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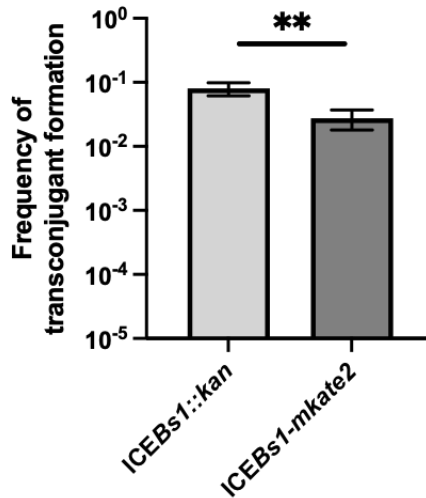


Figure S1. ICEBs1::mkate2 transfer at a slightly lower efficiency than ICEBs1::kan. Donor cells carrying ICEBs1::kan or ICEBs1-mkate2 were mated on MSgg medium at 30 °C for 20 h. The statistical analysis shows that ICEBs1-mkate2 transfer is slightly less efficient than the WT ICEBs1::kan but remains high (t-test; ** $P < 0.01$). Error bars represent the standard deviation (SD). The result is representative of at least three

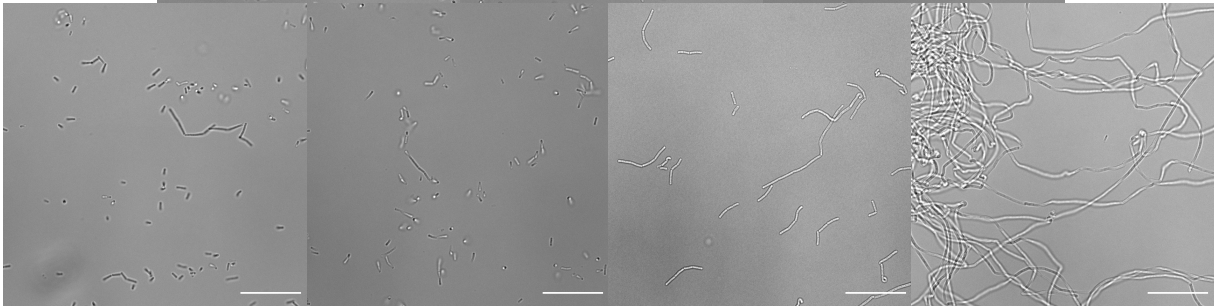
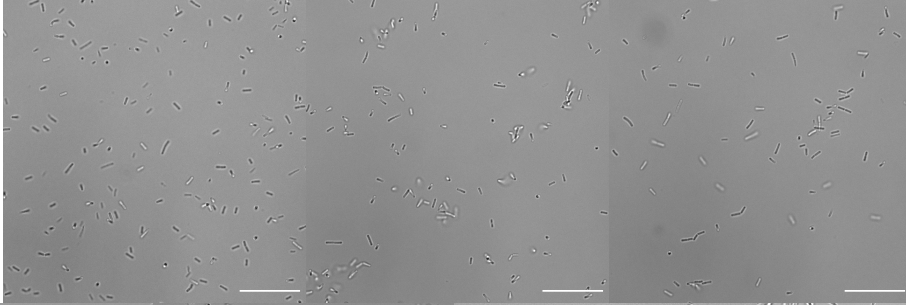
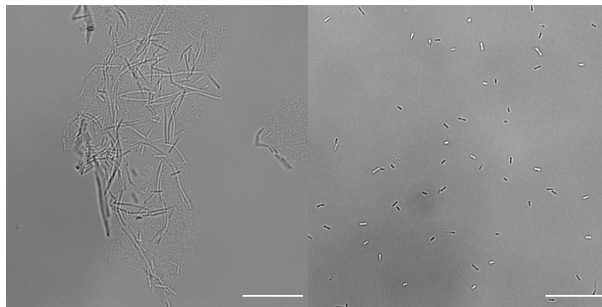
A**MSNc****WT*****lytC******lytD******lytE******lytF******lytD lytE lytF******lytABC lytD lytE lytF*****B****MSgg****WT*****P_{hyperspank}-lytABC***

Figure S2. Lytic enzyme mutant phenotype. (A) Lytic mutants were grown in liquid MSNc medium and incubated at 37 °C for 16 h with agitation prior to transferring on an agarose pad and imaged using epifluorescence microscopy. (B) Overexpression of *lytABC* was grown in liquid MSgg medium containing 50 μM IPTG and incubated at 30 °C for 16 h without agitation prior to transferring on an agarose pad and imaged using epifluorescence microscopy. The scales bars represent a size of 25 μm. Images are representative of the phenotype observed in at least 3 independent replicates°

Video S1. Multiple transconjugant bacteria are not adjacent to donor cells. Donor bacteria bearing *ICEBs1-mKate2* and expressing a CFP fluorescent protein were mated with recipients expressing a GFP fluorescent protein. Pictures were taken in a Z-stack of 10 μM ticks and overlaid using ImageJ 3D Project. The movie shows only donor (blue) and *ICEBs1* (magenta) to have an easier view of the transconjugants. Donor bacteria appear blue to purple and transconjugants appear bright red. The scale bar represents a size of 10 μm . The video shown is representative of conjugative clusters observed.

Table S1. Strains

Strain	Genotype	Construction
168	Domesticated strain	Lab stock
NCIB 3610	Wild type Undomesticated strain	Lab stock
JSB86	JH642 ICEBs1ΔconG:: <i>kan trpC2 pheA1</i>	This study
JSB93	ICEBs1ΔconG:: <i>kan</i>	This study
FL62 (1)	ICEBs1- <i>kan ycbU-lmrB::spec</i>	Lab stock
MG25 (1)	ICEBs1 ⁰ <i>ylnF/yboA::Tn917 ::amyE::cat</i>	Lab stock
DL875 (2)	<i>amyE::Pskf-yfp lacA::PtapA-cfp (erm)</i>	Lab stock
MG13 (1)	ICEBs1 ⁰	Lab stock
JSB103	168 <i>amyE::P_{hypersnak}-conG (spec)</i>	This study
JSB105	ICEBs1 ⁰ <i>amyE::P_{hypersnak}-conG (spec) lacA::P_{tapA}-cfp (erm)</i>	DL875 (2) and JSB103 in MG13 (1)
JSB106	ICEBs1ΔconG:: <i>kan amyE ::P_{hyperspank}-conG (spec)</i>	JSB103 in JSB93
CAL51 (3)	JH642 opp::(Tn917lac::pTV21?2 cat) Δ(<i>rapI phrI</i>)342:: <i>kan amyE::[(Pspank(hy)-rapI) spc]</i>	Lab stock
MG28 (1)	ICEBs1- <i>kan</i>	Lab stock
JSB107	ICEBs1- <i>kan amyE::P_{spank(hy)}-rapI (spec)</i>	CAL51 (3) in MG28 (1)
VCL28 (1)	<i>ycbU-lmrB::spec</i>	Lab stock
JSB97	ICEBs1ΔconG:: <i>kan ycbU-lmrB::spec</i>	VCL28 (1) in JSB93
JSB61	168 ICEBs1- <i>mkate2 (spec)</i>	This study
JSB62	ICEBs1- <i>mkate2 (spec)</i>	JSB61 in NCIB 3610
JSB68	168 <i>amyE::P_{hyperspank}-cfp (spec)</i>	This study
JSB69	ICEBs1 ⁰ <i>amyE::P_{hyperspank}-cfp (spec)</i>	JSB68 in MG13
JSB71	ICEBs1- <i>mkate2 amyE::P_{hyperspank}-cfp (spec)</i>	JSB62 in JSB69
PB69 ^a	<i>amyE ::Physpank-gfp (cat)</i>	Lab stock
JSB79	ICEBs1 ⁰ <i>amyE::P_{hyspank}-gfp (cat)</i>	PB69 ^a in MG13
PB232 ^a	<i>kinD::tet 317amE::kinD WT (spc) lacA::PtapA-yfp (erm) ywrK::Tn917::amyE::cat</i>	Lab stock
JSB66	ICEBs1- <i>mkate2 lacA::P_{tapA}-yfp (erm)</i>	PB232 ^a in JSB62
JSB70	ICEBs1 ⁰ <i>lacA ::P_{tapA}-yfp amyE::P_{hyperspank}-cfp (spec)</i>	JSB68 and PB232 in MG13 (1)
BJM396 ^b	PY79 <i>lytC::cat</i>	Lab stock
GJ4	ICEBs1 ⁰ <i>lytC::cat</i>	BJM396 ^b in MG13 (1)
BJM402 ^b	PY79 <i>lytD::cat</i>	Lab stock
GJ5	ICEBs1 ⁰ <i>lytD::cat</i>	BJM402 ^a in MG13 (1)

BJM76 ^b	PY79 <i>lytE::cat</i>	Lab stock
GJ6	ICEBs1 ⁰ <i>lytE::cat</i>	BJM76 ^b in MG13 (1)
BJM104 ^b	PY79 <i>lytF::cat</i>	Lab stock
GJ7	ICEBs1 ⁰ <i>lytF::cat</i>	BJM104 ^b in MG13 (1)
L16648	168 <i>lytABC::neo lytD::tet lytE::cat lytF::spc</i>	BGSC
BKE09420	<i>trpC2 ΔlytE::erm</i>	BGSC
GJ32	ICEBs1 ⁰ <i>lytD::tet lytE::erm lytF::cat</i>	L16648 and BJM104 in MG13 (1)
GJ42	ICEBs1 ⁰ <i>lytABC::neo lytD::tet lytE::erm lytF::cat</i>	L16648 and BJM104 in MG13 (1)
JSB100	168 <i>amyE::P_{hyperspank}-lytABC (spec)</i>	This study
JSB101	ICEBs1 ⁰ <i>amyE::P_{hyperspank}-lytABC (spec)</i>	JSB100 in MG13 (1)
FL94 (1)	ICEBs1 ⁰ <i>ylnF/yboA::Tn917::amyE::cat sinR::spec</i>	Lab stock
PB514 ^a	PY79 <i>ylnF/yloA::Tn917::amyE::cat</i>	Lab stock
JSB19	ICEBs1- <i>kan ylnF/ybaO::tn917::amyE::cat</i>	PB514 ^a in MG28 (1)
SSB569 ^a	<i>tasA::erm epsA-O::tet</i>	Lab stock
JSB125	ICEBs1 ⁰ <i>epsA-O::tet</i>	SSB569 ^a in MG13
PB194 ^a	<i>sinR::spc epsA-O::tet</i>	Lab stock
JSB127	ICEBs1 ⁰ <i>epsA-O::tet sinR::spec</i>	PB194 ^a in JSB125
JSB130	168 <i>tapA-sinR::erm</i>	This study
JSB132	ICEBs1 ⁰ <i>tapA-sinR::erm</i>	JSB130 in MG13
JSB133	ICEBs1 ⁰ <i>tapA-sinR::erm epsA-O::tet</i>	JSB130 in JSB125

*Unless indicated, all strains are derivatives of *B. subtilis* NCIB 3610. Antibiotic: spectinomycin (spec), chloramphenicol (cat), erythromycin/lincomycin (erm), kanamycin (kan), tetracycline (tet), neomycin (neo)

^aThese strain come from the Kolter and/or Beaugard lab stock.

^bThese strains were gift of the Rudner's Lab

1. Lécuyer F, Bourassa J-S, Gélinas M, Charron-Lamoureux V, Burrus V, Beaugard PB. 2018. Biofilm Formation Drives Transfer of the Conjugative Element ICEBs1 in *Bacillus subtilis*. *mSphere* 3:e00473-18.
2. López D, Vlamakis H, Losick R, Kolter R. 2009. Cannibalism enhances biofilm development in *Bacillus subtilis*. *Mol Microbiol* 74:609–618.

3. Auchtung JM, Lee CA, Monson RE, Lehman AP, Grossman AD. 2005. Regulation of a *Bacillus subtilis* mobile genetic element by intercellular signaling and the global DNA damage response. *Proc Natl Acad Sci U S A* 102:12554–12559.

Table S2. Plasmids

Strain	Plasmid name	Genotype	Antibiotics resistance
GJ55	pGJ1	pDR111- <i>cfp</i>	ampicillin and spectinomycin
JSB59	pJSB12	pDR111- <i>yddM-mkate2-attR</i>	ampicillin and spectinomycin
JSB83	pJSB18	pDR111- <i>lytABC</i>	ampicillin and spectinomycin
JSB84	pJSB19	pminiMAD2- Δ <i>conG</i>	erythromycin and lincomycin
JSB102	pJSB22	pDR111- <i>conG</i>	ampicillin and spectinomycin

Table S3. Primers

Primer	Sequence (5'→3')
P27	GATCC TTTAA CTCTG GCAAC CCTC
P28	GCCGA CTGCG CAAAA GACAT AATCG
P342	TGT <u>GG ATCCG</u> AATCT CAGAT TGTTA ATCCT GC
P343	TTAGG <u>ATCCA</u> ATCCA GTACA TTACG ATCTC
P344	AATAG <u>GCGAT CGCGG</u> CGAAC TATGA GTTTG CTG
P496	CACTG <u>AGCTC</u> GAGCA ACTTT AGAAA TCGAG TC
P507	GCATG <u>AATTC</u> GACTC TCTAG CTTGA GGCAT C
P508	CATCG <u>CATGC</u> TCATC TGTGC CCCAG TTTG
P719	AGCTG <u>TCGAC</u> TAAAG GAGGA AGATT AGGAG GTGGT TGTA T
P673	GCTAG <u>CATGC</u> GCCGA GAATA CCAAA GGAAG AAC
P615	GCCTC CAAGT CTCTA TGTT C AGC
P616	CGAAG <u>CATGC</u> CCAAC TGGTA ATGGT AGCGA C
P647	GTAGG <u>TCGAC</u> ACATA AGGAG GAACT ACTCA TCCTT TGCAC CTCGT CTG
P648	AGATG <u>CATGC</u> GACCT ATGCC AGTCA GTTTA GC
P668	CATCG <u>GATCC</u> AACCA AACGA GCGTG AAGAA G
P669	GTTCT TTCAT CCCTT CTCAG TCGAC AACCA CCTCC TAATC TAACC
P670	GGTTA GATTA GGAGG TGGTT GTCGA CTGAG AAGGG ATGAA AGAAC
P674	GTCCG <u>AATTC</u> ACATG GTCAA CATAA CTTGG GTC
P746	CAAAA TCCGC TATGC AGGAG
P747	GAGGG TTGCC AGAGT TAAAG GATCG GCCAA TCAAT GTCAT CACC
P748	CGATT ATGTC TTTTG CGCAG TCGGC CTGAA AGCTG AAAGA TAAGC AGAAC
P749	AACCG CAAAT AACG AATAA GGTCC

*Underlined sequence represents restriction site.