

## **Supplementary Information for Conservation at the uterine-placental interface.**

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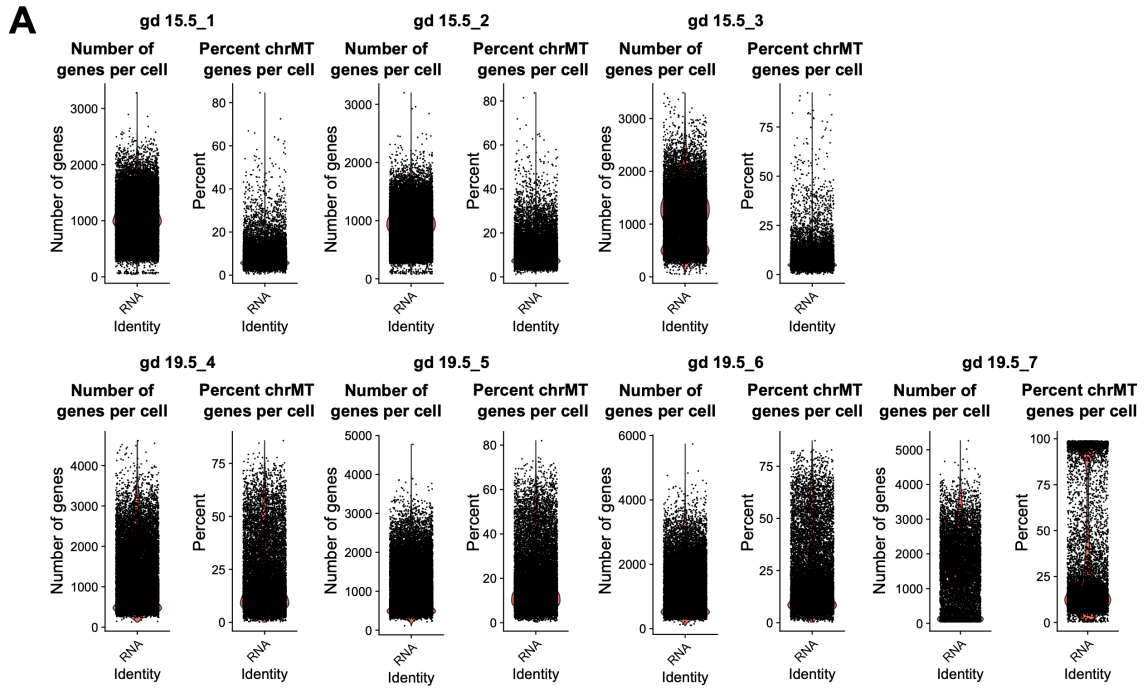
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### **This PDF file includes:**

Supplementary text  
Figures S1 to S9

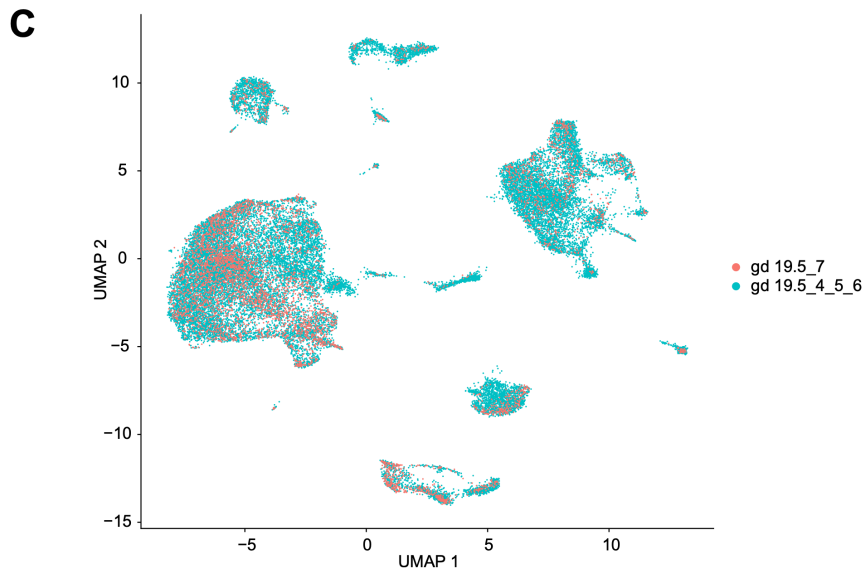
### **Other supplementary materials for this manuscript include the following:**

Datasets S1 to S5



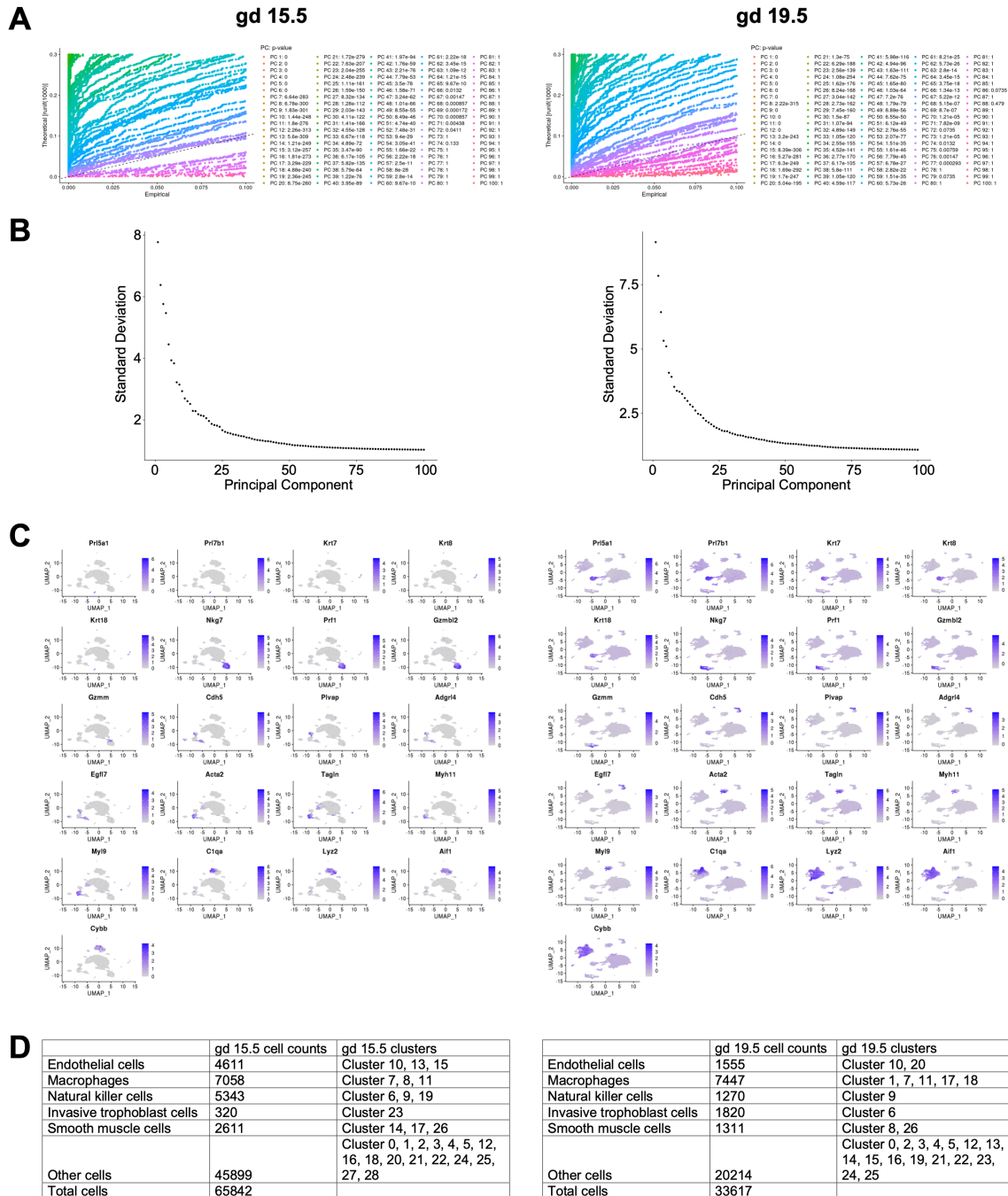
**B**

Sample	Before filtering	After filtering
gd 15.5_1	17769 genes across 29487 cells	17769 genes across 25384 cells
gd 15.5_2	17663 genes across 30757 cells	17663 genes across 24845 cells
gd 15.5_3	17649 genes across 18338 cells	17649 genes across 15613 cells
gd 19.5_4	18547 genes across 15401 cells	18547 genes across 8664 cells
gd 19.5_5	18283 genes across 18388 cells	18283 genes across 10641 cells
gd 19.5_6	18606 genes across 15444 cells	18606 genes across 8750 cells
gd 19.5_7	17727 genes across 10391 cells	17727 genes across 5562 cells



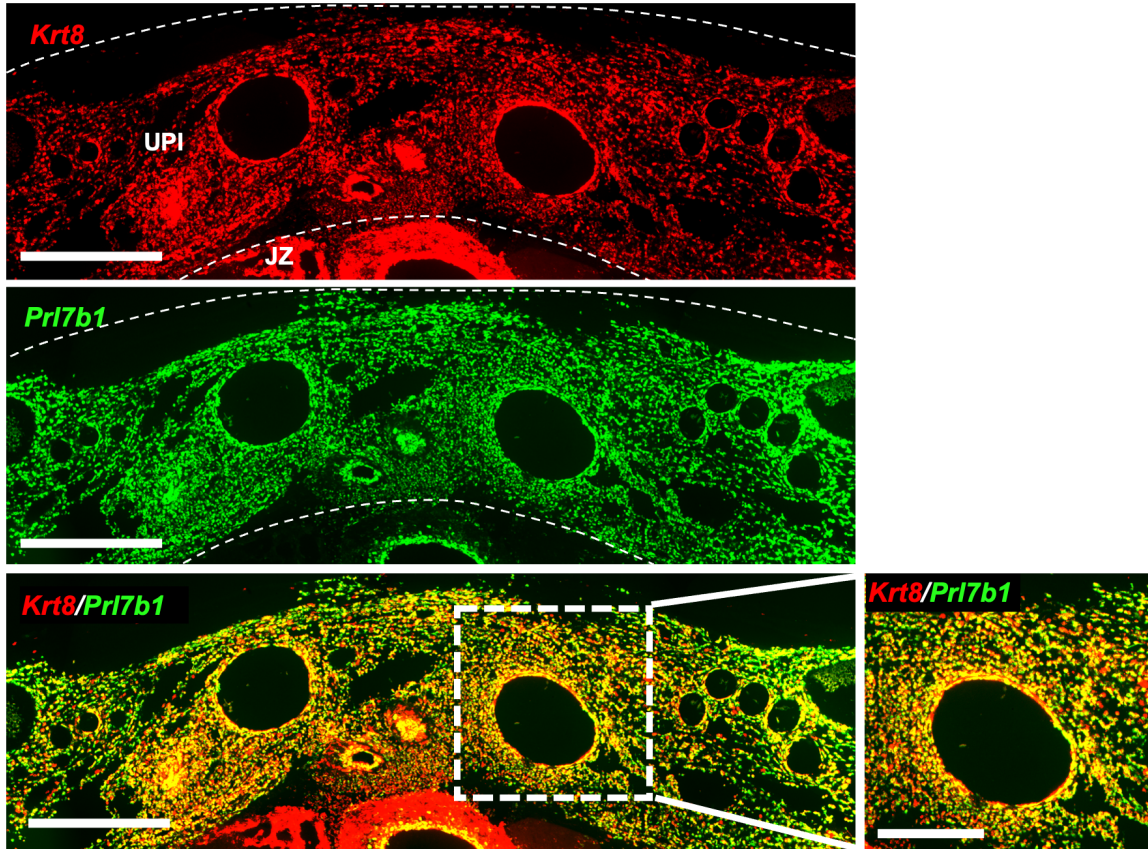
**Fig. S1. Quality control and processing of single cell RNA sequencing (scRNA-seq) data.**

**A)** Violin plots showing the distributions of number of genes and percentage of mitochondrial chromosome (chrMT) genes expressed per cell. Cells with the number of unique genes between 500 and 3500, and less than 20% of mitochondrial genes were retained. **B)** Summary table of the number of genes and cells retained in each sample after preprocessing. **C)** UMAP plot showing the cells from samples gd 19.5\_4\_5\_6 and sample gd 19.5\_7, which were generated in different batches, are integrated and well blended after the integration process.

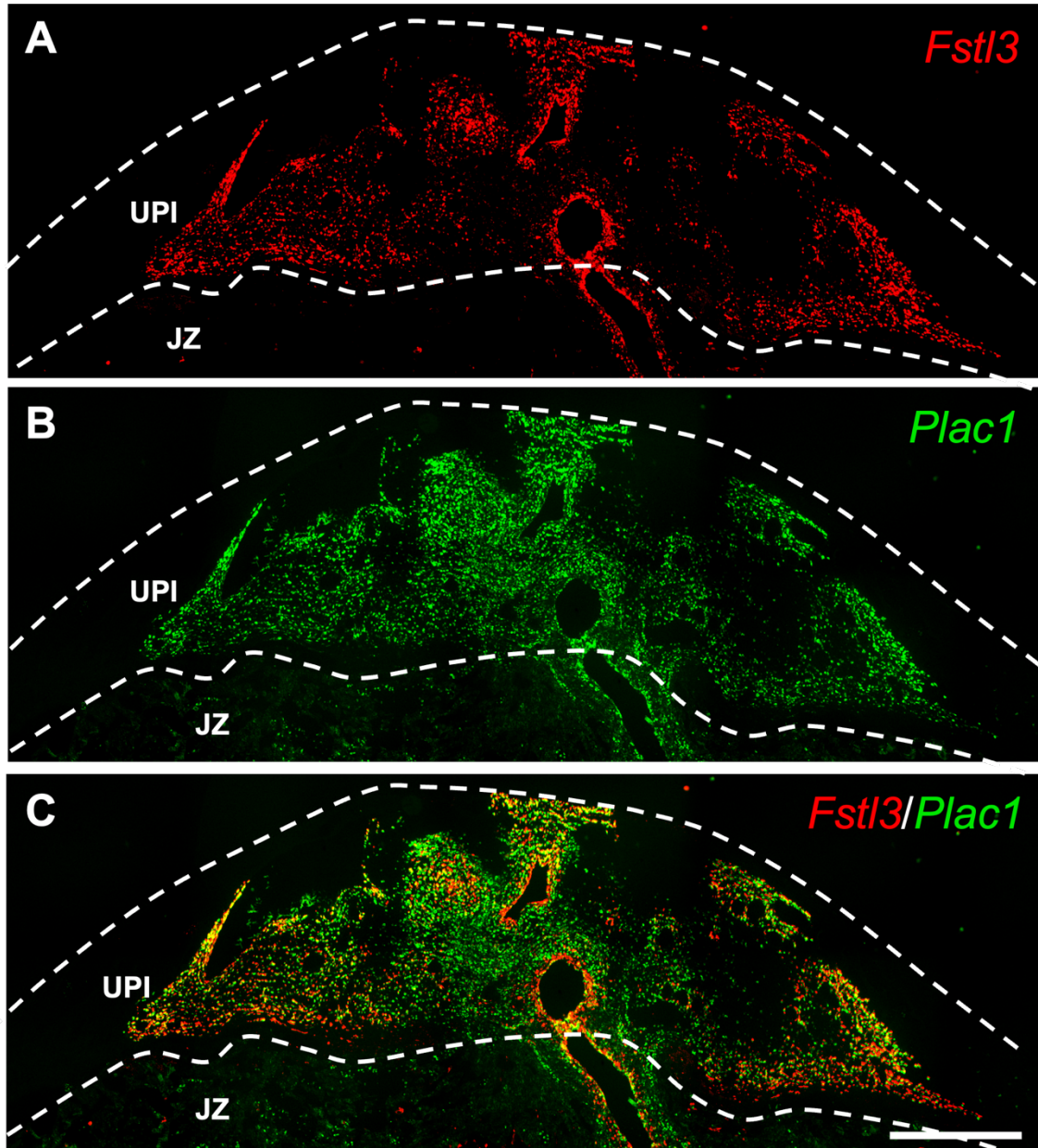


**Fig. S2. Clustering and cluster annotations.**

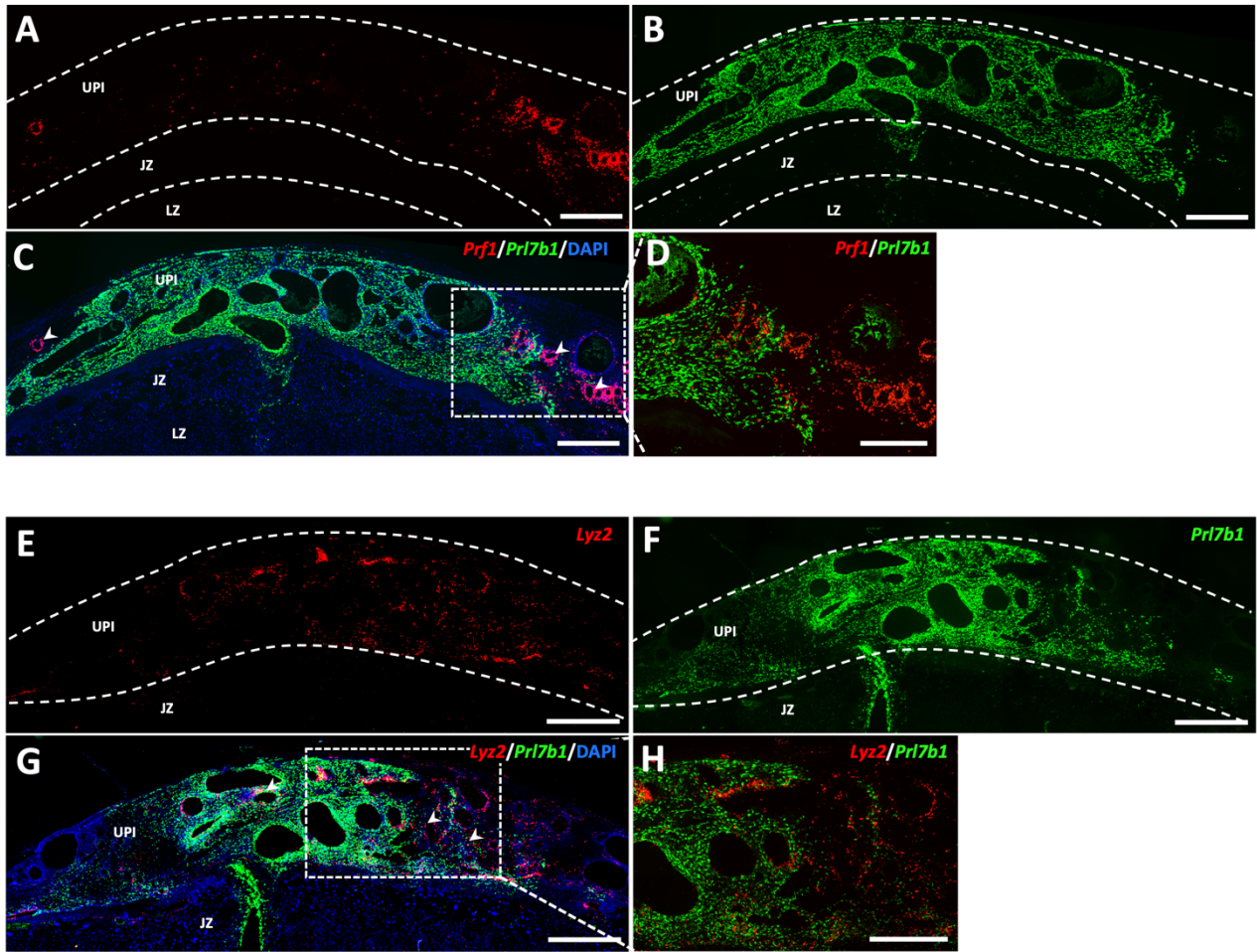
**A)** QQ plots showing the JackStraw analysis for principal component (PC) analysis significance. P-values indicating the significance of each PC were also shown. Based on the p-values, the first 72 PCs were used for the analyses on gd 15.5 samples, and the first 77 PCs were used for gd 19.5. **B)** Elbow plots showing the amount of standard deviation each PC represents. The first 72 PCs of gd 15.5 samples, and 77 PCs of gd 19.5 samples, captured the majority of the variation in the data. **C)** UMAP plots showing expression level of marker genes across cell clusters. **D)** Summary table of the cell cluster identities and number of cells in each cell group.



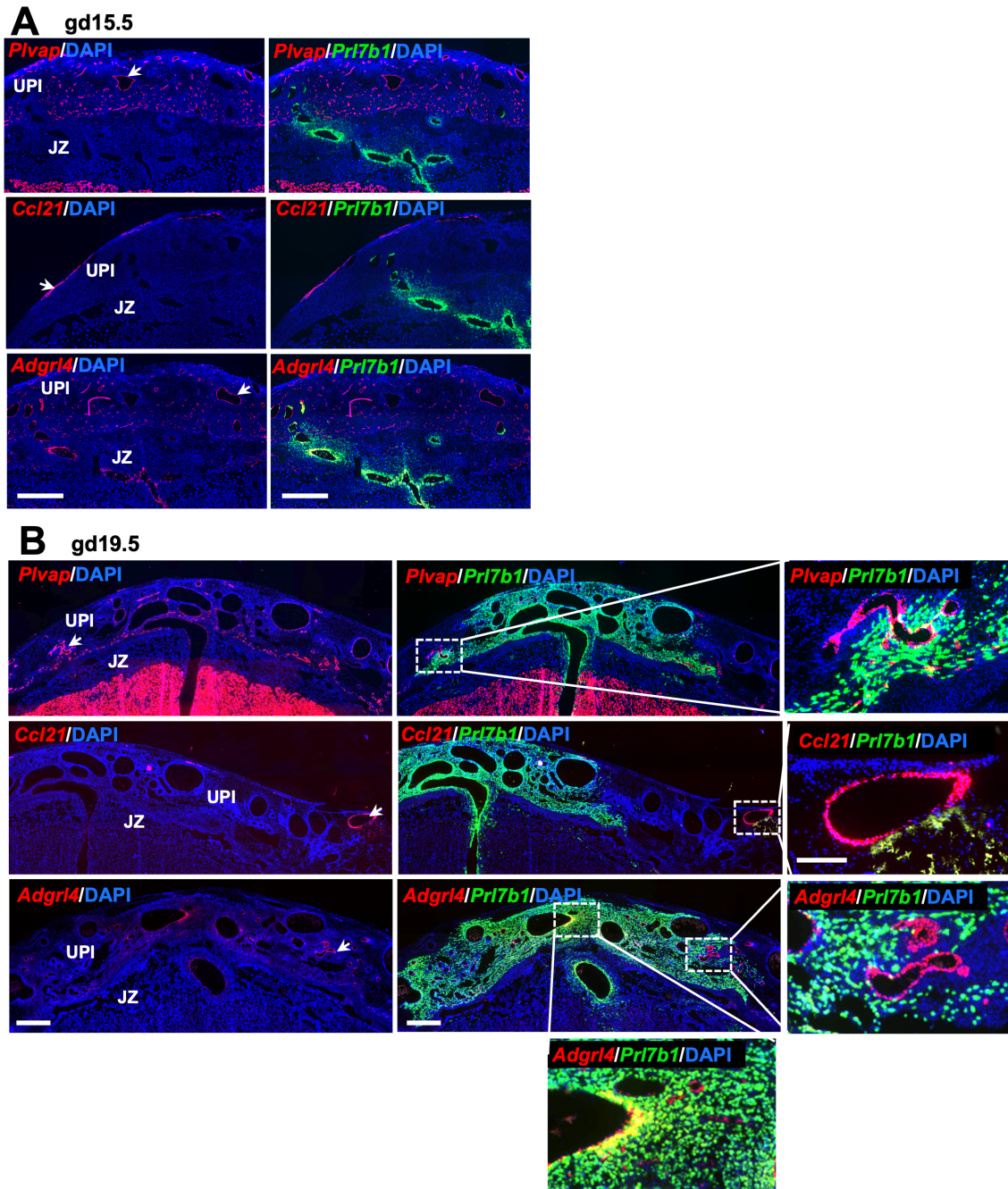
**Figure S3. Colocalization of *Krt8* and *Prl7b1* in invasive trophoblast cells at the uterine-placental interface.** In situ hybridization of gd 19.5 rat placentation site for *Krt8* (red; top panel), *Prl7b1* (green; center panel), and co-localization of *Krt8* and *Prl7b1* (bottom panels). Scale bar (left panels)=1000  $\mu\text{m}$ , scale bar (right panel)=500  $\mu\text{m}$ . Abbreviations: UPI, uterine-placental interface; JZ, junctional zone.



**Figure S4. Colocalization of *Fstl3* and *Plac1* in invasive trophoblast cells at the uterine-placental interface.** In situ hybridization of gd 19.5 rat placentation site for **A)** *Fstl3* (red), **B)** *Plac1* (green) and **C)** colocalization of *Fstl3* (red) and *Plac1* (green) . Scale bar=1000  $\mu$ m. Abbreviations: UPI, uterine-placental interface; JZ, junctional zone.

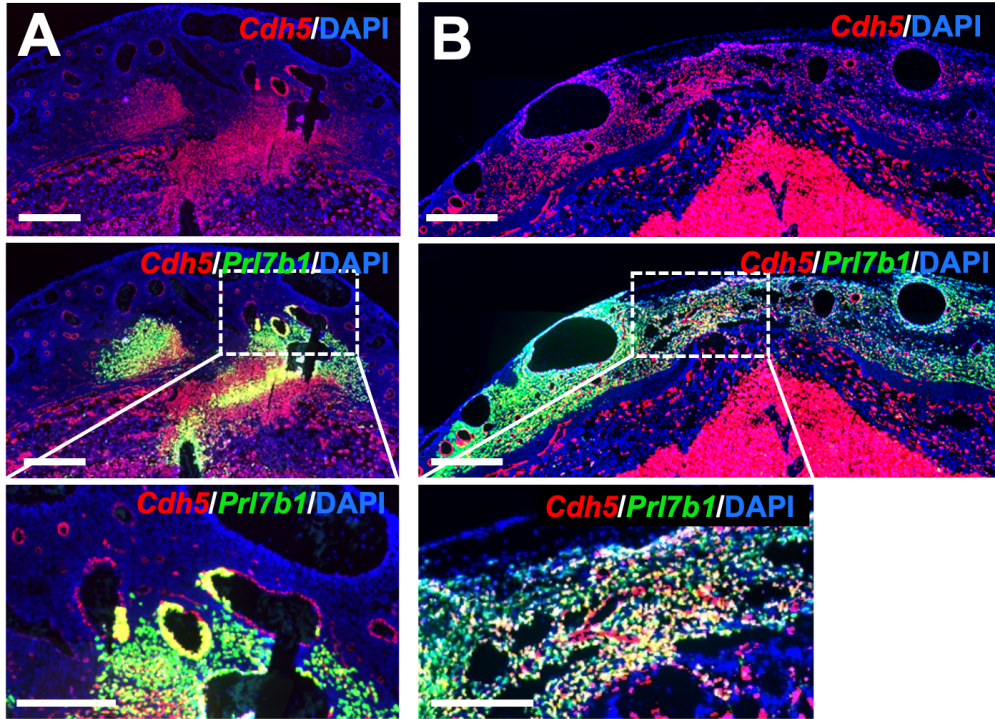


**Figure S5. Distribution of NK cells and macrophages within the gestation day (gd) 19.5 rat uterine-placental interface.** **A-D)** NK cell and invasive trophoblast cells were identified in gd 19.5 placentation sites using in situ hybridization for **A)** *Prf1* (red) and **B)** *Prl7b1* (green), respectively. **C)** Image from **Figure 3**, top right panel. **D)** High magnification image from **C**. **E-H)** Macrophages and invasive trophoblast cells were identified in gd 19.5 placentation sites using in situ hybridization for **E)** *Lyz2* (red) and **F)** *Prl7b1* (green), respectively. **G)** Image from **Figure 3**, bottom right panel. **H)** High magnification image from **G**. Scale bar (A-C, E-G)=1000  $\mu\text{m}$ , Scale bar (D, H)=500  $\mu\text{m}$ . Abbreviations: UPI, uterine-placental interface; JZ, junctional zone, LZ, labyrinth zone.

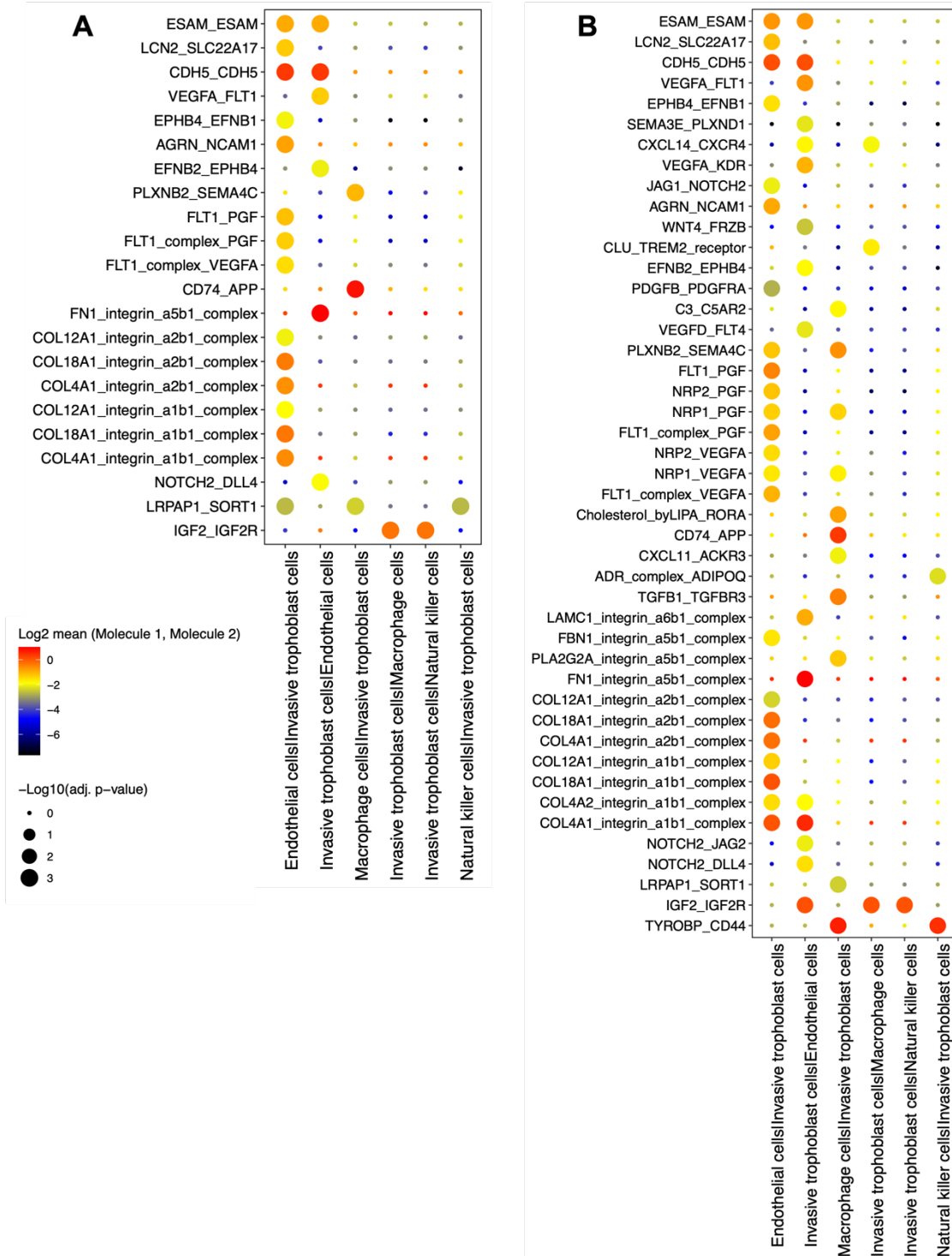


**Figure S6. Endothelial cells at the uterine-placental interface. A)** In situ hybridization of gd 15.5 rat placental site for endothelial cell-specific transcripts *Plvap*, *Ccl21*, and *Adgrl4* (red; left panels) and localization of each endothelial cell marker with an invasive trophoblast cell-specific transcript (*Prl7b1*; right panels). Scale bar=1000  $\mu$ m. **B)** In situ hybridization of gd 19.5 rat placental sites for *Plvap*, *Ccl21*, and *Adgrl4*, and (red; left panels) and localization of each endothelial cell marker with an invasive trophoblast cell-specific transcript (*Prl7b1*; center, right, and bottom panels). Panels on the right and bottom are high magnification images of regions shown in the center panels. Scale bar (left and center panel)=1000  $\mu$ m, scale bar (right and bottom panels)=500  $\mu$ m. Abbreviations: UPI, uterine-placental interface; JZ, junctional zone.

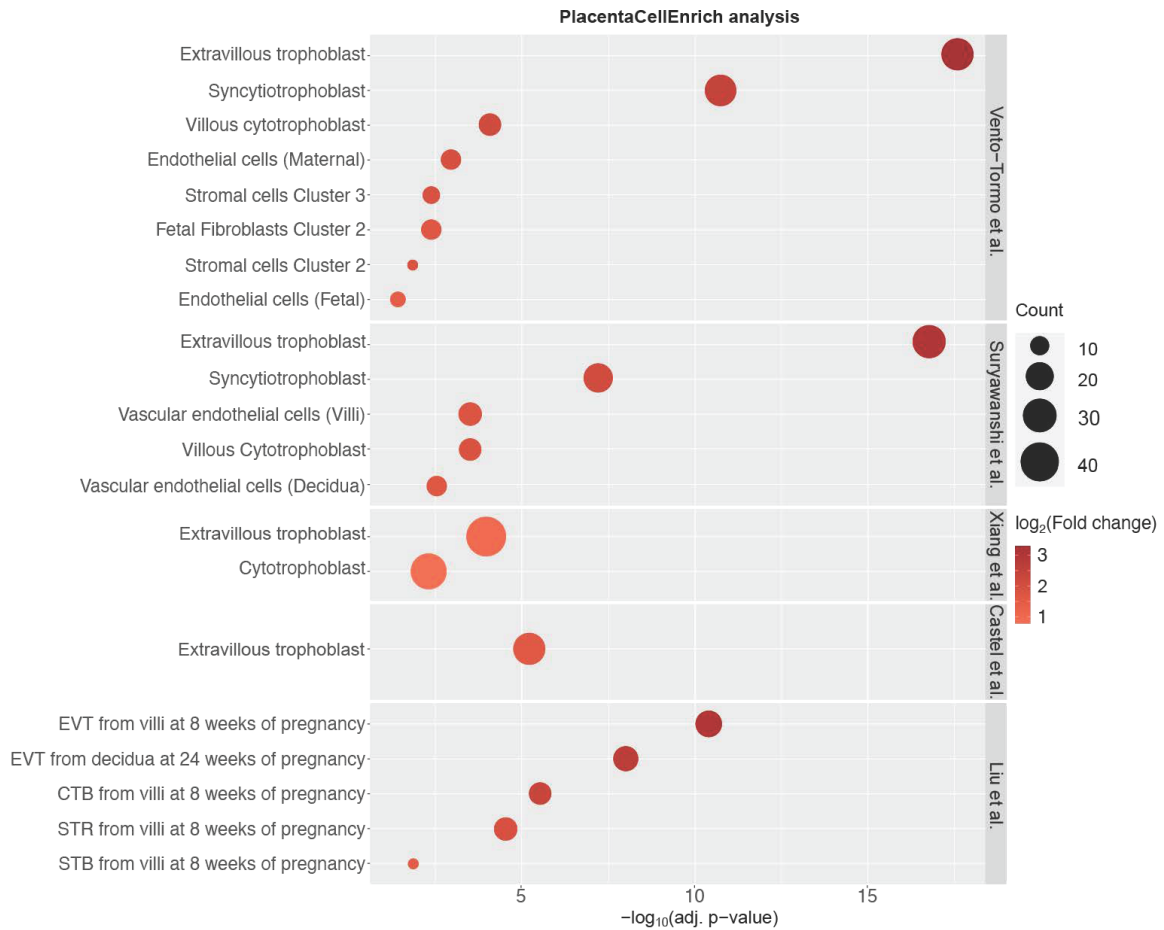




**Figure S7. Localization of *Cdh5* in endothelial cells and invasive trophoblast cells.** **A)** In situ hybridization localization of transcripts for *Cdh5* (upper) and *Cdh5* and *Prl7b1* (center and lower) within gd 15.5 placental sites. **B)** In situ hybridization localization of transcripts for *Cdh5* (upper) and *Cdh5* and *Prl7b1* (center and lower) within gd 19.5 placental sites. The lower panels are high magnification images of regions shown in the central panels. Scale bar=1000  $\mu$ m.



**Figure S8. Ligand-receptor pair analysis in the uterine-placental interface using CellPhoneDB.** Dot plots showing mean expression level of curated ligand – receptor pairs for invasive trophoblast cells with endothelial cells, macrophages, or natural killer cells from gestation day (gd) 15.5 (A) and gd 19.5 (B) uterine-placental interface. Only curated interactions are shown (adj. p-value  $\leq 0.05$ ). The x-axis shows the cell type pairs. The y-axis shows the ligand – receptor pairs. Dot size corresponds to  $-\log_{10}(\text{adj. p-value})$  of the tests for interaction significance. Dot color corresponds to  $\log_2(\text{mean expression})$  of the molecules in each pair.



**Figure S9. Full PlacentaCellEnrich results.** Dot plots showing cell type enrichment when using marker genes from the invasive trophoblast cell gene clusters in rats as input to PlacentaCellEnrich. A significant enrichment has an adjusted p-value  $\leq 0.05$ , fold change  $\geq 1.5$ , and number of observed genes  $\geq 5$ . Dot size represents the number of observed genes specific to a cell type; dot colors correspond to  $\log_2(\text{Fold change})$  of the enrichment. Abbreviations: EVT, extravillous trophoblast; SCT, syncytiotrophoblast; CTB, cytotrophoblast; STR, villous stromal cells.

**Dataset S1 (separate file).**

**Dataset S2 (separate file).**

**Dataset S3 (separate file).**

**Dataset S4 (separate file).**

**Dataset S5 (separate file).**