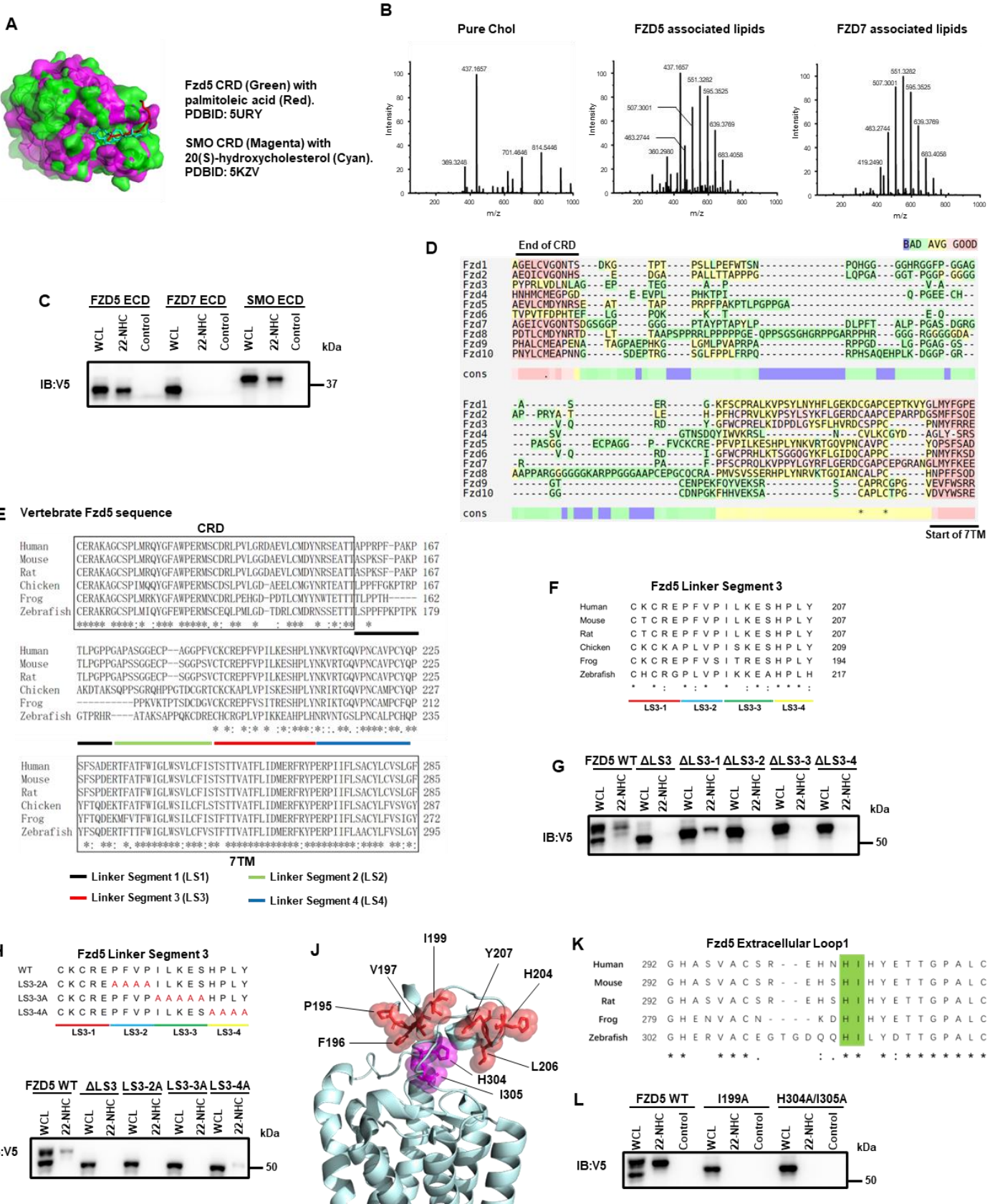


Supporting Information

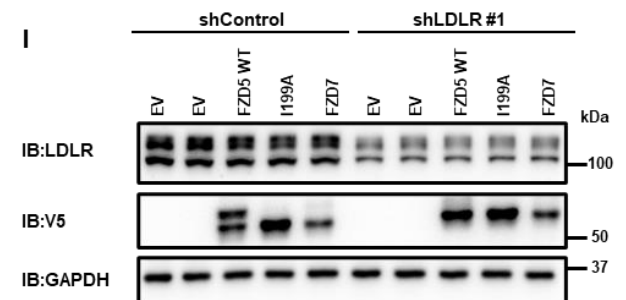
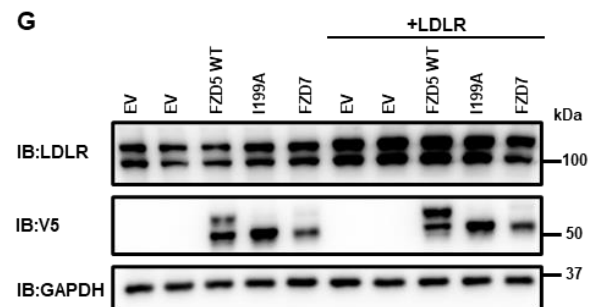
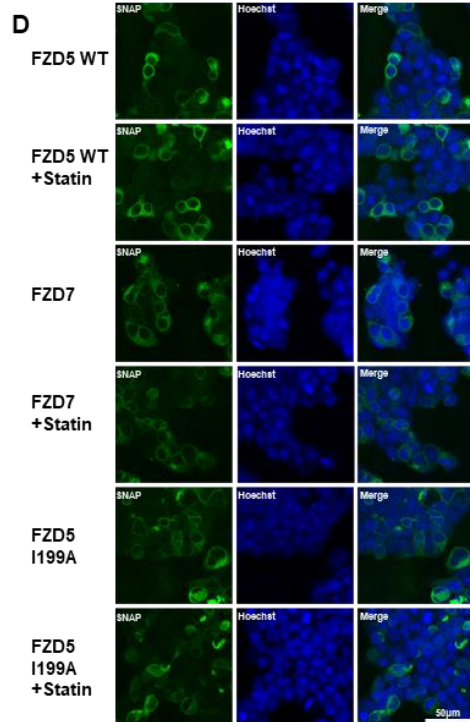
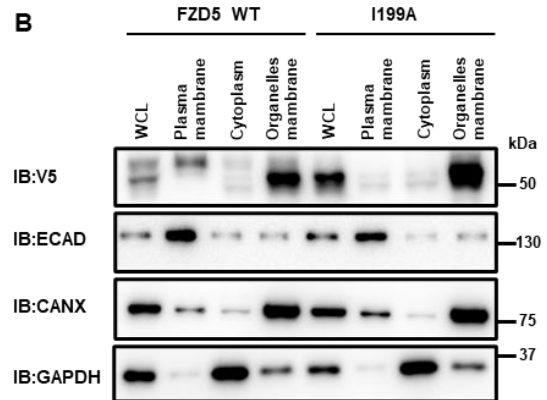
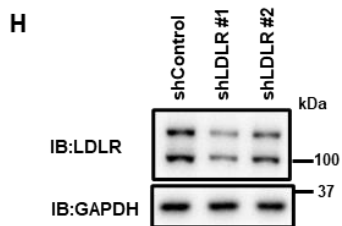
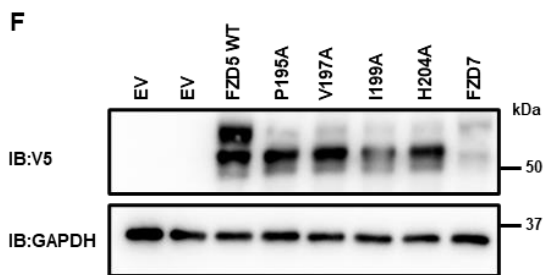
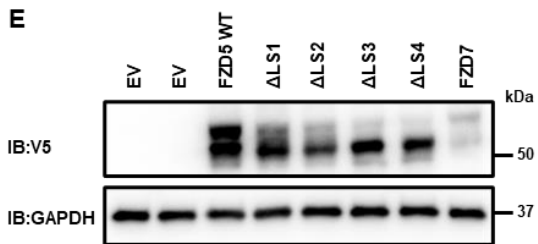
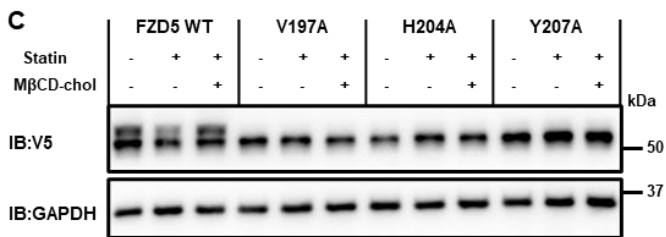
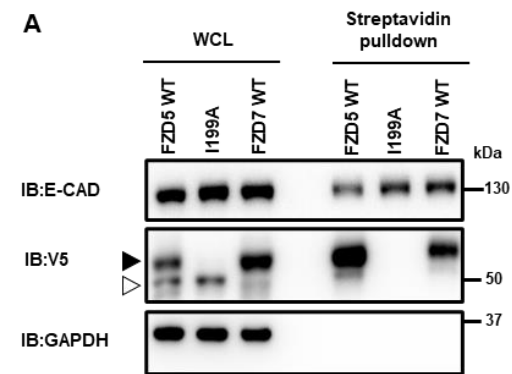
**Aberrant cholesterol metabolism and Wnt/ β -catenin signaling coalesce via
Frizzled5 in supporting cancer growth**

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Haijiao Zhang, Xi Zhang, Maorong Chen, Xinjun Zhang, Chao Zhao, Jun Qi, Liu Cao,
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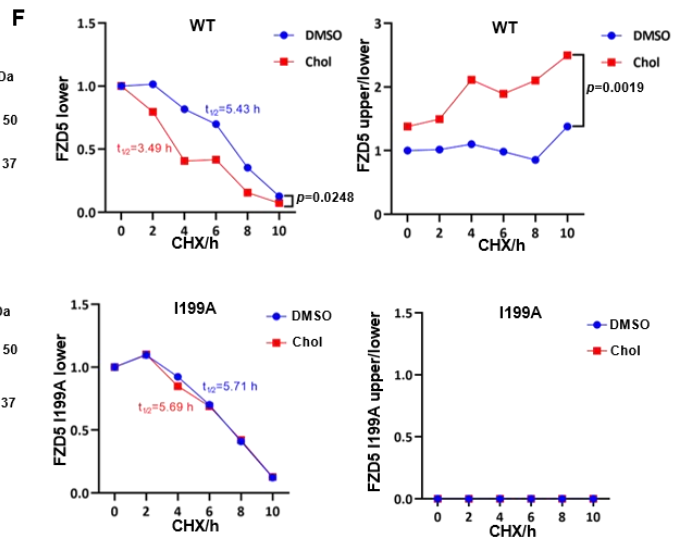
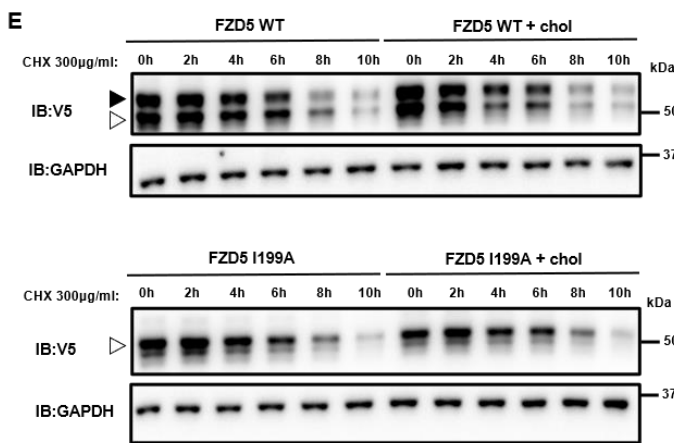
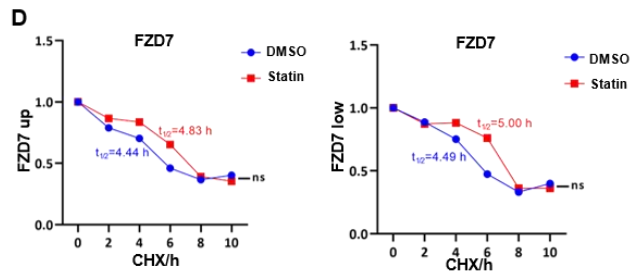
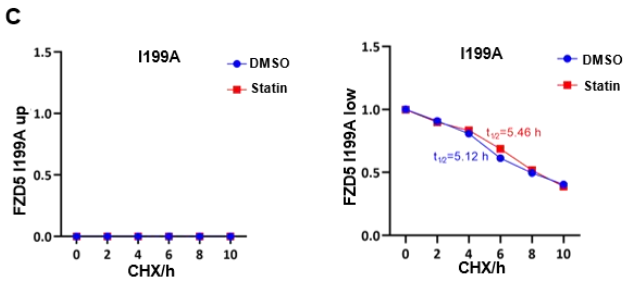
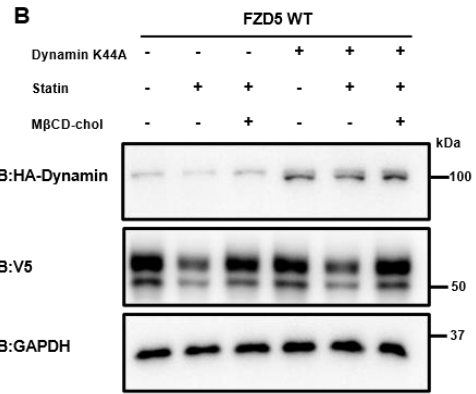
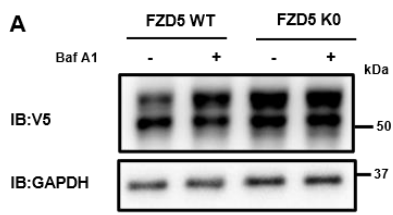
Supplemental figures and legends



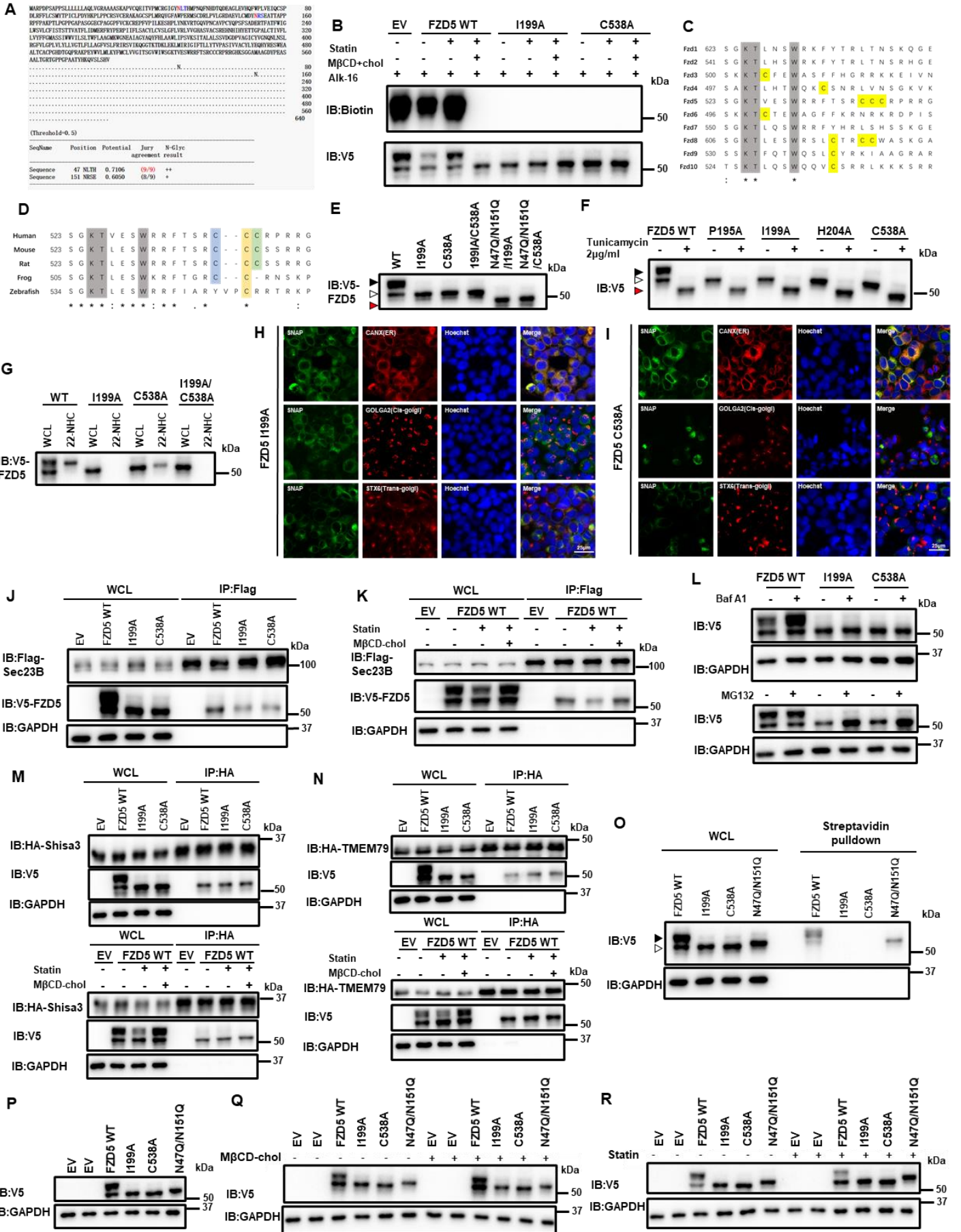
Supplemental Figure 1. Supporting information and assays of Fzd5-cholesterol bindings. A) Overlay of Fzd5 and Smo CRDs by Pymol (PDBID: 5URY and 5KZV). B) Full spectra (m/z range: 50-1000) of mass-spectrometry assay of pure cholesterol, Fzd5-associated lipids and Fzd7-associated lipids. C) 22-NHC pulldown and WB assay of secreted Fzd5, Fzd7 and Smo ECDs. D) Amino acid sequence alignment of human Fzd1-10 extracellular linker region by T-coffee (<http://tcoffee.crg.cat/>). The different conservation levels are highlighted by different colors. The result shows that the extracellular linker region is not conserved among Fzds. E) Amino acid sequence alignment by clustalW of the extracellular linker region of vertebrate Fzd5 and demonstration of LS1, LS2, LS3 and LS4. F) Amino acid sequence alignment of Fzd5 LS3 in vertebrate and demonstration of LS3-1, LS3-2, LS3-3 and LS3-4. G) 22-NHC beads pulldown assay of Fzd5 WT, Δ LS3 and smaller truncations (Δ LS3-1, Δ LS 3-2, Δ LS 3-3 and Δ LS 3-4). H) Demonstration of LS3-2A, LS3-3A and LS3-4A (Replacing all amino acids with Ala). I) 22-NHC beads pulldown and WB assay of Fzd5 WT and mutations in LS3. J) Demonstration of conserved residues on LS3 that involves in cholesterol binding (red) and adjacent aromatic/hydrophobic conserved residues in ECL1 (pink) based on the Fzd5 structure model (PDBID: 6WW2). K) Sequence alignment of Fzd5 ECL1 among vertebrates. The conserved His and Ile are highlighted in green. L) 22-NHC beads pulldown assay of Fzd5 WT, I199A and H304A/I305A.



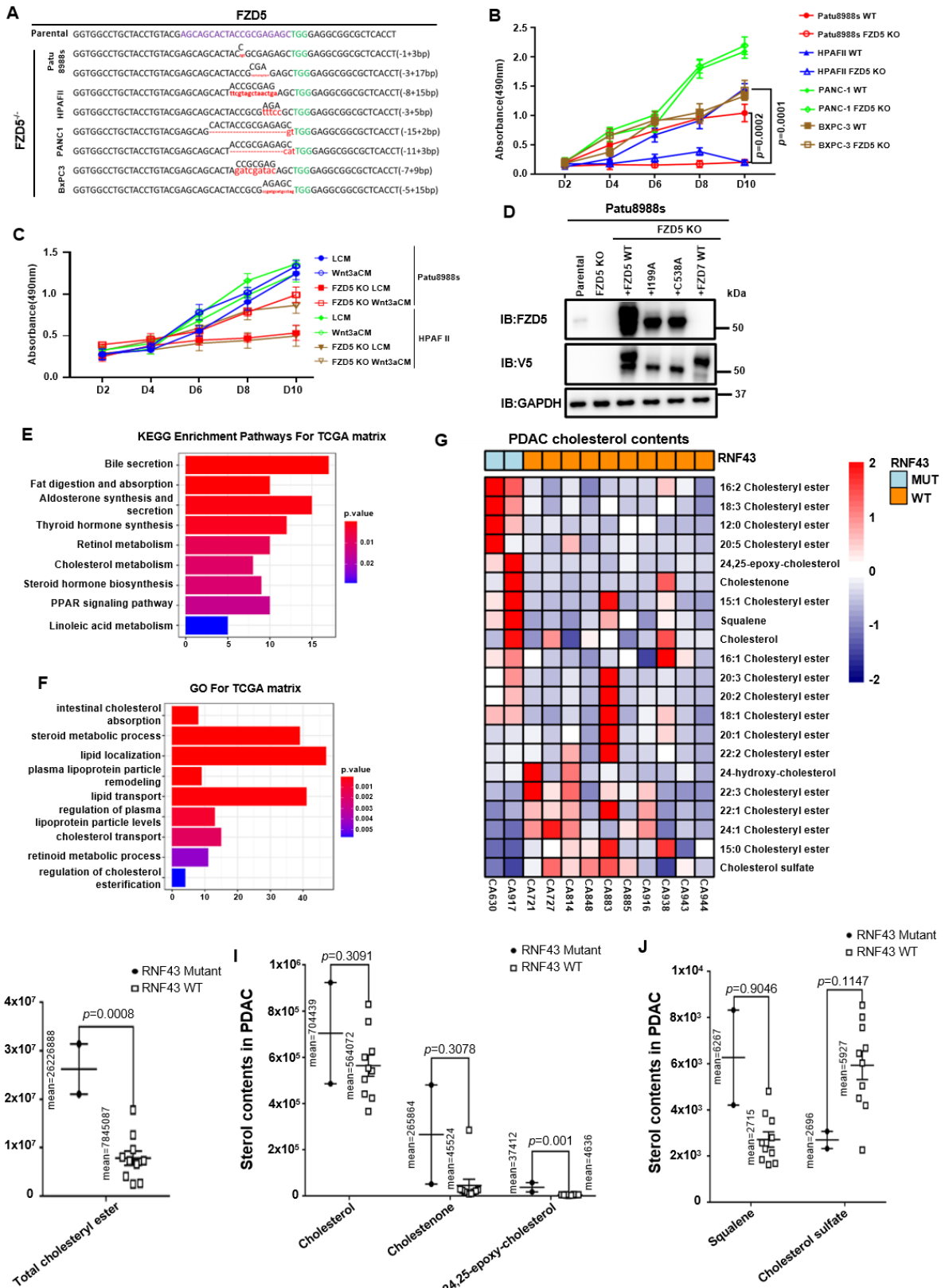
Supplemental Figure 2. Cholesterol promotes Fzd5 PM localization and Wnt signaling. A) Surface biotin labeling assay of Fzd5 WT, I199A and Fzd7. B) Cellular fractionation assay of Fzd5 WT and I199A in HEK293T cells. E-CAD: PM marker. CANX: inner membrane marker. GAPDH: cytoplasm marker. C) Mature/immature band pattern of Fzd5 WT and other cholesterol binding LoF mutations besides I199A in HEK293T cells. D) Confocal fluorescent imaging of total SNAP-tagged Fzd5, Fzd7 and I199A under normal and statin treatment conditions in HEK293T cells. SNAP-tagged proteins are labelled with cell-permeable SNAP Cell Oregon Green. Hoechst labels the nucleus. All images in the panel are in the same scale. E) Protein expression of the TOPFlash assay in Figure 2D in Fzd-null HEK293T. F) Protein expression of the TOPFlash assay in Figure 2E in Fzd-null HEK293T. G) Protein expression of the TOPFlash assay in Figure 2K in Fzd-null HEK293T. H) Validation of LDLR knockdown by WB in Fzd-null HEK293T. I) Protein expression of the TOPFlash assay in Figure 2L in Fzd-null HEK293T.



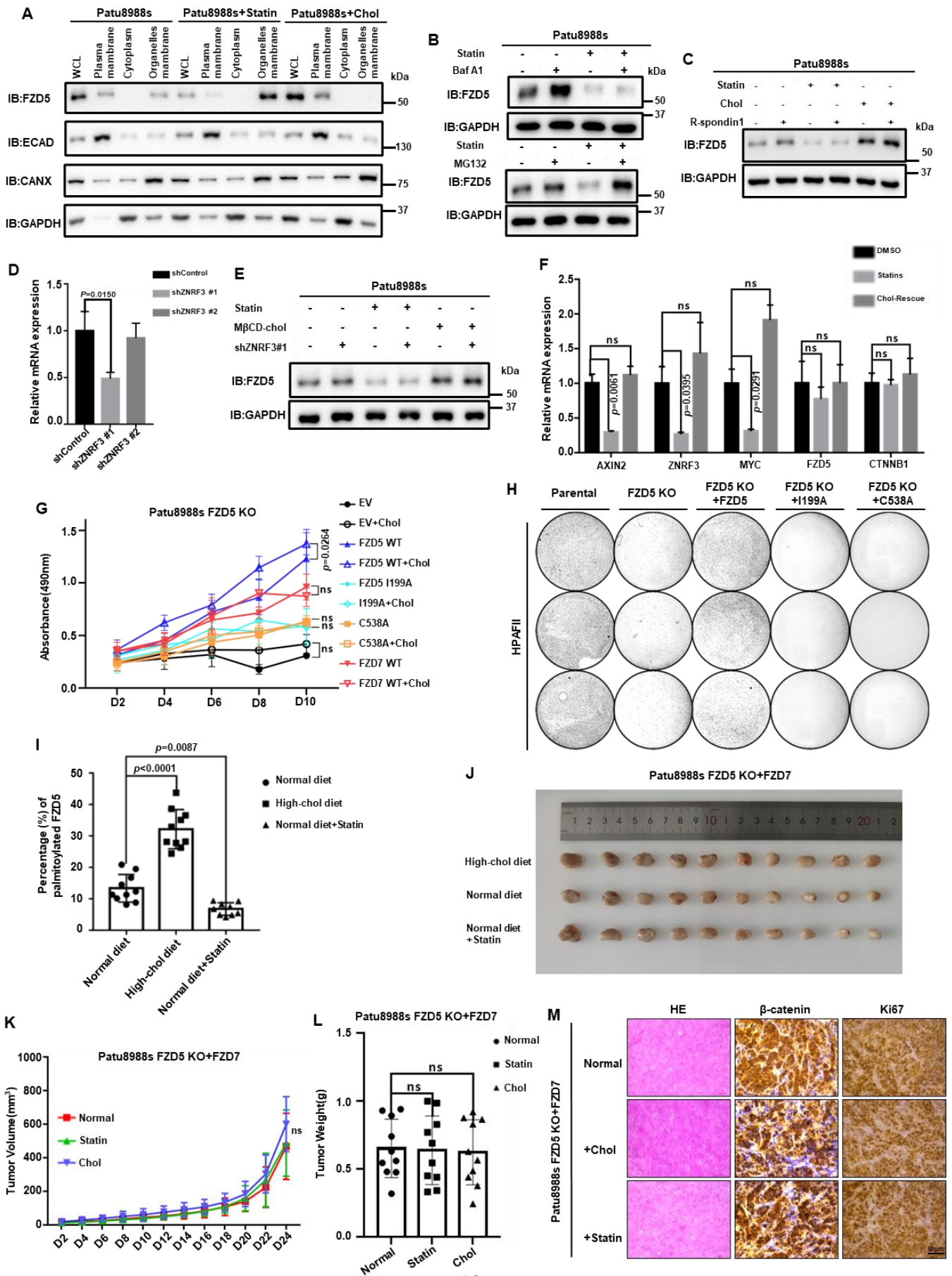
Supplemental Figure 3. Supporting assays of cholesterol regulating Fzd5 independent of ubiquitination-lysosomal degradation. A) Treatment of Baf A1 on Fzd5 WT and K0 in HEK293T. B) WB assay showing cholesterol does not affect endocytosis by employing dominant negative dynamin in HEK293T. C) The quantification by ImageJ of the upper/lower protein and the half-lives in Figure 3H. D) The quantification by ImageJ of the upper/lower protein and the half-lives in Figure 3I. By two-tailed unpaired student's t-test analysis. E) Pulse chase assay of Fzd5 WT and I199A under normal and additional cholesterol treatment conditions. The solid arrow head represents the upper mature Fzd and the hollowed arrow head represents the lower immature Fzd. F) The quantification by ImageJ of the protein and upper/lower ratio of the blots in E). By two-tailed unpaired student's t-test analysis. ns: not significant.



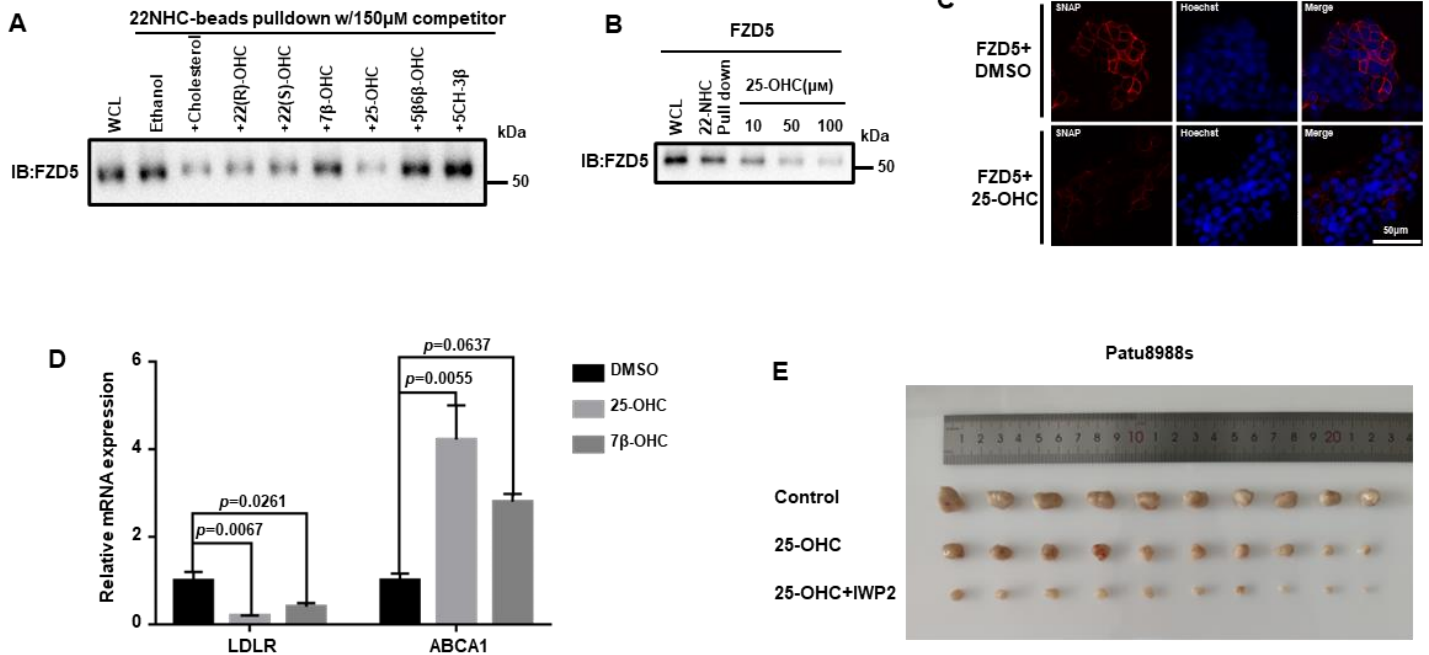
Supplemental Figure 4. Supporting analysis and assays of the cholesterol binding dependent S-palmitoylation of Fzd5, maturation and Wnt signaling. A) N-glycosylation prediction of Fzd5. B) Click chemistry and pulldown assay showing Fzd5 palmitoylation by Alkyne-palmitic acid. C) Sequence alignment of the cytoplasmic tail of 10 Fzd receptors. KTxxxW motif is highlighted in grey. Cysteines are highlighted in yellow. The three-in-a-row Cys pattern is unique for Fzd5. D) Sequence alignment of the cytoplasmic tail of Fzd5 among vertebrates. KTxxxW motif is highlighted in grey. Three-in-a-row Cys pattern is conserved among mammals and at least one Cys at the cytoplasmic tail is conserved in vertebrates. E) Band pattern of Fzd5 WT and mutants. The solid arrow head represents the upper mature Fzd. The hollowed arrow head represents the lower immature Fzd. The red solid arrow head represents the naïve Fzd without modifications. F) Band pattern of Fzd5 WT and mutants under the treatment of tunicamycin. The solid arrow head represents the upper mature Fzd. The hollowed arrow head represents the lower immature Fzd. The red solid arrow head represents the naïve Fzd without modifications. G) 22-NHC beads pulldown assay of Fzd5 WT, I199A, C538A, I199A/C538A. H, I) Confocal fluorescent microscopic images showing Fzd5 WT subcellular localizations of Fzd5 I199A H) and Fzd5 C538A I) in HEK293T. SNAP-Fzd5 is labelled by cell-permeable dye. CANX, GOLGA2 and STX6 staining are immunofluorescent. All images are in the same scale. J) Co-immunoprecipitation of Fzd5 WT, I199A and C538A with COPII complex subunit Sec23B. K) Co-immunoprecipitation of Fzd5 under normal, statin treatment and cholesterol rescue conditions with Sec23B. L) Fzd5 WT, I199A and C538A stability test under Baf A1 and MG132 treatment. M) Co-immunoprecipitation assay of Fzd5 WT and mutants with Shisa (upper) or Fzd5 WT with Shisa during cholesterol perturbations (lower). N) Co-immunoprecipitation assay of Fzd5 WT and mutants with TMEM79 (upper) or Fzd5 WT with TMEM79 during cholesterol perturbations (lower). O) Surface biotin labeling of Fzd5 WT, I199A, C538A and N47Q/N151Q. P) Protein expression of the TOPFlash assay in Figure 4J in Fzd-null HEK293T. Q) Protein expression of the TOPFlash assay in Figure 4K in Fzd-null HEK293T. R) Protein expression of the TOPFlash assay in Figure 4L in Fzd-null HEK293T.



Supplemental Figure 5. Supporting information and assays showing RNF43-mutant PDAC has Fzd5 dependence and aberrant cholesterol metabolic pattern. A) Sanger sequence of Fzd5 KO cell lines. B) MTT cell growth assay for parental PDAC and their corresponding Fzd5 KO stable cell lines. Error bars mean \pm SD, n = 3 replicates, by two-way ANOVA analysis. C) MTT cell growth assays of PDAC cell lines under LCM or Wnt3a CM treatment. n = 3 replicates. D) WB validation of the protein expressions in Figure 5A. E, F) KEGG E) and GO F) enrichments of the differentially expressed genes between RNF43-mutant and RNF43-WT PDAC in TCGA database. G) Lipid content heatmap of cholesterol metabolites showing that RNF43-mutant PDAC has a distinct cholesterol metabolic pattern compared to RNF43-WT PDAC. Each row represents a specific metabolite and each column represents a specific patient. H) Total cholesteryl ester content in RNF43-mutant PDAC is significantly higher than that in RNF43-WT PDAC. Error bars mean \pm SEM, by two-tailed unpaired student's t-test analysis. I, J) Comparison of unesterified cholesterol metabolite contents between RNF43-mutant and -WT groups by histograms. Error bars mean \pm SEM, by two-tailed unpaired student's t-test analysis.



Supplemental Figure 6. Supporting assays showing cholesterol regulating RNF43-mutant PDAC growth via Fzd5-mediated Wnt/ β -catenin signaling. A) Cellular fractionation assay of endogenous Fzd5 under different treatments in Patu8988s cells. B) Endogenous Fzd5 level test under Baf A1 and MG132 treatment in Patu8988s cells. C) Endogenous Fzd5 level test upon Rspo1 treatment in Patu8988s cells. D) Knockdown of ZNRF3 validation by RT-qPCR. Error bars mean \pm SD, n = 3 replicates, by one-way ANOVA analysis. E) Endogenous Fzd5 level test upon ZNRF3 knockdown in Patu8988s cells. F) RT-qPCR assays of Wnt target genes, FZD5 and CTNNB1 under cholesterol starvation or rescue conditions in Patu8988s cells. Error bars mean \pm SD, n = 3 replicates, by one-way ANOVA analysis. G) MTT cell growth assays of different Fzds rescued Patu8988s Fzd5 KO cell under normal or additional cholesterol conditions. Error bars mean \pm SD, n = 3 replicates, by two-way ANOVA analysis. H) Colony formation assays of HPAF-II parental and Fzd5-KO cells with different rescue conditions. n = 3 replicates. I) Quantification of immunoblotting of S-palmitoylated endogenous Fzd5 percentage in the transplanted tumors shown in Figure 5K after APE-assay. Each data point represents an individual tumor. For high cholesterol diet and normal diet groups, n=10; for stain treated group, n=9. Error bars mean \pm SD, by one-way ANOVA analysis. J) The PDAC tumors formed by subcutaneous implantations of Patu8988s Fzd5-KO Fzd7-rescue cells. From top to bottom, the rows represent high-cholesterol diet fed group, normal diet fed group and normal diet plus pravastatin fed group, respectively. n = 10 for each group. K) Subcutaneously implanted tumor size measurement by days. Error bars mean \pm SD, n = 10 for each group, by two-way ANOVA analysis. L) Subcutaneously implanted tumor weight measurements after section at Day24. Error bars mean \pm SD, n = 10 for each group, by one-way ANOVA analysis. M) Immunohistochemistry of subcutaneously implanted tumors and hematoxylin-eosin staining. ns: not significant.



Supplemental Figure 7. Supporting data showing 25-OHC treatment effects on Fzd5 PM localization and Fzd5-mediated PDAC growth. A) 22-NHC beads pulldown and competition assay. Endogenous Fzd5 is pulled down by 22-NHC and competed by various oxysterols such as 25-OHC. B) 22-NHC beads pulldown and competition assay of endogenous Fzd5 by different doses of 25-OHC. C) Confocal fluorescent imaging of SNAP-tagged Fzd5 on PM under normal and 25-OHC treatment conditions in HEK293T. All images are in the same scale. D) RT-qPCR assay of cholesterol metabolism genes under the treatment of 25-OHC and 7 β -OHC in Patu8988s cells. Error bars mean \pm SD, n = 3 replicates, by one-way ANOVA analysis. E) The PDAC tumors formed by subcutaneous implantations of Patu8988s cells. From top to bottom, the rows represent control group, 25-OHC treated group and 25-OHC+IWP2 treated group, respectively. n = 10 for each group.

Supplemental Table 1. PDAC patients' basic information

Supplemental Table 2. Mass spectrometry original data for the untargeted liposome analysis