Supporting Information

Structure-Activity Relationships of Styrylquinoline and Styrylquinoxaline

Derivatives as α-Synuclein Imaging Probes

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Synthesis method for known compounds

1. Materials and Methods

General Remarks

All reagents were commercial products used without further purification unless indicated otherwise. All compounds were purified by Smart Flash EPCLC W-Prep 2XY (Yamazen Corporation, Osaka, Japan) unless otherwise stated. ¹H and ¹³C NMR spectra were recorded on a JEOL JNM-ECS400 and JEOL JNM-ECA500 (JEOL, Tokyo, Japan) with tetramethylsilane as an internal standard. Coupling constants are reported in Hertz. Multiplicity was defined as singlet (s), doublet (d), triplet (t), quartet (q), or multiplet (m). ESI mass spectrometry was conducted with a Shimadzu LCMS-2020 EV (Shimadzu, Kyoto, Japan). High-resolution mass spectrometry (HRMS) was carried out with a JEOL JMS-700 (JEOL). HPLC was performed with a Shimadzu system (an LC-20AD pump with SPD-20A UV detector, $\lambda = 254$ nm) using a Cosmosil C₁₈ column (5C₁₈-AR-II 4.6 mm I.D. × 150 mm, Nacalai Tesque, Kyoto, Japan) and MeCN/H₂O (with 0.1% TFA) or MeCN/H₂O as the mobile phase at a flow rate of 1.0 mL/min. All key compounds were proven by this method to show > 95% purity. No unexpected or unusually high safety hazards were encountered.

Chemistry

7-Bromo-2-methylquinoxaline (1)/ 6-Bromo-2-methylquinoxaline (1')¹

To a stirred solution of 4-bromo-1,2-benzenediamine (600 mg, 3.21 mmol) in EtOH (6.00 mL) was added pyruvic aldehyde (35-45%, w/w aq. soln) (2.40 mL, 11.7-15.0 mmol) at 25 °C. After being stirred for 10 min, the mixture was diluted with H₂O. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-EtOAc (2:1) to give mixture of **1** and **1**' as a yellow solid (621 mg, 87%). ¹H NMR (400 MHz, CDCl₃): δ 2.70–2.79 (m, 3H), 7.69–7.96 (m, 2H), 8.11–8.27 (m, 1H), 8.64–8.78 (m, 1H); LC-MS *m/z*: [M + H]⁺ calcd for C₉H₈BrN₂, 223.0, 225.0; found, 223.0, 225.0.

(E)-N,N-Dimethyl-4-(2-(7-(tributylstannyl)quinoxalin-2-yl)vinyl)aniline (2)

To a stirred solution of a mixture of **1** and **1'** (408 mg, 1.83 mmol, 1.00 eq.), *p*-dimethylaminobenzaldehyde (1030 mg, 7.31 mmol), and piperidine (4.10 mL) in dry toluene (15.0 mL) was added AcOH (4.10 mL) dropwisely at 25 °C under Ar. After being stirred for 24 h at 120 °C, the mixture was diluted with saturated aqueous NaHCO₃. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried

over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) to give a crude styryl compound as a red solid (1.12 g). To a stirred solution of the crude styryl compound (48.1 mg, 0.136 mmol) and Bu₄Sn (323 mg, 0.557 mmol) in dry toluene (3.30 mL) was added Pd(PPh₃)₄ (15.7 mg, 0.0136 mmol) under Ar. After being stirred for 8 h at 110 °C, the mixture was diluted with H₂O. The whole solution was extracted with CHCl₃. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) to give **2** as a red solid (19.0 mg, 3.4% yield, 2 steps): ¹H NMR (400 MHz, CDCl₃): δ 0.90 (m, 9H), 1.16 (m, 6H), 1.36 (m, 6H), 1.59 (m, 6H), 3.03 (s, 6H), 6.74 (d, *J* = 8.7 Hz, 2H), 7.18 (d, *J* = 16.0 Hz, 1H), 7.56 (d, *J* = 8.7 Hz, 2H), 7.72–7.82 (m, 2H), 7.97 (d, *J* = 8.2 Hz, 2H), 8.14 (s, 1H), 8.99 (s, 1H); LC-MS *m/z*: [M + H]⁺ calcd for C₃₀H₄₄N₃Sn, 566.3; found, 566.2.

(E)-4-(2-(7-Iodoquinoxalin-2-yl)vinyl)-N,N-dimethylaniline (3, ISQ)

To a stirred solution of **2** (55.4 mg, 0.0981 mmol) in CHCl₃ (0.750 mL) was added I₂ (24.9 mg, 0.0981 mmol) in CHCl₃ (0.750 mL). After being stirred for 2 h at 25 °C, the mixture was quenched with saturated aqueous NaHSO₃. The whole solution was extracted with CHCl₃. The extract was washed with brine and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) to give **3** (ISQ) as a red solid (20.8 mg, 53% yield). The structure of **3** was confirmed by X-ray crystallography: ¹H NMR (400 MHz, CDCl₃): δ 3.03 (s, 6H), 6.72 (d, *J* = 8.2 Hz, 2H), 7.11 (d, *J* = 16.0 Hz, 1H), 7.54 (d, *J* = 8.2 Hz, 2H), 7.71 (d, *J* = 8.7 Hz, 1H), 7.80 (d, *J* = 16.0 Hz, 1H), 7.84–7.89 (m, 1H), 8.41–8.44 (m, 1H), 8.94 (s, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃): δ 40.2 (2C), 96.0, 112.0 (2C), 119.8, 123.7, 129.1 (2C), 130.3, 137.1, 137.7, 137.8, 140.2, 143.4, 145.1, 151.2, 152.1; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₁₈H₁₇N₃I, 402.0467; found, 402.0463.

(E)-4-(2-(7-(2-Fluoroethoxy)quinoxalin-2-yl)vinyl)-N,N-dimethylaniline (4, SQ1) (E)-4-(2-(6-(2-Fluoroethoxy)quinoxalin-2-yl)vinyl)-N,N-dimethylaniline (5, SQ2)

The same reaction as described above to prepare the crude styryl compound (synthesis of **2**) was used. To a stirred solution of the crude styryl compound (1.12 g), Cu(acac)₂ (79.1 mg, 0.302 mmol), LiOH·H₂O (436 mg, 10.4 mmol), and N^1 , N^2 -bis(4-hydroxy-2,6-dimethylphenyl)oxalamide (99.2 mg, 0.302 mmol) in dry DMSO (6.00 mL) was added sonicated H₂O (1.50 mL) under Ar. After being stirred for 24 h at 80 °C under Ar, the mixture was dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) and CHCl₃-MeOH

(10:1) following flash chromatography over amino silica gel with *n*-hexane-AcOEt (6:1) and CHCl₃-MeOH (10:1) to give a crude phenol compound as a red liquid (224 mg). To a stirred solution of the crude phenol compound (169 mg) and Cs₂CO₃ (860 mg, 2.64 mmol) in dry DMF (6.00 mL) was added 2-fluoroethyl p-toluenesulfonate (444 µL, 2.60 mmol) under Ar. After being stirred overnight at 95 °C under Ar, the mixture was diluted with H₂O. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration in vacuo, the residue was purified by flash chromatography over amino silica gel with *n*-hexane-AcOEt 4:1 to 2:1 to give 4 (SQ1) as an orange solid (45.0 mg, 6.2% yield, 3 steps) and 5 (SQ2) as an orange solid (38.0 mg, 7.3% yield, 3 steps). The structures of 4 and 5 were confirmed by X-ray crystallography. [4]: ¹H NMR (400 MHz, CDCl₃): δ 3.01 (s, 6H), 4.33 (t, J = 4.1 Hz, 1H), 4.40 (t, J = 4.1 Hz, 1H), 4.78 (t, J = 4.1 Hz, 1H), 4.90 (t, J = 4.1 Hz, 1H), 6.71 (d, J = 8.7Hz, 2H), 7.11 (d, J = 16.2 Hz, 1H), 7.28 (d, J = 2.3 Hz, 1H), 7.33 (dd, J = 9.3, 2.3 Hz, 1H), 7.53 (d, J = 8.7 Hz, 2H), 7.76 (d, J = 16.2 Hz, 1H), 7.91 (d, J = 9.3 Hz, 1H), 8.83 (s, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.2 (2C), 67.3, 67.5, 80.7, 82.4, 107.1, 112.1 (2C), 120.4, 121.6, 124.0, 128.9 (2C), 130.2, 136.6, 137.3, 142.3, 143.9, 151.0, 151.6, 159.6; HRMS (ESI-TOF) m/z: $[M + H]^+$ calcd for C₂₀H₂₁FN₃O, 338.1669; found, 338.1670. [5]: ¹H NMR (400 MHz, CDCl₃): δ 3.00 (s, 6H), 4.29–4.34 (m, 1H), 4.36–4.42 (m, 1H), 4.73-4.79 (m, 1H), 4.85-4.91 (m, 1H), 6.71 (d, J = 8.7 Hz, 2H), 7.11-7.20 (m, 2H), 7.32–7.36 (m, 1H), 7.45–7.54 (m, 3H), 7.55–7.65 (m, 2H), 7.96 (d, J = 8.1, 3.5 Hz, 1H); ${}^{13}C{}^{1}H$ NMR (100 MHz, CDCl₃): δ 40.2 (2C), 67.0, 67.2, 80.8, 82.5, 107.7, 112.1 (2C), 117.2.5, 118.7. 122.4, 124.3, 124.6, 128.5 (2C), 128.6, 134.6, 135.6, 149.7, 150.7, 157.2, 159.5; HRMS (ESI-TOF) m/z: $[M + H]^+$ calcd for C₂₀H₂₁FN₃O, 338.1669; found, 338.1670.

7-Bromo-2-methylquinoline (6)²

To a stirred solution of 3-bromoaniline (1170 µL, 10.7 mmol) in AcOH (10.0 mL) was added ethyl vinyl ether (3.08 mL, 32.0 mmol). After being stirred for 5 h at 25 °C, the temperature was increased to 100 °C. After being stirred for 4 h at 100 °C, the mixture was quenched with aqueous NaOH. The whole solution was extracted with EtOAc. The extract was washed with brine, and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-EtOAc (8:1) to give **6** as a yellow solid (431 mg, 18% yield): ¹H NMR (400 MHz, CDCl₃): δ 2.70 (s, 3H), 7.23 (d, *J* = 8.7 Hz, 1H), 7.49 (dd, *J* = 8.7, 1.2 Hz, 1H), 7.55 (d, *J* = 8.7 Hz, 1H), 7.92 (dd, *J* = 8.7, 2.9 Hz, 2H), 8.17 (d, *J* = 1.2 Hz, 1H); LC-MS *m/z*: [M + H]⁺ calcd for C₁₀H₉NBr, 221.0, 223.0; found, 221.1, 223.1.

6-Bromo-2-methylquinoline (7)²

By a procedure identical to that described for synthesis of **6**, 4-bromoaniline (860 mg, 5.00 mmol) and ethyl vinyl ether (1.44 mL, 15.0 mmol) were converted into **7** as a yellow solid (432 mg, 39% yield): ¹H NMR (400 MHz, CDCl₃): δ 2.67 (s, 3H), 7.17 (d, J = 8.7 Hz, 1H), 7.66 (dd, J = 8.7, 1.7 Hz, 1H), 7.75–7.85 (m, 3H); LC-MS m/z: [M + H]⁺ calcd for C₁₀H₉NBr, 221.0, 223.0; found, 221.1, 223.1.

5-Bromo-2-methylquinoline (8)³

To a stirred solution of 2-methylquinoline (2.19 g, 15.3 mmol) in conc. H₂SO₄ (15.0 mL) was added NBS (2.72 g, 15.3 mmol) under Ar. After being stirred overnight at 25 °C, the mixture was diluted with iced H₂O. After being quenched with saturated aqueous Na₂SO₃, the whole solution was extracted with CHCl₃. The extract was washed with brine, and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-EtOAc (10:1) to give **8** as a yellow solid (195 mg, 6% yield): ¹H NMR (400 MHz, CDCl₃): δ 2.73 (s, 3H), 7.29 (d, *J* = 8.7 Hz, 1H), 7.48 (d, *J* = 8.1 Hz, 1H), 7.70 (dd, *J* = 8.1, 8.1 Hz, 1H), 7.96 (d, *J* = 8.7 Hz, 1H), 8.33 (d, *J* = 8.1 Hz, 1H), 7.92 (d, *J* = 8.7 Hz, 1H); LC-MS *m/z*: [M + H]⁺ calcd for C₁₀H₉NBr, 221.0, 223.0; found, 221.1, 223.1.

8-Bromo-2-methylquinoline (9)³

By a procedure identical to that described for synthesis of **8**, 2-methylquinoline (2.19 g, 15.3 mmol) and NBS (2.72 g, 15.3 mmol) were converted into **9** as a yellow solid (493 mg, 15% yield): ¹H NMR (400 MHz, CDCl₃): δ 2.73 (s, 3H), 7.15–7.23 (m, 2H), 7.56–7.61 (m, 1H), 7.86 (d, J = 8.7 Hz, 1H), 7.92 (d, J = 8.7 Hz, 1H); LC-MS m/z: [M + H]⁺ calcd for C₁₀H₉NBr, 221.0, 223.0; found, 221.1, 223.1.

6-(Dimethylamino)nicotinaldehyde (10)⁴

To a stirred solution of NHMe₂ in H₂O (abt. 50%, 10.0 mL) was added 2-chloro-5pyridinecarbaldehyde (1040 mg, 7.34 mmol). After being stirred for 4 h at 90 °C, the mixture was diluted with H₂O. The whole solution was extracted with EtOAc. The extract was washed with brine, and dried over Na₂SO₄. Evaporate the solvent *in vacuo* to give **10** as a white solid (1170 mg, quant.): ¹H NMR (400 MHz, CDCl₃): δ 3.21 (s, 6H), 6.55 (d, *J* = 8.7 Hz, 1H), 7.89 (dd, *J* = 2.3, 8.7 Hz, 1H), 8.55 (d, *J* = 2.3 Hz, 1H), 9.76 (s, 1H); LC-MS *m/z*: [M + H]⁺ calcd for C₈H₁₁N₂O, 151.1; found, 151.1.

4-(1H-1,2,3-Triazol-1-yl)benzaldehyde (11) ⁵

To a stirred solution of 4-fluorobenzaldehyde (2.00 g, 16.0 mmol) and 1, 2, 3 triazole (1.32 g, 19.0 mmol) in DMF (50.0 mL) was added K₂CO₃ (3.30g, 23.9 mmol). After being stirred for 5 h at 100 °C, the mixture was diluted with H₂O. The whole solution was extracted with EtOAc. The extract was washed with brine, and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-EtOAc (1:1) to give crude phenol compound as a yellow solid (792 mg, 29% yield): ¹H NMR (400 MHz, CDCl₃): δ 3.21 (s, 6H), 6.55 (d, *J* = 8.7 Hz, 1H), 7.89 (dd, *J* = 2.3, 8.7 Hz, 1H), 8.55 (d, *J* = 2.3 Hz, 1H), 9.76 (s, 1H).

2-(2-(Tosyloxy)ethoxy)ethoxy)ethyl hypofluorite (12) ⁶

To a stirred solution of triethylene glycol bis(*p*-toluenesulfonate) (608 mg, 1.21 mmol) in dry THF (5.00 mL) was added TBAF in THF (1.0 M, 2.42 mL) under Ar. The mixture was stirred for 15 min at 70 °C. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-EtOAc 4:1 to EtOAc only to give **12** as a colorless liquid (350 mg, 90% yield): ¹H NMR (400 MHz, CDCl₃): δ 2.46 (s, 3H), 3.50–3.80 (m, 8H), 4.09–4.21 (m, 2H), 4.49 (t, *J* = 4.1 Hz, 1H), 4.61 (t, *J* = 4.1 Hz, 1H), 7.36 (d, *J* = 7.5 Hz, 2H), 7.80 (d, *J* = 7.5 Hz, 2H); LC-MS *m/z*: [M + H]⁺ calcd for C₁₃H₁₉FO₆SNa, 345.1; found, 345.1.

1-Chloro-3-fluoropropan-2-ol (13)⁷

The solution of epichlorohydrin (1.00 g, 10.8 mmol) in TREAT HF (1.77 mL, 10.8 mmol) was stirred for 2 h at 130 °C in a sealed tube. The mixture was stirred for 15 min at 70 °C. After being quenched with iced H₂O, the whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. Evaporate the solvent *in vacuo* to give crude **13** as a colorless liquid (1000 mg).

(E)-4-(2-(7-(2-Fluoroethoxy)quinolin-2-yl)vinyl)-N,N-dimethylaniline (14, SQ3)

To a stirred solution of **6** (353 mg, 1.59 mmol, 1.00 eq.), *p*dimethylaminobenzaldehyde (949 mg, 6.36 mmol), and piperidine (4.10 mL) in dry toluene (15.9 mL) was added AcOH (4.10 mL) dropwisely at 25 °C under Ar. After being stirred for 24 h at 120 °C, the mixture was diluted with saturated aqueous NaHCO₃. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) to give a crude styryl compound as a yellow solid (920 mg). To a stirred solution of the crude styryl compound (920 mg), Cu(acac)₂ (65.0 mg, 0.248 mmol), LiOH·H₂O (359 mg, 8.56 mmol), and N¹, N²-bis(4-hydroxy-2,6-dimethylphenyl)oxalamide (81.5 mg, 0.248 mmol) in dry DMSO (6.00 mL) was added sonicated H₂O (1.50 mL). After being stirred for 24 h at 80 °C under Ar, the mixture was dried over Na₂SO₄. After concentration in vacuo, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) and CHCl₃-MeOH (10:1) following flash chromatography over amino silica gel with n-hexane-AcOEt (6:1) and CHCl₃-MeOH (10:1) to give a crude phenol compound as a black liquid (127 mg). To a stirred solution of the crude phenol compound (126 mg), and NaH (31.0 mg, 1.30 mmol) in dry DMF (6.00 mL) was added 2-fluoroethyl p-toluenesulfonate (222 μ L, 1.30 mmol). After being stirred overnight at 95 °C, the mixture was diluted with H₂O. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration in vacuo, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) and CHCl₃-MeOH (10:1) following flash chromatography over amino silica gel with *n*-hexane-AcOEt (6:1) and CHCl₃-MeOH (10:1) to give 14 as a yellow solid (92.4 mg, 17% yield): ¹H NMR (400 MHz, CDCl₃): δ 3.01 (s, 6H), 4.32 (t, J = 4.1 Hz, 1H), 4.39 (t, J = 4.1 Hz, 1H), 4.77 (t, J = 4.1 Hz, 1H), 4.89 (t, J = 4.1 Hz, 1H), 6.72 (d, J = 8.7 Hz, 2H), 7.12 (d, J = 16.2 Hz, 2H), 7.32 (d, J = 2.3 Hz, 1H), 7.42 (dd, J = 9.3, 2.3 Hz, 1H), 7.53 (d, J = 8.7 Hz, 2H), 7.72 (d, J = 16.2 Hz, 1H), 7.92 (d, J = 9.3 Hz, 1H), 8.91 (s, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.2 (2C), 67.3, 67.5, 80.7, 82.4, 107.6, 112.1 (2C), 120.5, 123.1, 124.2, 128.7 (2C), 130.0, 135.5, 138.7, 142.3, 144.5, 149.7, 150.9, 158.5; HRMS (ESI-TOF) m/z: [M + H]⁺ calcd for C₂₁H₂₂FN₂O, 337.1716; found, 337.1718.

(E)-4-(2-(6-(2-Fluoroethoxy)quinolin-2-yl)vinyl)-N,N-dimethylaniline (15, SQ4)

To a stirred solution of a mixture of 7 (432 mg, 1.95 mmol, 1.00 eq.), *p*dimethylaminobenzaldehyde (1370 mg, 9.72 mmol), and piperidine (4.31 mL) in dry toluene (16.0 mL) was added AcOH (4.31 mL) dropwisely at 25 °C under Ar. After being stirred for 24 h at 120 °C, the mixture was diluted with saturated aqueous NaHCO₃. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) to give a crude styryl compound as a yellow solid (622 mg). To a stirred solution of the crude styryl compound (622 mg), Cu(acac)₂ (46.0 mg, 0.176 mmol), LiOH·H₂O (359 mg, 9.13 mmol), and N^1 , N^2 -bis(4-hydroxy-2,6-dimethylphenyl)oxalamide (57.4 mg, 0.175 mmol) in dry DMSO (6.00 mL) was added sonicated H₂O (1.50 mL). After being stirred for 24 h at 80 °C under Ar, the mixture was dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over silica gel with *n*-hexane-AcOEt (6:1) and CHCl₃-MeOH (10:1) following flash chromatography over amino silica gel with n-hexane-AcOEt (6:1) and CHCl₃-MeOH (10:1) to give a crude phenol compound as a black liquid (394 mg). To a stirred solution of the crude phenol compound (394 mg), Cs₂CO₃ (1330 mg, 4.07 mmol) in dry DMF (6.00 mL) was added 2-fluoroethyl p-toluenesulfonate (347 μ L, 2.04 mmol). After being stirred overnight at 95 °C, the mixture was diluted with H₂O. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration in vacuo, the residue was purified by flash chromatography over amino silica gel with n-hexane-AcOEt 4:1 to 1:1 and silica gel with *n*-hexane-AcOEt 3:1 to 2:1 to give **15** as a yellow solid (92.4 mg, 17% yield): ¹H NMR $(400 \text{ MHz}, \text{CDCl}_3)$: $\delta 3.00 \text{ (s, 6H)}, 4.29 \text{ (t, } J = 4.1 \text{ Hz}, 1\text{H}), 4.36 \text{ (t, } J = 4.1 \text{ Hz}, 1\text{H}), 4.76$ (t, J = 4.1 Hz, 1H), 4.88 (t, J = 4.1 Hz, 1H), 6.72 (d, J = 8.7 Hz, 2H), 7.04 (d, J = 2.9 Hz, 10.1 Hz)1H), 7.18 (d, J = 16.2 Hz, 1H), 7.37 (dd, J = 8.7, 2.9 Hz, 1H), 7.47–7.57 (m, 3H), 7.60 (d, J = 8.7 Hz, 1H), 7.93–7.98 (m, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.3 (2C), 67.2, 67.4, 81.0, 82.7, 106.4, 112.2 (2C), 119.4. 122.0, 124.4, 124.8, 127.7, 128.4 (2C), 130.5, 133.8, 134.9, 144.4, 150.6, 155.0, 156.0; HRMS (ESI-TOF) m/z: [M + H]⁺ calcd for C₂₁H₂₂FN₂O, 337.1716; found, 337.1711.

(E)-4-(2-(5-(2-Fluoroethoxy)quinolin-2-yl)vinyl)-N,N-dimethylaniline (16, SQ5)

By a procedure identical to that described for synthesis of **15**, quinolone **8** (195 mg, 0.882 mmol, 1.00 eq.), *p*-dimethylaminobenzaldehyde (896 mg, 6.01 mmol), and 2-fluoroethyl *p*-toluenesulfonate (347 µL, 2.04 mmol) were converted into **16** as a red solid (8.90 mg, 3.0% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 3.01 (s, 6H), 4.33 (t, *J* = 4.1 Hz, 1H), 4.40 (t, *J* = 4.1 Hz, 1H), 4.77 (t, *J* = 4.1 Hz, 1H), 4.89 (t, *J* = 4.1 Hz, 1H), 6.66 (d, *J* = 8.72 Hz, 2H), 6.89 (d, *J* = 8.1 Hz, 1H), 7.37 (t, *J* = 16.2 Hz, 1H), 7.52 (t, *J* = 8.7 Hz, 2H), 7.66–7.83 (m, 5H), 8.75 (d, *J* = 8.7 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.3 (2C), 68.1, 68.3, 80.5, 82.2, 107.4, 112.4 (2C), 112.6, 113.2, 116.6, 118.4, 122.6, 131.3 (2C), 134.6, 137.7, 139.1, 145.7, 152.5, 154.0, 154.2; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₁H₂₂FN₂O, 337.1716; found, 337.1718.

(E)-4-(2-(8-(2-Fluoroethoxy)quinolin-2-yl)vinyl)-N,N-dimethylaniline (17, SQ6)

By a procedure identical to that described for synthesis of **15**, quinoline **9** (493 mg, 2.22 mmol, 1.00 eq.), *p*-dimethylaminobenzaldehyde (1260 mg, 8.45 mmol), and 2-fluoroethyl *p*-toluenesulfonate (347 µL, 2.04 mmol) were converted into **17** as a red solid (3.06 mg, 0.4% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 3.02 (s, 6H), 4.53 (t, *J* = 4.6 Hz, 1H), 4.60 (t, *J* = 4.6 Hz, 1H), 4.93 (t, *J* = 4.6 Hz, 1H), 5.05 (t, *J* = 4.6 Hz, 1H),

6.73 (d, J = 8.7 Hz, 2H), 7.11 (d, J = 8.7 Hz, 1H), 7.29 (d, J = 16.8 Hz, 1H), 7.34–7.40 (m, 2H), 7.51–7.61 (m, 3H), 7.70 (d, J = 8.7 Hz, 1H), 8.05 (d, J = 8.7 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.4 (2C), 68.5, 68.7, 81.3, 83.0, 110.9, 112.2 (2C), 119.2, 120.6, 124.8, 125.1, 125.5, 128.3, 128.6 (2C), 134.4, 136.0, 140.5, 150.7, 154.0, 156.2; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₁H₂₂FN₂O, 337.1716; found, 337.1714.

(E)-2-(4-(1H-1,2,3-Triazol-1-yl)styryl)-7-(2-fluoroethoxy)quinolone (18, SQ7)

By a procedure identical to that described for synthesis of **15**, quinoline **6** (300 mg, 1.35 mmol, 1.00 eq.), aldehyde **11** (792 mg, 4.57 mmol), and 2-fluoroethyl *p*-toluenesulfonate (338 µL, 1.98 mmol) were converted into **18** as a colorless solid (27.3 mg, 5.6% yield): ¹H NMR (400 MHz, DMSO-*d*⁶): δ 4.40 (t, *J* = 4.1 Hz, 1H), 4.47 (t, *J* = 4.1 Hz, 1H), 4.77 (t, *J* = 4.1 Hz, 1H), 4.89 (t, *J* = 4.1 Hz, 1H), 7.25 (dd, *J* = 8.7, 2.9 Hz, 2H), 7.40 (d, *J* = 2.9 Hz, 1H), 7.55 (d, *J* = 16.2 Hz, 1H), 7.71 (d, *J* = 8.7 Hz, 1H), 7.85–7.92 (m, 3H), 7.93–8.01 (m, 5H), 8.29 (d, J = 8.7 Hz, 1H); ¹³C {¹H} NMR (100 MHz, DMSO-*d*⁶): δ 107.9, 118.3, 119.0, 120.4 (2C), 122.5, 123.8, 128.6, 129.1, 129.9, 132.3, 134.5, 136.27, 136.32, 136.5, 149.3, 155.4, 159.3; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₁H₁₈FN₄O, 361.1465; found, 361.1465.

(E)-2-(4-(1H-1,2,3-Triazol-1-yl)styryl)-6-(2-fluoroethoxy)quinolone (19, SQ8)

By a procedure identical to that described for synthesis of **15**, quinoline **7** (323 mg, 1.46 mmol, 1.00 eq.), aldehyde **11** (756 mg, 4.36 mmol), and 2-fluoroethyl *p*-toluenesulfonate (338 µL, 1.98 mmol) were converted into **19** as a colorless solid (3.9 mg, 0.7% yield): ¹H NMR (400 MHz, DMSO-*d*⁶): δ 4.42 (t, *J* = 3.7 Hz, 1H), 4.50 (t, *J* = 3.7 Hz, 1H), 4.84 (t, *J* = 3.7 Hz, 1H), 4.95 (t, *J* = 3.7 Hz, 1H), 7.47 (d, *J* = 2.7 Hz, 1H), 7.51 (dd, *J* = 9.2, 2.7 Hz, 1H), 7.62 (d, *J* = 16.0 Hz, 1H), 7.85–7.94 (m, 2H), 7.97–8.08 (m, 6H), 8.32 (d, *J* = 9.2 Hz, 1H), 8.95 (d, *J* = 1.4 Hz, 1H); ¹³C{¹H} NMR (100 MHz, DMSO-*d*⁶): δ 67.3, 67.5, 81.2, 82.9, 106.8, 120.3 (2C), 122.3, 123.0, 128.1, 128.4 (2C), 130.0, 130.3, 131.5, 134.5, 135.4, 136.2, 136.7, 143.7, 153.1, 156.1; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₁H₁₇FN₄O, 361.1465; found, 361.1465.

(E)-5-(2-(7-(2-Fluoroethoxy)quinolin-2-yl)vinyl)-N,N-dimethylpyridin-2-amine (20, SQ9)

By a procedure identical to that described for synthesis of **15**, quinoline **6** (432 mg, 1.96 mmol, 1.00 eq.), aldehyde **10** (1020 mg, 6.79 mmol), and 2-fluoroethyl *p*-toluenesulfonate (304 μ L, 1.78 mmol) were converted into **20** as a yellow solid (79.3 mg, 12% yield, 3steps): ¹H NMR (400 MHz, CDCl₃): δ 3.14 (s, 6H), 4.34 (t, *J* = 4.6 Hz, 1H),

4.41 (t, J = 4.6 Hz, 1H), 4.78 (t, J = 4.6 Hz, 1H), 4.90 (t, J = 4.6 Hz, 1H), 6.56 (d, J = 8.7 Hz, 1H), 7.09–7.19 (m, 2H), 7.36 (d, J = 2.3 Hz, 1H), 7.49 (d, J = 8.7 Hz, 1H), 7.57 (d, J = 16.2 Hz, 1H) 7.65 (d, J = 8.7 Hz, 1H), 7.80 (dd, J = 8.7, 2.3 Hz, 1H), 8.00 (d, J = 8.7 Hz, 1H), 8.35 (d, J = 2.3 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 38.1 (2C), 67.0, 67.2, 80.9, 82.6, 106.0, 107.7, 117.4, 119.0, 120.5, 122.6, 124.9, 128.7, 131.6, 134.3, 135.9, 148.8, 149.8, 156.8, 159.1, 159.6; HRMS (ESI-TOF) *m*/*z*: [M + H]⁺ calcd for C₂₀H₂₁FN₃O, 338.1669; found, 338.1672.

(E)-5-(2-(6-(2-Fluoroethoxy)quinolin-2-yl)vinyl)-N,N-dimethylpyridin-2-amine (21, SQ10)

By a procedure identical to that described for synthesis of **15**, quinoline **7** (430 mg, 1.94 mmol, 1.00 eq.), aldehyde **10** (1170 mg, 7.76 mmol), and 2-fluoroethyl *p*-toluenesulfonate (338 µL, 1.98 mmol) were converted into **21** as a colorless solid (12.2 mg, 1.9% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 3.06 (s, 6H), 4.22 (t, *J* = 4.1 Hz, 1H), 4.29 (t, *J* = 4.1 Hz, 1H), 4.69 (t, *J* = 4.1 Hz, 1H), 4.80 (t, *J* = 4.1 Hz, 1H), 6.49 (d, *J* = 9.2 Hz, 1H), 6.97 (d, *J* = 2.7 Hz, 1H), 6.97 (d, *J* = 16.5 Hz, 1H), 7.31 (dd, *J* = 9.2, 2.7 Hz, 1H), 7.88 (d, *J* = 16.5 Hz, 1H), 7.52 (d, *J* = 8.7 Hz, 1H), 7.73 (dd, *J* = 8.7, 2.7 Hz, 1H), 7.88 (d, *J* = 2.3 Hz, 1H), 7.91 (d, *J* = 2.7 Hz, 1H), 8.26 (d, *J* = 2.3 Hz, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃): δ 38.1 (2C), 67.2, 67.4, 81.0, 82.7, 106.0 106.4, 119.4, 120.5, 122.2, 124.8, 127.8, 130.5, 130.8, 134.2, 135.1, 144.3, 148.6, 154.5, 156.1, 159.0; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₀H₂₁FN₃O, 338.1669; found, 338.1672.

(E)-7-(2-Fluoroethoxy)-2-(4-nitrostyryl)quinolone (22, SQ11)

By a procedure identical to that described for synthesis of **15**, quinoline **6** (294 mg, 1.32 mmol, 1.00 eq.), *p*-nitrobenzaldehyde (999 mg, 6.61 mmol), and 2-fluoroethyl *p*-toluenesulfonate (147 µL, 0.861 mmol) were converted into **22** as a colorless solid (25.0 mg, 5.6% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 4.37 (t, *J* = 4.6 Hz, 1H), 4.44 (t, *J* = 4.6 Hz, 1H), 4.80 (t, *J* = 4.6 Hz, 1H), 4.92 (t, *J* = 4.6 Hz, 1H), 7.23–7.26 (m, 1H), 7.40 (d, *J* = 2.3 Hz, 1H), 7.48 (d, *J* = 16.2 Hz, 1H), 7.54 (d, *J* = 8.7 Hz, 1H), 7.68–7.79 (m, 4H), 8.11 (d, *J* = 8.7 Hz, 1H), 8.26 (d, *J* = 8.7 Hz, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 67.1, 67.3, 80.8, 82.5, 107.9, 118.1, 120.2, 123.3, 124.2 (2C), 127.7 (2C), 128.8, 131.5, 133.2, 136.4, 143.0, 147.4, 149.9, 154.9, 160.0; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₁₉H₁₆FN₂O₃, 339.1145; found, 339.1148

(E)-6-(2-Fluoroethoxy)-2-(4-nitrostyryl)quinolone (23, SQ12)

By a procedure identical to that described for synthesis of **15**, quinoline **7** (377 mg, 1.71 mmol, 1.00 eq.), *p*-nitrobenzaldehyde (1040 mg, 6.86 mmol), and 2-fluoroethyl *p*-toluenesulfonate (196 µL, 1.15 mmol) were converted into **23** as a colorless solid (38.6 mg, 8.3% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 4.31 (t, *J* = 4.6 Hz, 1H), 4.38 (t, *J* = 4.6 Hz, 1H), 4.78 (t, *J* = 4.6 Hz, 1H), 4.90 (t, *J* = 4.6 Hz, 1H), 7.08 (d, *J* = 2.3 Hz, 1H), 7.40–7.51 (m, 2H), 7.60 (d, *J* = 8.7 Hz, 1H), 7.64–7.75 (m, 3H), 7.98–8.07 (m, 2H), 8.22 (d, *J* = 8.7 Hz, 2H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 67.3, 67.5, 80.9, 82.6, 106.2, 120.2, 122.8, 124.1 (2C), 127.5, 128.6, 130.6, 131.1, 133.1, 135.4, 143.1, 144.5, 147.2, 152.6, 156.9; HRMS (ESI-TOF) *m*/*z*: [M + H]⁺ calcd for C₁₉H₁₆FN₂O₃, 339.1145; found, 339.1142.

(E)-4-(2-(7-(2-(2-(2-Fluoroethoxy)ethoxy)ethoxy)quinolin-2-yl)vinyl)-N,Ndimethylaniline (24, SQ13)

By a procedure identical to that described for synthesis of **15**, quinoline **6** (252 mg, 1.14 mmol, 1.00 eq.), *p*-dimethylaminobenzaldehyde (677 mg, 4.54 mmol), and fluorine **12** (171 mg, 0.558 mmol) were converted into **25** as a yellow solid (2.7 mg, 0.6% yield): ¹H NMR (400 MHz, CDCl₃): δ 3.02 (s, 6H), 3.64–3.76 (m, 6H), 3.89 (t, *J* = 4.1 Hz, 2H), 4.29 (t, *J* = 4.1 Hz, 2H), 4.44 (t, *J* = 4.1 Hz, 1H), 4.56 (t, *J* = 4.1 Hz, 1H), 6.65 (d, *J* = 8.7 Hz, 2H), 7.21(dd, *J* = 8.7, 2.3 Hz, 1H), 7.28 (d, *J* = 16.2 Hz, 1H), 7.51 (d, *J* = 8.7 Hz, 2H), 7.63 (d, *J* = 8.7 Hz, 1H), 7.67 (d, *J* = 8.7 Hz, 1H), 7.70–7.77 (m, 2H), 8.23 (d, *J* = 8.7 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.3 (2C), 68.5, 69.2, 70.3, 70.5, 70.8(2C), 82.3, 84.0, 100.4, 112.3 (2C), 112.8, 121.45, 121.55, 122.6, 129.1, 131.0 (2C), 140.9, 142.0 (2C), 144.4, 152.3, 153.0, 163.6; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₅H₃₀FN₂O₃, 425.2240; found, 425.2237.

(E)-4-(2-(6-(2-(2-(2-Fluoroethoxy)ethoxy)ethoxy)quinolin-2-yl)vinyl)-N,Ndimethylaniline (25, SQ14)

By a procedure identical to that described for synthesis of **15**, quinoline **7** (333 mg, 1.50 mmol, 1.00 eq.), *p*-dimethylaminobenzaldehyde (895 mg, 6.00 mmol), and fluorine **12** (138 mg, 0.394 mmol) were converted into **25** as a yellow solid (40.0 mg, 6.3% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 2.91 (s, 6H), 3.60–3.73 (m, 6H), 3.84 (t, *J* = 4.6 Hz, 2H), 4.16 (t, *J* = 4.6 Hz, 2H), 4.42 (t, *J* = 4.6 Hz, 1H), 4.54 (t, *J* = 4.6 Hz, 1H), 6.64 (d, *J* = 8.7 Hz, 2H), 6.95 (d, *J* = 2.9 Hz, 1H), 7.10 (d, *J* = 16.2 Hz, 1H), 7.28 (dd, *J* = 8.7, 2.9 Hz, 1H), 7.40–7.48 (m, 3H), 7.50 (d, *J* = 8.7 Hz, 1H), 7.85 (d, *J* = 4.6 Hz, 1H), 7.87 (d, *J* = 4.6 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.2 (2C), 67.6, 69.7, 70.3, 70.5, 70.80, 70.84, 82.2, 83.9, 106.3, 112.2 (2C), 119.2, 122.2, 124.4, 124.8, 127.7, 128.4

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(2C), 130.2, 133.7, 134.9, 144.2, 150.6, 154.7, 156.3; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₅H₃₀FN₂O₃, 425.2240; found, 425.2237.

(E)-1-((2-(4-(Dimethylamino)styryl)quinolin-7-yl)oxy)-3-fluoropropan-2-ol (26, SQ15)

By a procedure identical to that described for synthesis of **15**, quinoline **6** (500 mg, 2.25 mmol, 1.00 eq.), *p*-dimethylaminobenzaldehyde (1340 mg, 9.00 mmol), and crude fluorine **13** (1.00 g) were converted into **26** as a red solid (65.2 mg, 6.8% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 2.92 (s, 6H), 4.16 (d, *J* = 4.6 Hz, 2H), 4.22–4.32 (m, 1H), 4.48–4.53 (m, 1H), 4.60–4.65 (m, 1H), 6.64 (d, *J* = 8.7 Hz, 2H), 7.00 (dd, *J* = 8.7, 2.3 Hz, 1H), 7.10 (d, *J* = 16.2 Hz, 1H), 7.33 (d, *J* = 2.3 Hz, 1H), 7.39–7.55 (m, 5H), 7.88 (d, *J* = 8.7 Hz, 1H); ¹³C{¹H} NMR (100 MHz, CDCl₃): δ 40.2 (2C), 67.96, 68.03, 68.5, 68.7, 83.0, 84.7, 107.8, 112.2 (2C), 117.1, 118.4, 122.4, 124.0, 124.5, 128.6 (2C), 134.9, 135.8 (2C), 149.5, 150.7, 157.2, 159.4; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₂H₂₄FN₂O₂, 367.1822; found, 367.1821.

(E)-1-((2-(4-(Dimethylamino)styryl)quinolin-6-yl)oxy)-3-fluoropropan-2-ol (27, SQ16)

By a procedure identical to that described for synthesis of **15**, quinoline **7** (474 mg, 2.13 mmol, 1.00 eq.), *p*-dimethylaminobenzaldehyde (1270 mg, 8.52 mmol), and crude fluorine **13** (1.00 g) were converted into **27** as a red solid (35.0 mg, 3.9% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 3.08 (s, 6H), 4.20 (d, *J* = 4.6 Hz, 2H), 4.26–4.37 (m, 1H), 4.55–4.58 (m, 1H), 4.66–4.70 (m, 1H), 6.79 (d, *J* = 8.7 Hz, 2H), 7.17–7.21 (m, 1H), 7.41 (d, *J* = 16.2 Hz, 1H), 7.50 (d, *J* = 8.7 Hz, 1H), 7.53–7.59 (m, 2H), 7.71 (d, *J* = 8.7 Hz, 1H), 7.83–7.91 (m, 1H), 8.24 (d, *J* = 8.7 Hz, 1H), 8.27–8.34 (m, 1H); ¹³C {¹H} NMR (100 MHz, CDCl₃): δ 40.2 (2C), 68.54, 68.57, 68.77, 69.8, 82.5, 84.2, 107.4, 112.2 (2C), 112.8, 118.2, 122.5, 122.7, 125.7, 127.4, 130.9 (2C), 134.1, 141.2, 144.0, 151.6, 152.5, 157.8; HRMS (ESI-TOF) *m/z*: [M + H]⁺ calcd for C₂₂H₂₄FN₂O₂, 367.1822; found, 367.1819.

(*E*)-2-((2-(4-(Dimethylamino)styryl)quinolin-7-yl)oxy)ethyl methylbenzenesulfonate (28)

The same reaction described above to prepare crude phenol compound (synthesis of 14) was used. To a stirred solution of crude phenol compound (54.6 mg), and NaH (13.5 mg, 0.564 mmol) in dry DMF (1.88 mL) was added 1,2-bis(tosyloxy)ethane (348 mg, 0.939 mmol). After being stirred for 2 h at 100 °C, the mixture was diluted with H₂O. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried

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over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over amino silica gel with *n*-hexane-AcOEt 2:1 to 1:1 to give **28** as an orange solid (58.3 mg, 18% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 2.42 (s, 3H), 3.01 (s, 6H), 4.25–4.30 (m, 2H), 4.43–4.48 (m, 2H), 6.72 (d, *J* = 8.7 Hz, 2H), 6.97 (dd, *J* = 8.7, 1.7 Hz, 1H), 7.15 (d, *J* = 15.7 Hz, 1H), 7.22 (d, *J* = 1.7 Hz, 1H), 7.32 (d, *J* = 8.2 Hz, 2H), 7.47–7.63 (m, 5H), 7.83 (d, *J* = 8.2 Hz, 2H), 7.96 (d, J = 8.7 Hz, 1H); LC-MS *m/z*: [M + H]⁺ calcd for C₂₈H₂₉N₂O₄S, 489.2; found, 489.2.

(*E*)-2-((2-(4-(Dimethylamino)styryl)quinolin-6-yl)oxy)ethyl methylbenzenesulfonate (29)

The same reaction described above to prepare crude phenol compound (synthesis of **15**) was used. To a stirred solution of crude phenol compound (131 mg), and Cs₂CO₃ (1000 mg, 3.07 mmol) in dry DMF (9.00 mL) was added 1,2-bis(tosyloxy)ethane (1420 mg, 3.84 mmol). After being stirred for 2.5 h at 90 °C, the mixture was diluted with H₂O. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration in vacuo, the residue was purified by flash chromatography over amino silica gel with *n*-hexane-AcOEt 2:1 to 1:1 to give **29** as an orange solid (25.0 mg, 5% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 2.94 (s, 3H), 3.01 (s, 6H), 4.18–4.21 (m, 2H), 4.36–4.39 (m, 2H), 6.66 (d, *J* = 8.6 Hz, 2H), 6.86 (d, *J* = 2.9 Hz, 1H), 7.10 (d, *J* = 2.9 Hz, 1H), 7.12 (d, *J* = 2.9 Hz, 1H), 7.26 (d, *J* = 8.6 Hz, 2H), 7.43–7.56 (m, 4H), 7.76 (d, *J* = 8.6 Hz, 2H), 7.86 (d, *J* = 8.6 Hz, 2H); LC-MS *m/z*: [M + H]⁺ calcd for C₂₈H₂₉N₂O₄S, 489.2; found, 489.2.

The same reaction described above to prepare crude phenol compound (synthesis of **15**) was used. To a stirred solution of crude phenol compound (157 mg) and Cs₂CO₃ (1000 mg, 3.07 mmol) in dry DMF (4.00 mL) was added triethylene glycol bis(*p*-toluenesulfonate) (962 mg, 2.10 mmol). After being stirred for 3 h at 95 °C, the mixture was diluted with H₂O. The whole solution was extracted with AcOEt. The extract was washed with brine, and dried over Na₂SO₄. After concentration *in vacuo*, the residue was purified by flash chromatography over amino silica gel with *n*-hexane-AcOEt 2:1 to 1:1 to give **28** as an orange solid (7.6 mg, 1.3% yield, 3 steps): ¹H NMR (400 MHz, CDCl₃): δ 2.48 (s, 3H), 3.15 (s, 6H), 3.68–3.78 (m, 6H), 3.95–4.01 (m, 2H), 4.18–4.23 (m, 2H) 4.30–4.35 (m, 2H), 6.91 (d, *J* = 8.7 Hz, 2H), 7.38 (d, *J* = 8.2 Hz, 2H), 7.50 (d, *J* = 16.2 Hz, 1H), 7.60 (dd, *J* = 2.9, 8.7 Hz, 1H), 7.64 (d, *J* = 8.7 Hz, 2H), 7.74–7.86 (m, 4H), 7.94

(d, J = 8.7 Hz, 1H), 8.26 (d, J = 8.7 Hz, 1H), 8.42 (d, J = 8.7 Hz, 1H); LC-MS m/z: [M + H]⁺ calcd for C₃₂H₃₇N₂O₆S, 577.2; found, 577.2.

¹²⁵I-labeling reaction

Total of 100 μ L of 3% H₂O₂ aq. and 100 μ L of 1 N HCl aq. were added to a mixture of tributyltin precursor **2** (150 μ g/150 μ L EtOH) and [¹²⁵I]NaI (1.85-7.40 MBq, specific activity: 81.4 TBq/mmol) in 150 μ L EtOH in a sealed vial. The reaction was allowed to proceed at 25 °C for 10 min and terminated by the addition of saturated NaHSO₃ aq. (200 μ L). After neutralization with saturated NaHCO₃ aq. (200 μ L) and extraction with ethyl acetate (600 μ L ×3), the organic phase was dried by passing through a column filled with anhydrous Na₂SO₄. The solution was gas-dried with a stream of nitrogen gas. The crude ligands were purified by HPLC on a Cosmosil C₁₈ column (Nacalai Tesque) with an isocratic solvent of acetonitrile/H₂O = 85/15 at a flow rate of 1.0 mL/min.

Binding saturation assays using the aggregated α-syn and Aβ in solution

α-Syn and Aβ were purchased from rPeptide (Bogart, GA, USA) and the Peptides Institute, Inc. (Osaka, Japan), respectively. α-Syn aggregates were prepared by incubating recombinant α-syn monomer (1.67 mg/mL in 20 mM Tris–HCl, 100 mM NaCl, pH 7.48) at 37 °C for 144 h with shaking at 1000 rpm. Aggregation of Aβ was carried out by gently dissolving the peptide (0.25 mg/mL) in PBS solution (pH 7.4). The solution was incubated at 37 °C for 42 h with gentle and constant shaking. The reaction mixture contained increasing concentrations of [¹²⁵I]ISQ (15.6 nM–20 μM, 100 mL, EtOH), a fixed concentration of α-syn aggregates (33.3 μg/mL, 50 μL, 20 mM Tris–HCl, 100 mM NaCl) or Aβ aggregates (5.00 μg/mL, 50 μL, PBS), and 10% EtOH (850 μL). Nonspecific binding was determined in the presence of 1 μM nonradioactive ISQ. After incubating for 3 h at 25 °C, the solution was filtered through GF/B filters using an M-24 cell harvester. The radioactivity of the bound [¹²⁵I]ISQ was measured with a γ counter (Wizard 2480, PerkinElmer). The dissociation constants (K_d) and Bmax were determined by Scatchard analysis using GraphPad Prism 6 (GraphPad Software, San Diego, CA, USA).

Binding inhibition assays using the aggregated α -syn and A β in solution

Aggregated α -syn and A β were prepared by the same method as the saturation assay. The reaction mixture contained increasing concentrations of nonradioactive SQ derivative (1.28 nM-100 μ M, 50 μ L, EtOH or DMSO), a fixed concentration of [¹²⁵I]ISQ (6.25 pM-12.5 pM, 50 μ L, EtOH), α -syn aggregates (33.3 μ g/mL, 50 μ L, 20 mM Tris–HCl, 100 mM NaCl, pH 7.48) or A β_{1-42} aggregates (5.00 μ g/mL, 50 μ L, PBS, pH 7.4),

and 10% EtOH (850 µL). After incubating for 3 h at 25 °C, the mixture was then filtered through Whatman GF/B filters using a Brandel M-24 cell harvester, and the radioactivity of the filters containing the bound [¹²⁵I]ISQ were measured in a γ counter. Values for the half maximal inhibitory concentration (IC50) were determined from displacement curves using GraphPad Prism 5.0, and those for the inhibition constant (K_i) were calculated using the Cheng-Prusoff equation: $K_i = IC50/(1 + [L]/K_d)$, where [L] is the concentration of [¹²⁵I]ISQ used in the assay and K_d is the dissociation constant of ISQ (α -syn: 25.1 nM, A β : 8.53 nM).

Calculation of CNS MPO score

CNS MPO score was calculated using the drug discovery software, StarDrop (Optibrium Ltd., Cambridge, UK).

¹⁸F-Labeling reaction

Kryptofix 222 (10.0 mg) was added to the reaction vessel, and the mixture was evaporated azeotropically with dry acetonitrile (300 μ L) under a nitrogen gas stream at 120 °C, three times. A solution of Kryptofix 222 (3.00 mg) and tosyl precursor (**28**, **29**, and **30**) (1.00 mg) in anhydrous DMF (200 μ L) was added to the reaction vessel. After being heated at 120 °C for 20 min, the mixture was diluted with H₂O (1.00 mL). The whole solution was extracted with AcOEt (600 μ L ×3). The organic phase was dried by passing through a column filled with anhydrous Na₂SO₄. The solution was gas-dried with a stream of nitrogen gas. The crude ¹⁸F-labeled probes were purified by HPLC on a Cosmosil C₁₈ column (Nacalai Tesque) with a gradient solvent of acetonitrile/H₂O (with 0.1% TFA) 25:75 (0 min) to 50:50 (30 min) at a flow rate of 1.0 mL/min.

In vivo biodistribution study using normal mice

A saline solution (100 μ L) of [¹⁸F]**SQ3**, **4**, and **14** (10.0, 8.78, and 5.00 kBq) containing EtOH (10.0 μ L) and Tween 80 (0.100 μ L) was injected intravenously directly into the tails of ddY mice (5 weeks old, male, n = 5). The mice were sacrificed at various timepoints post injection. The brains were removed and weighed, and the radioactivity was measured with an automatic γ counter (Wallac WIZARD 1470, PerkinElmer Inc., Massachusetts, USA). The animal experiment was approved by the Animal Experimentation Committee of Graduate School of Pharmaceutical Sciences, Kyoto University.

2. ¹H and ¹³C NMR Spectrum

¹H NMR Spectrum for Compound **3** (ISQ).



¹³C NMR Spectrum for Compound **3** (ISQ)





¹H NMR Spectrum for Compound 4 (SQ1)

¹³C NMR Spectrum for Compound 4 (SQ1)





¹H NMR Spectrum for Compound **5** (SQ2)

¹³C NMR Spectrum for Compound **5** (SQ2)





¹H NMR Spectrum for Compound **14** (SQ3)

¹³C NMR Spectrum for Compound **14** (SQ3)





¹H NMR Spectrum for Compound **15** (SQ4)

¹³C NMR Spectrum for Compound **15** (SQ4)





¹H NMR Spectrum for Compound **16** (SQ5)

¹³C NMR Spectrum for Compound **16** (SQ5)





¹H NMR Spectrum for Compound **17** (SQ6)

¹³C NMR Spectrum for Compound **17** (SQ6)





¹H NMR Spectrum for Compound **18** (SQ7)

¹³C NMR Spectrum for Compound **18** (SQ7)





¹H NMR Spectrum for Compound **19** (SQ8)

¹³C NMR Spectrum for Compound **19** (SQ8)





¹H NMR Spectrum for Compound **20** (SQ9)

¹³C NMR Spectrum for Compound **20** (SQ9)





¹H NMR Spectrum for Compound **21** (SQ10)

¹³C NMR Spectrum for Compound **21** (SQ10)





¹H NMR Spectrum for Compound **22** (SQ11)

¹³C NMR Spectrum for Compound **22** (SQ11)





¹H NMR Spectrum for Compound **23** (SQ12)

¹³C NMR Spectrum for Compound **23** (SQ12)





¹H NMR Spectrum for Compound **24** (SQ13) solvent

¹³C NMR Spectrum for Compound **24** (SQ13)





¹H NMR Spectrum for Compound **25** (SQ14)

¹³C NMR Spectrum for Compound **25** (SQ14)





¹H NMR Spectrum for Compound **26** (SQ15)

¹³C NMR Spectrum for Compound **26** (SQ15)





¹H NMR Spectrum for Compound **27** (SQ16)

¹³C NMR Spectrum for Compound **27** (SQ16)



3. References

(1) Preparation of fused pyrazines as phosphoinositide 3-kinase (PI3K) inhibitors. **2010**, WO2010052448 A2010052442.

(2) Chandrashekarappa, K. K. H.; Mahadevan, K. M.; Manjappa, K. B. High throughput one pot synthesis of 2-methylquinolines. *Tetrahedron Lett* **2013**, *54* (11), 1368-1370. DOI: 10.1016/j.tetlet.2012.12.094.

(3) Eros, G.; Nagy, K.; Mehdi, H.; Papai, I.; Nagy, P.; Kiraly, P.; Tarkanyi, G.; Soos, T. Catalytic hydrogenation with frustrated Lewis pairs: selectivity achieved by size-exclusion design of Lewis acids. *Chemistry* **2012**, *18* (2), 574-585. DOI: 10.1002/chem.201102438.

(4) Zhou, K.; Bai, H.; Feng, L.; Dai, J.; Cui, M. Smart D-π-A Type Near-Infrared Abeta Probes: Effects of a Marked pi Bridge on Optical and Biological Properties. *Anal Chem* **2017**, *89* (17), 9432-9437. DOI: 10.1021/acs.analchem.7b02246.

(5) Dibenzothiazepinone derivatives as hepatitis B core protein allosteric modulators and their preparation. **2015**, WO2015138895 A2015138891.

(6) Moldovan, R. P.; Teodoro, R.; Gao, Y.; Deuther-Conrad, W.; Kranz, M.; Wang, Y.; Kuwabara, H.; Nakano, M.; Valentine, H.; Fischer, S.; et al. Development of a High-Affinity PET Radioligand for Imaging Cannabinoid Subtype 2 Receptor. *J Med Chem* **2016**, *59* (17), 7840-7855. DOI: 10.1021/acs.jmedchem.6b00554.

(7) Chaabouni, M. M.; Baklouti, A. Ring-Cleavage Reactions of F-Alkyl and Cl-Alkyl Epoxides by Action of Amines Hydrofluorides. *B Soc Chim Fr* **1989**, (4), 549-553.