

A Gene dosage-dependent effect unveils NBS1 as both an haploinsufficient tumor suppressor and an essential gene for SHH-medulloblastoma

^{1,6*} Marialaura Petroni, ^{1*} Francesca Fabretti, ^{1*} Stefano Di Giulio, ¹ Vittoria Nicolis Di Robilant, ¹ Veronica La Monica, ² Marta Moretti, ¹ Francesca Belardinilli, ¹ Francesca Bufalieri, ² Anna Coppa, ^{3,4} Paola Paci, ¹ Alessandro Corsi, ² Enrico De Smaele, ¹ Sonia Coni, ¹ Gianluca Canettieri, ¹ Lucia Di Marcotullio, ⁵ Zhao-Qi Wang, ^{1,6} Giuseppe Giannini.

Supplementary Materials and Methods

Genomic DNA isolation from mouse tails

Mouse tissues were digested at 55 C° O/N in DNA extraction buffer (50mM Tris-HCl pH 8.0, 100mM NaCl, 100mM EDTA pH 8.0, 1% SDS) and Proteinase K (Sigma Aldrich); DNA was isolated by Phenol extraction technique and quantified by spectrophotometry (Nanodrop, ThermoFisher Scientific, Waltham, MA, USA). Approximately 100-200 ng of DNA was used for PCR analysis using Taq-Platinum polymerase (Invitrogen, Carlsbad, CA, USA). SmoA1 transgene copy number was calculated using qPCR analysis. Probes and primer sequences are given in Supplementary Tables 1 and 2.

GCPs adhesion medium

Neurobasal medium supplemented with 0.6% glucose, 25 µg/ml insulin (Sigma Aldrich), 60 µg/ml N-acetyl-L-cystein (Santa Cruz Biothecchnology), 2 µg/ml heparin (Sigma Aldrich), penicillin-streptomycin, 2% B27 supplement without vitamin A (Life technologies), 2 µg/ml Heparin (Sigma Aldrich), 2 mM L-glutamine (Invitrogen) and SAG.

RNA sequencing

The quantification of transcripts was performed using HTSeq-count algorithm (Anders et al., Bioinformatics, 2015, <https://doi.org/10.1093/bioinformatics/btu638>). R was used to create a matrix of all transcripts expressed in all samples with the corresponding read-counts and the Bioconductor package DESeq2⁴⁷ was used to normalize the data and then to perform the differential expression analysis. DESeq2 package was used with standard parameters (Love et al., Genome Biol, 2014, <https://doi.org/10.1186/s13059-014-0550-8>).

Supplementary Table 1

Primer sequences for PCR genotyping.

Nbs1 exon 6 fw	5'-CAG GGC GAC ATG AAA GAA AAC-3'
Nbs1 LoxP rv	5'-AAT ACA GTG ACT CCT GGA GG-3'
Nbs1 intron 5 fw	5'-ATA AGA CAG TCA CCA CTG CG-3'
Cre fw	5'-CGG TCG ATG CAA CGA GTG ATG-3'
Cre rv	5'-CCA GAG ACG GAA ATC CAT CGC-3'
Norm fw	5'-CAC CGG AGA ATG GGA AGC CGA-3'
Norm rv	5'-TCC ACA CAG ATG GAG CGT CCA-3'

Supplementary Table 2

Primer and probe sequences for qPCR genotyping.

SmoA1 probe (FAM)	5'-CAGTGTCAGAACCCGCTGTTCACC-3'
SmoA1 Fw	5'-CAGACAACCCCAAGAGCTG-3'
SmoA1 Rv	5'-GTGCATGTCCTGGTGCTC-3'
Norm probe (VIC)	5'-CCAATGGTCGGGCACTGCTCAA-3'
Norm Fw	5'-CACGTGGGCTCCAGCATT-3'
Norm Rv	5'-TCACCAGTCATTTCTGCCTTTG-3'

Table 3

List of qPCR assays used for mRNA analysis.

Gli1: Mm00494654_m1
Gli2: Mm01293111_m1
MycN: Mm00476449_m1
Cyclin D2: Mm00438070_m1
Ptch1: Mm00436026_m1
Hes1: Mm01342805_m1
Hes5: Mm00439311_g1
B2M: Mm00437762_m1
HPRT: Mm00446966_m1

Table 4

List of CARD Assays used for mRNA analysis.

Atoh1: Mm00476035_s1
Gli1: Mm00494654_m1
MycN: Mm00476449_m1
Gli2: Mm01293117_m1
Gli3: Mm00492337_m1
Foxm1: Mm00514924_m1
Ptch1: Mm00436026_m1
Ptch2: Mm01187196_g1
Jag1: Mm00496902_m1
Jag2: Mm00439947_g1
Hey1: Mm00468865_m1
Hey2: Mm01180513_m1
Hes1: Mm01342805_m1
Hes5: Mm00439311_g1
Myc: Mm00487804_m1
Numb: Mm00477927_m1
Nestin: Mm01223405_g1
Prom1: Mm01211402_m1
Msi1: Mm01203522_m1
Sox2: Mm03053810_s1
Oct4 (Pou5f1): Mm03053917_g1
Pgk1: Mm00435617_m1
Hprt: Mm00446966_m1
Gusb: Mm00446954_g1
Tfrc: Mm00441941_m1

Table 5

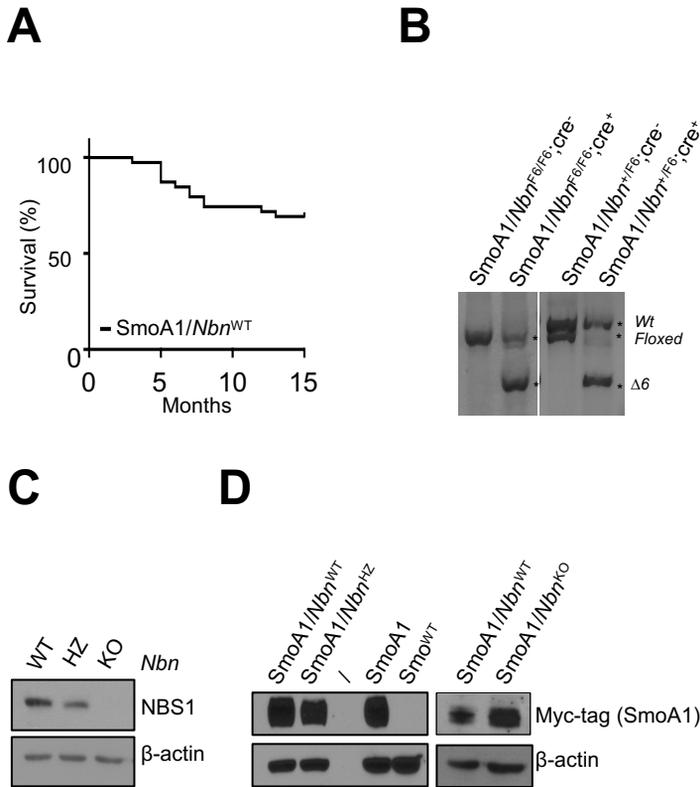
List of WB antibodies.

anti-MycN #SC-53993 (Santa Cruz Biotechnology)
anti-vinculin #SC-73614 (Santa Cruz Biotechnology)
anti-Cyclin D1 #SC-717 (Santa Cruz Biotechnology)
anti-Myc #C3956 (Sigma Aldrich)
anti- β -actin #A5441 (Sigma Aldrich)
anti-NBS1 #NB110-57272 (Novus Biologicals),
anti-NBS1 #ab32074 (Abcam, Cambridge, UK)
anti-ZIC1 ab72694 (Abcam, Cambridge, UK)
anti-Mre11 #ab65131 (Abcam, Cambridge, UK)
anti Rad50 #ab119708 (Abcam, Cambridge, UK)
anti-Gli1 #L42B10 (Cell Signaling Technology)
anti-phospho-histone H2AX (ser 139) #2577 (Cell Signaling Technology)
anti-phospho-p53 (Ser 15) #9284 (Cell Signaling Technology)
anti-p53 (IC12) #2524 (Cell Signaling Technology)
anti-PARP1 #9542 (Cell Signaling Technology)
anti-pKAP1(S824) #A300-767A (Bethyl laboratories, Montgomery, TX, USA)

Table 6Primer sequences for *Nbn* sequencing.

NBS A Fw: CCACCCATTGATGAACCAGC
NBS A Rv: GGTGACTTTAACTGCCGACA
NBS B Fw: TGAATCTAAGAAACAGCCTCCAG
NBS B Rv: TCTTCGTCGTCTTCTGCCAT
NBS C Fw: CTGCACGTTGAAGACAGGAG
NBS Cbis Fw: GGTCTTTCCTGCACGTTGAA
NBS C Rv: TGGGAGTGTGAAATTATGAGCTG
NBS D Fw: AACAGTAAACTTCCCTGTAACCA
NBS Dbis Fw: GGTCTTTCCTGCACGTTGAA
NBS E Fw TTACCGCAAACA ACTGGACG
NBS F Fw TACCCACCTTGTCATGTCGG

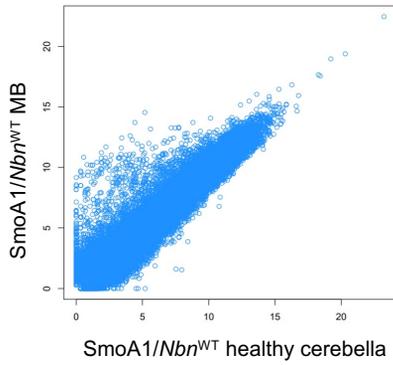
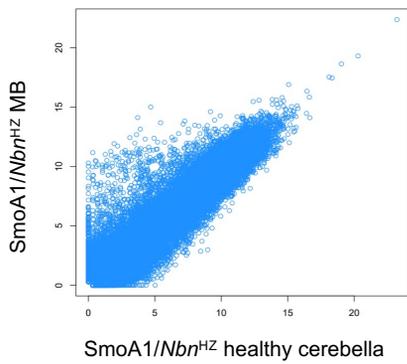
Supplementary Figures



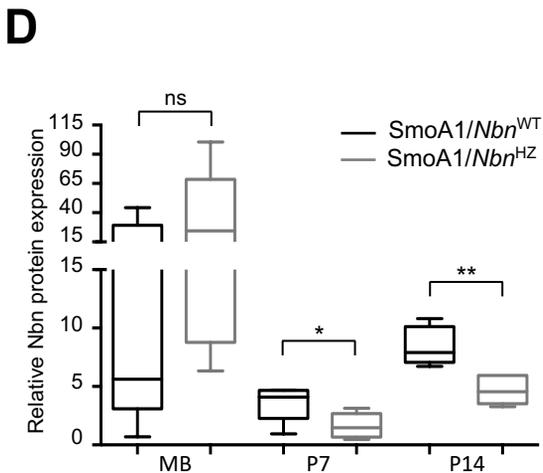
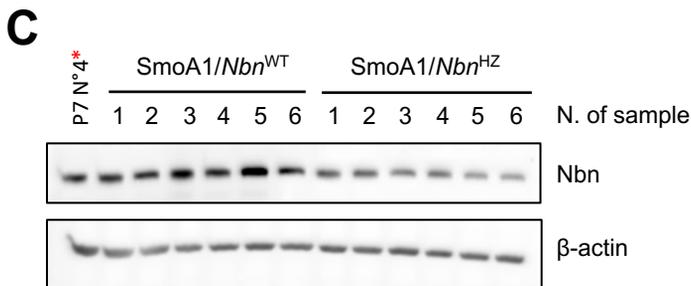
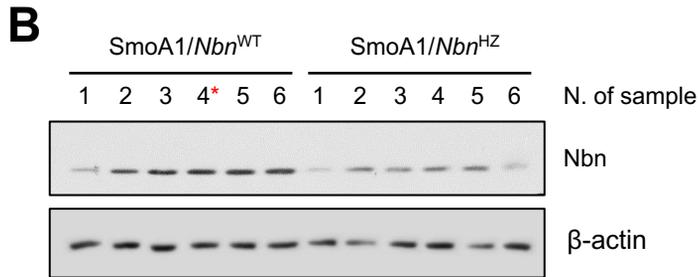
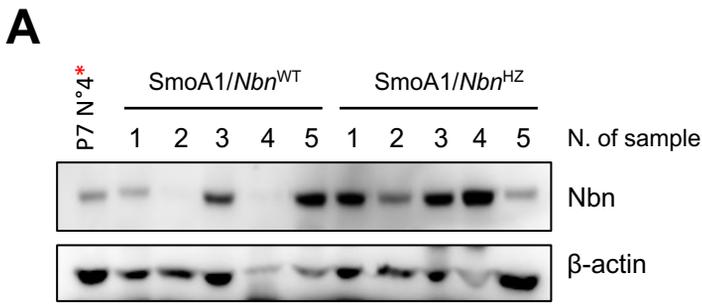
Petroni et al. Supplementary Figure 1.

Generation of the SmoA1/Nbn^{KO} and SmoA1/Nbn^{HZ} mice models.

A, Kaplan-Meier survival curve of SmoA1 homozygous mice after repeated inter crossing. **B**, **C** PCR analyses (**B**) showing *Nbn* gene deletion and Western Blot (**C**) showing NBS1 protein down-regulation in P1 cerebella. *Wt*: Wild type allele; *Floxed*: allele modified by flanking exon 6 with two loxP sites; $\Delta 6$: cleaved allele. β -actin was used as loading control. WT: Wild type; HZ: mono-allelic KO; KO: bi-allelic KO. **D**, Western Blot analysis confirming the expression of SmoA1 transgene in the new mouse models by the Myc-tag detection in P1 cerebella. Cerebella from SmoA1 and SmoWT mice were used as positive and negative controls for SmoA1 expression, respectively.

A**B****Petroni et al. Supplementary Figure 2*****Global gene expression analysis of MBs and healthy cerebella in SmoA1/Nbn^{WT} and SmoA1/Nbn^{HZ} mice.***

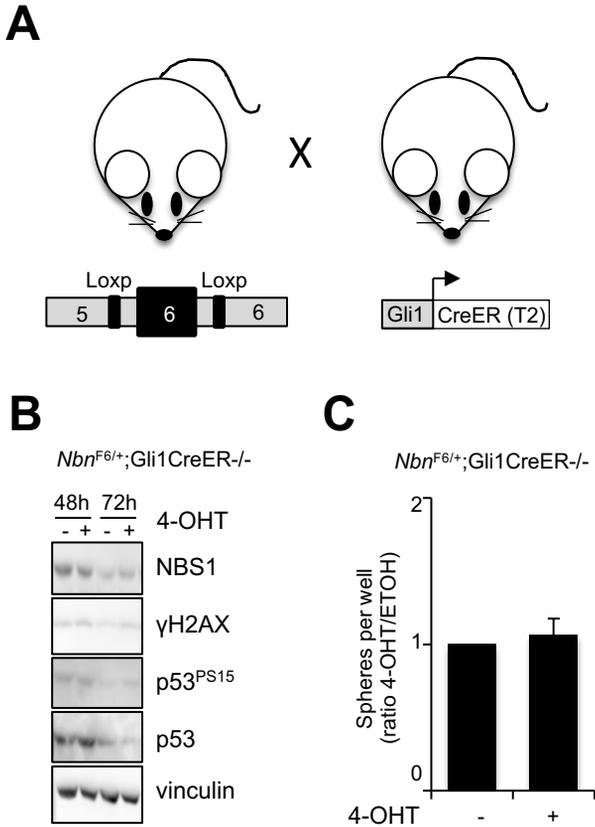
A, Scatter plot of the RNA-Seq expression data in SmoA1/Nbn^{WT} MB versus healthy cerebella. Data are reported as the mean of the gene expression levels (in log-scale) of three independent experiments. **B**, Scatter plot of the RNA-Seq expression data in SmoA1/Nbn^{HZ} MB versus healthy cerebella. Data are reported as the mean of the gene expression levels (in log-scale) of three independent experiments.



Petroni et al. Supplementary Figure 4

Analysis of the *NBS1* expression in pre-tumor and MBs originated from *SmoA1/Nbn*^{WT} and *SmoA1/Nbn*^{HZ} mice.

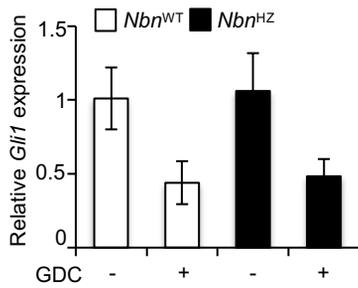
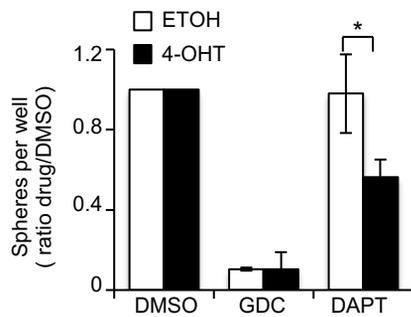
A, WB analysis of MBs (n=5) explanted from symptomatic mice with the indicated genotypes at around 7 months. Blot was probed with a Nbn antibody and β-actin was used as loading control **B**, **C**, WB analysis of developing cerebella (n=6) explanted from mice with the indicated genotypes at P7 (**A**) and P14 (**B**). Blots were probed with a Nbn antibody and β-actin was used as loading control. *P7 N°4 sample was loaded in all three blots to allow a proper comparison of NBS1 expression levels in the different blots. **D**, Box Blot representing the median Nbn protein expression in the indicated samples. Densitometry was performed from WB in A, B and C using ImageJ software (version 2.0.0-rc-69/1.52n). p value was calculated by two-sided Student's t-test (*p < 0.05, **p < 0.01, n.s. not significant).



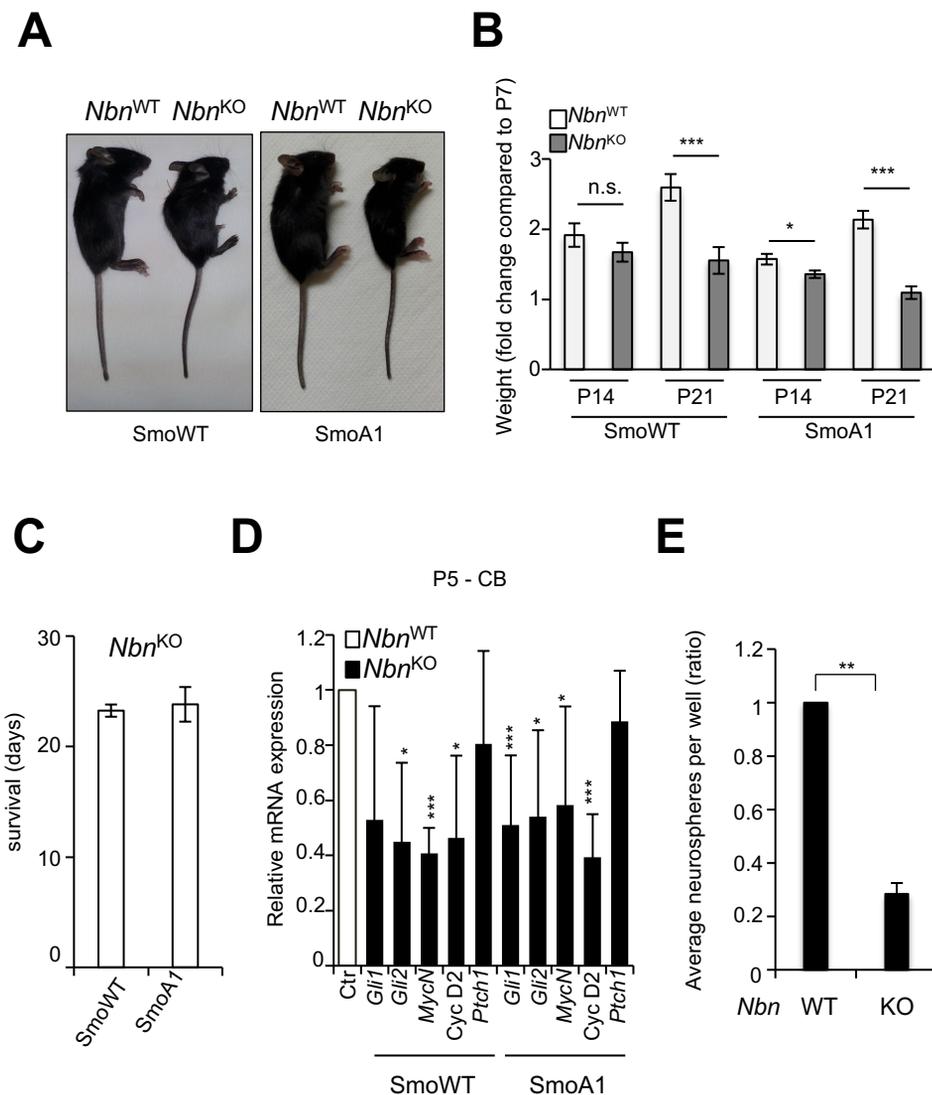
Petroni et al. Supplementary Figure 5

Generation and characterization of the *Nbn*^{F6/+};Gli1CreER^{+/-} cell model.

A, Schematic representation of the constructs used for the generation of the *Nbn*^{F6/+};Gli1CreER^{+/-} mice. **B**, WB analysis of the *Nbn*^{F6/+};Gli1CreER^{-/-} cells treated with ETOH or 4-OHT for 24 hours and collected after 48 and 72 hours from the wash out. Blots were probed with the indicate antibodies and vinculin was used as loading control. **C**, Neurosphere formation assay from P7 *Nbn*^{F6/+};Gli1CreER^{-/-} GCPs. Cultures were treated with ETOH or 4-OHT for 24 hours, dissociated and plated for the analysis. Data are expressed as ratio of 4-OHT versus ETOH treated samples and reported as mean (\pm SD).

A**B****Petroni et al. Supplementary Figure 6*****Involvement of the Notch pathway in the enhanced clonogenicity of the *SmoA1/Nbn*^{HZ} S-cNS.***

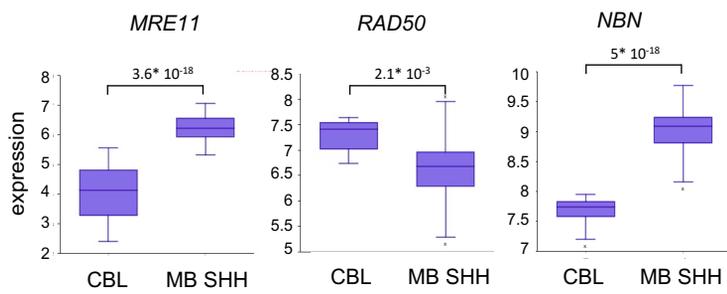
A, qPCR analysis of Gli1 mRNA expression in GCP neurospheres treated with GDC-0449 for 24 hours. **B**, Neurosphere formation assay from P7 *Nbn*^{F6/+};Gli1CreER^{+/-} GCPs. Neurospheres were treated with ETOH or 4-OHT for 24 hours and then dissociated and plated for the analysis, in the presence of DMSO, DAPT or GDC-0449. Data are expressed as ratio of 4-OHT versus ETOH treated samples and reported as mean (\pm SD) of three independent experiments. p value was calculated by two-sided Student's t-test (*p < 0.05).



Petroni et al. Supplementary Figure 7

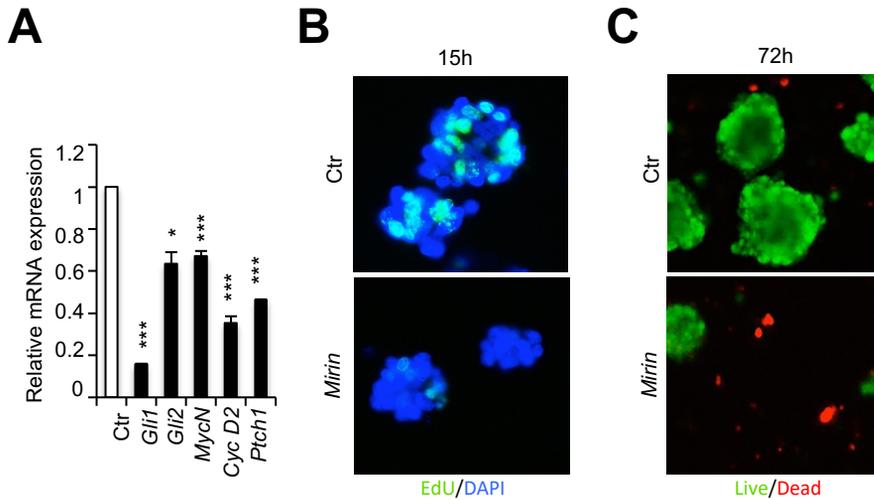
Additional features of *SmoA1/Nbn*^{KO} and *SmoWT/Nbn*^{KO} mice.

A, Representative pictures of the mice with the indicated genotypes at P21. **B**, Relative weight analysis of mice with the indicated genotypes at P14 and P21. Data are reported as mean (\pm SE) of at least 6 independent biological replicates. The values for each condition are represented as FC with respect to the corresponding weights at P7. p value was calculated by two-sided Student's t-test (* $p < 0.05$, *** $p < 0.001$, ns: not significant). **C**, Survival analysis of the *Nbn*^{KO} mice in the SmoWT and SmoA1 backgrounds. Data are reported as mean (\pm SE) of at least 4 animals. **D**, qPCR analysis of the indicated transcripts in cerebella explanted from P5 mice with the indicated genotype. mRNA expression levels were normalized on the mean of expression of two reference genes (*β 2-microglobulin*, *Hprt*) and relative mRNA quantification was expressed as FC. Data are reported as mean (\pm SD). p values were calculated by two-sided Student's t-test (* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$). **E**, Neurosphere formation assay from P5 *Nbn*^{WT} and *Nbn*^{KO} cerebellar explants grown with SAG. Data are expressed as ratio of *Nbn*^{KO} versus *Nbn*^{WT} samples and reported as mean (\pm SD). (** $p < 0.01$).



Petroni et al. Supplementary Figure 8

Expression of MRN complex genes in human samples. Box plots representing *MRE11*, *RAD50* and *NBN* mRNA expression in healthy cerebella and SHH-MBs (Roth-9-MAS5.0 and SHH-Pfister -73-MAS5.0 datasets, respectively) obtained from the R2-Genomics platform.



Petroni et al. Supplementary Figure 9

Mirin impairs both SHH pathway and survival in MB-neurospheres.

A, *q*PCR analysis of the indicated transcripts in MB-neurospheres treated with mirin or DMSO for 15 hours. mRNA expression levels were normalized on the mean of expression of two reference genes (*β2-microglobulin*, *Hprt*) and relative mRNA quantification was expressed as FC. Data are reported as mean (\pm SD). *p* values were calculated by two-sided Student's *t*-test (***p* < 0.01, ****p* < 0.001). **B**, Representative images of EdU staining in MB-neurospheres treated as in **A**. **C**, Representative images of alive (green) or dead (red) MB-neurospheres treated with mirin or DMSO for 72 hours.