SUPPLEMENTAL FIGURE LEGENDS

Figure S1. PQBP1 association with incoming HIV-1 virus particles, Related to Figure 1. (A) Specificity of PQBP1 antibody (Sigma) and lack of effect on overall cGAS expression was confirmed by IF and/or western blot of THP-1 cells expressing indicated shRNAs (top) or MDDCs treated with indicated siRNAs (bottom). Scale bars: 10 μ m (top), 25 μ m (bottom). (B) Identification of viral particles from infected THP-1 cells was done with python with skimage, scipy, pims, and trackpy tools. In short, cells were subjected to a binary threshold so that no extracellular signals (viruses or cell debris) would confound the analysis. Viral particles were detected using thresholding and watershed segmentation methods or by using trackpy. Each panel is 221.8 µm squareThe masks of the pixels that are positive for viral particles are then guantified for each independent channel (PQBP1, cGAS, iGFP, IN label, etc.) and (C) for each field of view that was imaged, the fraction of total virions (IN) showing positive PQBP1 signal (compared to controls using secondary antibodies only) were graphed. (D) Quantification of PQBP1 and IN distance distribution analysis. Left, probability distribution of distances, P(d), from each virion (IN) to the nearest PQBP1 (blue). P(d) for uniformly distributed jitter J=1 μm (yellow) becomes similar to the distribution obtained by random shuffling of puncta (purple). Arrows indicate reasonable arbitrary thresholds (δ) to be applied to d values in determining associations of PQBP1 puncta with corresponding IN. Data were obtained from 24 independent images (N=24,471). Right, the percentage of IN spots that have a PQBP1 within a chosen threshold, δ (0.1 µm < δ < 0.75 µm). See PQBP1 and IN distance distribution analysis section under Star Method for detail.

Figure S2. Mapping of the capsid interaction domain in PQBP1, Related to Figure 2. (A) PQBP1₁₋₄₆ and PQBP1₄₇₋₂₆₅ with N-terminal eGFP or C-terminal sfGFP produced in a cell-free protein expression mixture before (left) and after (middle) purification by Ni-NTA

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chromatography followed by dialysis. Recombinant PQBP1 was labelled at C60 with AF488maleimide (right). (B) Top panels, representative fluorescence traces of either AF488 labelled PQBP1, CPSF6 peptide or dATP (red traces) and AF568-labelled CA A204C particles (blue traces). Inclusion of hexacarboxybenzene (HCB) or use of either N74D or R18G double mutants with CA A204C is indicated. Bottom panels, analogous to the top, except eGFP-fused PQBP1 truncation constructs were utilized. (C) PQBP1 co-pellets preferentially with CA assemblies containing the R18 ring. Left, assembled CAA204C and CAR18G/A204C were incubated with PQBP1, centrifuged and supernatant (S) and pellet (P) fractions were analyzed using reducing SDS-PAGE with Coomassie staining. Right, bar graph of the intensity of the PQBP1 band relative to the intensity of the CA band in the pellet fraction. Two independent co-pelleting experiments were performed with two repeats per experiment. (D) Fluorescence resonance energy transfer (FRET) assay was used to measure interactions between PQBP11-46-YFP and CypA-dsRed. THP-1 cells, either wild type or stably expressing eYFP, PQBP1₁₋₄₆-eYFP or PQBP1₁₋₁₀₄-eYFP, were infected with HIV-1 virus packaged with CypA-DsRed for 1.5 hrs, followed by PFA fixation and FRET analysis. Signal intensity distribution of CypA-dsRed positive foci for indicated THP-1 clones infected with the virus (left) and a western blot depicting expression of both eYFP and PQBP1-eYFP proteins are shown.

Figure S3. PQBP1 is required for cGAS recruitment to incoming virus particles, Related to Figure 3. (A) The specificity of the anti-cGAS antibody was confirmed by IF and western blot on cGAS knock out in THP-1 cells. Scale bar, 10 μ m. (B) Time lapsed imaging to monitor the structural integrity of an incoming viral core. MDDCs were infected with HIV-1 virus with iGFP fluid phase marker (iGFP+IN-mRuby3 at low MOI) where iGFP and IN-mRuby signal intensities of each virion were monitored for one hour. See Video S1. A representative kinetic profile of the signal intensities of a virion where the loss of GFP signal is linked to virion fusion and capsid integrity loss (top). Panels of each virion, identified by both time-lapsed and corresponding IF x-

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v-z position of the respective foci point label were categorized based on capsid integrity and are color coded for the signals of PQBP1 (green), cGAS (blue), IN-mRuby3 (red). The RGB merged images and iGFP (gray in the individual boxes) are also shown (middle, particle B to G). Scale bars, 5 and 0.2 μ m as shown. 3D reconstruction of point E and point F foci are shown where we see the proximity and overlapping of PQBP1/cGAS complexes with the viral particle. 3D Ruler/ScaleBar indicates size in µm in the pictures. (C) The knockdown efficiency of siRNAs against PQBP1 is demonstrated by both RT-qPCR and IF analysis (left and right respectively). MFI of cGAS signal intensity per THP-1 cell treated with either non-targeting (NT) or PQBP1 specific siRNAs and infected with HIV-1 virus, as described in Figure 3C. Scale bar, 60 μ m. Mean and error bar (-/+ 1.5*IQR) are shown. Boxes denote the interguartile range. T-test (twotailed, equal variation), *p<0.05; Dunn's column comparison **** p<0.0001, * p<0.05. (D) Increase in cGAMPs in the cytosol of infected cells. PMA-THP-1 cells, either mock infected or infected with HIV-1 luciferase virus or transfected with either HT-DNA or cGAMP for 3 hours, were stained for cGAMP (green), p24 (red), IRF3 (magenta) and dapi (blue). The white contour line indicates the boundary of the nucleus. Scale bar, 10 µm. (E) PMA-THP-1, transfected with non-targeting siRNAs (NT) or siRNA targeting PQBP1, were challenged with HIV-1, stained for cGAMP (green), cGAS (red), and DAPI (blue). Knockdown efficiencies of both mRNA and protein level of PQBP1 were shown below. Scale bar, 5 μm. (F) The cells treated as in (E) were analyzed IFN β mRNAs induction post 16 hours of the infection. Averages and SEM are shown, One-way ANOVA, ** p<0.01, * p<0.05.

Figure S4. NONO is not required for PQBP1-dependent cGAS sensing during the early steps of infection, Related to Figure 4. (A) Knockdown efficiencies of siRNA-targeted genes in MDDCs were quantified by RT-qPCR. (B) MFIs of cGAS signal per infected MDDC, subjected to indicated siRNA treatments, are shown. (C) PMA-differentiated THP-1 cells, subjected to siRNA-mediated targeting were infected with were with either mock of HIV-1 in the presence of VLP-Vpx for 2 hrs, followed by post-fixation IF imaging. The levels of NONO and PQBP1 proteins (left) and expression levels of Flag-cGAS (red), p24 (green) and DAPI (blue) in cells used for proximal ligation assays, as in Figure 4 (right), are shown. Scale bars, 20 μ m (A table at the bottom of the graph shows # of INs analyzed for each condition. (D) Representative images of the infected MDDCs treated with indicated siRNAs (left) and MFI of indicated protein signals per cells (right), analyzed in Figure 4D and 4F, are shown. Scale bar, 10 μ m. Mean and SEMs are shown. One-way ANOVA, ***p<0.001, **p<0.01, *p<0.05. ns denotes no significance.

Figure S5. Distinct domain of PQBP1 interact with the capsid and cGAS, Related to

Figure 5. (A) HIV-1 infection (+) enhances co-immunoprecipitation of cGAS or Flag-cGAS with PQBP-IP in MDDCs or PMA-THP-1, respectively. The original uncropped images of the blots in Figure 5B are shown. See Figure 5B for details. Cells were infected with HIV-1 luciferase virus in the presence of VLP-Vpx and subjected to co-IPs at 3 hrs post infection. Endogenous cGAS or Flag-cGAS co-precipitating with PQBP1 was assayed. Normal IgG (NS) or antibody against PQBP1 were used as indicated. Non-specific (ns) bands and the cGAS band (arrow) are indicated. The lines between cGAS and PQBP1 blots indicate corresponding samples. 1x and 3x inputs indicate relative amount of input lysates. Sources of cGAS antibodies are indicated. (B) Either full-length or truncated PQBP1-YFP proteins were co-expressed with MBP-cGAS in 293T cells and subjected to anti-MBP pull down. *ns denotes non-specific protein. A schematic of the PQBP1 protein is shown. WWD and E/D denote the WW domain and the acidic residue-rich domain, respectively.















Scale bar = 20µm



Scale bar= 10 µm





