## nature portfolio

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## Reporting Summary

Nature Portfolio wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Portfolio policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For	all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.
n/a	Confirmed
	$oxed{x}$ The exact sample size $(n)$ for each experimental group/condition, given as a discrete number and unit of measurement
	🕱 A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.
x	A description of all covariates tested
	🕱 A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>
x	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings
x	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes
x	Estimates of effect sizes (e.g. Cohen's <i>d</i> , Pearson's <i>r</i> ), indicating how they were calculated
,	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.

## Software and code

Policy information about availability of computer code

Data collection Topspin 4.1.1 (Bruker), SpectraManager (JASCO), Image Lab 6.1, Gaussian09, Amber 20

Data analysis Topspin 4.1.1 (Bruker), Image Lab 6.1, ChemDraw 18.2, Pymol 2.3.2, VMD 1.9.4, Xplor-NIH 2.48, NMRFAM-SPARKY, GraphPad Prism 8.2

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Portfolio guidelines for submitting code & software for further information.

## Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A description of any restrictions on data availability
- For clinical datasets or third party data, please ensure that the statement adheres to our policy

Source data are provided with this paper. The data that support the findings of this study are available from the corresponding authors upon reasonable request. The coordinates and experimental details generated in this study have been deposited in the Protein Data bank under accession codes 7X8N [https://doi.org/10.2210/pdb7X8N/pdb] (free KRAS-G4), 7X8M [https://doi.org/10.2210/pdb7X8M/pdb] (2:1 berberine-KRAS-G4 complexes), and 7X8O [https://doi.org/10.2210/pdb7X8O/pdb] (2:1 coptisine-KRAS-G4 complexes).

Field-specific reporting				
Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.				
<b>x</b> Life sciences	□В	sehavioural & social sciences		
For a reference copy of th	ne document with	all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>		
Life scien	ces stu	udy design		
-		points even when the disclosure is negative.		
Sample size	All experiments	ints were run in 2 to 3 independent replicates.		
Data exclusions	Not applied.			
Replication	Three biologica	Il replicates were performed and experiments were repeated at least twice, resulting in similar results.		
Randomization	N.A.			
Blinding	N.A.			
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		pecific materials, systems and methods		
We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.				
Materials & exp	erimental s	ystems Methods		
n/a Involved in the	e study	n/a Involved in the study		
Antibodies		ChIP-seq		
Eukaryotic o		Flow cytometry		
=1=	ogy and archaeo d other organisn			
=1=	earch participan			
<b>✗</b> ☐ Clinical data				
Dual use res	search of conce	m		
<u>Eukaryotic ce</u>	ell lines			
Policy information a	bout <u>cell lines</u>			
Cell line source(s)		The A549, H460, and BEAS-2B cell lines were obtained from the cell bank of the Chinese Academy of Sciences (Shanghai, China).		
Authentication		The A549, H460, and BEAS-2B cell lines were authenticated and checked regularly for preservation of their typical morphology and behavior.		
Mycoplasma contam	nination	There was no Mycoplasma contamination		
Commonly miside (See <u>ICLAC</u> register)	entified lines	Commonly misidentified cell lines were not used in the study		