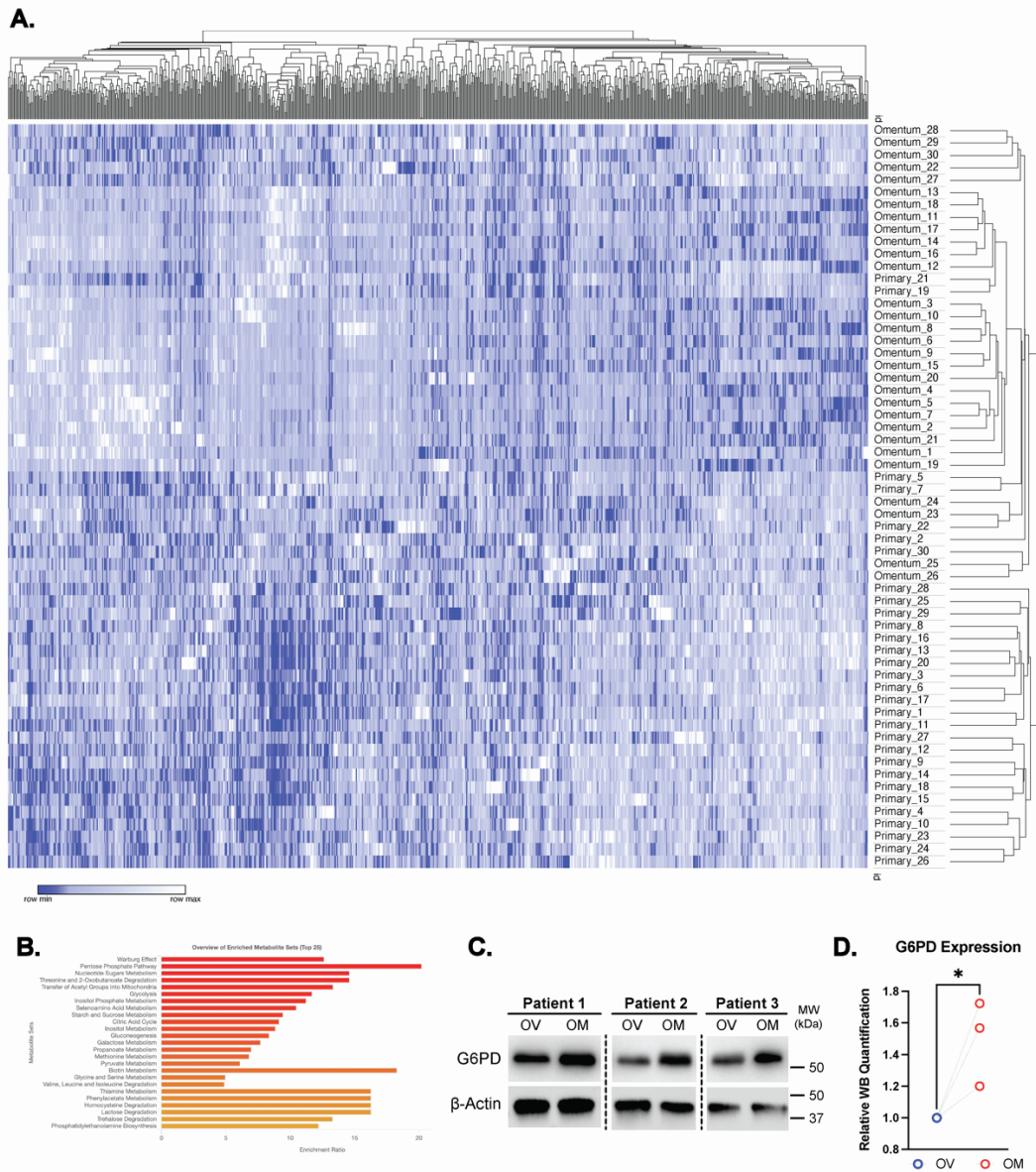
**Supplemental information**

**G6PD inhibition sensitizes ovarian cancer cells  
to oxidative stress in the metastatic  
omental microenvironment**

**Shree Bose, Qiang Huang, Yunhan Ma, Lihua Wang, Grecia O. Rivera, Yunxin Ouyang, Regina Whitaker, Rebecca A. Gibson, Christopher D. Kontos, Andrew Berchuck, Rebecca A. Previs, and Xiling Shen**

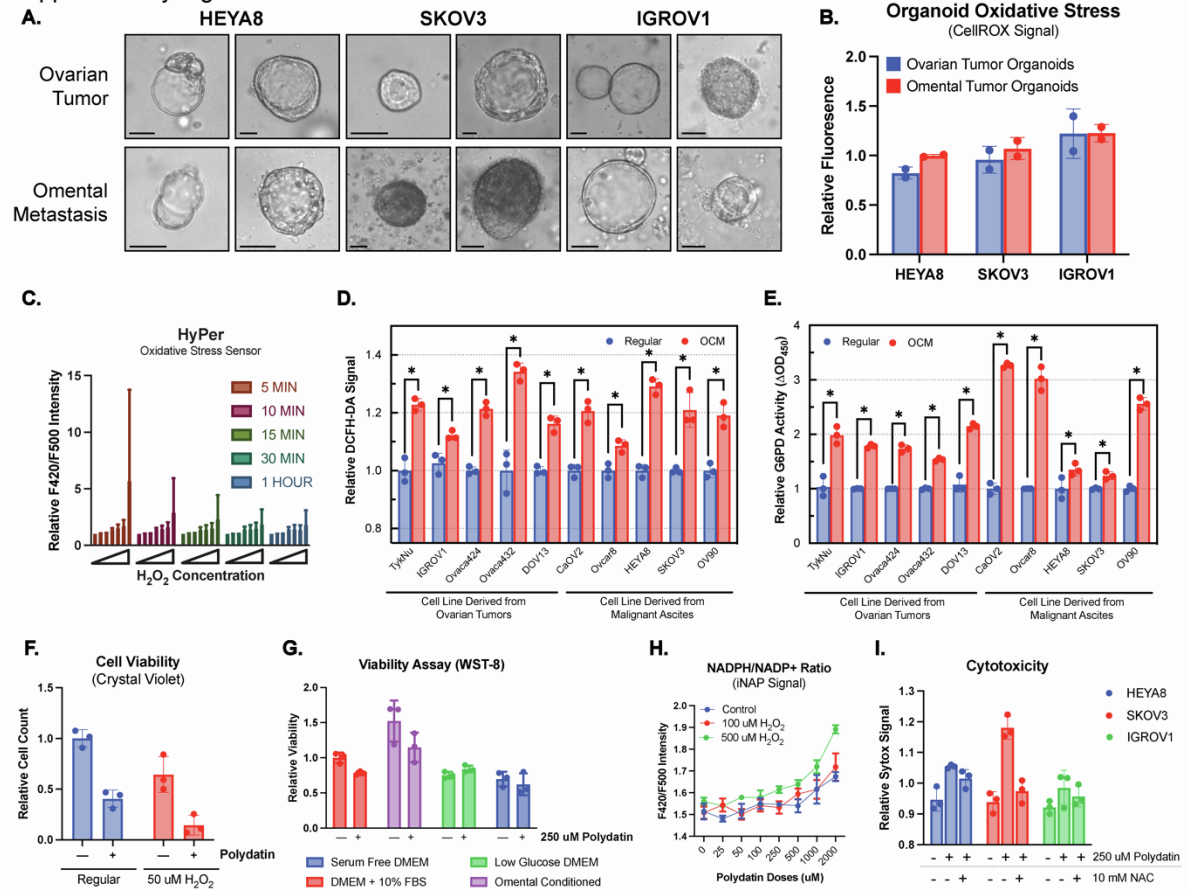
## Supplementary Figures

### Supplementary Figure 1



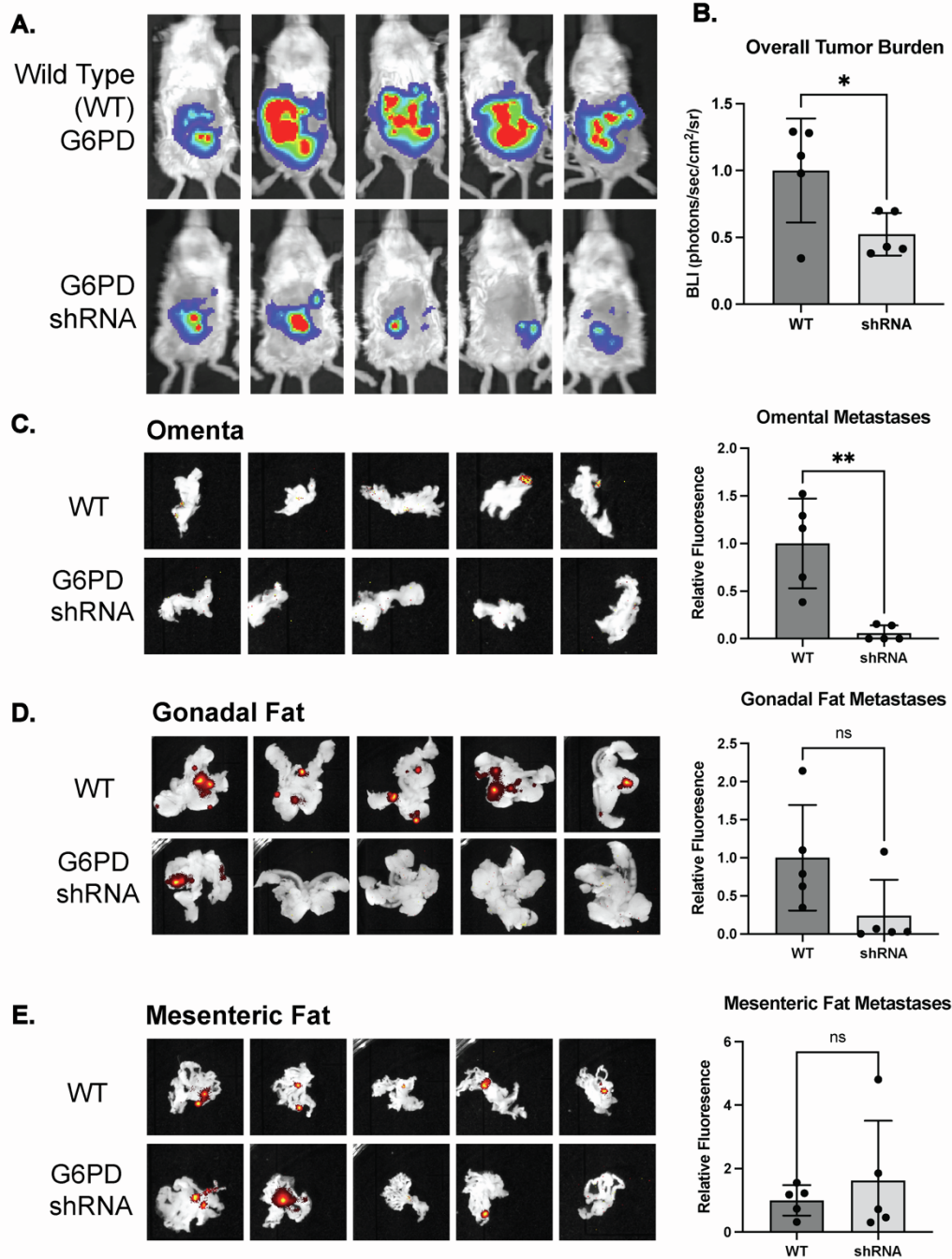
**Figure S1: Gene expression and metabolomics analysis highlight PPP changes in matched human ovarian and omental tumors, Related to Figure 1. (A)** Hierarchical clustering of heatmap of metabolic gene expression data from ovarian and omental tumors (n=30) show clustering of primary tumors and metastatic tumors independently. **(B)** Pathway analysis of metabolite abundance differences from LC-MS metabolomics analysis of matched primary ovarian tumor vs. omental metastases (n=8) highlight the pentose phosphate pathway. **(C)** Immunoblotting for G6PD in ovarian and omental tumors (n=3). **(D)** Quantification was performed by normalizing protein expression to the loading  $\beta$ -actin control and the primary tumor expression. Omental metastases exhibited increased G6PD expression (p-val<0.05, Student's *t*-Test).

Supplementary Figure 2



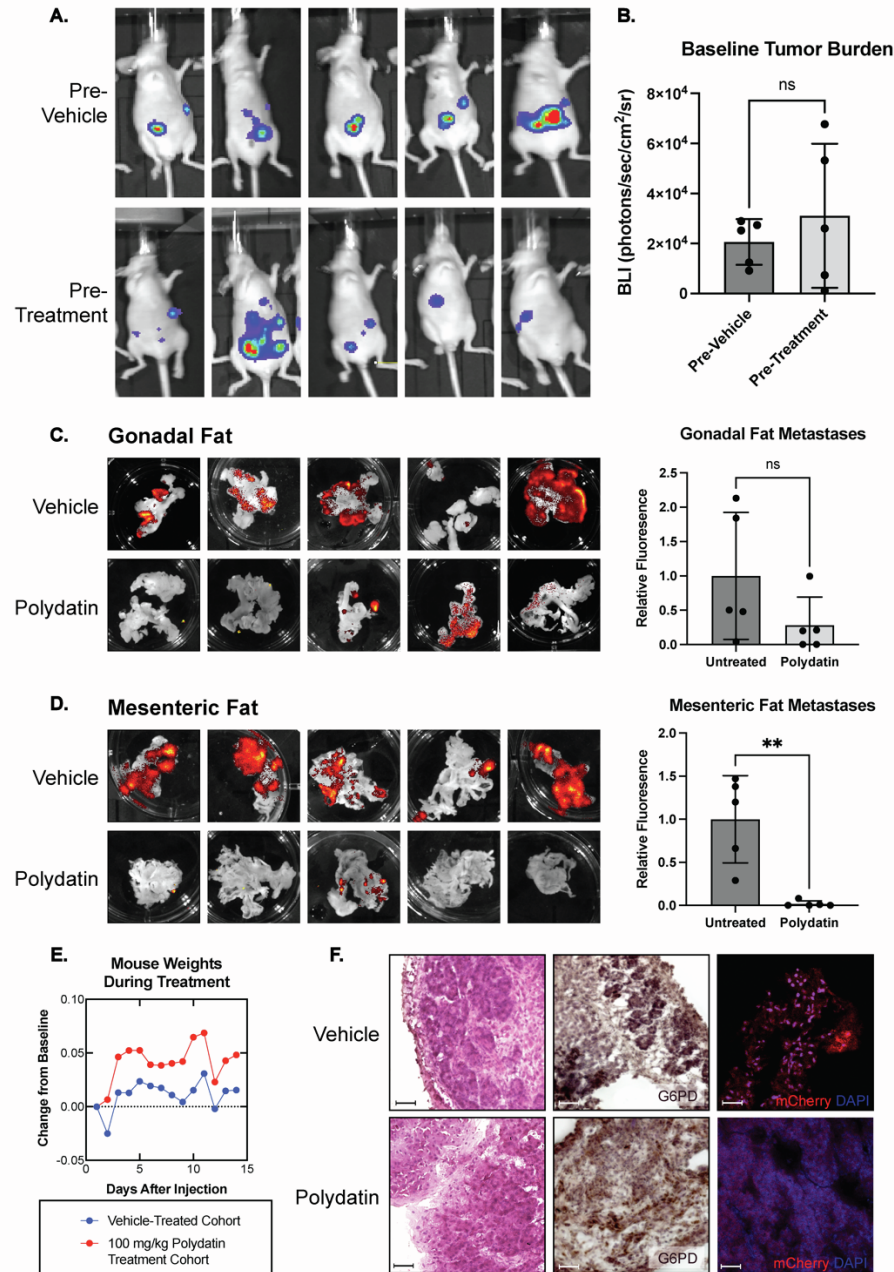
**Figure S2: *In vitro* models of OC reflect changes in oxidative stress, Related to Figure 3. (A)** Organoids were derived from murine ovarian tumors and omental metastases (n=2 per cell line). Scale bars represent 100  $\mu$ M. **(B)** Oxidative stress of organoids cultured in organoid media measured using CellROX reveal minimal differences (n=2). **(C)** Fluorimetry of OC cells expressing HyPer was responsive to hydrogen peroxide exposure (n=3). Growth in OCM induced increases in **(D)** oxidative stress as measured via DCFH-DA incubation (n=3) and **(E)** G6PD activity as measured via enzymatic assay in 10 OC cell lines (n=3). **(F)** Crystal Violet and **(G)** WST-8 cell viability assays revealed less viable cells evident in OCM-polydatin treated samples (n=3). **(H)** iNAP fluorescence revealed increased NADPH/NADP<sup>+</sup> in cells treated with polydatin (n=3). **(I)** N-acetylcysteine (NAC) treatment reversed these cytotoxic effects *in vitro* (n=3). The sample size (n) represents the number of technical replicates. Where indicated, statistical significance is noted using \* p<0.05, \*\* p<0.01 by two-tailed Student's *t*-Test.

Supplementary Figure 3



**Figure S3: G6PD-shRNA knockdown reduces omental metastases *in vivo*, Related to Figure 4.** (A) IVIS imaging and (B) quantification of tumor burden in NSG mice injected with wild-type and G6PD-shRNA expressing HEYA8-mCherry cells (n=5 per cohort). Fluorescence imaging and quantification of mCherry-expressing tumor burden in (C) omenta, (D) gonadal, and (E) mesenteric fat deposits (n=5 per cohort).

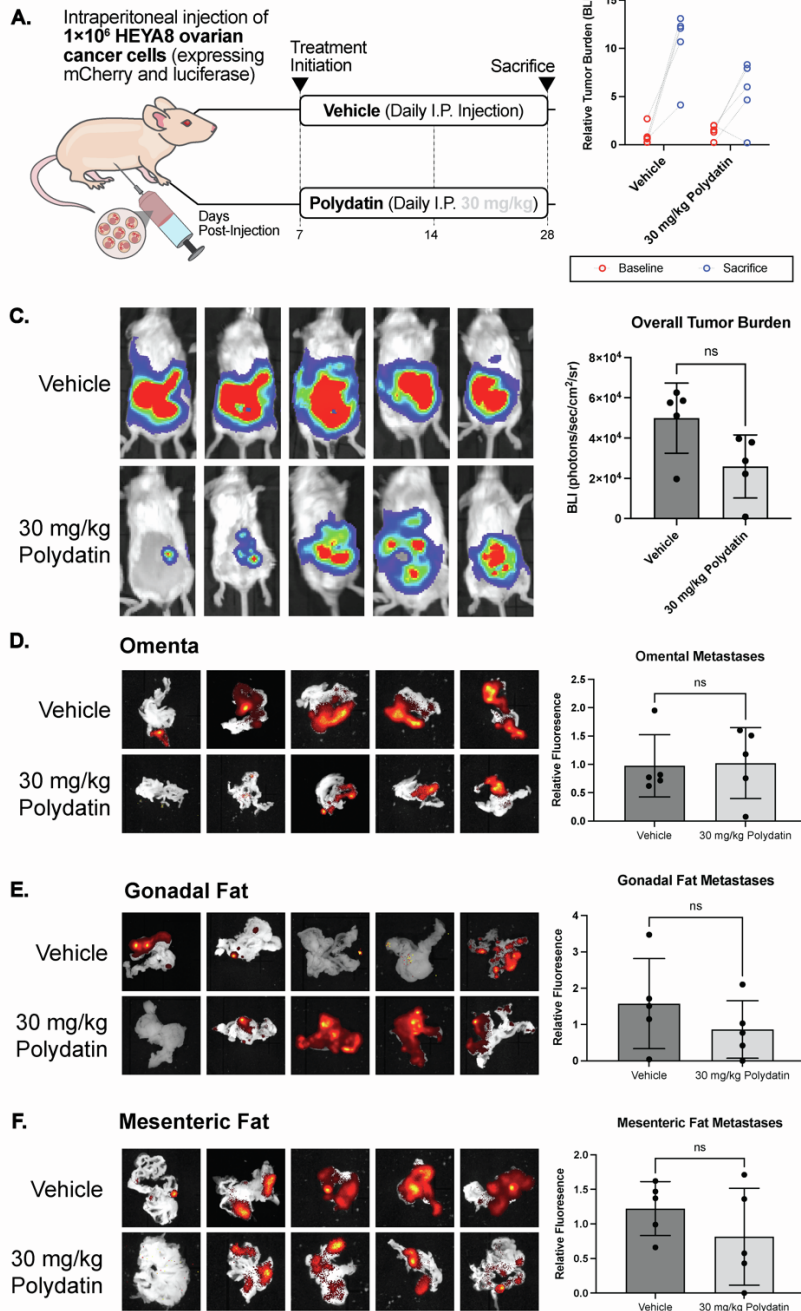
Supplementary Figure 4



**Figure S4: High-dose polydatin treatment (100 mg/kg) reduces mesenteric metastases *in vivo*, Related to Figure 4. (A) IVIS imaging and (B) quantification of baseline tumor engraftment taken 3 days post-injection of HEYA8-mCherry injected mice (n=5 per cohort). Fluorescence imaging and quantification of mCherry-expressing tumor burden in (C) gonadal and (D) mesenteric fat deposits. (E) Weights of mice did not vary significantly during course of treatment (each symbol is average of 5 mice per cohort, no statistically significant differences seen). (F) H&E, G6PD staining, and mCherry microscopy of omental tumors revealed similar tumor morphology between vehicle and polydatin treated tumors. Scale bars represent 100  $\mu$ M.**



Supplementary Figure 5



**Figure S5: Low-dose polydatin (30 mg/kg) treatment does not achieve therapeutic threshold *in vivo*, Related to Figure 4. (A) Schema for treatment. (B) Bioluminescence (BLI) was measured at baseline and prior to sacrifice using IVIS (n=5 per cohort). (C) IVIS imaging and BLI quantification prior to sacrifice shows tumor burden in 30 mg/kg polydatin treated and vehicle treated mice (n=5 per cohort). Fluorescence imaging and quantification of mCherry-expressing tumor burden in (D)**

omenta, (E) gonadal, and (F) mesenteric fat deposits (n=5 per cohort). Labels of n.s. indicate “not significant” (p-val>0.05)

<b>Media Component</b>	<b>Final Concentration</b>
<b>DMEM/F12</b>	
<b>GlutaMax</b>	1x
<b>Penicillin/Streptomycin</b>	100 U/mL
<b>17-B-Estradiol</b>	10 nM
<b>A083-01</b>	250 nM
<b>B27 (without Vitamin A)</b>	1x
<b>EGF</b>	50 ng/mL
<b>HGF</b>	10 ng/mL
<b>IGF1</b>	20 ng/mL
<b>N2 Supplement</b>	1x
<b>N-Acetylcysteine</b>	5 mM
<b>Neuregulin I</b>	10 ng/mL
<b>Nicotinamide</b>	5 mM
<b>Noggin</b>	100 ng/mL
<b>R-Spondin 1</b>	50 ng/mL
<b>SB203580 (p38i)</b>	1 uM
<b>Y-27632</b>	10 uM

**Table S1: OC Organoid Media Composition, Related to STAR Methods.**

<b>Patient Number</b>	<b>Ovarian Tumor ID</b>	<b>Omental Tumor ID</b>	<b>FIGO Stage</b>	<b>Age</b>
<b>1</b>	1579	1577	4	37
<b>2</b>	1716	1714	4	70
<b>3</b>	1844	1842	3C	67
<b>4</b>	1925	1924	4	73
<b>5</b>	3427	3428	3C	45
<b>6</b>	3454	3451	3C	57
<b>7</b>	3474	3475	3C	45
<b>8</b>	6053	6054	3C	68

**Table S2: Information About Human Tumor Samples, Related to STAR Methods.**