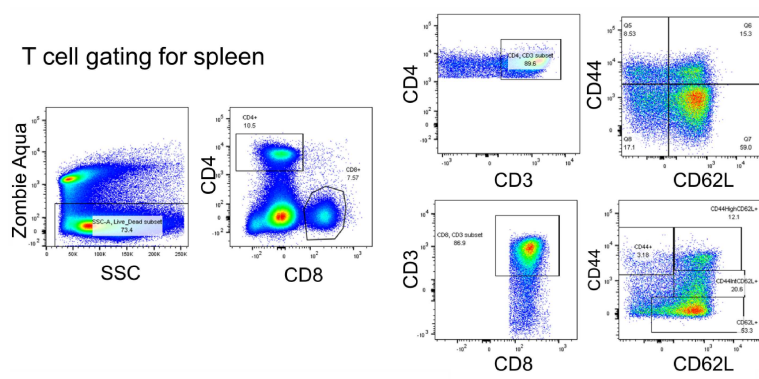
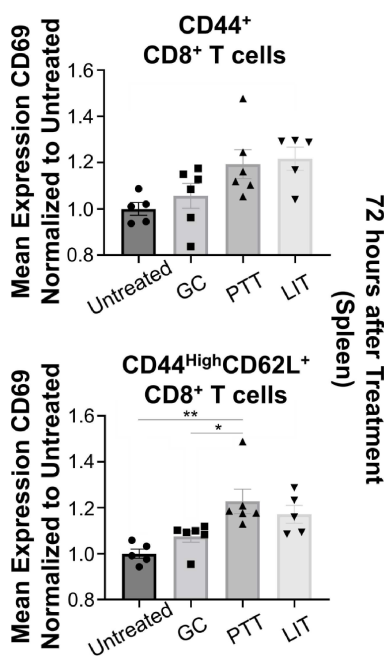


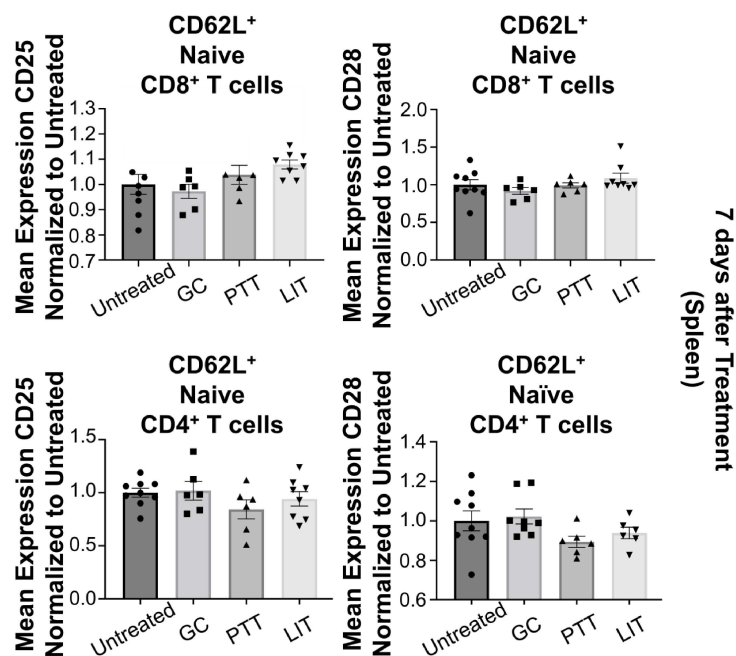
A



B



C



D

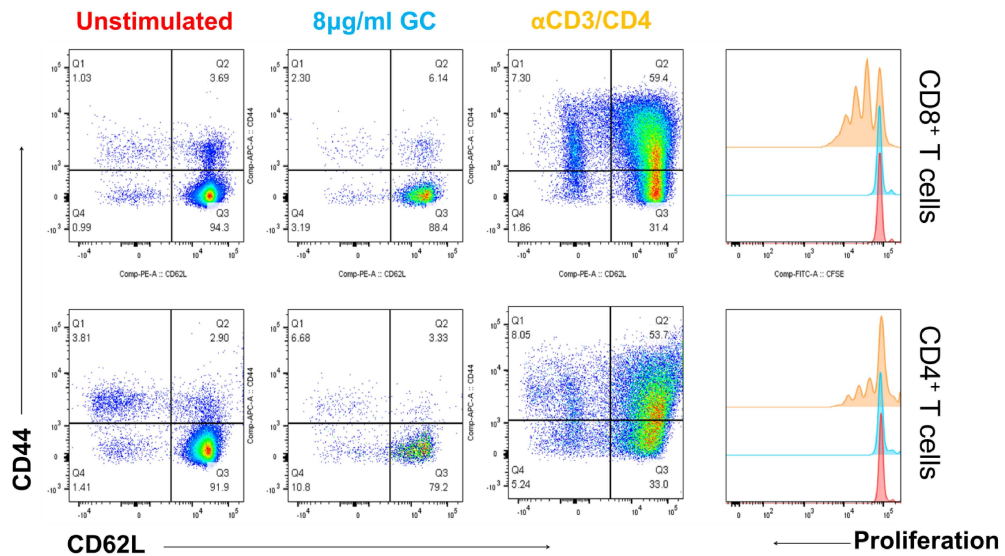


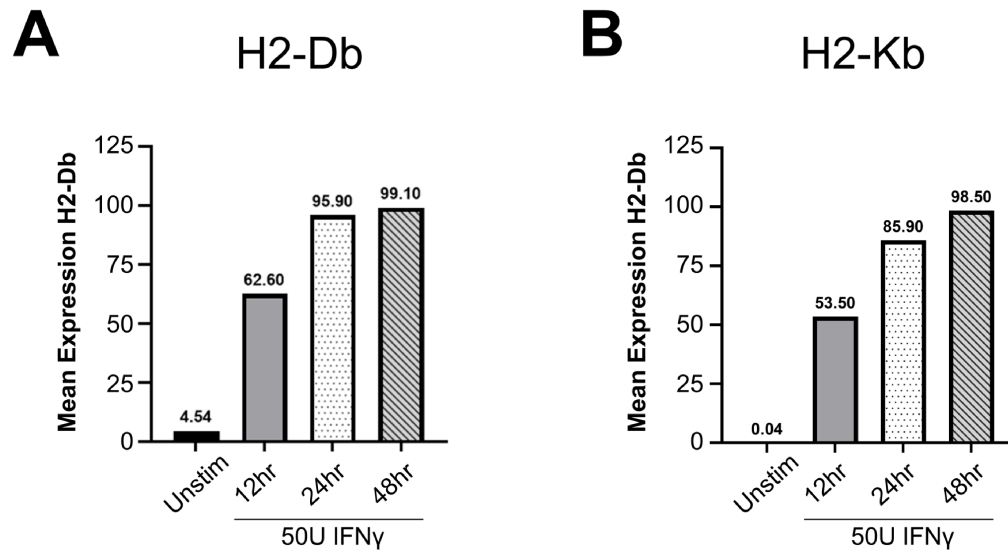
Figure S2

Figure S3

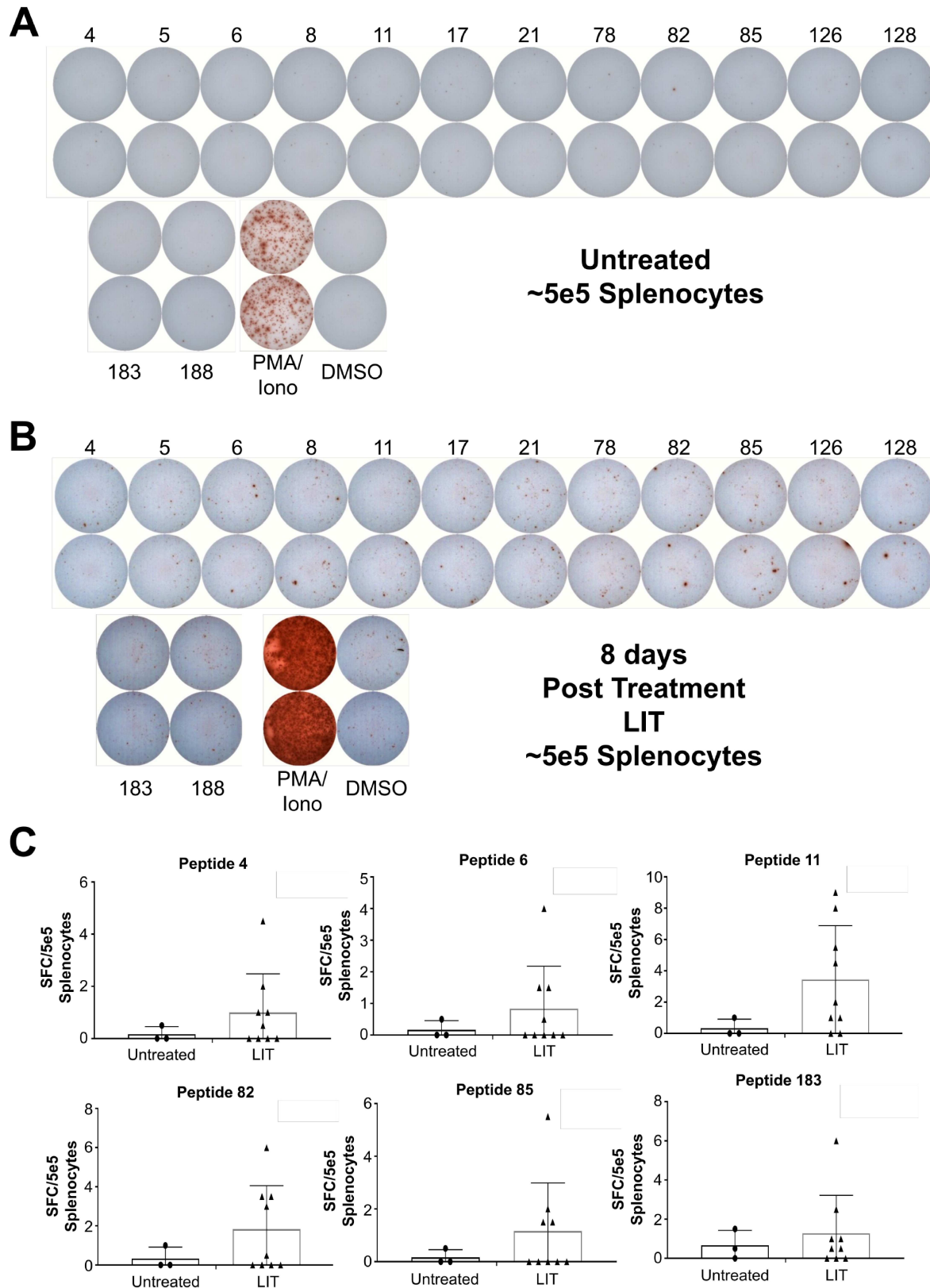
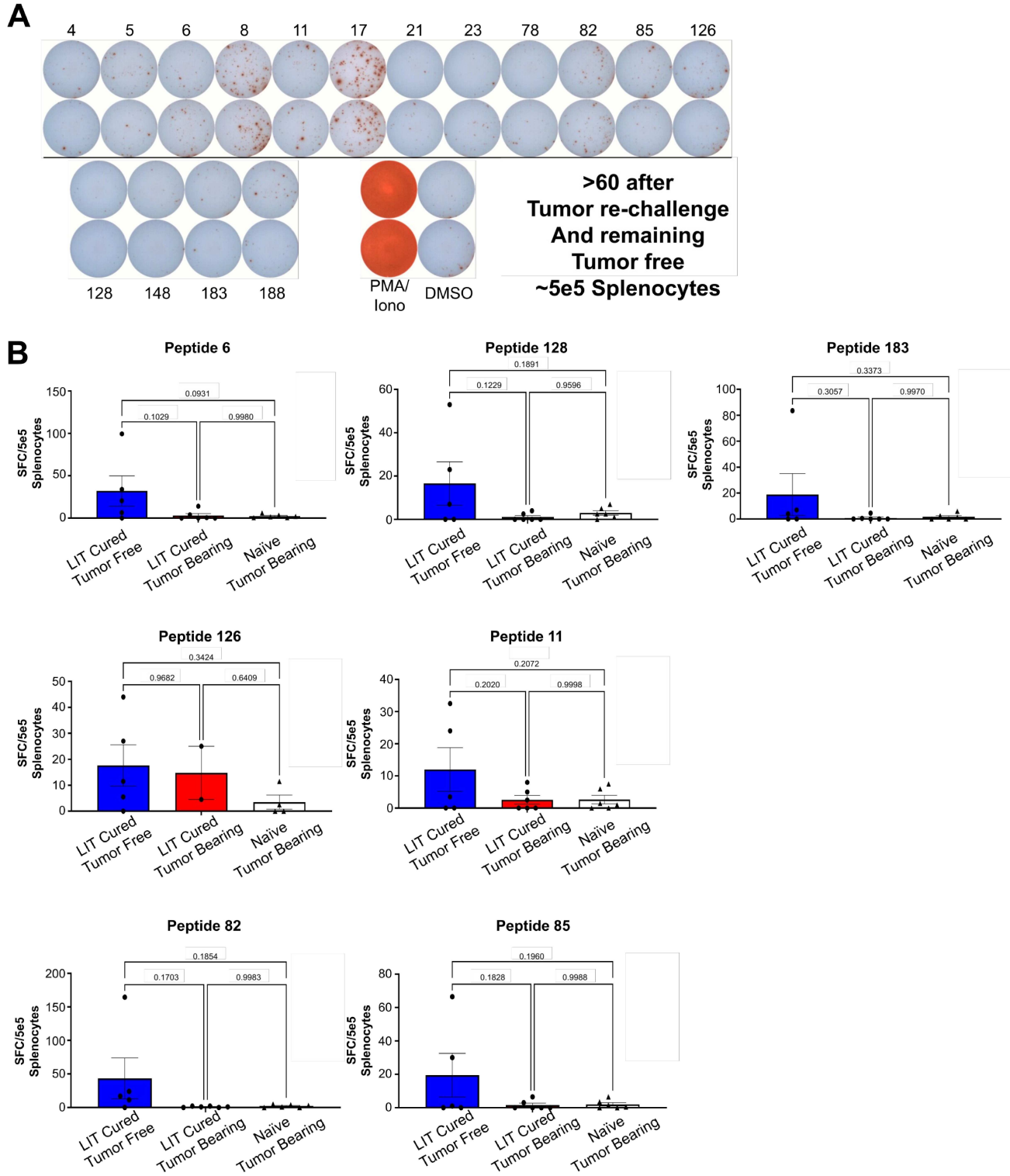


Figure S4



Supplemental Figure 1

LIT activation of T cells within the spleen. (A) Flow cytometry gating used for analysis of the CD4⁺ and CD8⁺ T cells. (B) Flow cytometry analysis of the normalized mean expression of CD69 on the surface of CD8⁺ effector and memory T cells from the spleens of animals in different groups 72 hours after treatment. (C) Flow cytometry analysis of the normalized mean expression of CD25 and CD25 on the surface of naïve CD4 and CD8 naïve T cells from the animal spleens seven days after treatment. One-way ANOVA was used for statistical analysis of the mean expression data. P values = *0.5, **0.05, **0.005, ****0.0005. (D) Flow cytometry analysis of CD44, CD62L, and CFSE expression of either purified CD4⁺ or CD8⁺ T cells from the spleens. T cells were incubated with either GC or anti-CD3/anti-CD28 for 72 hours prior to flow analysis.

Supplemental Figure 2

In vitro characterization of B16-F10 H2-Db and H2-Kb expression following 50 U/mL of IFN γ stimulation (A-B). Analysis of the expression of (A) H2Db and (B) H2Kb MHC-I molecules on B16-F10 cells grown in vitro. The mean expression of H2-Db or H2-Kb was examined after 12, 24, and 48 hours via flow cytometry. Expression of H2-Db or H2-Kb is represented as a bar graph.

Supplemental Figure 3

ELISpot screening of tumor specific peptides in untreated and LIT B16-F10 tumor-bearing animals. (A-C) C57BL/6 mice of 6-8-week-old were injected with 1e5 B16-F10 tumor cells. Once the tumors reached 0.5cm³, the tumors were treated with LIT or left untreated. Eight days after treatment splenocytes were isolated and used for ELISPOT. Approximately 5e5 total splenocytes were used for the ELISPOTs with ~1 μ M peptide. ELISPOTs were incubated for ~36 hours prior to developing the ELISPOT. Spots were counted using the default settings on the ELISPOT reader. Assays were performed in duplicate. The average number of spots in the negative control was then subtracted from the average number of spots in the peptide incubated wells to get the final spot count. (A) ELISPOT using splenocytes from untreated tumor-bearing animals. (B) ELISPOT using splenocytes from LIT treated animals 8 days after treatment. (C) Spot-forming cells (SFC) per well were calculated using the following formula for six selected peptides, Average spots peptide wells – Average spots DMSO wells = final spot count per well and graphed.

Supplemental Figure 4

Tumor specific peptide T cell reactivity in LIT-cured animals that resisted tumor rechallenge. (A-B) C57BL/6 mice of 6–8-week-old were injected with 1e5 B16-F10 tumor cells. Once the tumors reached 0.5cm³, the tumors were treated with LIT or left untreated. LIT-cured survivors were then rechallenged on the opposite flank after being tumor free for >60days. Once the animals were tumor free for >60days following tumor rechallenge, splenocytes were isolated and incubated with B16-F10 specific peptides for ~36 hours before being developed. Approximately 5e5 total splenocytes from LIT-cured tumor rechallenged non-tumor-bearing animals were used to ELISpots with ~1 μ M peptide. ELISpots were incubated for ~36 hours prior to developing the ELISpot. (A) ELISPOT using splenocytes from LIT-cured animals that resisted tumor rechallenge. (B) Spot-forming cells (SFC) were counted using the default settings on the ELISPOT reader for seven selected peptides. Assays were performed in duplicate. The average number of spots in the negative control was then subtracted from the average number of spots in the peptide incubated wells to get the final spot count. The number was graphed, and each dot represents an individual animal. One-Way ANOVA was used for statistical analysis. P values = *0.5, **0.05, **0.005, ****0.0005.