

# **Enhancing Gasdermin-Induced Tumor Pyroptosis Through Preventing ESCRT-Dependent Cell Membrane Repair Augments Anti-Tumor Immune Response**

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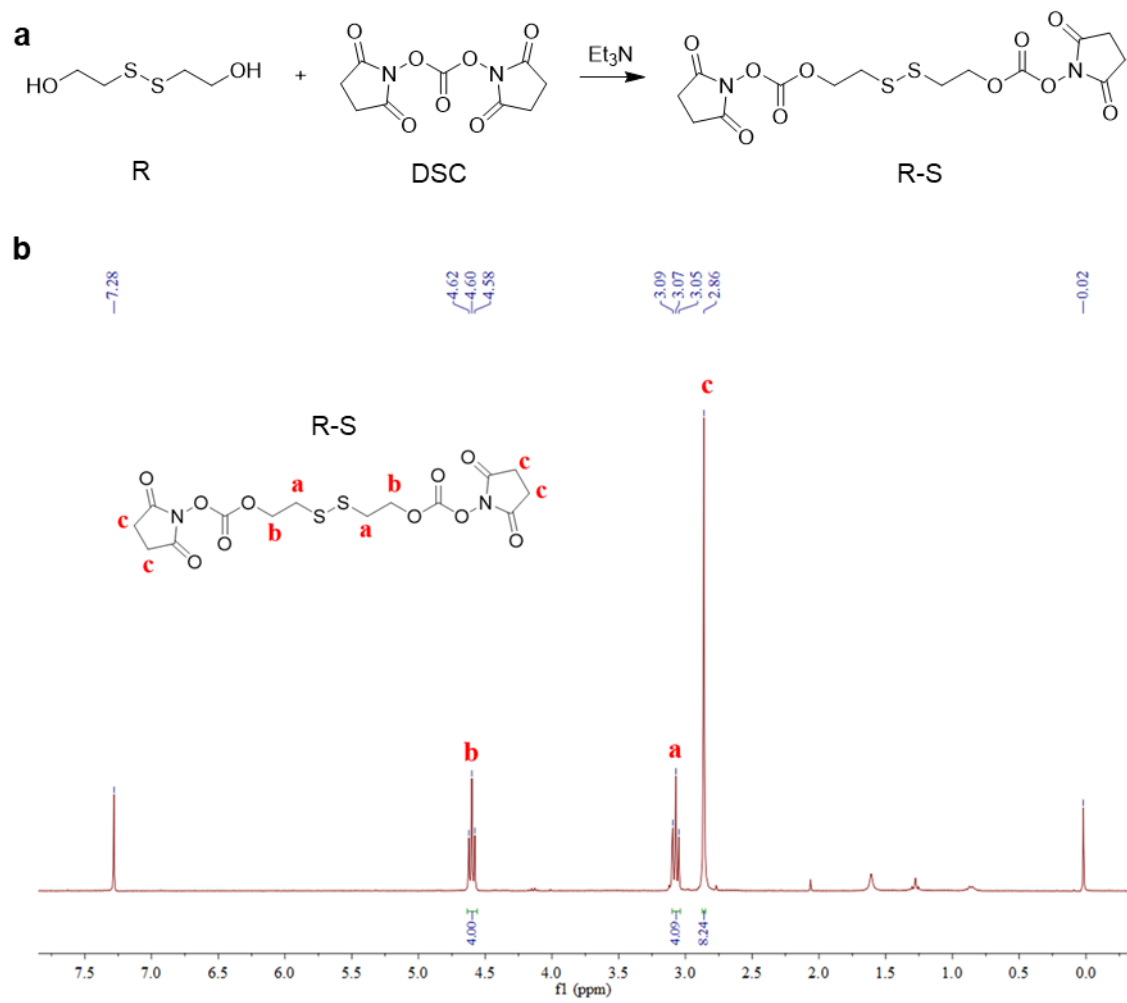
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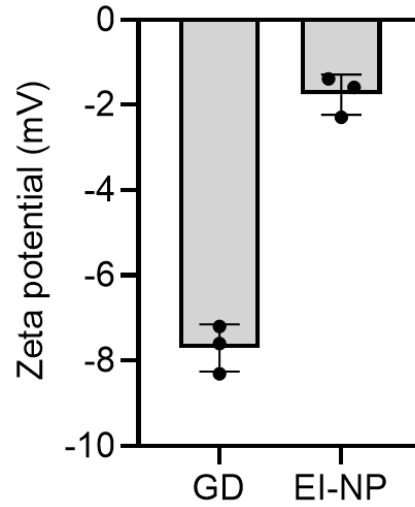
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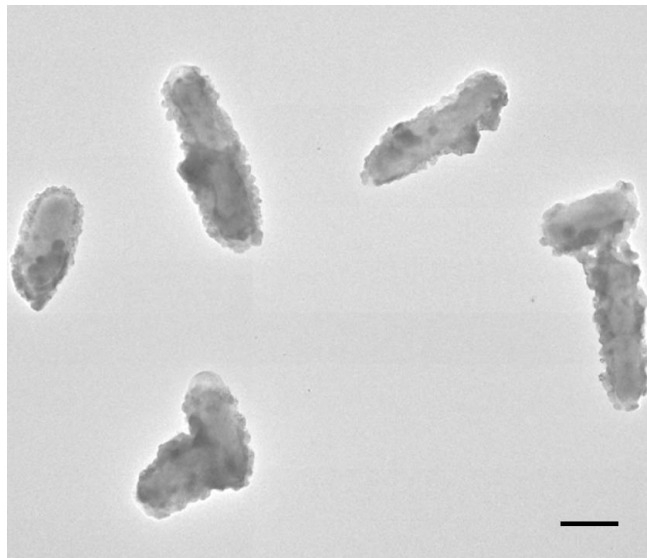
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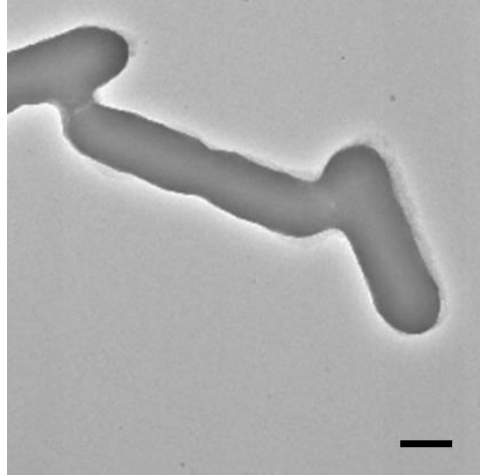
**Supplementary Fig. 1 | Synthesis route and <sup>1</sup>H NMR characterization of the GSH responsive linker. a,** Synthesis route of the GSH responsive linker (R-S). **b,** <sup>1</sup>H NMR characterization of the GSH responsive linker (R-S).



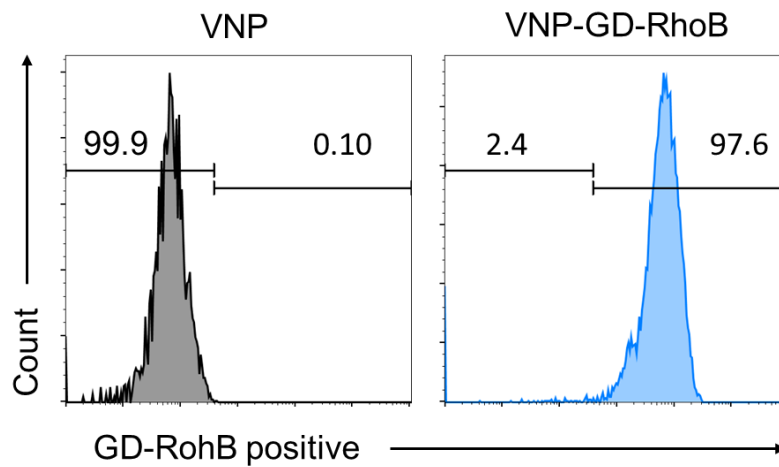
**Supplementary Fig. 2** | Zeta potential of the Gasdermin protein cage GD and ESCRT inhibitor loaded nanoparticle EI-NP. Data are shown as mean  $\pm$  s.d. (n = 3 biologically independent samples).



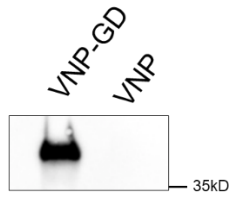
**Supplementary Fig.3** | Representative TEM image of VNP-GD. Scale bar = 1  $\mu$ m. The experiments were repeated three times independently.



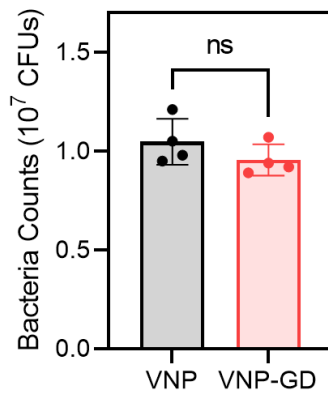
**Supplementary Fig. 4** | Representative TEM image of VNP. Scale bar = 500 nm. The experiments were repeated three times independently.



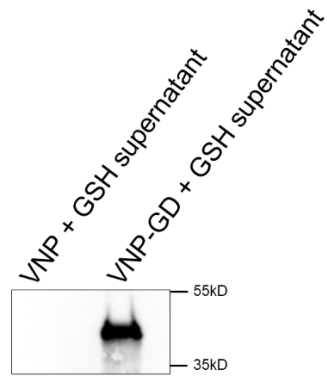
**Supplementary Fig. 5** | Flow cytometry assay of the conjugation efficiency of Rhodamine B-labeled Gasdermin D protein cages on the surface of VNP (n = 3 biologically independent samples).



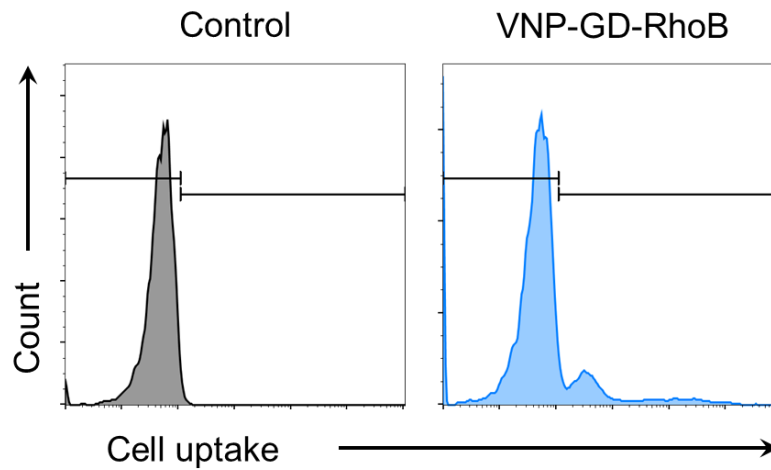
**Supplementary Fig. 6** | Western blot assay of Gasdermin D in VNP-GD and VNP. The experiments were repeated three times independently.



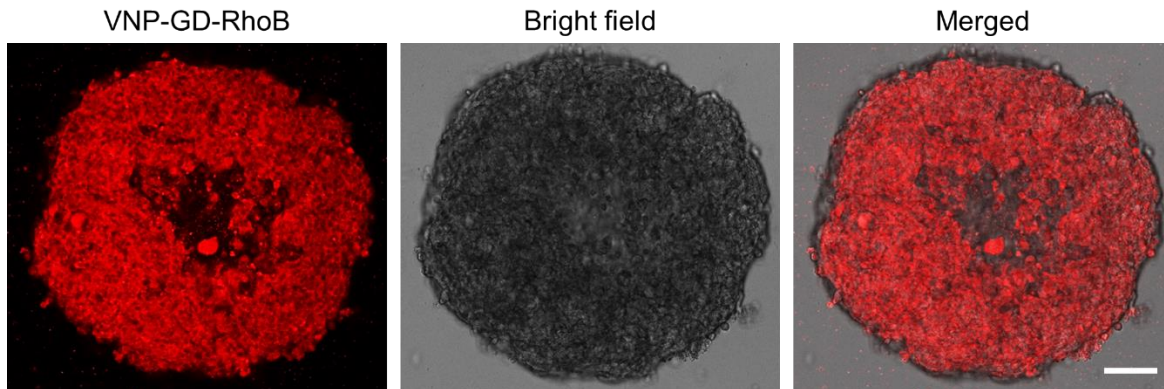
**Supplementary Fig. 7** | Bacterial counts of VNP and VNP-GD after cultured on LB solid medium. Data are shown as mean  $\pm$  s.d. (n = 4 biologically independent samples) and analyzed with two-tailed unpaired Student-*t* test.



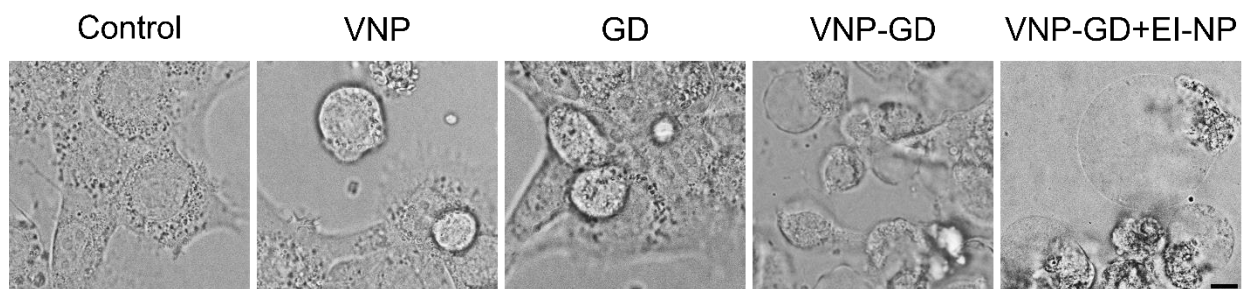
**Supplementary Fig. 8** | Western blot verification of Gasdermin D releasing from VNP-GD after GSH (10 mM) treatment. The experiments were repeated three times independently.



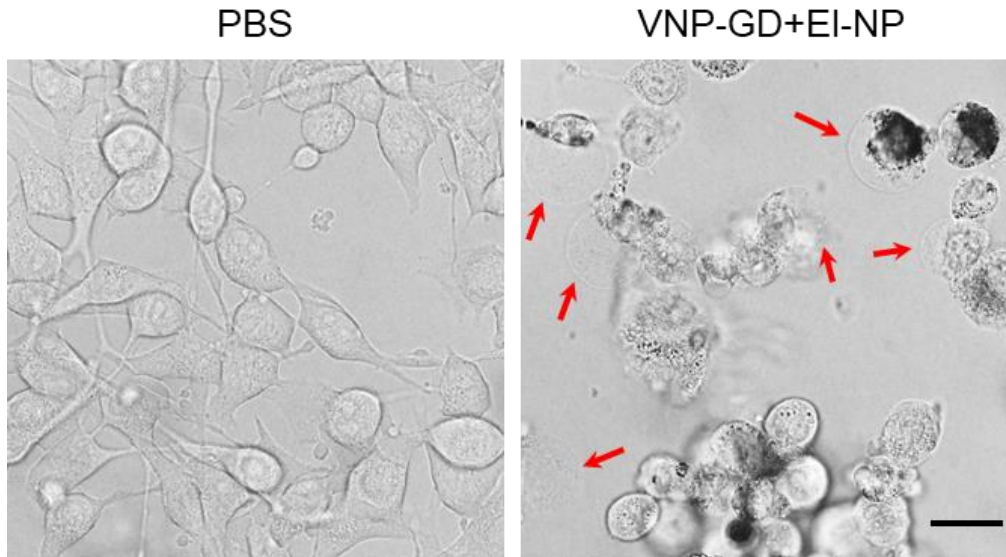
**Supplementary Fig. 9** | Flow cytometry assay of the cell uptake of VNP-GD-RhoB in 4T1 cells (n = 3 biologically independent samples). The protein cages were labeled with NHS-Rhodamine B before conjugation onto the bacteria. Untreated 4T1 cells served as control.



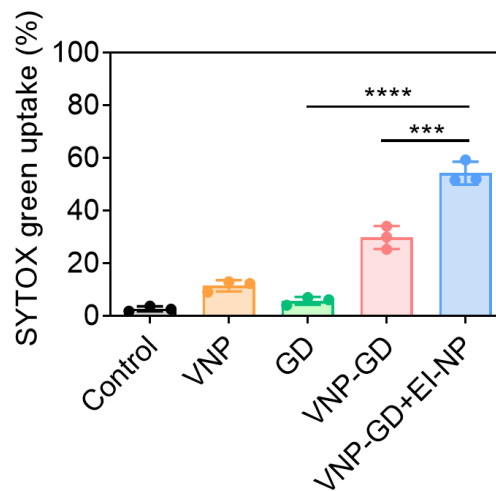
**Supplementary Fig. 10** | Confocal images of penetration of VNP-GD. The protein cages were labeled with NHS-Rhodamine B before conjugation onto VNP. Scale bar = 100  $\mu\text{m}$ . The experiments were repeated three times independently.



**Supplementary Fig. 11** | Enlarged pictures of 4T1 cell pyroptosis in **Fig. 2a** after different treatments (scale bar = 5  $\mu\text{m}$ ). The experiments were repeated three times independently.

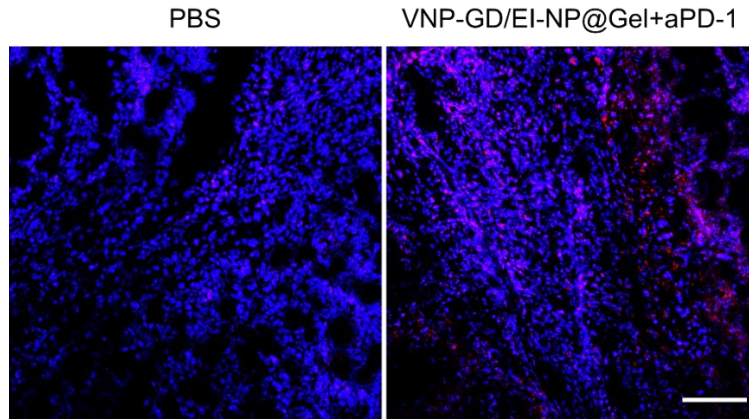


**Supplementary Fig. 12** | Representative images of B16F10 undergoing pyroptosis after treatment of PBS and VNP-GD+EI-NP for 24 h. Scale bar = 20  $\mu$ m. The experiments were repeated three times independently.

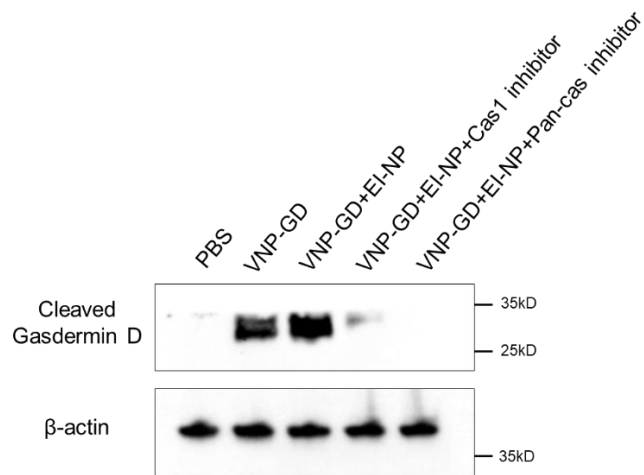


**Supplementary Fig. 13** | Data analysis of the cell uptake of SYTOX green in 4T1 tumor cells after incubation with PBS, VNP, GD (GSDMD protein cage), VNP-GD (GSDMD protein cage-conjugated VNP), and VNP-GD+EI-NP (GSDMD protein cage-conjugated VNP + EI-NP) for 24 hours. Data are presented as mean  $\pm$  s.d. (n = 3 biologically independent samples) and analyzed with one-way ANOVA followed by Dunnett's multiple comparisons test. VNP-GD+EI-NP vs. VNP-GD: \*\*\*  $P = 0.0003$ ; VNP-GD+EI-NP vs. GD: \*\*\*\*  $P < 0.0001$ .

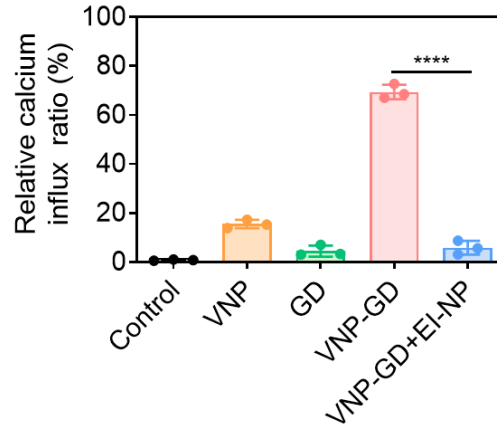




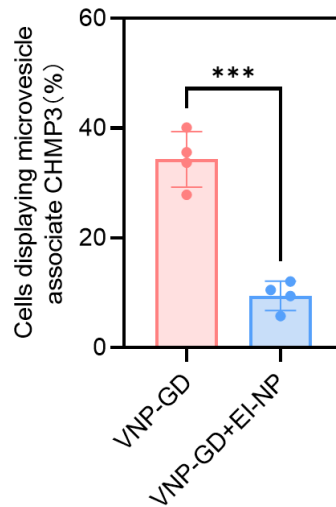
**Supplementary Fig. 14** | Immunofluorescence assay of cleaved caspase-1 after PBS and VNP-GD/EI-NP@Gel+aPD-1 treatments (n = 3 mice, scale bar = 100  $\mu$ m).



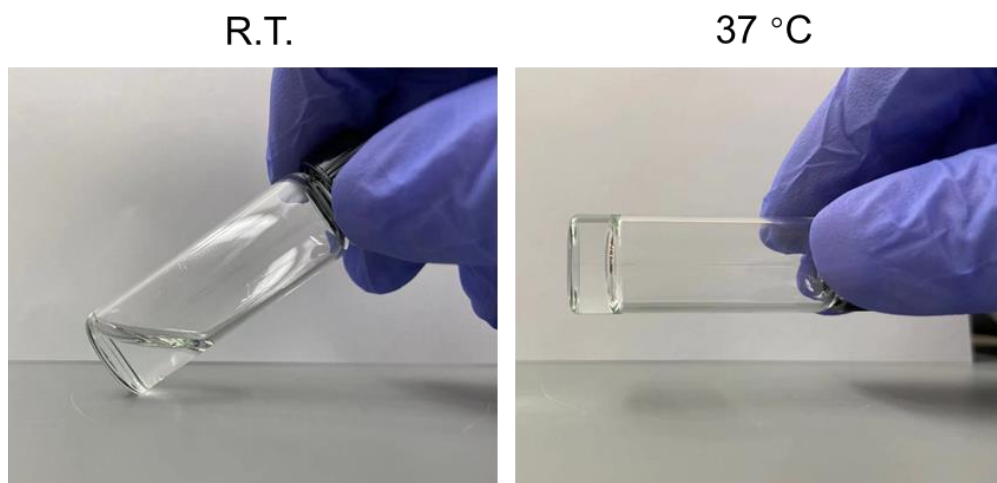
**Supplementary Fig. 15** | Western blot assay of cleaved Gasdermin D after different treatments with caspase inhibitors. The experiments were repeated three times independently.



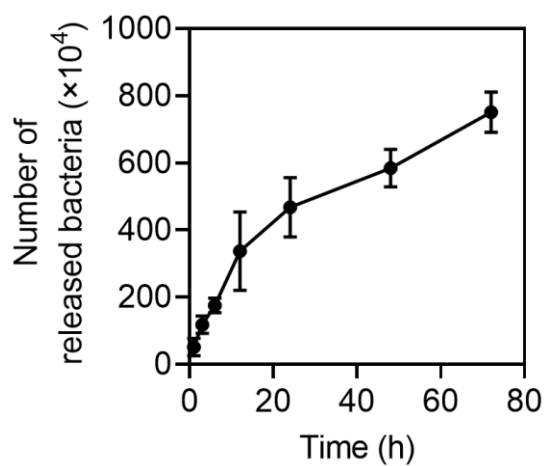
**Supplementary Fig. 16** | Quantified relative calcium influx ratio. Data are presented as mean  $\pm$  s.d. and analyzed with one-way ANOVA followed by Dunnett's multiple comparisons test ( $****P < 0.0001$ ,  $n = 3$  biologically independent samples).



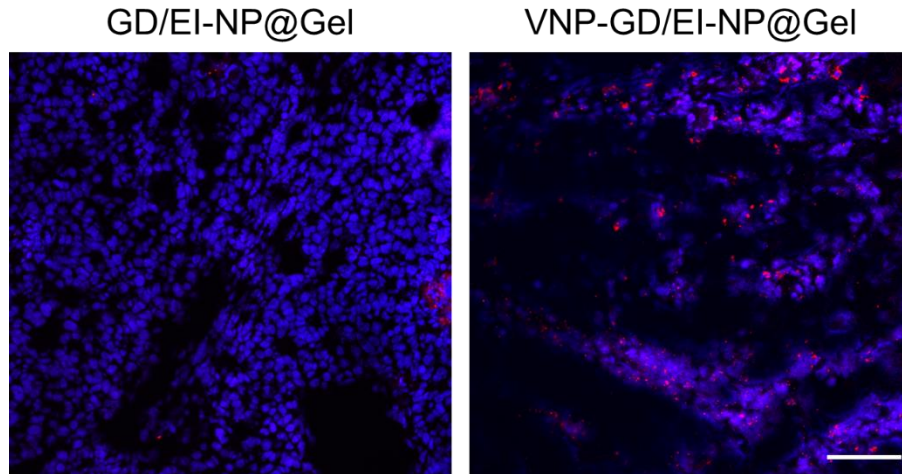
**Supplementary Fig. 17** | Data analysis of the percentage of cells displaying microvesicle-associated CHMP3 in Fig. 3e (Data are presented as mean  $\pm$  s.d. and analyzed with two-tailed unpaired Student-*t* test,  $***P = 0.0001$ ,  $n = 4$  biologically independent samples).



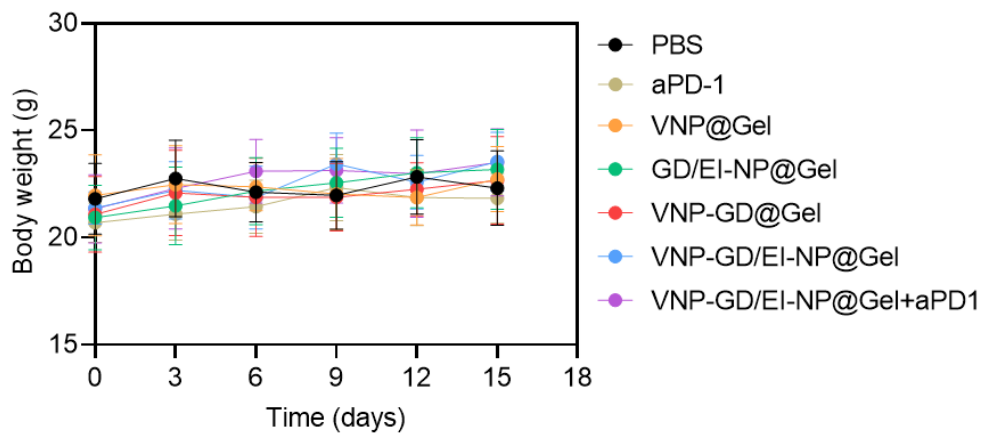
**Supplementary Fig. 18** | Gel formation of the Pluronic® F-127 solution at room temperature and 37 °C.



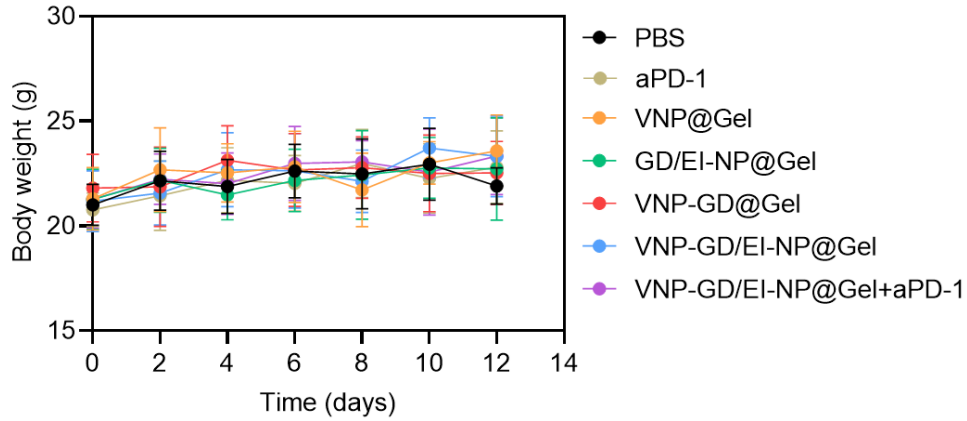
**Supplementary Fig. 19** | Number of the bacteria released from hydrogel.  $1 \times 10^7$  CFU bacteria were loaded in the hydrogel. Data are shown as mean  $\pm$  s.d. (n = 3 biologically independent samples).



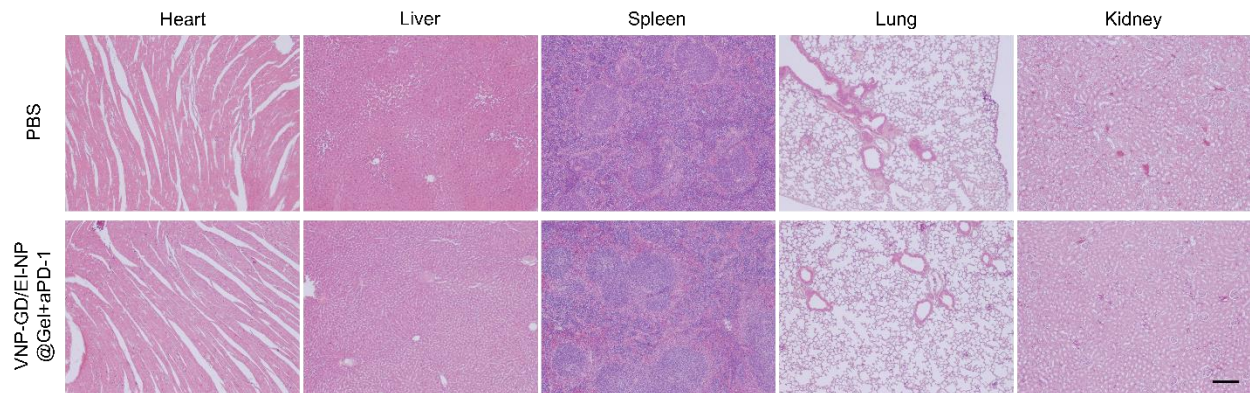
**Supplementary Fig.20** | Representative confocal images of the distribution of Rhodamine B-labeled GSDMD in the tumor tissue after peritumoral injection of GD/EI-NP@Gel and VNP-GD/EI-NP@Gel (n = 3 mice). Scale bar = 100  $\mu$ m. The experiments were repeated three times independently.



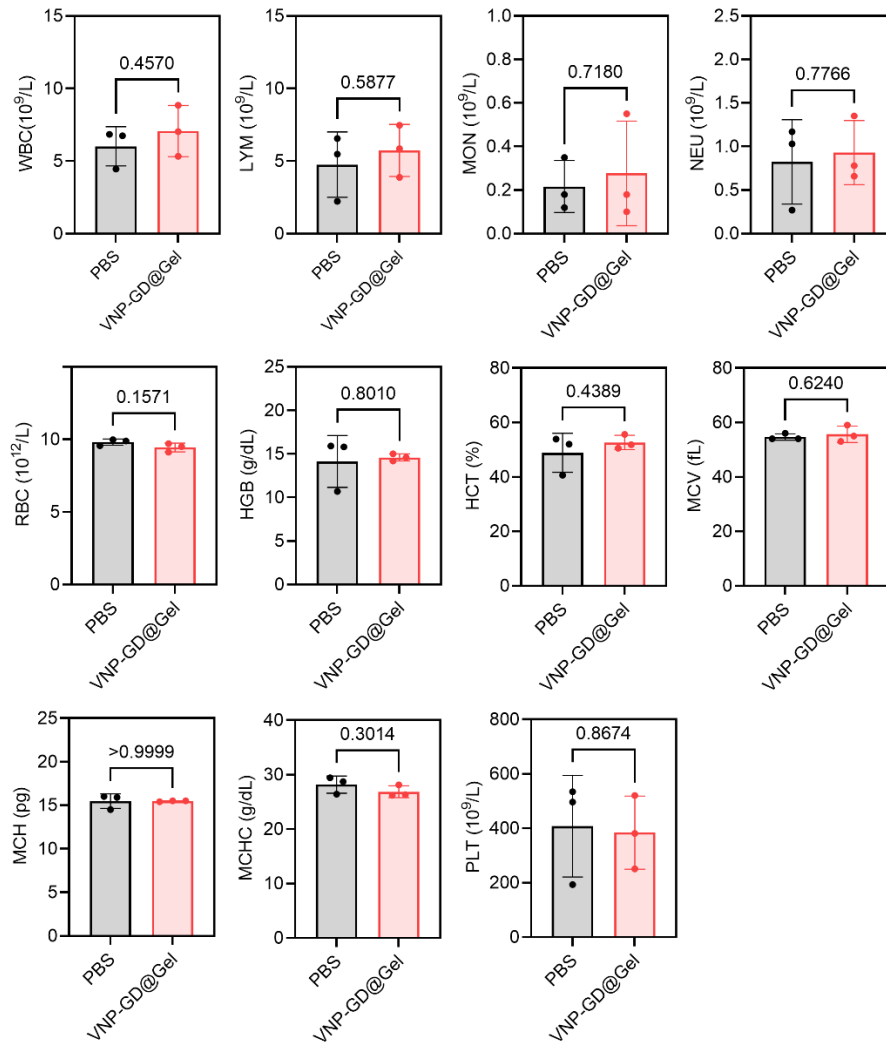
**Supplementary Fig. 21** | Body weight changes of the 4T1 breast tumor-bearing mice after treatment with PBS, aPD-1, VNP@Gel (hydrogel loaded with VNP), GD/EI-NP@Gel (GSDMD protein cage and EI-NP co-loaded in the hydrogel), VNP-GD@Gel (GSDMD protein cage-armed VNP loaded in the hydrogel), VNP-GD/EI-NP@Gel (GSDMD protein cage-armed VNP and EI-NP co-loaded in the hydrogel) and VNP-GD/EI-NP@Gel+aPD-1 (GSDMD protein cage-armed VNP and EI-NP co-loaded in the hydrogel with systemic injection of aPD-1 antibodies). GSDMD = 2 mg/kg, EI = 5 mg/kg, VNP =  $10^7$  CFU per mouse, aPD-1 = 2.5 mg/kg (three doses on day 0, day 2 and day 4). Data are presented as mean  $\pm$  s.d. (n = 6 mice).



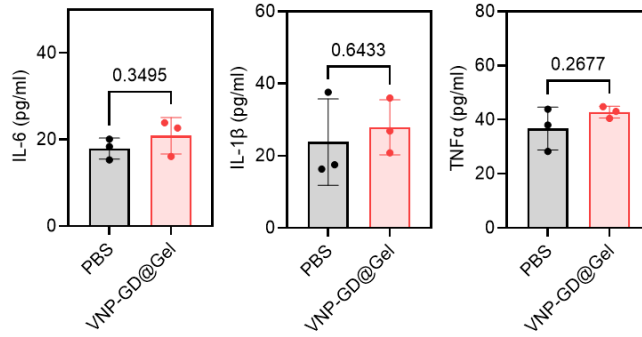
**Supplementary Fig. 22** | Body weight changes of the B16F10 tumor-bearing mice after treatment with PBS, aPD-1, VNP@Gel (hydrogel loaded with VNP), GD/EI-NP@Gel (GSDMD protein cage and EI-NP co-loaded in the hydrogel), VNP-GD@Gel (GSDMD protein cage-armed VNP loaded in the hydrogel), VNP-GD/EI-NP@Gel (GSDMD protein cage-armed VNP and EI-NP co-loaded in the hydrogel) and VNP-GD/EI-NP@Gel+aPD-1 (GSDMD protein cage-armed VNP and EI-NP co-loaded in the hydrogel with systemic injection of aPD-1 antibodies). GSDMD = 2 mg/kg, EI = 5 mg/kg, VNP =  $10^7$  CFU per mouse, aPD-1 = 2.5 mg/kg (three doses on day 0, day 2 and day 4). Data are presented as mean  $\pm$  s.d. (n = 6 mice).



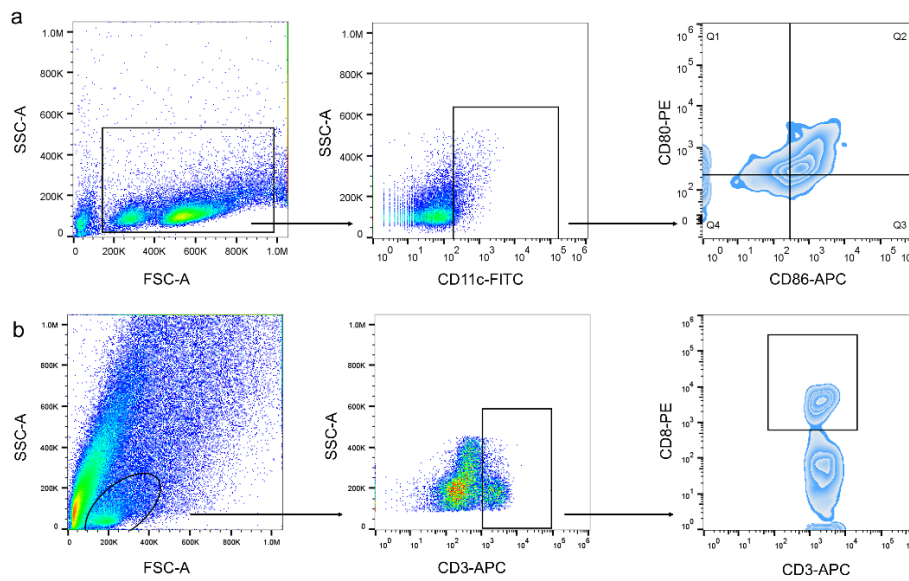
**Supplementary Fig. 23** | H&E assay of different organs including heart, liver, spleen, lung and kidney of C57BL/6 mice after treatments with PBS and VNP-GD/EI-NP@Gel+aPD-1. GSDMD = 2 mg/kg, EI = 5 mg/kg, VNP =  $10^7$  CFU per mouse, aPD-1 = 2.5 mg/kg (three doses on day 0, day 2 and day 4) (n = 3 mice). Scale bar = 200  $\mu$ m. The experiments were repeated three times independently.



**Supplementary Fig. 24** | Complete blood count analysis after PBS or VNP-GD@Gel treatments in Balb/c mice. Data are presented as mean  $\pm$  s.d. and analyzed with two-tailed unpaired Student-*t* test ( $n = 3$  mice). WBC: White Blood Cell Count; LYM: Lymphocytes; MON: Monocytes; NEU: Neutrophils; BAS: Basophils; RBC: Red Blood Cell Count; HGB: Hemoglobin; HCT: Hematocrit; MCV: Mean Corpuscular Volume; MCH: Mean Corpuscular Hemoglobin; MCHC: Mean Corpuscular Hemoglobin Concentration; PLT: Platelet Count.



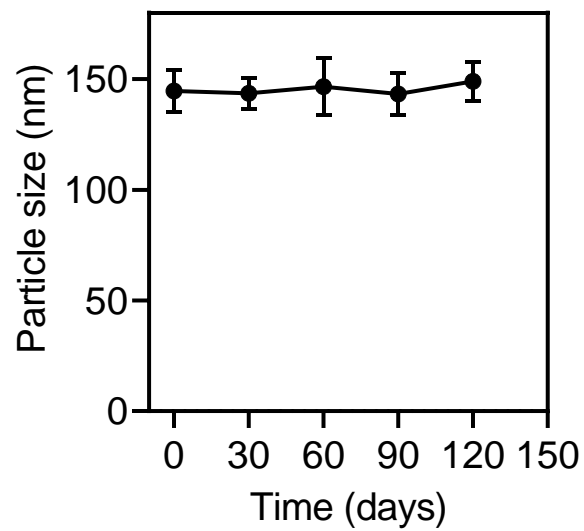
**Supplementary Fig. 25** | Systemic cytokine detections after PBS or VNP-GD@Gel treatments in Balb/c mice. Data are presented as mean  $\pm$  s.d. and analyzed with two-tailed unpaired Student-*t* test (n = 3 mice).



**Supplementary Fig. 26** | Gating strategy for flow cytometry assay of dendritic cells in lymph nodes and CD8<sup>+</sup> T cell infiltration in the tumor tissue. **a.** Representative gating strategy for dendritic cell detection in Fig. 5a, 5c. **b.** Representative gating strategy for CD8<sup>+</sup> T cell detection in Fig. 5b, 5d, 5e.



**Supplementary Fig. 27** | Representative image of lyophilized hydrogel patches loaded with EI-NP.



**Supplementary Fig. 28** | Stability of re-suspended dextran nanoparticle (EI-NP) after lyophilization. Data are shown as mean  $\pm$  s.d. (n = 3 biologically independent samples).