Simultaneous quantification of all B vitamins and certain biosynthetic precursors in seawater and bacteria by means of different mass spectrometric approaches

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Vitamin	Concentration (ng/l)
B_1	5000
B_2	5000
B_3	8000
B_5	4000
B_6	1000
\mathbf{B}_7	2000
B 9	600000
CB_{12}	125000
AB_{12}	80000
MB_{12}	20000
HB_{12}	25000
HMP	2500
HET	1000
DB ₇	1000
DMB	5000
Cbi	100000

Т

Table S1 Concentrations of the vitamins in a standard mixture of the dilution series

Table S2 Source parameters for both mass spectrometers

	TSQ	Orbitrap
Vaporizer Temperature (°C)	400	350
Transfer Tube Temperature (°C)	340	300
Sheath Gas Pressure (arbitrary units)	60	50
Aux Gas Pressure (arbitrary units)	20	10
	TSO	Orbitron

VitaminSpray Voltage (V) (+)Spray Voltage (V) (+) B_1 30002000 B_2 30002500 B_3 30003000 B_5 30003000 B_6 30003000 B_7 30003000 B_9 30003500 CB_{12} 20003500 AB_{12} 20003500 HB_{12} 20003500HMP30002000HET30002000DB730003000DMB30003000Cbi30003500		TSQ	Orbitrap
B_1 3000 2000 B_2 3000 2500 B_3 3000 3000 B_5 3000 3000 B_6 3000 3000 B_7 3000 3000 B_9 3000 3500 CB_{12} 2000 3500 AB_{12} 2000 3500 MB_{12} 2000 3500 HB_{12} 2000 3500 HB_{12} 2000 3500 HMP 3000 2000 HET 3000 2000 DB_7 3000 3000 DMB 3000 3500	Vitamin	Spray Voltage (V) (+)	Spray Voltage (V) (+)
B_2 3000 2500 B_3 3000 3000 B_5 3000 3000 B_6 3000 3000 B_7 3000 3000 B_9 3000 3500 CB_{12} 2000 3500 AB_{12} 2000 3500 MB_{12} 2000 3500 HB_{12} 2000 3500 HB_{12} 2000 3500 HB_{12} 2000 3500 HB_{12} 3000 2000 HET 3000 2000 DB_7 3000 3000 DMB 3000 3500	B ₁	3000	2000
B_3 3000 3000 B_5 3000 3000 B_6 3000 3000 B_7 3000 3000 B_9 3000 3500 CB_{12} 2000 3500 AB_{12} 2000 3500 MB_{12} 2000 3500 HB_{12} 2000 3500 HMP 3000 2000 HET 3000 2000 DB_7 3000 3000 DMB 3000 3500	B_2	3000	2500
B530003000B630003000B730003000B930003500CB1220003500AB1220003500MB1220003500HB1220003500HMP30002000HET30003000DB730003000Cbi30003500	B ₃	3000	3000
B630003000B730003000B930003500CB1220003500AB1220003500MB1220003500HB1220003500HMP30002000HET30002000DB730003000DMB30003500Cbi30003500	B ₅	3000	3000
B730003000B930003500CB1220003500AB1220003500MB1220003500HB1220003500HMP30002000HET30002000DB730003000DMB30003000Cbi30003500	B ₆	3000	3000
B930003500CB1220003500AB1220003500MB1220003500HB1220003500HMP30002000HET30002000DB730003000DMB30002000Cbi30003500	\mathbf{B}_7	3000	3000
CB1220003500AB1220003500MB1220003500HB1220003500HMP30002000HET30002000DB730003000DMB30002000Cbi30003500	B 9	3000	3500
AB_{12} 20003500 MB_{12} 20003500 HB_{12} 20003500 HMP 30002000 HET 30002000 DB_7 30003000DMB30002000Cbi30003500	CB_{12}	2000	3500
MB ₁₂ 2000 3500 HB ₁₂ 2000 3500 HMP 3000 2000 HET 3000 2000 DB ₇ 3000 3000 DMB 3000 2000 Cbi 3000 3500	AB_{12}	2000	3500
HB1220003500HMP30002000HET30002000DB730003000DMB30002000Cbi30003500	MB ₁₂	2000	3500
HMP30002000HET30002000DB730003000DMB30002000Cbi30003500	HB_{12}	2000	3500
HET30002000DB730003000DMB30002000Cbi30003500	HMP	3000	2000
DB7 3000 3000 DMB 3000 2000 Cbi 3000 3500	HET	3000	2000
DMB 3000 2000 Cbi 3000 3500	DB ₇	3000	3000
Cbi 3000 3500	DMB	3000	2000
	Cbi	3000	3500



Fig. S1 Fragmentation of B_3 with remaining molecular ion (top), fragmentation of AB_{12} into multiple fragment ions of moderate intensity (middle), fragmentation of B_7 into one intense fragment ion (bottom). All spectra were measured on the TSQ Quantum



Fig. S2 Summed mass spectrum of a full scan measurement of a seawater extract. The signal of a background contamination at m/z 214 was cut off to show the distribution of the remaining masses in the elution range of the vitamins



Fig. S3 Signals of HET from PRM measurements with resolutions of (a) 15.000, (b) 60.000, and (c) 120.000

	LOI	D TSQ SRM	LOD Orbitrap PRM		LOD C	brbitrap full scan
Vitamin	ng/l	pg on column	ng/l	pg on column	ng/l	pg on column
B ₁	1000	5.00	20000	100.00	20000	100.00
B_2	50	0.25	50	0.25	50	0.25
B_3	500	2.50	100	0.50	50	0.25
B ₅	10	0.05	50	0.25	100	0.50
B_6	20	0.10	20	0.10	50	0.25
\mathbf{B}_7	50	0.25	50	0.25	200	1.00
B 9	500	2.50	200	1.00	200	1.00
CB_{12}	500	2.50	100	0.50	100	0.50
AB_{12}	500	2.50	100	0.50	100	0.50
MB_{12}	200	1.00	200	1.00	50	0.25
HB_{12}	20000	100.00	20000	100.00	50000	250.00
HMP	20	0.10	100	0.50	50	0.25
HET	10	0.05	10	0.05	10	0.05
DB_7	10	0.05	20	0.10	20	0.10
DMB	10	0.05	20	0.10	50	0.25
Cbi	500	2.50	1000	5.00	1000	5.00

Table S3 Instrumental limits of detection in ng/l and pg on column with 5 μ l injection volume. Data represent thelowest concentration that gave a reproducible signal in the linear range



Fig. S4 EIC of CB₁₂ with concentrations of (a) 250 pg/ml, (b) 125 pg/ml, (c) 25 pg/ml with SRM at the TSQ; (d) 100 pg/ml, (e) 50 pg/ml, (f) 20 pg/ml with PRM at the Orbitrap and (g) 50 pg/ml, (h) 20 pg/ml, (i) 10 pg/ml with full scan at the Orbitrap. For the HRMS measurements, no noise is seen besides the analyte peak



Fig. S5 Extracted ion chromatograms of 16 different B vitamins and biosynthetic precursors from a North Sea water sample, concentrated by solid phase extraction, measured with three different MS methods (selected reaction monitoring (SRM), parallel reaction monitoring (PRM) and full scan (FS))

	Т	SQ	Orbitrap		
Vitamin	no Matrix	with Matrix	no Matrix	with Matrix	
B ₃	1.11	1.11	0.83	0.85	
B ₅	1.60	1.90	1.24	1.55	
B_6	2.26	2.29	1.76	1.84	
HMP	2.33	1.66	1.68	1.06	
\mathbf{B}_1	2.89	3.25	2.90	3.51	
\mathbf{B}_7	3.58	3.99	3.95	4.13	
B 9	3.61	3.84	4.02	3.95	
DB_7	4.78	5.01	4.91	5.02	
HET	5.13	5.22	4.99	4.98	
B_2	8.08	8.18	7.69	7.69	
CB_{12}	8.18	8.25	7.80	7.81	
AB_{12}	8.90	9.02	8.47	8.49	
Cbi	9.63	9.78	9.22	9.24	
DMB	11.00	11.13	10.27	10.36	
MB_{12}	11.03	11.09	10.61	10.62	
HB_{12}	6.59	6.91	6.00	6.12	

Table S4 Retention times of individual vitamins at TSQ and Orbitrap in the presence or absence of matrix



Fig. S6 Regression lines illustrating linear response and calibration behavior of the respective analytes. Furthermore, ion suppression via addition of known amounts of B_1 (left) and B_2 (right) to ultrapure water (red circles) and a processed seawater sample (black squares) is reflected by the decline in slope. The three methodological approaches compared are: SRM on the TSQ Quantum, PRM and FS on the Orbitrap Fusion, respectively. The regression lines in black with the linear equations and the squared correlation coefficients are displayed as well as the 95% confidence (dark red) and prediction (light red) bands



Fig. S7 Regression lines illustrating linear response and calibration behavior of the respective analytes. Furthermore, ion suppression via addition of known amounts of B_3 (left) and B_5 (right) to ultrapure water (red circles) and a processed seawater sample (black squares) is reflected by the decline in slope. The three methodological approaches compared are: SRM on the TSQ Quantum, PRM and FS on the Orbitrap Fusion, respectively. The regression lines in black with the linear equations and the squared correlation coefficients are displayed as well as the 95% confidence (dark red) and prediction (light red) bands



Fig. S8 Regression lines illustrating linear response and calibration behavior of the respective analytes. Furthermore, ion suppression via addition of known amounts of B_6 (left) and B_9 (right) to ultrapure water (red circles) and a processed seawater sample (black squares) is reflected by the decline in slope. The three methodological approaches compared are: SRM on the TSQ Quantum, PRM and FS on the Orbitrap Fusion, respectively. The regression lines in black with the linear equations and the squared correlation coefficients are displayed as well as the 95% confidence (dark red) and prediction (light red) bands



Fig. S9 Regression lines illustrating linear response and calibration behavior of the respective analytes. Furthermore, ion suppression via addition of known amounts of CB_{12} (left) and MB_{12} (right) to ultrapure water (red circles) and a processed seawater sample (black squares) is reflected by the decline in slope. The three methodological approaches compared are: SRM on the TSQ Quantum, PRM and FS on the Orbitrap Fusion, respectively. The regression lines in black with the linear equations and the squared correlation coefficients are displayed as well as the 95% confidence (dark red) and prediction (light red) bands



Fig. S10 Regression lines illustrating linear response and calibration behavior of the respective analytes. Furthermore, ion suppression via addition of known amounts of HB_{12} (left) and HMP (right) to ultrapure water (red circles) and a processed seawater sample (black squares) is reflected by the decline in slope. The three methodological approaches compared are: SRM on the TSQ Quantum, PRM and FS on the Orbitrap Fusion, respectively. The regression lines in black with the linear equations and the squared correlation coefficients are displayed as well as the 95% confidence (dark red) and prediction (light red) bands



Fig. S11 Regression lines illustrating linear response and calibration behavior of the respective analytes. Furthermore, ion suppression via addition of known amounts of HET (left) and DB₇ (right) to ultrapure water (red circles) and a processed seawater sample (black squares) is reflected by the decline in slope. The three methodological approaches compared are: SRM on the TSQ Quantum, PRM and FS on the Orbitrap Fusion, respectively. The regression lines in black with the linear equations and the squared correlation coefficients are displayed as well as the 95% confidence (dark red) and prediction (light red) bands



Fig. S12 Regression lines illustrating linear response and calibration behavior of the respective analytes. Furthermore, ion suppression via addition of known amounts of DMB (left) and Cbi (right) to ultrapure water (red circles) and a processed seawater sample (black squares) is reflected by the decline in slope. The three methodological approaches compared are: SRM on the TSQ Quantum, PRM and FS on the Orbitrap Fusion, respectively. The regression lines in black with the linear equations and the squared correlation coefficients are displayed as well as the 95% confidence (dark red) and prediction (light red) bands

Vitamin	SRM (pl	M)	PRM (pM)	full sc	an (j	oM)
B_1	26.8 ±	7.5	16.6 ±	5.3		nd	
B_2	35.9 ±	2.4	31.9 ±	3.5	18.6	±	0.9
B ₃	nd		26.0 ±	4.3	46.3	±	9.6
B ₅	97.2 ±	8.2	53.4 <u>+</u>	3.1	38.8	±	4.5
B ₆	40.2 ±	12.4	31.3 ±	2.1	91.0	±	8.6
B ₇	29.9 ±	2.5	0.8 ±	0.6		nd	
B ₉	nd		n	d		nd	
CB ₁₂	15.4 ±	1.7	22.1 ±	2.5	15.4	±	1.2
AB_{12}	0.6 ±	0.1	1.1 ±	0.2	0.4	±	0.2
MB ₁₂	nd		2.3 ±	1.3		nd	
HB_{12}	nd		n	d		nd	
HMP	15.8 ±	2.4	12.9 ±	1.8	27.3	±	4.2
HET	1.4 ±	0.2	2.8 ±	0.2	1.4	±	0.2
DB ₇	5.6 ±	0.7	1.4 ±	0.1		nd	
DMB	13.7 ±	0.8	13.7 ±	1.7	9.6	±	0.8
Cbi	$1.1 \pm$	0.2	0.9 ±	0.1	0.7	±	0.4
nd = not detected							

Table S5 Vitamin content in a seawater sample from the North Sea in pM

Phaeobacter inhibens (DSM 17395)



Fig. S13 Growth curve of *P. inhibens* (DSM 17395), determined by optical density (circles). The cell counts per ml at the time of sampling are shown

	I						
	molecules per cell \pm SD						
Vitamin	TSQ SF	RM	Orbit	trap	PRM	Orbitrap f	ull scan
B ₁	21393 ±	6394	16616	±	3962	$18701 \hspace{.1in} \pm \hspace{.1in}$	5077
B_2	$1829 \ \pm$	349	2879	±	954	2550 \pm	763
B ₃	160950 \pm	30245	140318	±	22659	$126079 \ \pm$	23554
B ₅	$2988 \pm$	988	6635	±	2387	$2841 \hspace{0.2cm} \pm \hspace{0.2cm}$	1069
B_6	690 ±	74	668	±	75	$673 \hspace{0.1in} \pm \hspace{0.1in}$	67
B ₇	$49 \hspace{0.2cm} \pm \hspace{0.2cm}$	4	14	±	2	14 ±	2
B 9	391 ±	106	475	±	158	$448 \hspace{0.1in} \pm \hspace{0.1in}$	148
CB ₁₂	nd		10	±	2	9 ±	2
AB_{12}	549 ±	79	623	±	58	574 \pm	55
MB_{12}	55 \pm	21	57	±	24	55 \pm	22
HB_{12}	nd			nd		nd	
HMP	1299 ±	616	1245	±	619	1260 \pm	589
HET	17 ±	6	15	±	6	$13 \pm$	5
DB ₇	$802 \pm$	90	584	±	66	645 \pm	69
DMB	$4976 \ \pm$	523	4824	±	367	$4723 \hspace{0.2cm} \pm \hspace{0.2cm}$	421
Cbi	nd			nd		nd	
αRib	194 ±	26	192	±	27	185 \pm	25
nd = not de	tected						

Table S6 Intracellular amount of 17 different B vitamins and biosynthetic precursors in molecules per cell P.

 inhibens

SD = standard deviation



Fig. S14 Percentage of all vitamins and precursors found in a seawater sample from the North Sea (top) and in an extract of cells of the bacterium *P. inhibens* (bottom). Display of all analytes found separately without B_1 and B_3 (bottom right)