

Supplementary information**Induced pluripotent stem cells of endangered avian species**

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Supplemental methods

Generation of iPSCs with PB-R6F reprogramming vector.

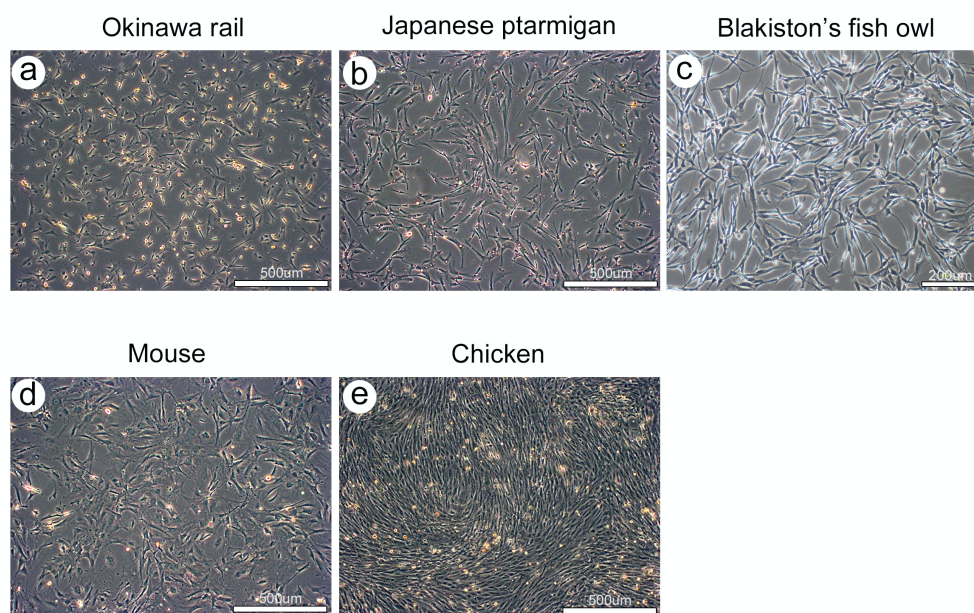
PB-R6F reprogramming vectors were transfected into mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived fibroblasts using Lipofectamine 2000 transduction reagent (11668019; Thermo Fisher Scientific, Waltham, MA, USA). The PB-R6F reprogramming vector has been described in detail in our previous reports^{1,2}. After hygromycin selection (Wako Pure Chemical Industries, Osaka, Japan), transduced cells were reseeded onto a mouse embryonic fibroblast (MEF) feeder layer. Primary iPS cell-like colonies were selected and seeded onto new MEF feeder cell plates.

Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl genome and amino acids.

Nucleic acid sequences derived from Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl DNA were obtained using a next-generation sequencer (HiSeq and IonPGM). To obtain target genes (e.g., *Pou5*, *Sox2*, and *Nanog*), we searched all Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl sequences using Blastn, with chicken mRNA sequences as queries. The sequences obtained were converted into amino acids, and a comparison of mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl was performed using the T-Coffee website (<http://tcoffee.vital-it.ch/apps/tcoffee/index.html>).

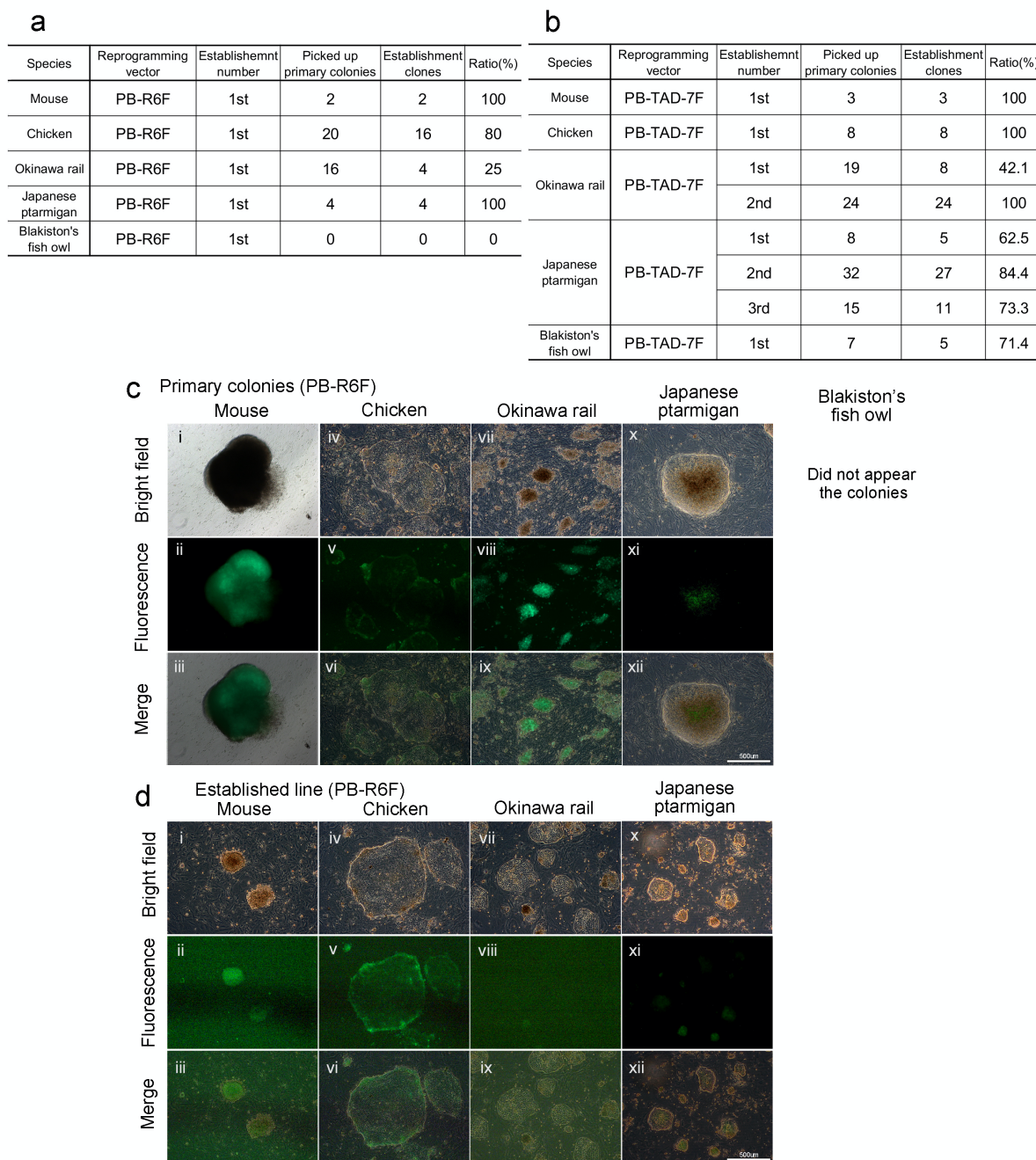
Comparison of characteristics of iPSCs with PB-R6F and PB-TAD-7F.

In addition to the PB-TAD-7F reprogramming vector, we used the PB-R6F vector (*M3O*, *Sox2*, *Klf4*, *c-Myc*, *Nanog*, and *Lin28*). There were no observable differences between iPSCs with PB-TAD-7F or PB-R6F in chicken and mouse iPSCs concerning morphology, pluripotency marker staining, and pluripotency-related gene expression (Supplementary Fig. 2 to 4). However, we observed differences between reprogramming vectors in Blakiston's fish owl-derived cells. In brief, the PB-TAD-7F reprogramming vector generated Blakiston's fish owl iPSCs, whereas the PB-R6F reprogramming vector was unable to generate Blakiston's fish owl iPSCs (Supplementary Fig. 2). Therefore, we conclude that the PB-TAD-7F reprogramming vector is more advantageous than the PB-R6F reprogramming vector for generating avian iPSCs. We analyzed the characteristics of iPSCs using the PB-TAD-7F reprogramming vector.



Supplementary Figure 1. Images of primary cells.

Images of the primary cells, including Okinawa rail-derived primary cells (a), Japanese ptarmigan-derived primary cells (b), Blakiston's fish owl-derived primary cells (c), mouse-derived primary cells (d), and chicken-derived primary cells (e). Bars represent 200 µm (c) or 500 µm (a, b, d, e).



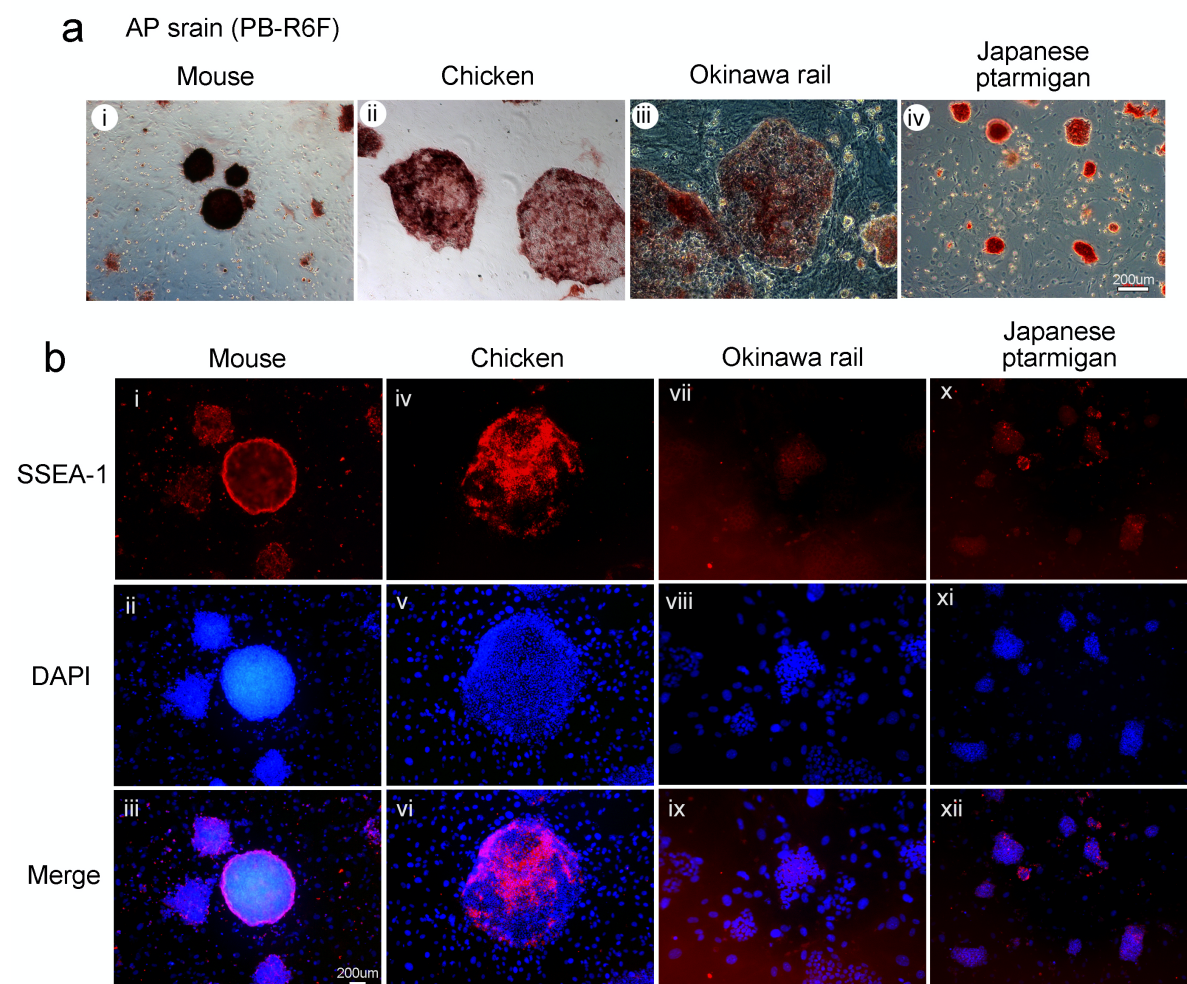
Supplementary Figure 2. Establishment of iPSCs derived from mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl using the PB-R6F vector.

a,b: Comparison between PB-R6F and PB-TAD-7F reprogramming vectors for the establishment ratio of avian-derived iPSCs. a: PB-R6F, b: PB-TAD-7F.

c: Primary colonies of iPSCs derived from mice, chickens, Okinawa rail, and Japanese ptarmigans. Panels are bright field images (i, iv, vii, and x), green fluorescence protein (GFP) images (ii, v, viii, xi), and merged images (iii, vi, ix, and xii). Panels show mice (i-iii), panels show chickens (iv-vi), Okinawa rail (vii-ix), and Japanese ptarmigans (x-xii). The bars represent 500 μ m.

d: Morphologies of our established iPS cell lines derived from mouse, chicken, Okinawa rail, and Japanese

ptarmigan. Panels are bright field images (i, iv, vii, and x), green fluorescence protein (GFP) images (ii, v, viii, xi), and merged images (iii, vi, ix, and xii). Panels show mice (i-iii), panels show chickens (iv-vi), Okinawa rail (vii-ix), and Japanese ptarmigans (x-xii). The bars represent 500 μ m.

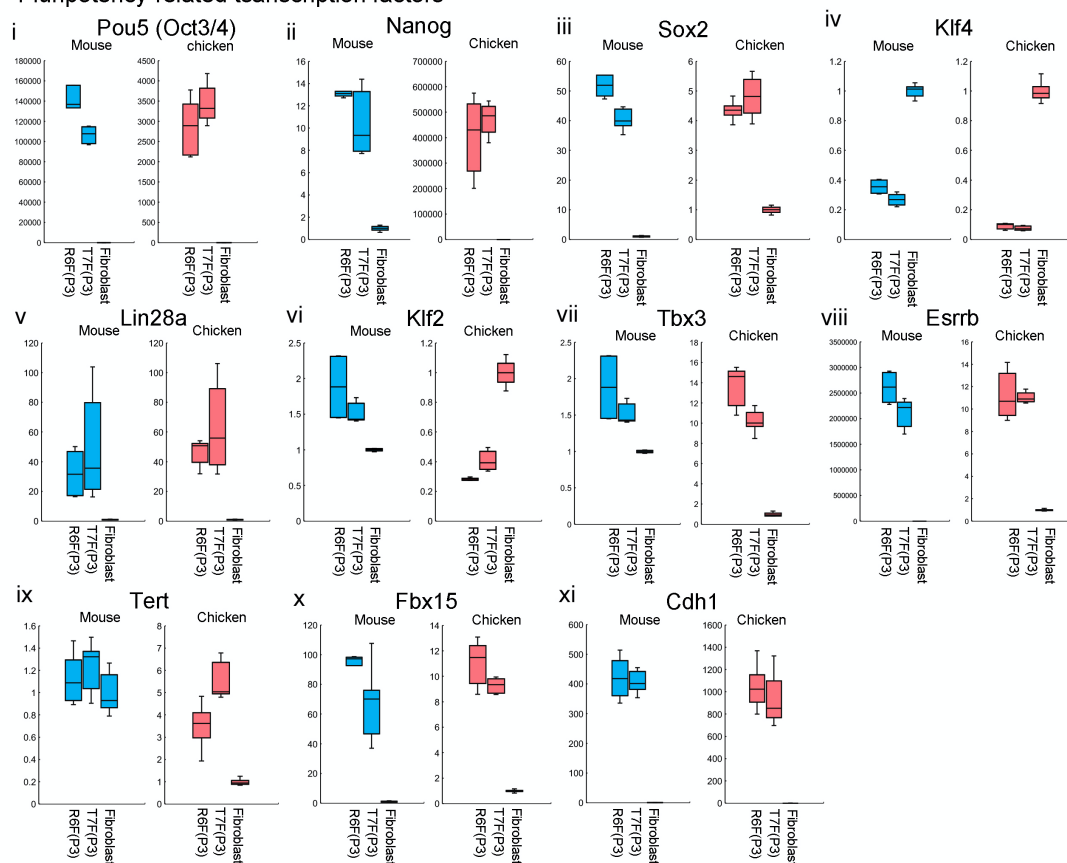


Supplementary Figure 3. Detection of stem cell markers in our established iPSCs derived from mouse, chicken, Okinawa rail, and Japanese ptarmigan using the PB-R6F reprogramming vector.

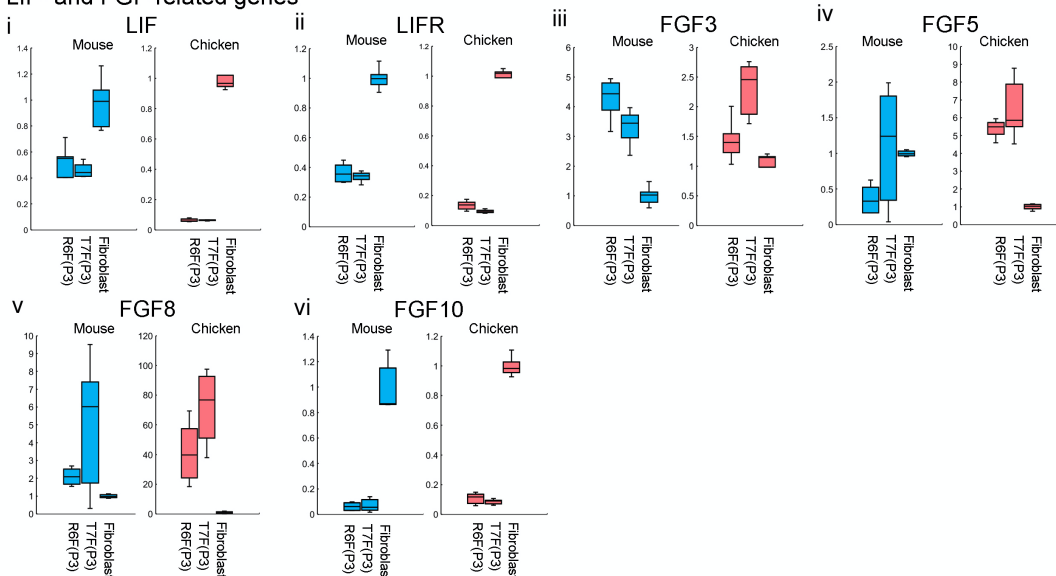
a: Alkaline phosphatase (AP) activity in iPSCs derived from mouse (i), chicken (ii), Okinawa rail (iii), and Japanese ptarmigan (iv). The bars represent 200 μ m.

b: Detection of stage-specific embryonic antigen (SSEA)-1 in mouse (i-iii), chicken (iv-vi), Okinawa rail (vii-ix), and Japanese ptarmigan (x-xii). Panels show SSEA-1 images (i, iv, vii, and x), 4',6-diamidino-2-phenylindole (DAPI) images (ii, v, viii, and xi), and merged images (iii, vi, ix, and xii). The bars represent 200 μ m.

a Pluripotency-related transcription factors



b LIF- and FGF-related genes



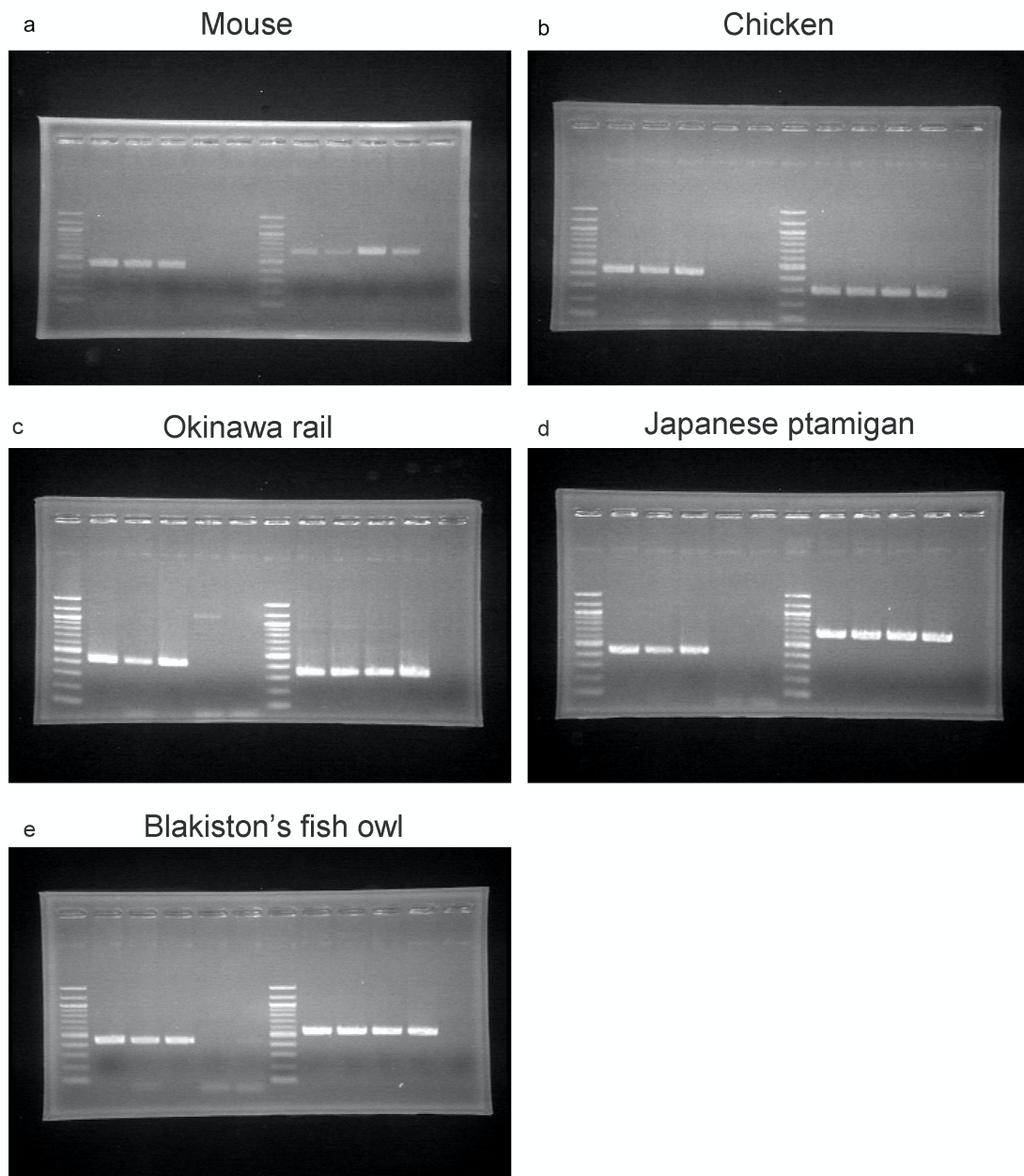
Supplementary Figure 4. Comparison of pluripotency-related gene expression in mouse and chicken iPSCs with PB-R6F, iPSCs with PB-TAD-7F, and fibroblasts.

a: Endogenous expression of pluripotency marker genes in mouse and chicken-derived cells. The mRNA expression of *Pou5 (Oct3/4)* (i), *Nanog* (ii), *Sox2* (iii), *Klf4* (iv), *Lin28a* (v), *Klf2* (vi), *Tbx3* (vii), *Esrrb* (viii), *Tert* (ix), *Fbx15* (x), *Cdh1* (xi) shows.

b: Endogenous expression of *LIF*- and *FGF*-related genes in mouse and chicken-derived cells. The mRNA

expression of *LIF* (i), *LIFR* (ii), *FGF3* (iii), *FGF5* (iv), *FGF8* (v), and *FGF10* (vi) shows.

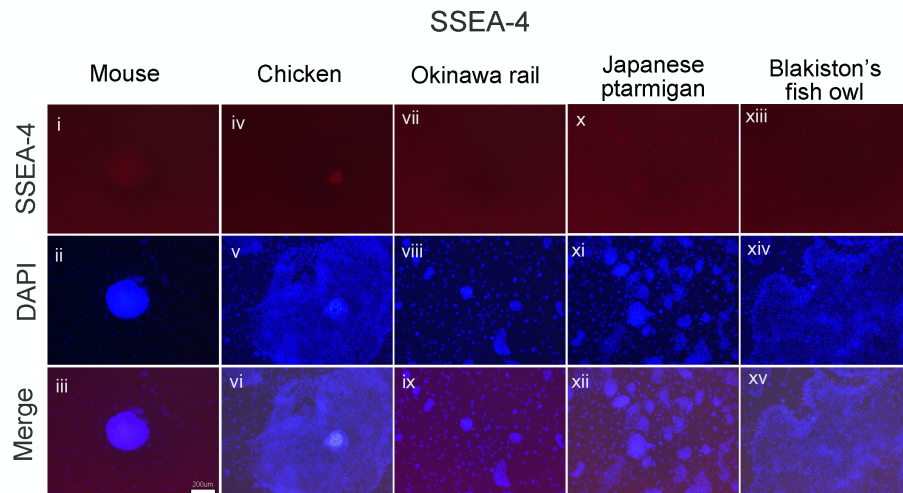
Gene expression was quantified relative to *GAPDH* (internal control), and showed that the fibroblast expression level was 1.0. Blue bars denote mouse-derived cells and red bars denote chicken-derived cells. Bars represent iPSCs with PB-R6F (passage number 3; P3), iPSCs with PB-TAD-7F (passage number 3; P3), and fibroblasts. Centerlines of box plots indicate medians; box limits indicate the 25th and 75th percentiles. n=4 (mouse PB-R6F and chicken fibroblast), n=6(other).



Supplementary Figure 5. Detection of transduced genes by genomic PCR (full scan blot).

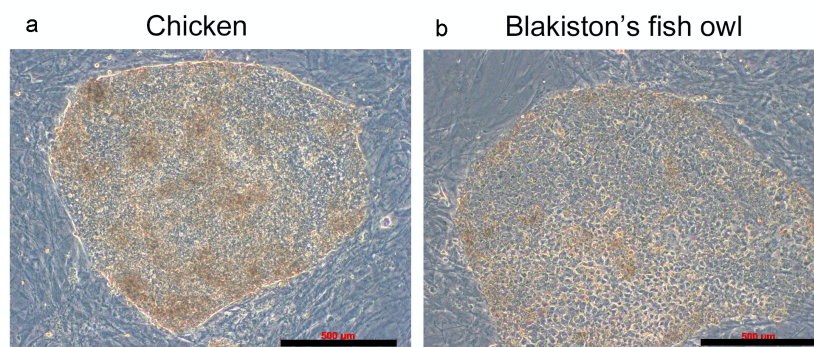
The gel photographs show mouse (a), chicken (b), Okinawa rail (c), Japanese ptarmigan (d), and Blakiston's fish owl (e). The six lanes on the left of each gel show the PB-TAD-7F reprogramming vector. The six

right lanes of each gel show *Tsc-2* (internal control). Detailed information is shown in Figure 2a (high-magnification image of these gel).



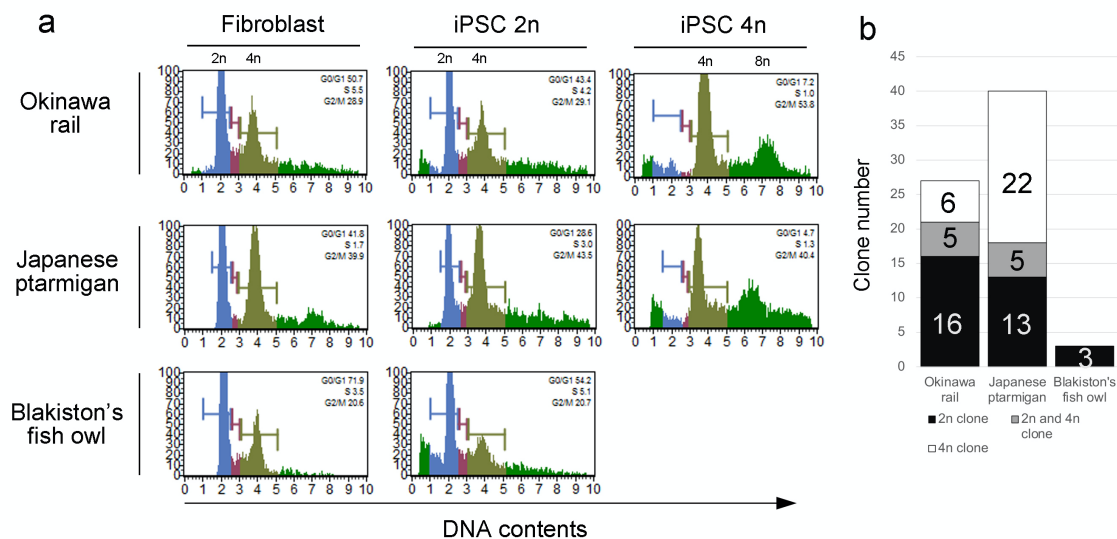
Supplementary Figure 6. Detection of stage-specific embryonic antigen (SSEA)-4.

Images are from mouse (i-iii), chicken (iv-vi), Okinawa rail (vii-ix), Japanese ptarmigan (x-xii), and Blakiston's fish owl (xiii-xv). Panels show SSEA-4 (i, iv, vii, x, xiii), DAPI (ii, v, viii, xi, xiv), and the merged (iii, vi, ix, xii, xv) images. The bars represent 200 μ m.



Supplementary Figure 7. Morphology of iPSCs from chick and Blakiston's fish owl at a late passage.

The images are a chicken iPSC colony at passage 20 (a) and a Blakiston's fish owl iPSC colony at passage 20 (b). The bars represent 500 μ m.

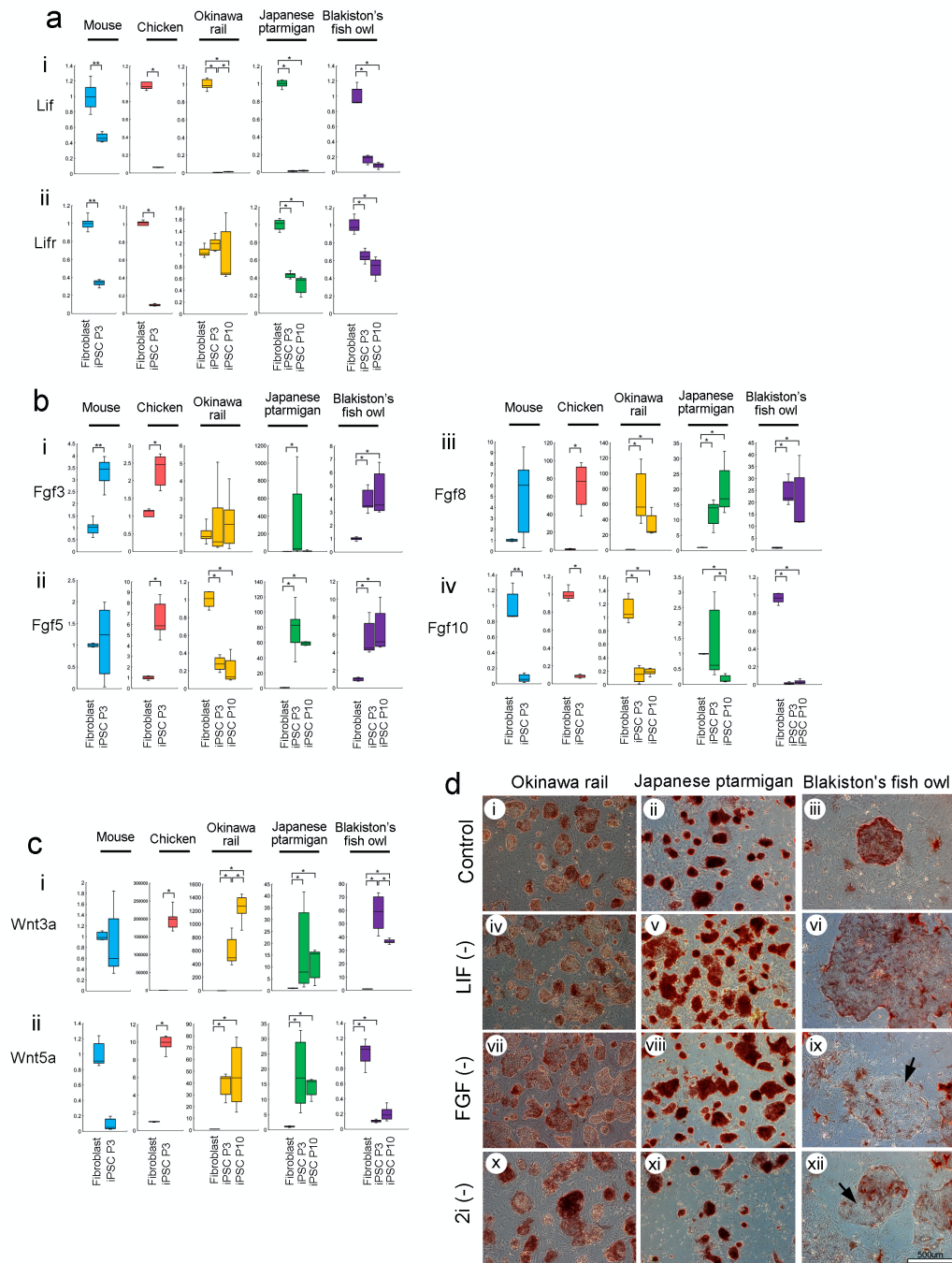


Supplementary Figure 8 Chromosomal analysis of Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived iPSCs.

a: DNA content profile analysis of Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived iPSCs by flow cytometry. The DNA content profile was analyzed by flow cytometry to evaluate chromosomal polyploidy and diploidy. Panels represent fibroblasts, diploid iPSCs, and polyploid iPSCs.

b: Number of diploid and polyploid clones derived from Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived iPSCs. The black area shows the number of 2n clones, the gray area shows the number of 2n and 4n clones, and the white area shows the number of 4n clones.

c: Chromosomal patterns of endangered Japanese avian-derived iPSCs. The panels show the chromosome pattern of Okinawa rail-derived iPSCs (i, ii), Japanese ptarmigan-derived iPSCs (iii, iv), and Blakiston fish owl-derived iPSCs (v, vi). Panels show the representative mitotic phase (i, iii, v), and the aligned chromosomes (ii, iv, vi).



Supplementary Figure 9 *Lif*, *Lifr*, *Fgf* family, and *Wnt* gene expression and Cell culture conditions of Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived iPSCs.

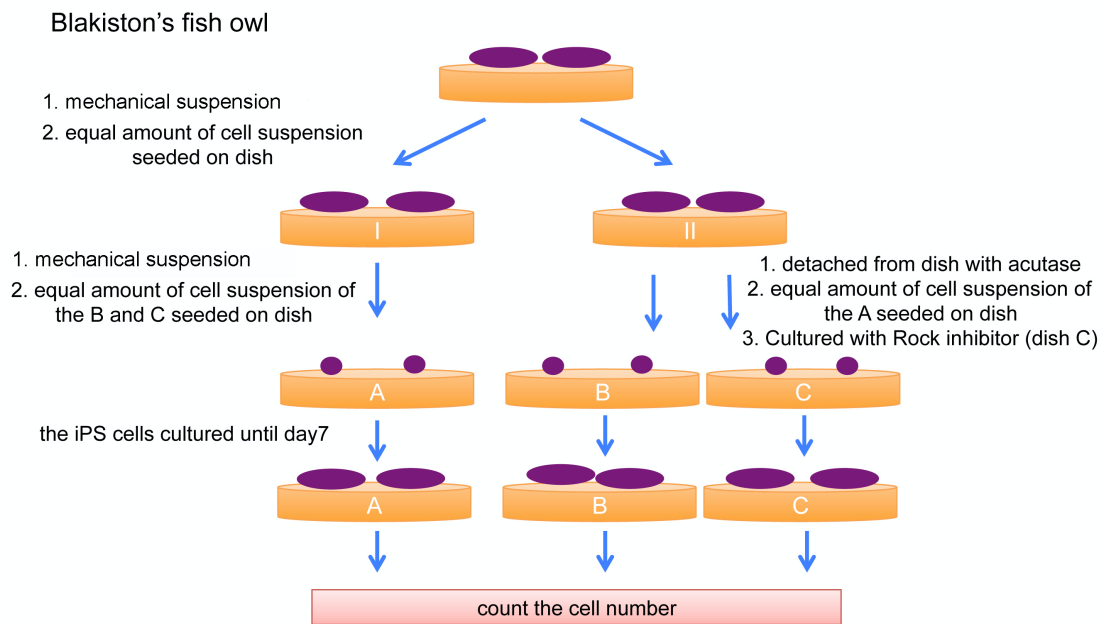
a: *Lif* (i) and *Lifr* (ii) gene expression in mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston fish owl-derived iPSCs.

b: *Fgf 3* (i), *Fgf 5* (ii), *Fgf 8* (iii), *Fgf 10* (iv) gene expression in mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston fish owl-derived iPSCs.

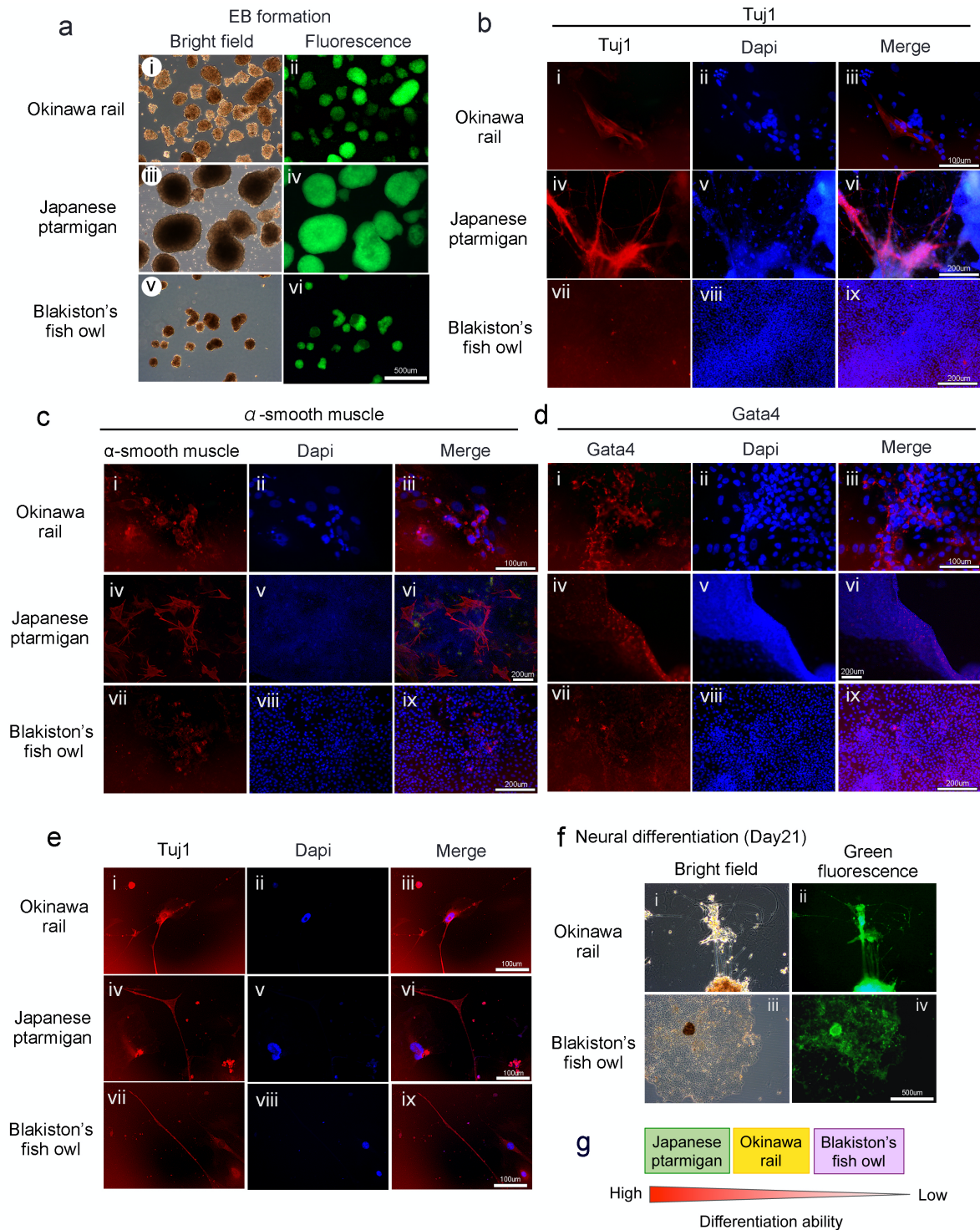
c: *Wnt 3a* and *Wnt 5a* gene expression in mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston fish owl-derived iPSCs.

Gene expression was quantified relative to that of the *GAPDH* internal control. n=4 (chicken fibroblast), n=6 (other). * P<0.05, ** P<0.01. Blue bars, mouse; red bars, chicken; yellow bars, Okinawa rail; green bars, ptarmigan; and purple bars, Blakiston's fish owl-derived cells. Mouse and chicken: bars represent fibroblasts and iPSCs (passage number 3; P3). Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl: bars represent fibroblasts, iPSCs (P3), and iPSCs (P10). Centerlines of box plots indicate medians; box limits indicate the 25th and 75th percentiles.

d: Alkaline phosphatase staining of Okinawa rail (i, iv, vii, x), Japanese ptarmigan (ii, v, viii, xi), and Blakiston's fish owl iPSCs (iii, vi, ix, xii) in standard medium (i-iii), LIF minus medium (iv-vi), FGF minus medium (vii-ix), and 2i minus medium (x-xii). The panels are Okinawa rail, Japanese ptarmigans, and Blakiston's fish owls iPSCs. The bars represent 500 μ m.



Supplementary Figure 10. Schematic of cell number counting for Blakiston's fish owl-derived iPSCs.



Supplementary Figure 11. *In vitro* differentiation of Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived iPSCs.

a: Cytomorphology of embryoid bodies (EBs). Panels show Okinawa rail-derived EBs (i, ii), Japanese ptarmigan-derived EBs (iii, iv), and Blakiston's fish owl-derived EBs (v, vi). The panels represent bright field images (i, iii, v) and GFP (ii, iv, vi). The bars represent 500 μ m.

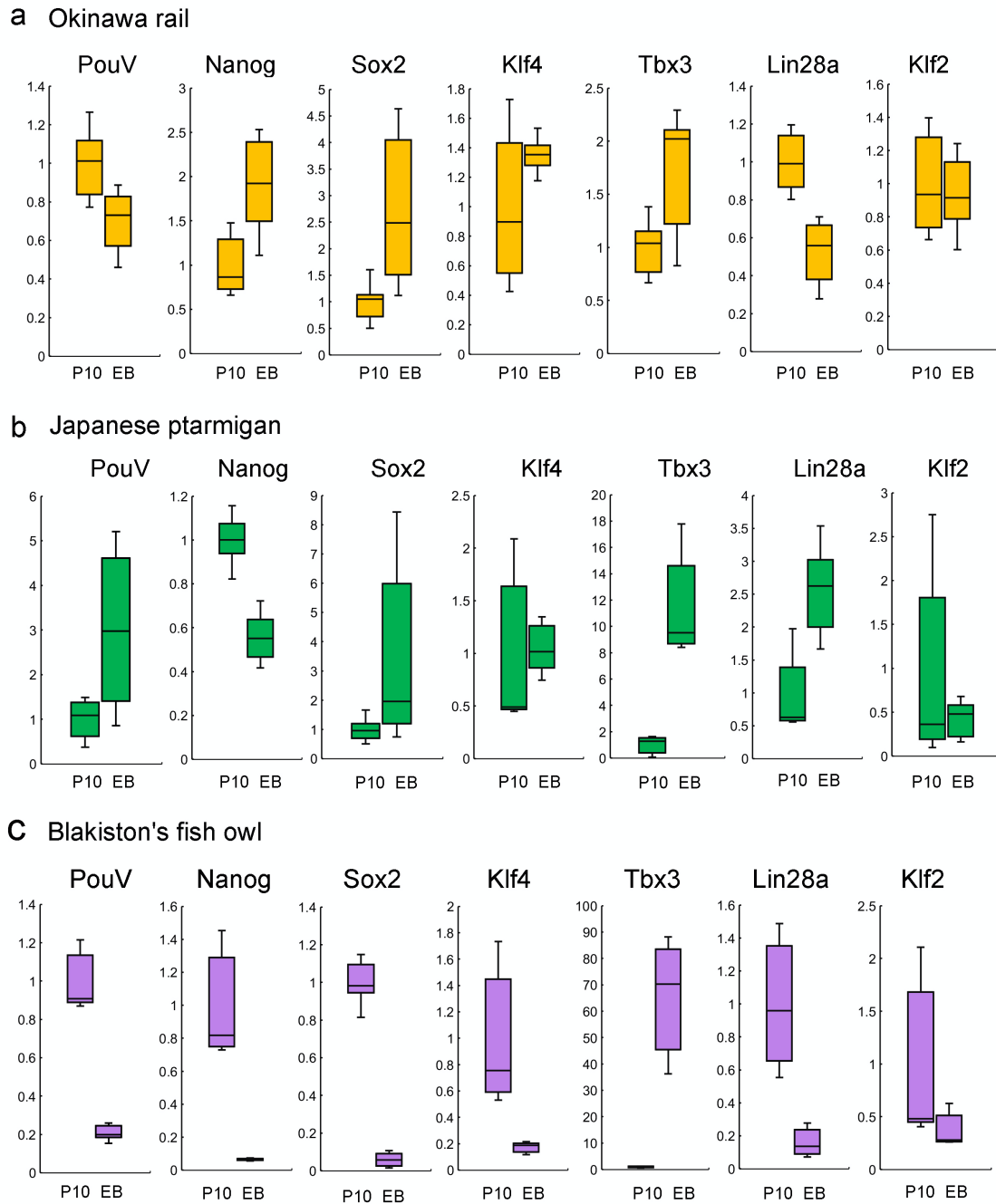
b, c, and d: Immunocytochemical evaluation of the ability to differentiate into the three germ layers *in vitro*. Ectodermal cells (Tuj1) (b), mesodermal cells (α -smooth muscle) (c), and endodermal cells

(Gata4) (d) are shown. Panels show Okinawa rail-derived cells (i-iii), Japanese ptarmigan-derived cells (iv-vi), and Blakiston's fish owl-derived cells (vii-ix). The bars represent 100 μm (Okinawa rail) or 200 μm (Japanese ptarmigan and Blakiston fish owl). Red fluorescence (i, iv,vii), dapi (ii, v, viii) and merge (iii, vi, ix).

e: Cytomorphology of dendrite-like structures derived from Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived iPSCs. Panels show Okinawa rail-derived cells (i-iii), Japanese ptarmigan-derived cells (iv-vi), and Blakiston's fish owl-derived cells (vii-ix). The bars represent 100 μm (Okinawa rail) or 200 μm (Japanese ptarmigan and Blakiston fish owl). Red fluorescence (i, iv,vii), dapi (ii, v, viii) and merge (iii, vi, ix). Bars represent 100 μm .

f: Differences in the *in vitro* differentiation potential of Okinawa rail and Blakiston fish owl-derived iPSCs at day 21. Panels show Okinawa rail (i, ii) and Blakiston's fish owls (iii, iv). Panels show bright field (i, iii) and green fluorescence (ii, iv). The bars represent 500 μm .

g: Differentiation ability of Okinawa rail, Japanese ptarmigan, and Blakiston fish owl-derived iPSCs. Differences in *in vitro* differentiation potential among Okinawa rail, Japanese ptarmigan, and Blakiston fish owl-derived iPSCs. Japanese ptarmigan-derived iPSCs have the highest differentiation ability, followed by Okinawa rail-derived iPSCs and Blakiston's fish owl-derived iPSCs.



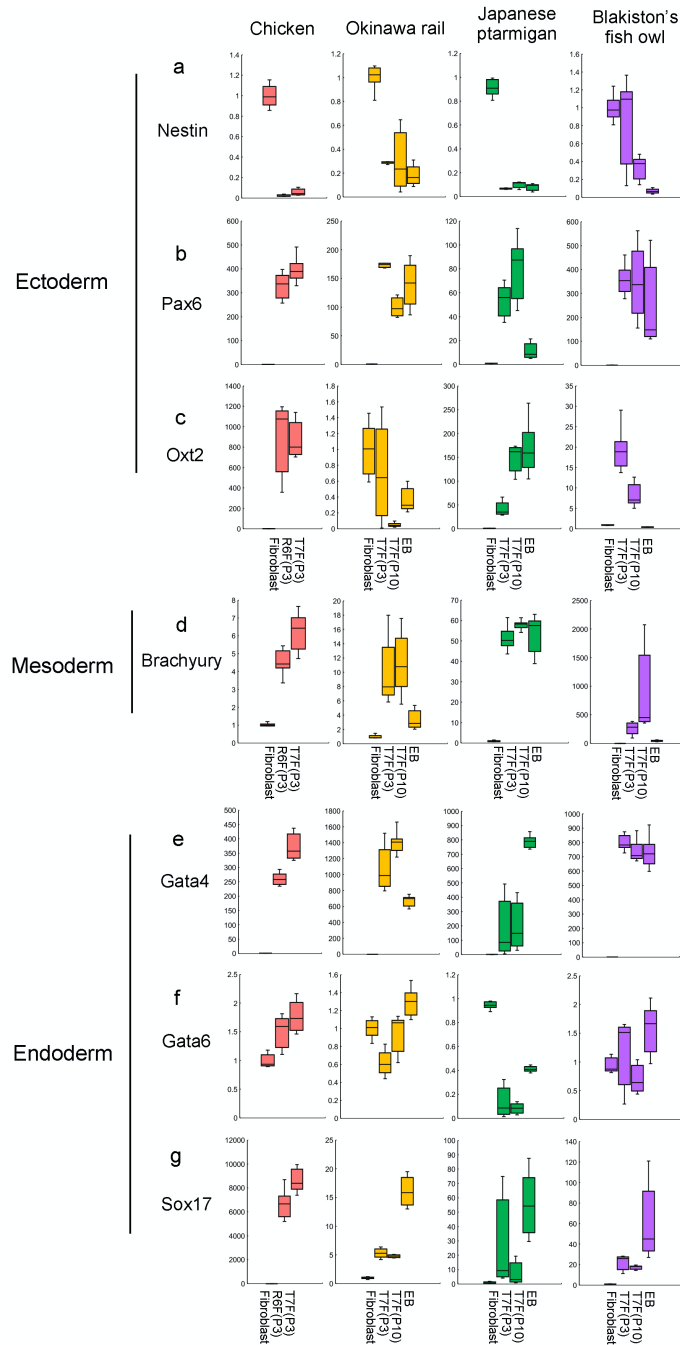
Supplementary Figure 12. Comparison of pluripotency-related gene expression of Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived cells between iPSCs with PB-TAD-7F and embryoid bodies (EBs).

a: Endogenous expression of pluripotency marker genes in Okinawa rail-derived iPSCs and EBs.

b: Endogenous expression of pluripotency marker genes in Japanese ptarmigan-derived iPSCs and EBs.

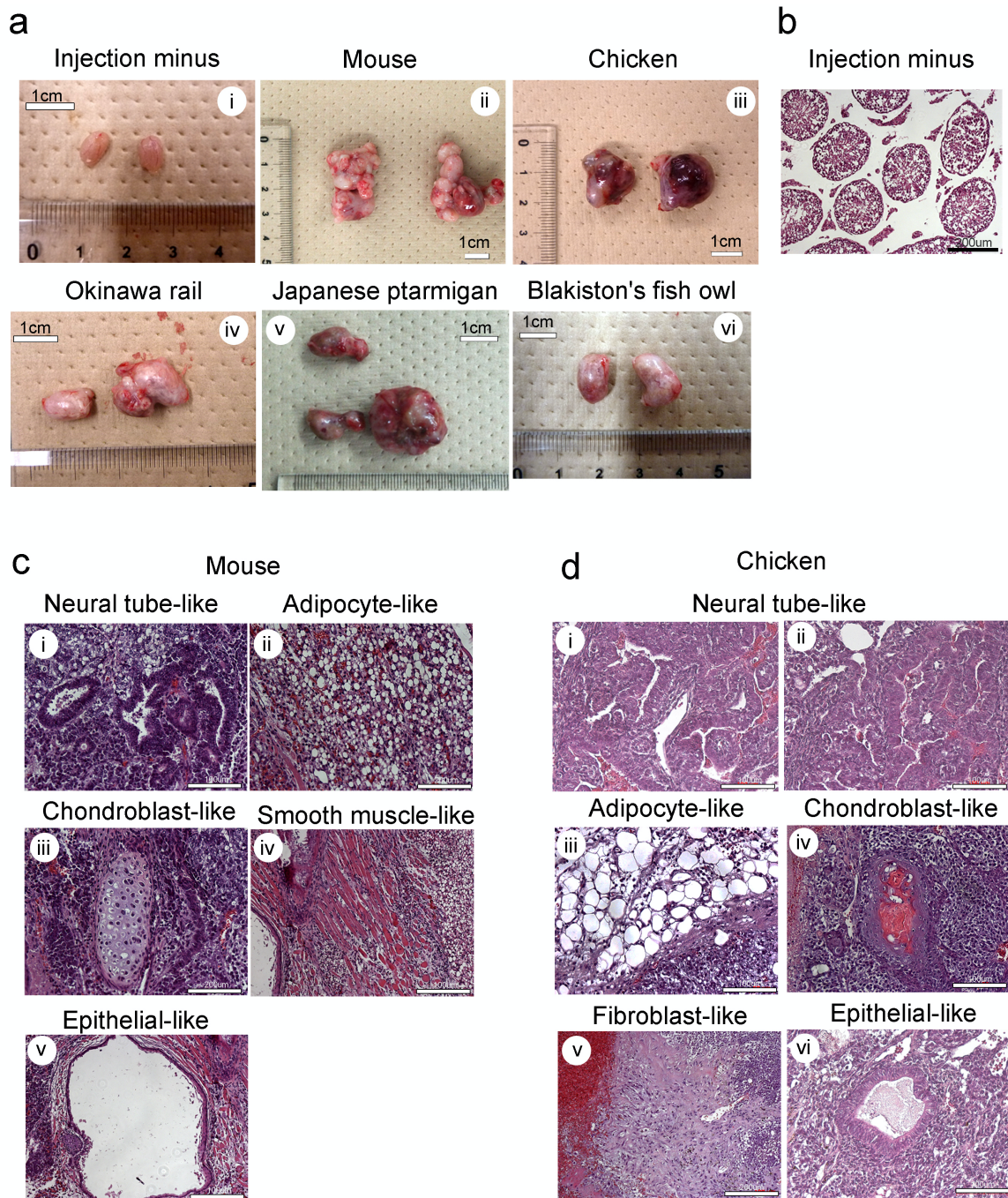
c: Endogenous expression of pluripotency marker genes in Japanese ptarmigan-derived iPSCs and EBs.

Gene expression was quantified relative to *GAPDH* (internal control). The expression level is 1.0 in iPSCs with PB-TAD-7F (passage 10; P10). The bars represents iPSCs with PB-TAD-7F (P10) and EBs. n=6. Centerlines of box plots indicate medians; box limits indicate the 25th and 75th percentiles.



Supplementary Figure 13. Comparison of the germ layer-related gene expression of chick, Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived cells among fibroblasts, iPSCs (passage 3; P3), iPSCs (passage 10, P10), and embryoid bodies (EBs).

The mRNA expression of Nestin (a), Pax6 (b), Oxt2 (c), Brachyury (d), Gata4 (e), Gata6 (f), Sox17 (g) shows. Chicken (red), Okinawa rail (yellow), Japanese ptarmigan (green), Blakiston's fish owl (purple). The bars represent fibroblasts, iPSCs (P3), iPSCs (P10), and EBs. $n=6$. Centerlines of box plots indicate medians; box limits indicate the 25th and 75th percentiles.



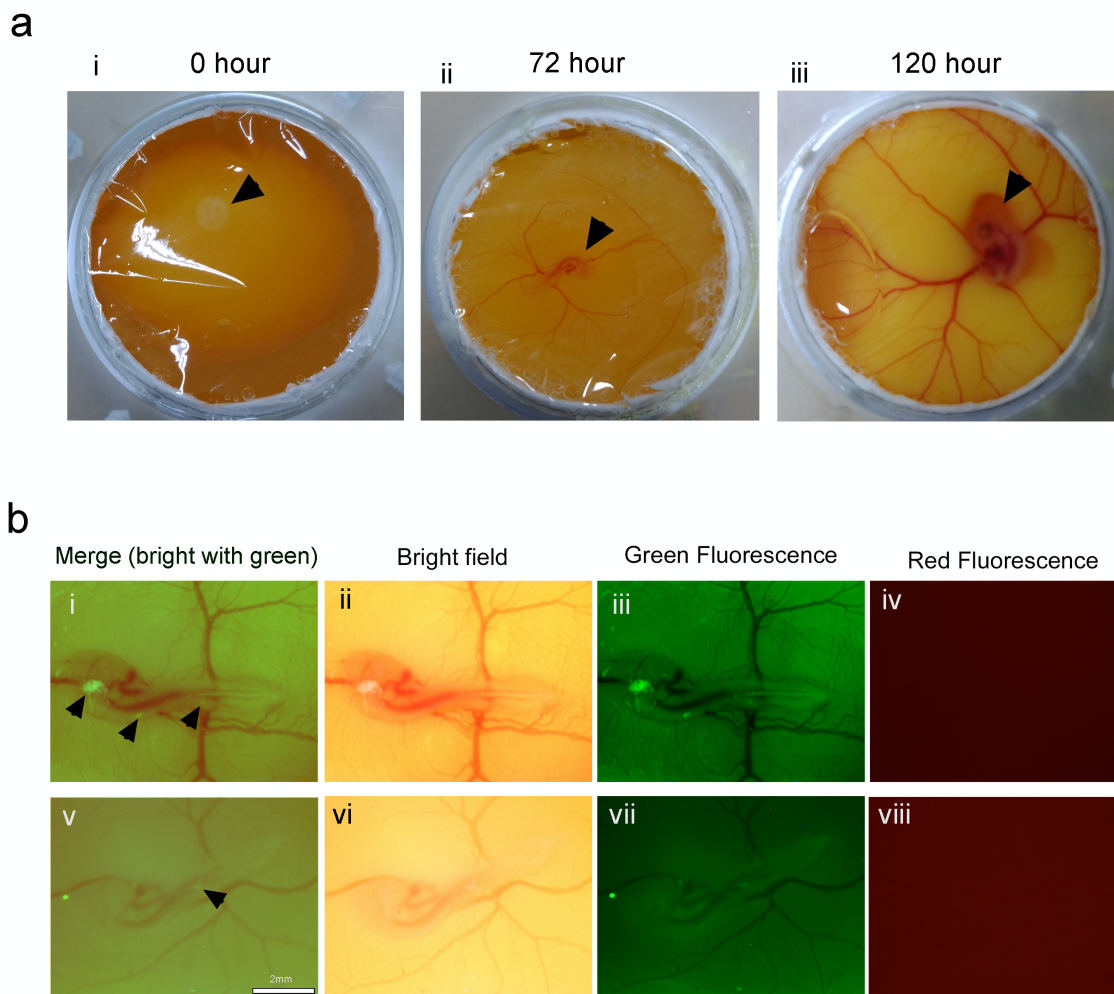
Supplementary Figure 14. Differentiation of the established mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl-derived iPSCs *in vivo*.

a: Teratoma formation from iPSCs generated with the PB-TAD-7F reprogramming vector.

Injection minus (control) (i), mouse iPSC-derived teratomas (ii); chicken iPSC-derived teratomas (iii); Okinawa rail iPSC-derived teratomas (iv); Japanese ptarmigan iPSC-derived teratomas (v); Blakiston's fish owl iPSC-derived teratomas (vi). Bars show 1cm.

b–d: Histological analysis of injection minus (b), mouse iPSC-derived teratomas (c), and chicken iPSC-derived teratomas (d). Various tissues originating from the three germ layers were identified, and include neural tube-like structures (ectoderm) (ci, di, dii), adipocyte-like (cii, diii), fibroblast-like (dv), chondroblast-

like (ciii, div), and smooth muscle-like structures (civ) (mesoderm), and an epithelial-like structure (cv, dvi) (endoderm). The tissues were stained with hematoxylin and eosin. Bars represent 100–200 μm .

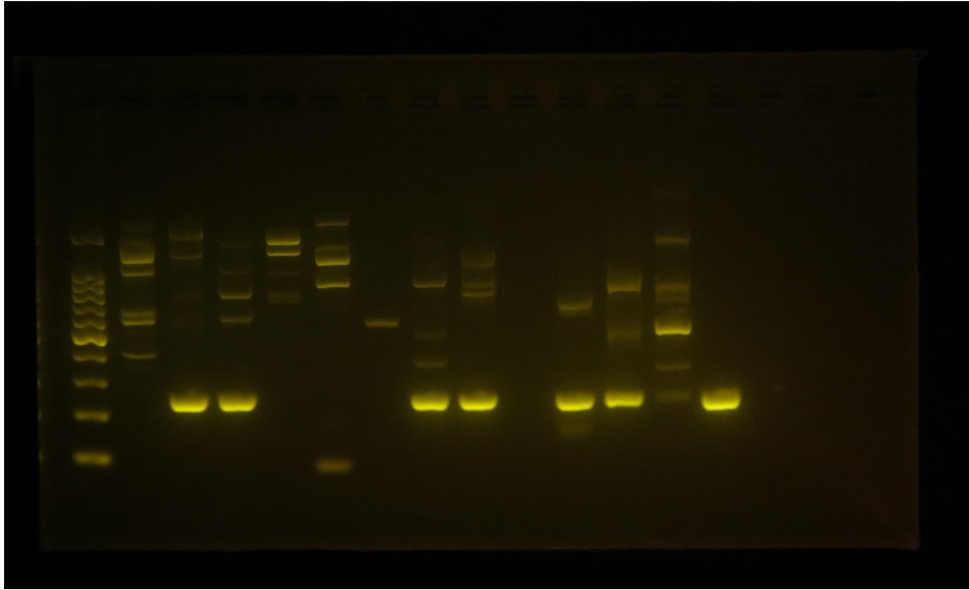


Supplementary Figure 15. Formation of the chimeric embryo after injection of Japanese ptarmigan-derived iPSCs into the chicken blastoderm.

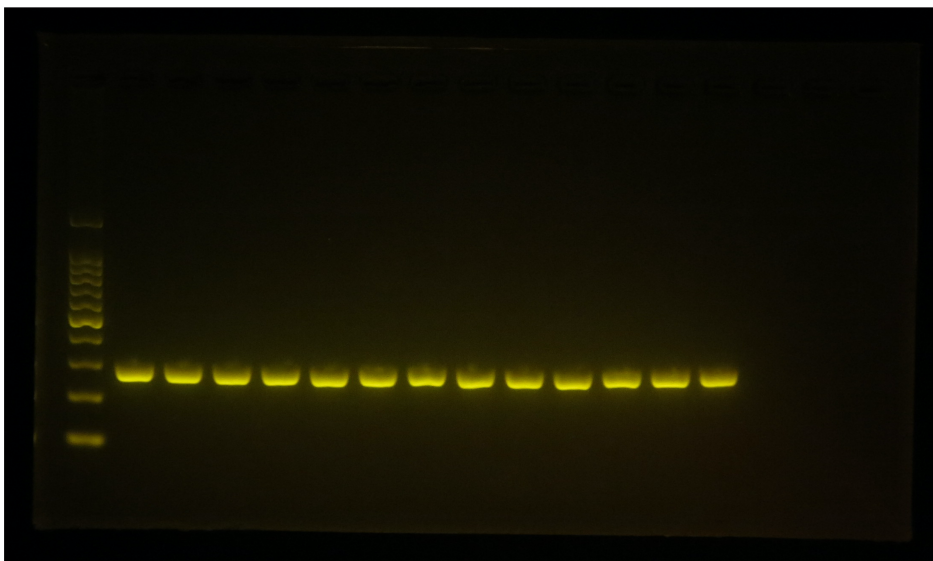
a: The developmental process of Japanese ptarmigan-derived iPSCs injected into chicken embryos. The panels show the time after injection: 0 h (i), 72 h (ii), and 120 h (iii).

b: Embryos exhibited iPSC contribution after injection. The arrow indicates green signal. Panels represent bright field and green fluorescence image (i, v), bright field (ii, vi), green fluorescence (iii, vii), and red fluorescence (iv, viii). Bars represent 2 mm.

a PB-TAD-7F

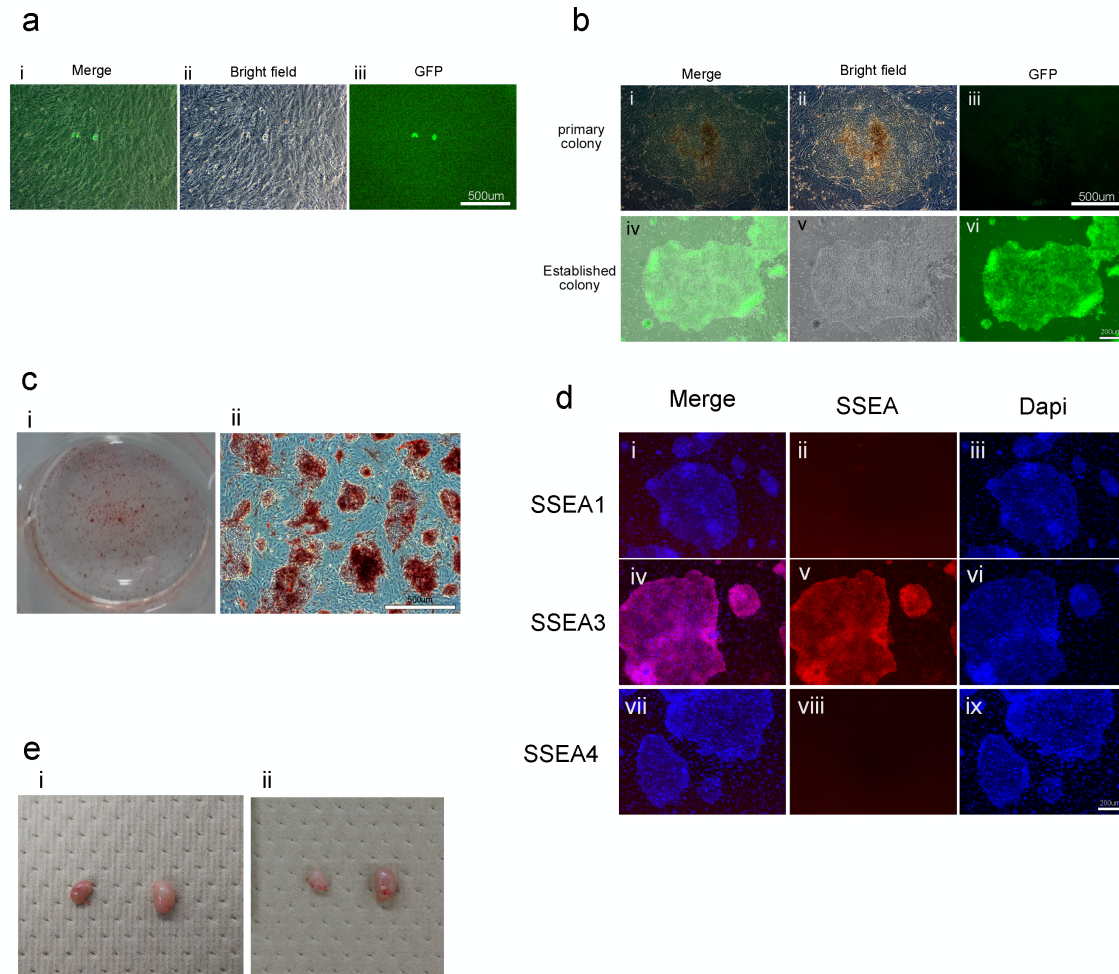


b Tsc-2 (internal control)



Supplementary Figure 16 Detection of transduced genes by genomic PCR (full scan blot).

a: Reprogramming vector for PB-TAD-7F; b: Tsc-2 gene (internal control). Detailed information can be found in Figure 8f (a high-magnification image of this gel).



Supplementary Figure 17 Establishment of Japanese golden eagle reprogrammed cells with PB-TAD-7F reprogramming vector.

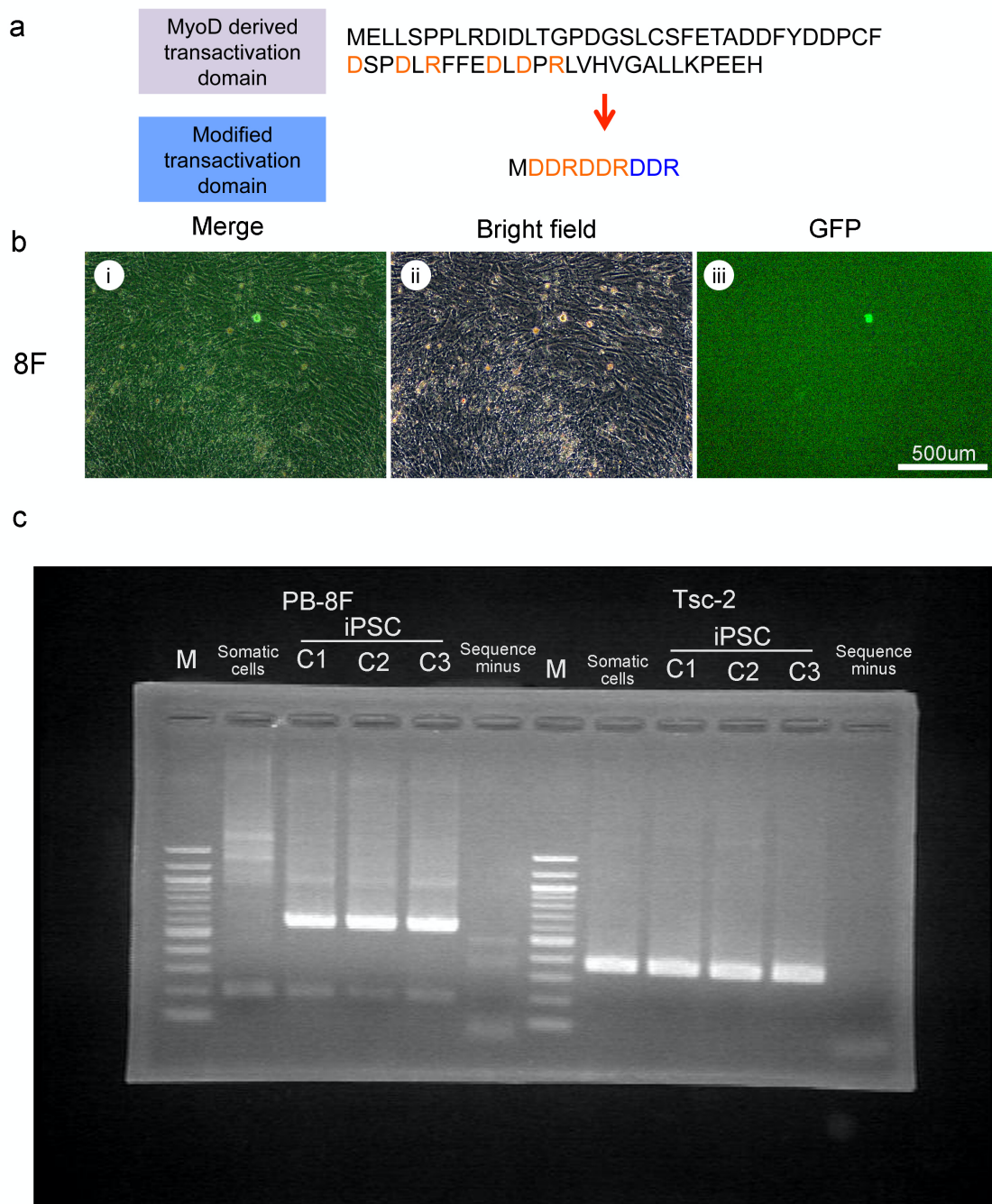
a: Transduction of the reprogramming vector into Japanese golden eagle cells after 72 h. The panels show a merged image (i), a bright field image (ii), and a GFP image (iii). The bar indicates 500 μm .

b: Morphological features of Japanese golden eagle-derived primary and established iPSC colonies using PB-TAD-7F. The panels show a primary colony of reprogrammed cells using PB-TAD-7F (i-iii) and iPSC colonies established using PB-TAD-7F (iv-vi). Panels show merge images (i, iv), bright field images (ii, v), and GFP images (iii, vi). The bars indicate 500 μm (primary colony) and 200 μm (established colony).

c: Alkaline phosphatase staining of Japanese golden eagle iPSCs established using PB-TAD-7F (i, ii). The bar indicates 500 μm .

d: Pluripotency marker staining. The panels show SSEA-1 (i-iii), SSEA-3 (iv-vi), and SSEA-4 (vii-ix). The panels show merged images (i, iv, vii), red fluorescence (iv, v, vi), and counterstained with DAPI (iii, vi, ix). Bar indicates 200 μm .

e: Macroscopic analysis of tumors formed in the testis tissue of SCID mice derived from Japanese golden eagle iPSCs. The images show the testicular tissues of SCID mice with iPSCs established from PB-TAD-7F (i, ii).



Supplementary Figure 18. Establishment of Japanese golden eagle iPSCs using PB-DDR-8F reprogramming vector.

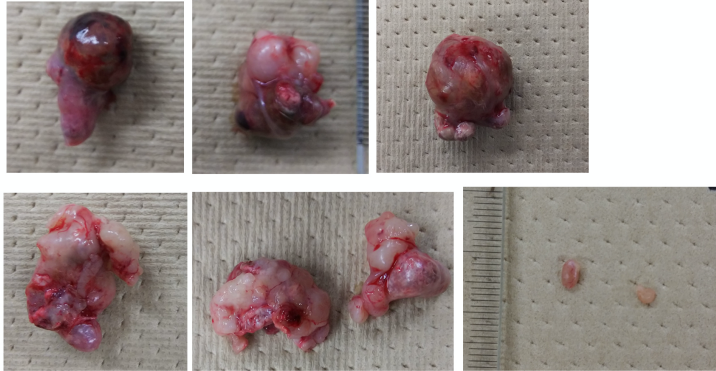
a: Amino acid sequence of the MyoD-derived transactivation domain (the domain used in the PB-TAD-7F vector) and modified transactivation domain (the domain used in the PB-DDR-8F vector).

b: Transduction of the reprogramming vector into Japanese golden eagle cells after 72 h. Panels show transduction of the PB-DDR-8F reprogramming vector. The panel shows the merged image (i), a bright fields image (ii), and GFP image (iii). The bar indicates 500 µm.

c: Detection of the reprogramming cassettes of PB-DDR-8F by genomic PCR. The left six lanes show

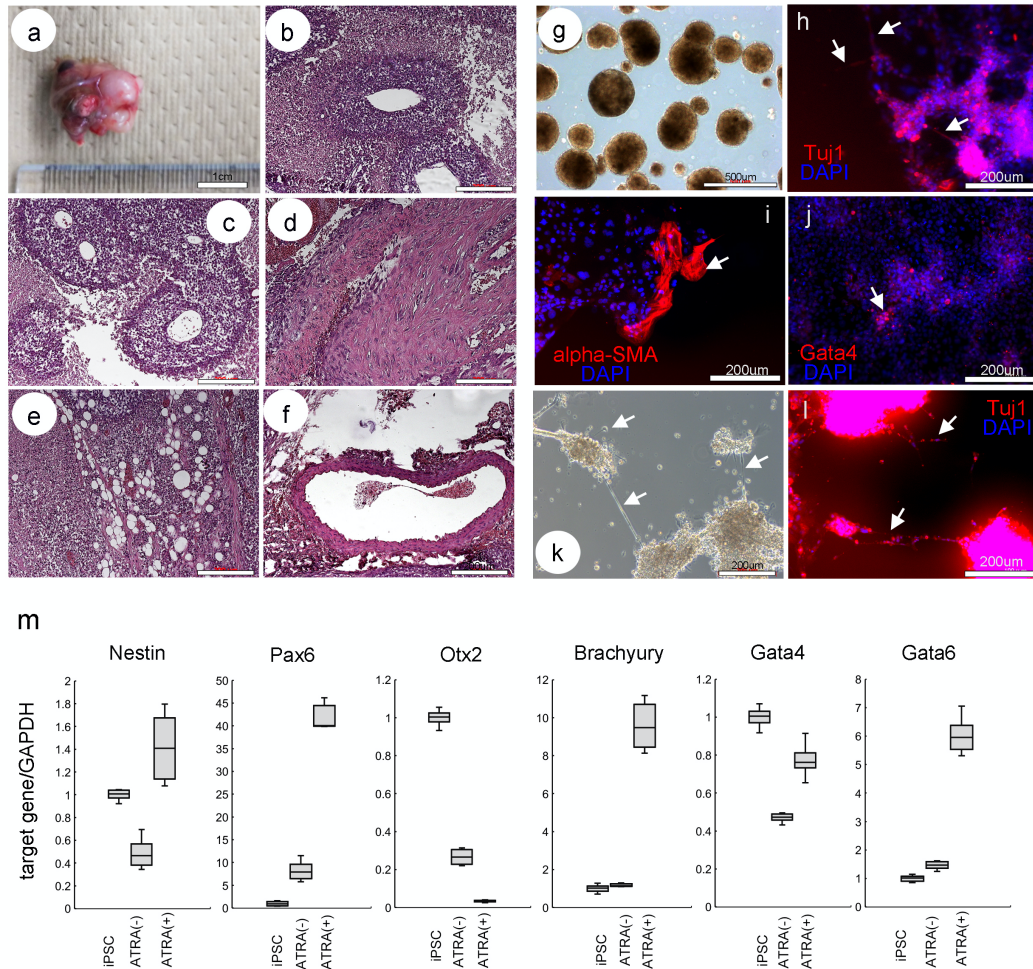
detection of the PB-DDR-8F vector. The six right lanes show the detection of the *Tsc-2* gene (internal control).

DDR-DDR-DDR-8F



Supplementary Figure 19. Macroscopic analysis of tumors formed in testis tissue of SCID mice from Japanese golden eagle iPSCs.

Images show testis tissues of SCID mice with iPSCs established from PB-DDR-8F.



Supplementary Figure 20. Differentiation ability of established Japanese golden eagle-derived iPSCs.

a: Representative appearance of tumors in the testes of SCID mice. The bar indicates 1 cm.

b–f: Histological analysis of the tumors formed in the testis tissue of SCID mice. Shown are neural tube-like structures (b and c; ectoderm), fibroblast-like tissue (d, mesoderm), adipocyte-like tissue (e, mesoderm), and epithelial-cell-like tissue (f, endoderm). Bars indicate 200 μ m.

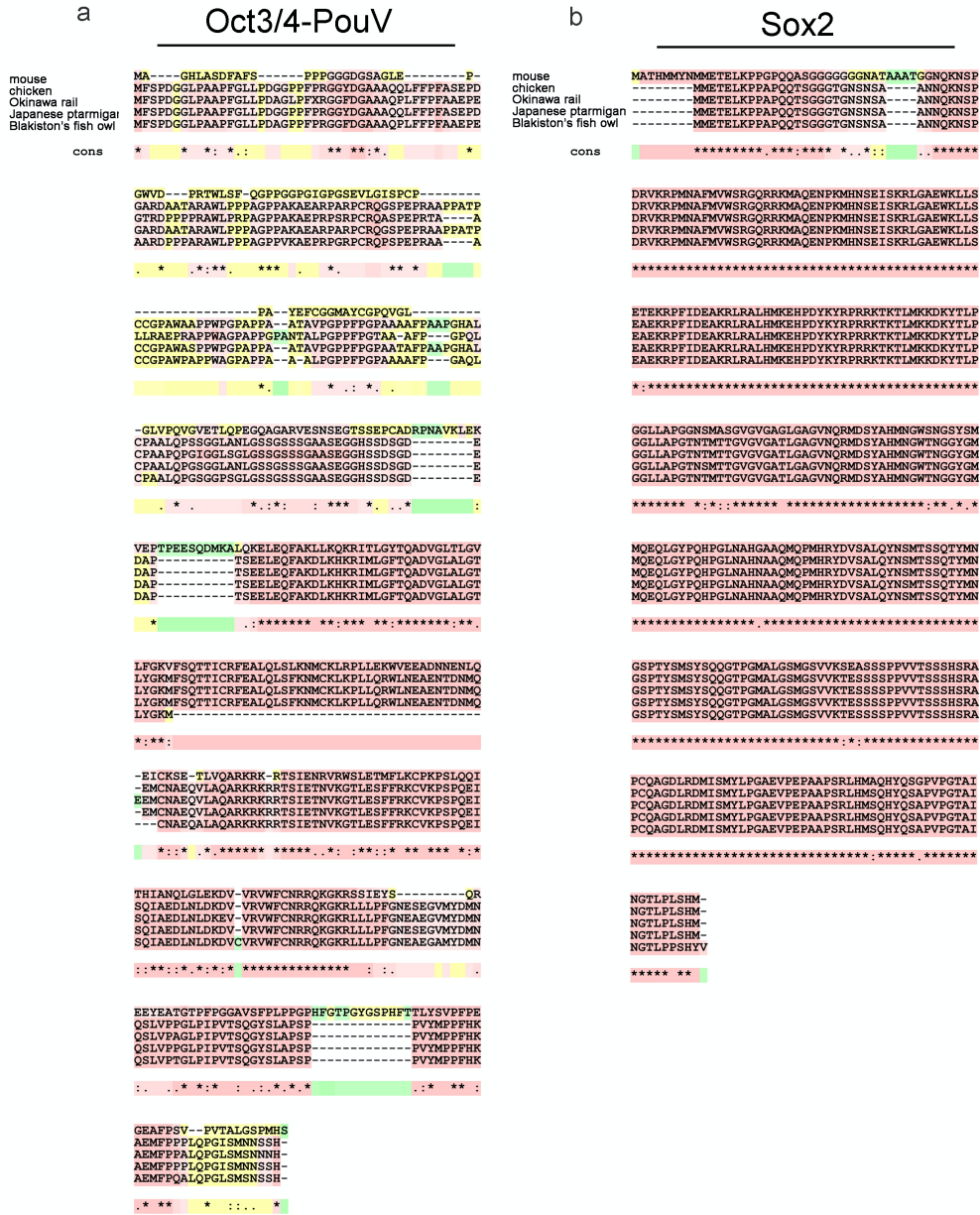
g–l: Immunocytochemical evaluation of the in vitro ability to differentiate into the three germ layers. Shown are ectodermal markers (Tuj1 and g), mesoderm markers (alpha-SMA, h), and endoderm marker (Gata4; i). Bars indicate 200 μ m. (j) Cytomorphology of the Japanese golden eagle iPSC-derived EBs. Bars indicate 500 μ m. (k l) In vitro differentiation of Japanese golden eagle iPSCs in ATRA+ medium. (k) Cytomorphology of neuron-like cells. (l) Immunological staining for the neuron-like cell marker (Tuj1). Bars indicate 200 μ m.

m: Relative mRNA expression of three germ layer differentiation-related genes evaluated by quantitative RT-PCR in differentiated cells. Bars show gene expression in the Japanese golden eagle iPSCs, differentiated cells in ATRA medium, and differentiated cells in ATRA+ medium. *: $P < 0.05$. Gene expression was quantified relative to that of the *GAPDH* internal control. n=6.

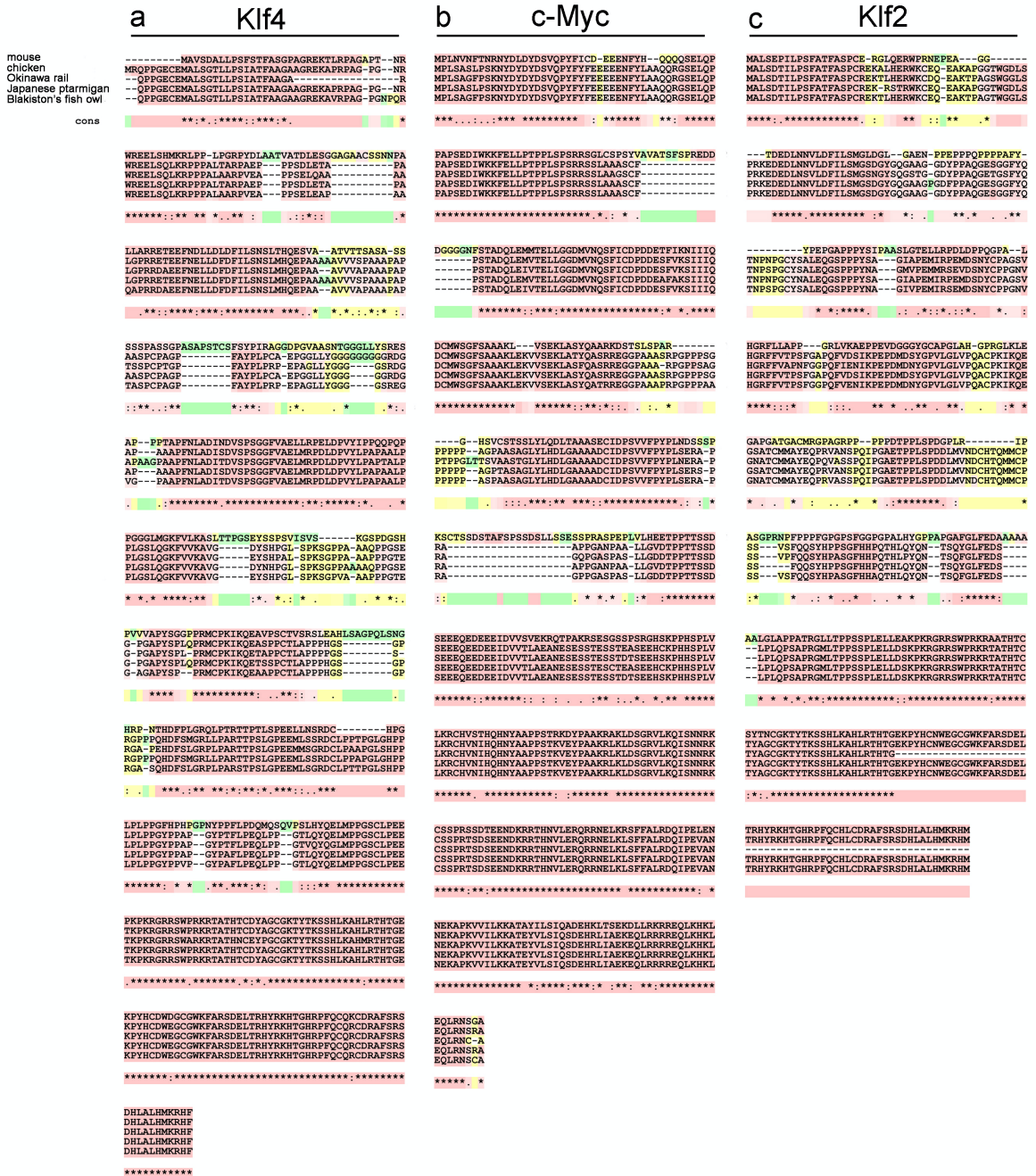


Supplementary Figure 21. Comparison of amino acid homologies of the DNA binding domains of the seven reprogramming factors.

a: Oct3/4 (avian POU5), b: Sox2, c: Klf4, d: c-Myc, e: Nanog, f: Lin28a, g: Klf2

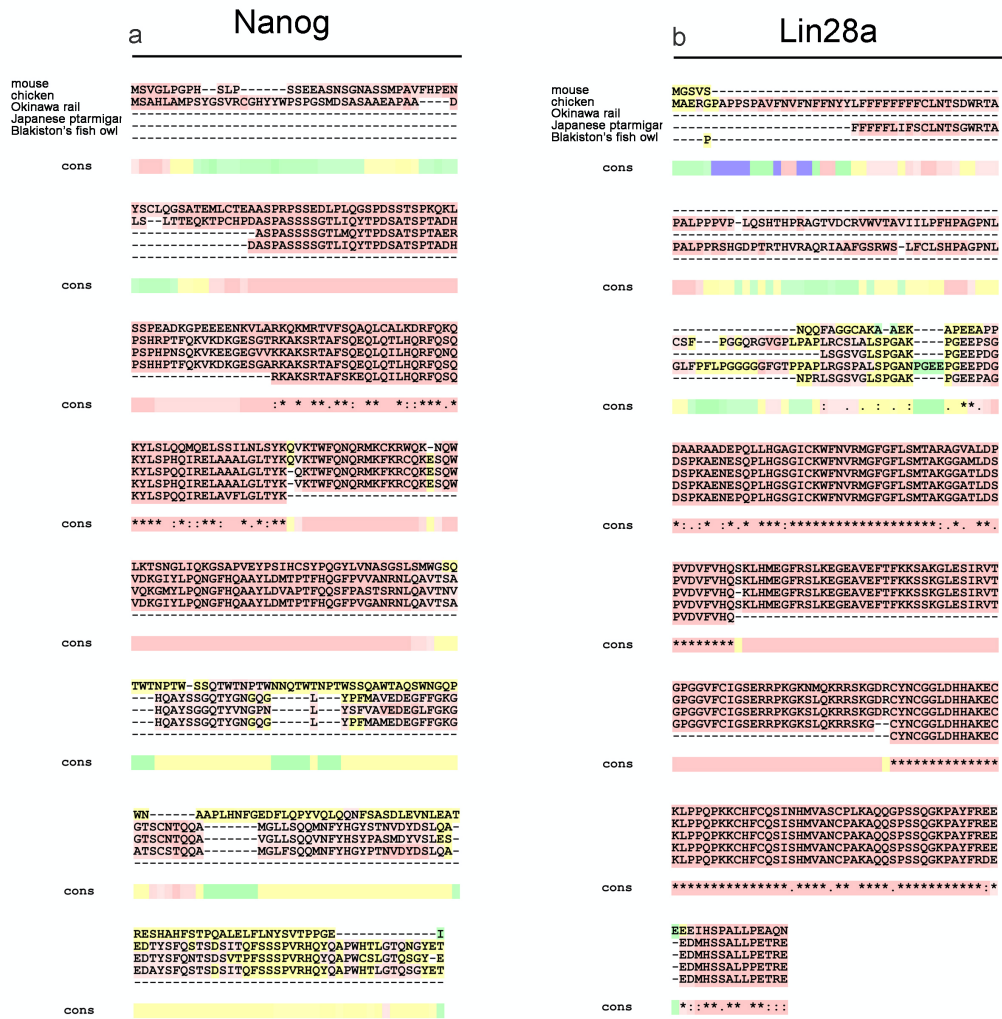


Supplementary Figure 22. Comparison of amino acid homologies for Oct3/4 (Avian: Pou5) and Sox2 among mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston’s fish owl.
 Comparison of amino acid homologies for Oct3/4 (a) and Sox2 (b).



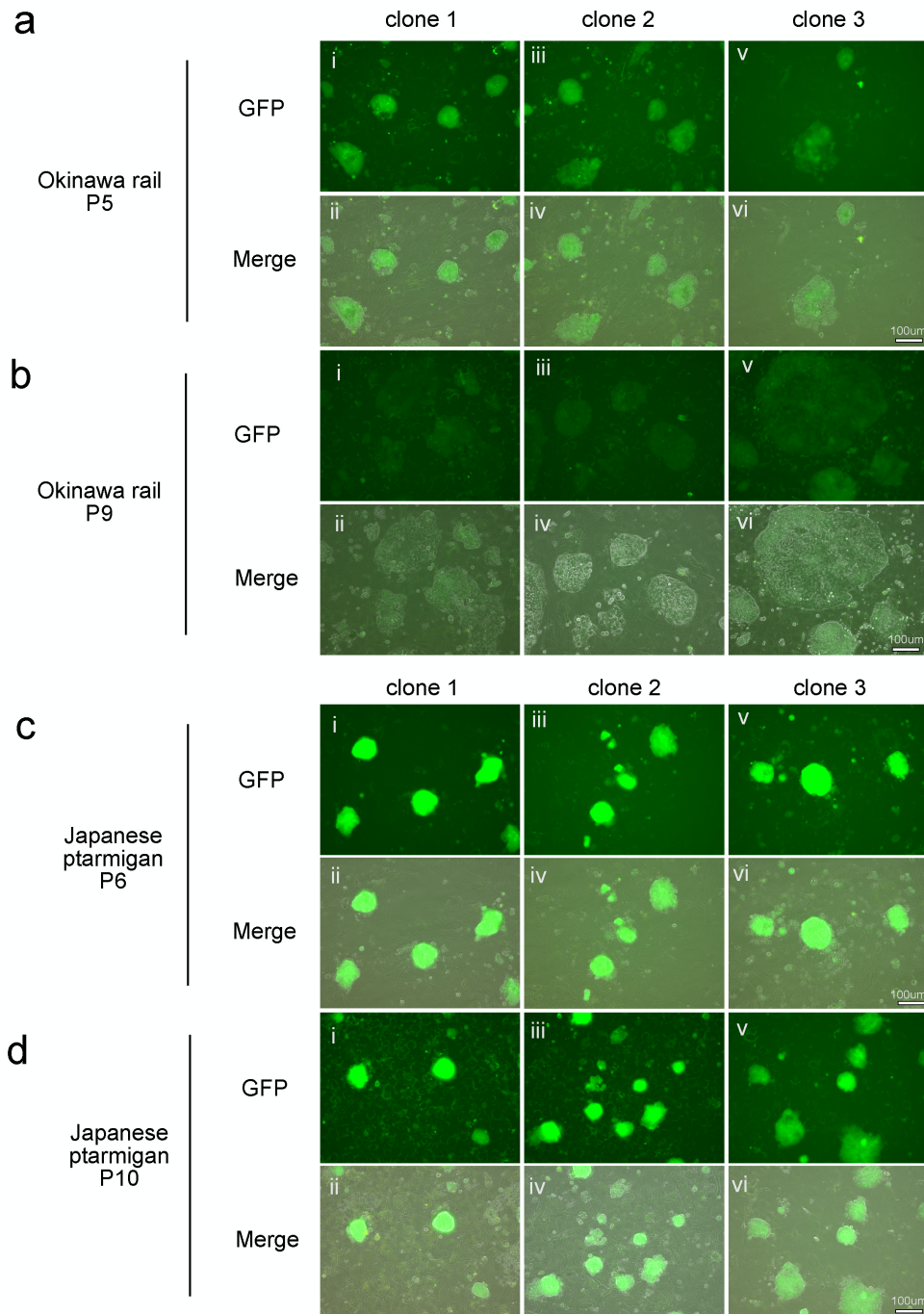
Supplementary Figure 23. Comparison of amino acid homologies for Klf4, c-Myc, and Klf2 among mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl.

Comparison of the amino acid homology of Klf4 (a), c-Myc (b), and Klf2 (c).



Supplementary Figure 24. Comparison of amino acid homologies for Nanog and Lin28a among mouse, chicken, Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl.

Comparison of amino acid homologies of Nanog (a), and Lin28a (b).



Supplementary Figure 25 Exogenous gene expression of Okinawa rail and Japanese ptarmigan.

a: Okinawa rail iPSCs passage 5 day7 after the previous reseeded. b: Okinawa rail iPSCs passage 9 day4 after the previous reseeded. c: Japanese ptarmigan iPSCs passage 6 day7 after the previous reseeded. d: Japanese ptarmigan iPSCs passage 10 day4 after the previous reseeded. Panels show clone 1 (i, ii), clone 2 (iii, iv), and clone 3 (v, vi). The images are GFP (i, iii, v), and merge image (ii, iv, vi). Bars show 100 µm.

Supplementary Table 1 Karyotype analysis of Okinawa rail, Japanese ptarmigan, and Blakiston's fish owl.

Species	Clone No.	Chromosome number																			total		
		70	71	72	73	74	75	76	77	78	79	80	81	82	83	84	85	86	87	88		89	90
Okinawa rail	1							1	5	9	13	10	5	3	3	1							50
	2	1	2	2	3	6	6	8	7	8	4	1		2									50
Japanese ptarmigan	1												5	10	6	9	6	5	1	6	2		50
	2											5	5	8	5	3	4	6	4	10			50
Blakiston's fish owl	1							2	9	17	14	5	2	1									50
	2										4	3	2	41									50

Supplementary Table 2 Antibody information

name	Company	Isotype	Cat No.	Dilution
SSEA-1	STEMGENT, Cambridge, MA, US	Mouse IgM	09-0005	1:200
SSEA-3	Bioss, Boston, MA, US	Rabbit IgG	bs-3575R	1:200
SSEA-4	STEMGENT, Cambridge, MA, US	Mouse IgG3	09-0006	1:200
anti-betaIII tubulin (TuJ-1)	R&D systems, Minneapolis, MN, US	Mouse IgG2A	55461211	1:200
Anti-alpha-Smooth Muscle Actin	Novus, Centennial, CO, US	Mouse IgG2A	NB120-18147	1:100 - 1:200
Anti-GATA4	LifeSpanBiosciences, seattle, WA, US	Rabbit IgG	LS-C352237-100	1:100 - 1:200
Anti-GFP	MBL, Nagoya, JAPAN	Rabbit IgG	598	1:100 - 1:200
anti-HPT 2	Biorbyt, cambridge, UK	Mouse IgG	ORB383723	1:200
anti-vimentin	Thermo fisher Scientific, Waltham, MA	Mouse IgG	MA5-11883	1:200
Goat anti-Mouse IgG, Alexa Fluor 488	Thermo fisher Scientific		A11001	1:200
Goat anti-Mouse IgM Alexa Fluor 568	Thermo fisher Scientific		A21043	1:200
Goat anti-Rabbit IgG, Alexa Fluor 568	Thermo fisher Scientific		A11011	1:200
Goat anti-Mouse IgG, Alexa Fluor 568	Thermo fisher Scientific		A11004	1:200
Anti-IgG (H+L chain) (Mouse) pAb-HRP	MBL		330	1:200
Anti-IgG (H+L chain) (Rabbit) pAb-HRP	MBL		458	1:200

Supplementary Table 3 Primer sequence information for genomic PCR (Figure2a)

Species	Gene designation	F/R	Sequence	Product length (bp)
Mouse	Tsc2	Forward	TGGCATCTCCAAAACACTGCAT	640
		Reverse	TATTCCTGCGACAACCTGACC	
Chicken	Tsc2	Forward	CCGTTTGCTTGCCCTCTGATGACT	294
		Reverse	CAGCTCAGTGTTGTTCTCGGCATT	
Okinawa rail	Tsc2	Forward	TCCAGCCATCTCAGCCGCTCA	345
		Reverse	CTGCTGTTCTGCTCTGAAGGCAT	
Japanese ptarmigan	Tsc2	Forward	CCTACCTTGTGCTGAGATGCT	653
		Reverse	TACAGACCGGCTCCATTGCTT	
Blakiston's fish owl	Tsc2	Forward	ACATCCCCACAGTTACAAAATACGC	535
		Reverse	GCTCGGGAACCCACCCTT	
Reprogramming vector	PB-TAD-7F	Forward	CTAGCGACACCGAAGAGAACGACA	432
		Reverse	CGTTTCTAGGCCACCGTTCCTGT	

Supplementary Table 4. Primer information for genomic PCR

Species	Gene name	F/R	Sequence	Product length (bp)
Japanese Golden eagle	Tsc2	Forward	CTCGCTGTTTTGTCCTCAGTGTTG	363
		Reverse	ACTATCCCCACCTTACTATGGTC	
Reprogramming vector	PB-DDR-8F	Forward	ACTGGACCCTAGGTTCCGCAT	591
		Reverse	TCTAGGGTCCCCAGGTCAACGTTT	

Supplementary Table 5 Primer sequence information for real-time PCR (mouse)

Species	Gene name	F/R	Sequence	Product length (bp)
Mouse	Oct3/4	Forward	CCATGCATTCAAAGTGGAGCACA	132
		Reverse	ACCCCTGTTGTGCTTTAATCCCT	
	Sox2	Forward	GCACATGGCCCAGCACTACCAG	148
		Reverse	TTGCACCCCTCCCAATTCCCT	
	Nanog	Forward	CCCTTACCCACGCCAC	132
		Reverse	TCGAGAGTAGCCACCATATCGTT	
	Tbx3	Forward	ATTCTTGTTTTGCCTTTGGGTCT	110
		Reverse	AGCCAGCTCTACTTGAAAGCA	
	Klf4	Forward	ATCACCTTGATGCTCTTTGCCT	136
		Reverse	ACTTTAGAACACGACTCACCA	
	Klf2	Forward	ACCTGGCCTTGACATGAAGC	177
		Reverse	AACTGGTGGCAGAGTCATTTTCCT	
	Lin28a	Forward	GACACTCCCTGGCTCTCCCA	113
		Reverse	CCCAGCACCCCAACAAGGTTT	
	Esrrb	Forward	GTATGCTATGCCTCCCAACGA	92
		Reverse	TGAGGAACACAAGCTCCCGAT	
	Fbx15	Forward	ATGAGGACAAGAAGCCATC	113
		Reverse	TCCTACGCTGTCCATGTACTCG	
	Prdm14	Forward	CCCATGAACCTCCGTGCTCACT	100
		Reverse	ACACCTTCCACAGCGTTCACA	
	CDH1	Forward	GCCCTCAATTACCATGTTTGCT	138
		Reverse	GCACCCACACAAGATACCTG	
	DDX4 (VASA)	Forward	CAGTACCTATGTGCCTCCAG	110
		Reverse	CCGCTGTATTCAACGTGTGCTT	
	Dazl	Forward	TGAGGCTCCAAATCAGTGT	86
		Reverse	CTTCTGGCAAAACATATCCTTGACT	
	TERT	Forward	TCATCATGTAGAGCGAGCCTT	105
		Reverse	ACAACTGCCTTACCATAAGTCA	
	LIF	Forward	CTTCTATACAGCCAGCACCA	149
		Reverse	TGTGCAGCTTGTTCATACACC	
	LIFR	Forward	GATTGAGCCAAAGAACCCTCC	120
		Reverse	CCATACAACGCTGAGAAAGCC	
	FGF3	Forward	ACAGCGCTATAGCATCCTGGAGA	119
		Reverse	ATCCGAAGCATAAGCCGCTCT	
	FGF5	Forward	TTCAAGCAGTCCGAGCAACCG	146
		Reverse	CATCAAAGCGAAACTTCAGTCT	
	FGF8	Forward	AGCCCACTCCCTGCTCCGA	113
		Reverse	CGTCACCAGGCTCTGCTCCCTC	
	FGF10	Forward	ATGTGCGGAGCTACAATCACC	96
		Reverse	CCTTGCCGTTCTTCTCAATCGT	
	Wnt3a	Forward	TGGCCCTTTCCAGTCTT	148
		Reverse	AGCTTTAAAAGAGTACCAGGCA	
Wnt5a	Forward	AGTTTTCTCTGGCTTGCTCT	149	
	Reverse	CCTCCGAGTCTTAGCTTGCTT		
Gapdh	Forward	CCCAATGTGTCCGTCGT	92	
	Reverse	CCTCAGATGCCTGCTTCACC		
c-myc	Forward	CCTCACTCAGCTCCCCTCC	105	
	Reverse	AGCTTCTTTTACTGCGACTCAGG		
Sox3	Forward	CCCCAAGAAGTCAATGCCTCCAC	142	
	Reverse	AAACACACGCACCTGGCTA		
Lin28b	Forward	AAATTGCGTTAGAAAGAACAAGGA	117	
	Reverse	ACATTGCGTACAAAGCAGTCC		
Tfcp2l1	Forward	AGATCAAGGTGTTCAAGCCCAA	90	
	Reverse	TCTCTTCTCTTGAGCCGTTT		
Sall4	Forward	CTTACCACGAAAGGCAACCTG	144	
	Reverse	GAAACACCTCGGGCGCTCT		
Rex1	Forward	CCCACAGCCATCACTCCAGA	132	
	Reverse	TGGACGAACAGAAGTGTGAGACT		
FGF2	Forward	AGAAGAGCGACCCACACGTC	125	
	Reverse	CTTAGAAGCCAGCAGCCGTTCA		
Dnmt3b	Forward	AGTACCCATCAGTTGACTTGAGC	146	
	Reverse	ATCTTCCCCACAGAGTCCAC		

Supplementary Table 6 Primer sequence information for real-time PCR (chicken)

Species	Gene name	F/R	Sequence	Product length (bp)
Chicken	Pou5	Forward	TCCGCAAAATGTGTGAAGCCAGT	114
		Reverse	CTTGCTTTCTGACGCCGGTT	
	Sox2	Forward	GCCCGCTCCGCTTC	82
		Reverse	TTTCTAGCTCGGTTTCCATCATGTTGT	
	Nanog	Forward	TCTGGGGCTCACCTACAAGCA	85
		Reverse	CCACTGACTCTCTTTGGCAAC	
	Tbx3	Forward	CATCGCAGTGACCCGATACCAG	146
		Reverse	CGTCGTACACCCGATGGAT	
	Klf4	Forward	CCGGACTACAAAATGCCAAGGAGT	89
		Reverse	CGTGAATATCCCCACCTCCGAAC	
	Klf2	Forward	AGCGATACCATCCTGCCCTCCTT	88
		Reverse	CCTCTGCTCGACTTCCACCT	
	Lin28a	Forward	CGGTAACAGCCACAACCTCGGAA	80
		Reverse	CTTCGCACCTGCAAACTGCT	
	Esrrb	Forward	AGTCTTACTCTGCTGGGCTCT	103
		Reverse	TCATCTGGTCCAAGCCCTG	
	Fbx15	Forward	TGTTTTCAGTGCCCGAGTTGCAT	127
		Reverse	ACCACACCAACTCAGCACAGAC	
	Prdm14	Forward	AGAGTTTTCAGCTGCATTCTCG	75
		Reverse	AGAAATCAACAGAAAGCCACACA	
	CDH1	Forward	CCTGGCAAGCCGTTTACCACA	103
		Reverse	ACCTGGCTGTCTTCAGGATCCG	
	DDX4 (VASA)	Forward	GACCAACAGCCATCCCTC	127
		Reverse	CCTTCATTAGCACCAAGTGAGC	
	Dazl	Forward	TCTGTTAACCTGAAAACCGTCT	86
		Reverse	TTCTGAAGTGATGCGCCCTCC	
	TERT	Forward	TGCGCAAAGTACATCTACGTGCCTT	95
		Reverse	AATGAACCGGAGCCTTGATGCAAT	
	LIF	Forward	CTCAACCGCTCCACAACACC	119
		Reverse	TTGCTGCTCTCCCGTAGCTC	
	LIFR	Forward	ATGGAGCAAAGATTACCCAC	144
		Reverse	CAACAAGACAGCTCCATCC	
	FGF3	Forward	CGCAGGAAGCTACTGTGCCACCA	132
		Reverse	AGCGAGATCCCGACATCAACAGCA	
	FGF5	Forward	CCCTCCTCCACGCCACCCAA	144
		Reverse	CCACGTCCATCCAGCGTACC	
	FGF8	Forward	CTCTCCATCCTCAGCGGTCT	100
		Reverse	ATCTGGGCTTCATCAACAGG	
	FGF10	Forward	GTGCGGAGCTACAATCACCT	97
		Reverse	TGACCTTGGCTTCTTCTCG	
	Wnt3a	Forward	AACTTTTGGAGGCCAACCCCT	93
		Reverse	AAGGTCAACCCGTCATCCCA	
	Wnt5a	Forward	ACCTGCCCATGACTTCTGGTT	94
		Reverse	ACCCCTTATTACAGAGCCGAGT	
	Nestin	Forward	ATCACATCCAAGAGCCAAACC	147
		Reverse	GCTCTGCACCTCGTCCCA	
	Brachyury	Forward	ACCACCTCCACATCCCACT	99
		Reverse	CCACATGGATGCCAACCGAG	
	Gata4	Forward	TTTCCCGCAGGCTTACCAG	122
		Reverse	ACCAGTTTATGCCGTTATGATGTCC	
	Gapdh	Forward	GAGGCCAGTTCTGTCCCTT	88
		Reverse	ATCAGTTTCTATCAGCCTCTCC	
	c-myc	Forward	AGGAGCACTGTAAGCCCAACAC	142
		Reverse	ACCTGCCACTGTCCAACCTTAGCC	
Sox3	Forward	CGCAACCGCTCCTAAAGCC	85	
	Reverse	GCGAACAAAACACTAGCCCAACCC		
Lin28b	Forward	TTTCAACAAGCCACTAGTTCTCA	104	
	Reverse	AGATGAATAGCCATACGTTCTTCCC		
Tfcp2l1	Forward	CAAAGGAAGAAATGTAAGGCCAA	103	
	Reverse	ACTGCCATCTCCATTCTCTCG		
Nanog-like	Forward	CAGCCAGTAGCAGTCTATCTGTGA	148	
	Reverse	AGCTGTTTGCATGAAGCACCGAA		
Sall4	Forward	CCCGAATCCTTTGCTACCTCAGTCC	148	
	Reverse	CTCCTCTGCGCTCGATTGTCT		
Rex1	Forward	AGGAGCCTCGTGCCCAATTCCC	142	
	Reverse	CCTCTGCCTTCAAGCTCCCTGACC		
Sox17	Forward	TGGCGCAGCAGAACCAGGCTT	72	
	Reverse	ACAGCGCTTCCACGATTACCCA		
Gata6	Forward	GCGTGACCCACTGCCAGCAA	85	
	Reverse	ACCCTCCGAGAAAGAAACCC		
Otx2	Forward	CCCTGTTCCCAAGACCCGCTA	94	
	Reverse	AACCACACCTGCACTCTGGAC		
Pax6	Forward	AGCACAAAGCTTTACCAGCCGAT	90	
	Reverse	CGCTGTGTCTGTCTGCCAAC		
FGF2	Forward	ATCCGCACATCAAAGTGCAGCTTC	75	
	Reverse	AGCGGTTTGCACCTTACGCCTT		
Dnmt3b	Forward	GGTCCCCTCAGTCACATCCTC	105	
	Reverse	CTTTGTTTGCTTCCCAAGTCTC		

Supplementary Table 7 Primer sequence information for real-time PCR (Okinawa rail)

Species	Gene name	F/R	Sequence	Product length (bp)
Okinawa rail	Pou5	Forward	GCCGGACCAGCATTGAAACCAAC	79
		Reverse	TCCTGGGGACTGGGCTTCACACA	
	Sox2	Forward	ACCCGGAGGAAAACCAAGACC	153
		Reverse	TGCGCGTAACTGTCCATCCTCT	
	Nanog	Forward	CAGAGCGCCATCTCCCAT	75
		Reverse	ACTTGGCCTTCTCACCACACC	
	Tbx3	Forward	AGAAGGCCAAGTACATTTTGCTGA	93
		Reverse	CCTTGCCAGCGACCATCCAG	
	Klf4	Forward	CCCGATCCGATGAACCTACTCGT	75
		Reverse	TGTCACACCGCTGGCACT	
	Klf2	Forward	ATTAAGCCCGAGCCTGACATGGAC	91
		Reverse	TCGCGTTGCCCTCCTGT	
	Lin28a	Forward	TGAAGCCGTGGAGTTCACCTT	112
		Reverse	CTTGCCCTTGGGTCTCCTCTCG	
	Esrrb	Forward	TGGAAATCCTCATCTGGGCAT	75
		Reverse	TGTAGTCTCAGCATAGACGAGCTT	
	Fbx15	Forward	AAATCTGAACAACAGAAACCGT	75
		Reverse	TTTCTCCAGTACCCAGGCTT	
	Prdm14	Forward	AGCAGAATCTGACGGCTATCCA	118
		Reverse	CCCAGGAATGCACGTAGCAA	
	CDH1	Forward	CGCTCAGCTCCCTCAACTCTCC	103
		Reverse	CCGCCATAGAGGTCTGCCAGTTC	
	DDX4 (VASA)	Forward	CAACTCAGTGGCAGCAA	129
		Reverse	TGCCGTGATTTCCACAACGA	
	Dazl	Forward	AGTCTCAATCTCTGCACCAC	103
		Reverse	ATACTGTGTTATGGGGCTCACCAC	
	TERT	Forward	TGAAGCTAACGACTCCAGCAC	133
		Reverse	TGCTCATGTTCTGTACCCAT	
	LIF	Forward	CATCTTCTCCCGCTTTCCG	111
		Reverse	CGCGTGATGTTCCCAACGA	
	LIFR	Forward	ACCCATGATACATCTGAGCAA	104
		Reverse	ATCTTCAGACAGTATGCACACC	
	FGF3	Forward	TGCCAAACAACATAAATGCTT	88
		Reverse	TTGCCCTTCTTTGTCTTCCA	
	FGF5	Forward	GAGCTTGCCCTCACCGTGACC	127
		Reverse	ACCCAAGCGAAACTCAGCCGGTA	
	FGF8	Forward	ACTGGCTTTCACAGAGATCGTCT	81
		Reverse	AGGCCATGTACCAGCCCTCGT	
	FGF10	Forward	TTCCCTCGTCTTCTCTCT	142
		Reverse	CGCTAACCTTGCCGTCTTCTCG	
	Wnt3a	Forward	AACTTTGTGAGCCCAACCT	103
		Reverse	CGCAGCACAAAGGTCCACAACCA	
	Wnt5a	Forward	TCCTGTAGCCTGAAGACCTGT	87
		Reverse	AGCAGCACTATCGTATTTCTCT	
	Nestin	Forward	TTCCCTGCTCCCGGTGGCTT	100
		Reverse	TCTGTAGGTCTCCACCGCCCTT	
	Brachyury	Forward	ACCACCTGCTGAGCGCCGTG	121
		Reverse	AGCTCTTGAACCGCAGCCACA	
	Gata4	Forward	TTTCCCGCAGGCTTACCAG	86
		Reverse	AGCTAAGACCAGGCTGTCCA	
	Gapdh	Forward	TTATCATCTCTGCCCCCTC	82
		Reverse	ATTTTCAGAGACTGTCTACTTGT	
c-myc	Forward	TGCTTCCCTCCACCGCCGACCA	142	
	Reverse	AGCCGCTCCACATGCAGTCCT		
Sox3	Forward	ACCTATATGAACCGCCTTCCACCT	103	
	Reverse	ATTTACCACCGAGCCATGGAG		
Lin28b	Forward	ACTTCTGCTTACCAGGGGA	136	
	Reverse	ACGCAGACAACCTACTCGAC		
Tfcp2l1	Forward	GCATCAGCACCGAATTTACTCCAC	141	
	Reverse	ACACTTGTATCTGGCAGCTTG		
Nanog-like	Forward	TGACGGCTGTAACCTCCACCAC	100	
	Reverse	GCCCCACATCCAGAGTGCTT		
Sall4	Forward	ATTGCAGCCCTGGAGAACCAA	88	
	Reverse	AGCCATTGCTCACTAGACTTCAAGC		
Rex1	Forward	CAGCCTTTGCCATCCAGT	105	
	Reverse	GCGTTTCACTTCTTATCCCT		
Sox17	Forward	AGCACATGCAGGACCAACCCAAAC	94	
	Reverse	TGCTGCAGGAAGCCGCTCTCCAC		
Gata6	Forward	ACCAGTCTGAAAAGCAGCAAC	85	
	Reverse	TCACTCAGCCGTGGAGGTCA		
Otx2	Forward	ACCTACACCCAGGCATCAGGCTA	149	
	Reverse	ACCGGTTGGCACCCAT		
Pax6	Forward	CGAAACTGGCTCCATCAGACC	93	
	Reverse	CGTTTACTGCGCTATTTTGTCT		
FGF2	Forward	TCTTACTGCAAGAACGGCGGCTT	82	
	Reverse	AGGGTCGCTTCTCCCGGACG		
Dnmt3b	Forward	TGAAGTACAGGCAATTTATGCTT	145	
	Reverse	ATTGGTTTCTGTATCCTGCAT		

Supplementary Table 8 Primer sequence information for real-time PCR (Japanese ptarmigan)

Species	Gene name	F/R	Sequence	Product length (bp)
Japanese Ptarmigan	Pou5	Forward	AAACCACCATCTGCCGCTTCG	103
		Reverse	TCCGTGTCTCTGCCTCGTTG	
	Sox2	Forward	CCCCGGAGGAAAACCAAGACCCTG	100
		Reverse	CCCTACCCCGGTCTGTCATGC	
	Nanog	Forward	GCACACCAGGTTACAGCAG	89
		Reverse	CCAAAGAACCCTCATCTCC	
	Tbx3	Forward	ACCGATGAGAGATCCAGTGATCCC	90
		Reverse	ACGGCGCTCATGGCAAAGTCC	
	Klf4	Forward	AACTCACTCGTCATTACCGAA	117
		Reverse	ATTAAAGTGCCTTTCATGTGT	
	Klf2	Forward	GGCCGCTCTCTCGTGACACC	87
		Reverse	TGGCCATAACTGTCCATGTCAGG	
	Lin28a	Forward	CTCCCCAGCCTTGCCCT	96
		Reverse	AGCTTTCATTGGCATTGGCTT	
	Esrrb	Forward	ACATGCTTAATGCCATCCCAG	89
		Reverse	TGCCTCACAGGAAGCCACACC	
	Fbx15	Forward	CTATTTCAGTTGTTTCCAGCA	118
		Reverse	TTTCTCCAGTATCCAGGCTT	
	Prdm14	Forward	TGTTTTCTCCTATGTAAAGATCCCC	139
		Reverse	GTGCAAGACAGCTAGTACCTT	
	CDH1	Forward	GCCITCAGCATTATACCGACCC	92
		Reverse	CCCACACTGGAGCCATCGACA	
	DDX4 (VASA)	Forward	TCAGTCTAGAAATTGGGCTT	127
		Reverse	TGACTGCCGTTACTTTGGTT	
	Dazl	Forward	ATAATAAAGTAAGGTGAGGCGTGA	100
		Reverse	CAAAATGGCCGCTCGCAGCAC	
	TERT	Forward	ACCAAAATACATCTTCCGCTCT	141
		Reverse	TCTATAAGTGCCTTCCACCT	
	LIF	Forward	TCGACCCGCTCCACAACACC	110
		Reverse	CTCCCCATAGCTCAGTCCAC	
	LIFR	Forward	CCGCAACAACAGACACC	144
		Reverse	CCACCTGAGATAACCCCA	
	FGF3	Forward	GCATCCCGTCTATACCGGACT	132
		Reverse	CTGTGCTCTGCGTGTITTAAGCC	
	FGF5	Forward	TTCAGATTGCAGCCTTCCA	91
		Reverse	ACAGCACAGAGTACCTCA	
	FGF8	Forward	GTTTGGCACCCTCACCCAT	85
		Reverse	ACCCGCTGCCTCACCGTA	
	FGF10	Forward	CCTTAAATGCTCATTCAAGCCTA	114
		Reverse	TCCATCTCCACAAGGCTTC	
	Wnt3a	Forward	ACAACCTCTTCAAGGCTCCGACT	148
		Reverse	AGGTCAAGCCATCAATCCCA	
	Wnt5a	Forward	TTCTGTAGCTGAAGACCTG	132
		Reverse	CTGTTCACTGCACCAGCTTG	
	Nestin	Forward	CCAACCAAGAGAGTCCAGCATC	89
		Reverse	ATCCTTCTGTCTTCTGTCTGCTT	
	Brachyury	Forward	GTTTCTCCCACTAGCTGGCAA	102
		Reverse	CTCCAGTACATTAGCACTTCG	
	Gata4	Forward	AAACGATAGAATTTGGCTAACTGG	142
		Reverse	ATTTCTACTATGAAAGGGCCGAGA	
	Gapdh	Forward	CCCTTTGTGGAGCCCTT	138
		Reverse	TTCTATCAGCCTCTCCACCT	
c-myc	Forward	AGGAGCACTGTAAGCCCCACCAC	142	
	Reverse	ACCCTGCCACTGTCCAACTTAGCC		
Sox3	Forward	GCACGACCGCTCCTAAAGCC	87	
	Reverse	AGCGAACAAAACACTACGCCCAACC		
Lin28b	Forward	GACATGAAGCTGAACCCCA	75	
	Reverse	GAGATGAATAGCCATACGTTCC		
Tfcp2l1	Forward	ATCTTCAAGCAGGAGGAACCAC	116	
	Reverse	AGGGTTTCTTCGTGACGTTT		
Nanog-like	Forward	AGTTCATGACCATTGGCAAG	127	
	Reverse	ATATATCTGAGTCTGGTGCCAT		
Sall4	Forward	TCCCGAATCCTTTGCTACCTCAGT	150	
	Reverse	GTCCTCCGATTTCGATTGCT		
Rex1	Forward	AGCAAAGCGTTCTAAGTTGTCC	134	
	Reverse	ATTCATTGAAAATCCTGAGGCACT		
Sox17	Forward	TGGCGCAGCAGAACCCGGACCT	72	
	Reverse	ACAGCGCCTTCCACGATTATCCAG		
Gata6	Forward	ACCTCTGTAAAGCCGCGAGT	78	
	Reverse	AATCCCGGTGCCATCCCTCC		
Otx2	Forward	CCCATGACCTACACCCAGGCATC	82	
	Reverse	ATCCACAGTCCATCCCTCCGAA		
Pax6	Forward	TAGCGACTCCCGAAGTTGTAAGCA	82	
	Reverse	CTGTCTCGAATCTCCACGCAAA		
FGF2	Forward	TCAAATCCAGTTCAAGCAG	90	
	Reverse	TGCCATCCTCTTCATAGCC		
Dnmt3b	Forward	ATTCACTACGTCCTTTTCCAAGCA	129	
	Reverse	CAGAACATTTTCCCTCCGGTCA		

Supplementary Table 9 Primer sequence information for real-time PCR (Blakiston's fish owl)

Species	Gene name	F/R	Sequence	Product length (bp)
Blakiston's fish owl	Pou5	Forward	AGAAAAACGACGACCAGCAT	78
		Reverse	ACTGGGCTTACGCACT	
	Sox2	Forward	CTCGCTTCTCTGCTCAAGGCTGGT	121
		Reverse	TTCGAGTTGCCGTGCCTCCC	
	Nanog	Forward	AGAGCCGCACAGCTTCTCCA	103
		Reverse	AAAACAGCCAGCTCCCGGAT	
	Tbx3	Forward	GGTATCCCCTTTTCTCCCTGGGTC	148
		Reverse	CCGTGCCTCTTTATGGAATTGCT	
	Klf4	Forward	GACCACCTTGCTTACACA	122
		Reverse	GTGTTCTCCCATCTTGC	
	Klf2	Forward	ACTTAGCCAGTCATCTCCT	99
		Reverse	CACAGAAGCTGCATAAATACCC	
	Lin28a	Forward	CCGCGAAAGCACAGCAGTCCC	75
		Reverse	AGTGTGCATATCCTCCTCGTCT	
	Esrrb	Forward	CGCCATCATGGAGGACTCAC	89
		Reverse	AAGCAATGTCCCACACACC	
	Fbx15	Forward	AATGAAATTGCTTTTGTATTGTGA	70
		Reverse	CAGAACCCAAAGTGTTCTC	
	Prdm14	Forward	CTCTGGCAACTGGATGTCCCT	92
		Reverse	CGTAGAAGATTGTCCCTGGCACT	
	CDH1	Forward	TGCTCCGCCCGAGGATGACA	75
		Reverse	CCTGATCTCCTCACGCCACCCT	
	DDX4 (VASA)	Forward	GAGAACTCGCTCTTACGAT	75
		Reverse	GACCTCTTGCTGCCACT	
	Dazl	Forward	AAGTGACTAATACTGACCAGC	76
		Reverse	TAAAATACAACTCTGGGCAAA	
	TERT	Forward	TGGAGCAAAGCACAAACCAG	82
		Reverse	TGTCTCATTGCCAGTAGCG	
	LIF	Forward	GGCCTCATCTCAAACCTCACCT	143
		Reverse	TGCACGTACTCTCTGAGCACCT	
	LIFR	Forward	GACTCAGAAACGGAGAACCAC	105
		Reverse	CACCTGAGATGACCCACA	
	FGF3	Forward	CGCAGGAACTCTACTGTGCCACCA	84
		Reverse	GCTGTTTTTCTCCAGGGTGCCGTTG	
	FGF5	Forward	ACACCTATGCCTCAGCTGCCACC	76
		Reverse	TTGCCCTCTTGTGAGTGCCACA	
	FGF8	Forward	ACTGTGTCTTACGGAGATCGTCT	82
		Reverse	AAGCCATGTACCCCTCG	
	FGF10	Forward	ATTAATATGCCATTACGAC	85
		Reverse	TCCTTATCTAGGTATCGTCT	
	Wnt3a	Forward	AACTTTGTGAGCCCAACCT	92
		Reverse	AGGTCGCAACCATCAATCCCA	
	Wnt5a	Forward	TTCAGTACATTTAGGGCTCTGCAA	117
		Reverse	GCTCTTACCCTCCGATGACA	
	Nestin	Forward	AGCAAGCAGCCGAGAAGAAGCCAA	139
		Reverse	TGCACCTCCAGCAAGGTTCCAC	
	Brachyury	Forward	ATCCCTCAATCTACGGTGTACTCT	80
		Reverse	ACATCCATGCTGTCAATGTGCC	
	Gata4	Forward	CACCGTCTTCTACTACCCAA	126
		Reverse	GCCCTAGACCATATCAATCCAC	
	Gapdh	Forward	CCTTTTACCACCGCTTAGCTCT	114
		Reverse	CTATCAGCCTCTCCACTCCC	
	c-myc	Forward	AGCTTCATCTGCGACCCGGACGAC	73
		Reverse	AGCCGCTCCACATGCAGTCT	
Sox3	Forward	TCACAGACCTTCTCCCTGC	89	
	Reverse	TACGGTACCATTGACGCCAGT		
Lin28b	Forward	GCGAAGAATGTAGTCTACCTCC	135	
	Reverse	TTCAGCTTCGTGCTTCCC		
Tfcp2l1	Forward	AACTTGACAGCATTTCTCCAC	80	
	Reverse	TTGCTCACTAATACATGGATCCCT		
Nanog-like	Forward	AACATCCAAACTGTGCCTCCC	87	
	Reverse	GACTCCTCATCTTCGCTGGT		
Sall4	Forward	CGTCATCTTCCAAATCCGCTT	104	
	Reverse	CGGTTGTCTCTCAGACACGCTA		
Rex1	Forward	ACCCACTTTCAAATTACTCAGCC	89	
	Reverse	ATAACTAGACACCTTACCCTT		
Sox17	Forward	GCAGCAAGAGCGAGGCGGGAT	70	
	Reverse	TTGCGCTCGTCTTCCGCCACACC		
Gata6	Forward	CACCAGTCCCAGAAAGCAGCAA	102	
	Reverse	TGCTCACAGATGCCGTCCT		
Otx2	Forward	GCCCTTTTCGCCAAAACCCGCTAC	70	
	Reverse	GCAGGTTGATTTTACGCGCCACTC		
Pax6	Forward	TAGCGACTCCAGAAGTTGAAGCA	116	
	Reverse	ATTATCGTTGGTACAGACCCC		
FGF2	Forward	AGTCAAAGTCAACTTCAAGC	92	
	Reverse	TGCCATCCTCTTCAATAGCC		
Dnmt3b	Forward	GCATGAAATACTCCGTTTTAGCC	108	
	Reverse	ATTCGCTATAGTTCAAGTTTGGC		

Supplementary Table 10. Primer information for qPCR

Species	Gene name	F/R	Sequence	Product length (bp)
Japanese Golden Eagle	Sox3 like	Forward	ACAACCTCCTGCCTCCTAGAAGCC	93
		Reverse	AGCGAATAAAACAGTGCCCAACC	
	Nanog like	Forward	AGCAGAAAACCTTCAGGCTGTGACC	107
		Reverse	CCCTCATCCTCCACAGCCACGAA	
	Lin28a	Forward	CGCCCGTCGATGTCTTCGTACACC	81
		Reverse	ACTCAACAGCTTCGCCCTCCTT	
	Lin28b	Forward	AGGAATCCCAGGTTTTGCAC	75
		Reverse	TGGAGATGAACCCGAATCCC	
	Sox2	Forward	CCCGGAGGAAAACCAAGACCCTCA	103
		Reverse	CCAACCCCTACCCCGGTCGTC	
	Tert	Forward	AGCAACAACCTACGCGTCCTC	100
		Reverse	GCTCCCAAGGGCTGAATCACAC	
	Cdh1	Forward	CACCGACCCAAAACCAACGA	102
		Reverse	ACAGCGTTCTCCACCGTTACCAG	
	Sall4	Forward	TGCCAGAAAACACCTGCGACA	145
		Reverse	TTTGAAGAACTCCTAGGGCCAT	
	Esrrb	Forward	ACCAAAATGAAATGCACACCAC	102
		Reverse	CTTAGGATCTTTTCTGCCCAT	
	Fbxo15	Forward	ATACCATCTTGCTAACGTGAC	90
		Reverse	ACTAAGAGTCTGGCTTCAGA	
	Gapdh	Forward	ACCATCTTCCAGGAGCGTGACCC	134
		Reverse	ACACGCTTAGCACCACCCTT	
	Nestin	Forward	AAGCAACCGAAGCAGGCAAA	83
		Reverse	TAAATGCCTCAAAGTGCCCTTCGT	
	Pax6	Forward	GGACCCACTATCCCAGTGTGT	141
		Reverse	CTGTCTCCGCTGGTTCCTCA	
	Otx2	Forward	ACGCTCCAGTTTITAGTCAGGT	124
		Reverse	AGTCTGAACGGGATCACACCA	
Brachyury	Forward	CAGCACCGGCACAGCTACCAG	101	
	Reverse	CGTTGGACATCCCGCTCGACT		
Gata4	Forward	GCCACCATCTTCTCATTACCCAA	114	
	Reverse	CAAATCCGCTAAAGCCACCAC		
Gata6	Forward	ACCAGTCCCAGAAAGCAGCAAC	103	
	Reverse	CCTGTCTCACCGATGCCGTCA		

Supplementary Table 11 Primer sequence information for genomic PCR (Figure 8f)

Species	Gene name	F/R	Sequence	Product length (bp)
Chicken	Tsc2	Forward	CCGTTTGCTTGCCCTCTGATGACT	294
		Reverse	CAGCTCAGTGTTGTTCTCGGCATT	
Reprogramming vector	PB-TAD-7F	Forward	CAGCCCCAGTCGAGTACCCAT	250
		Reverse	TCCTCCCCGAAGTTGTGCAGT	

Supplementary Table 12 Primer sequence information for real-time PCR (Figure 8g, h)

Species	Gene name	F/R	Sequence	Product length (bp)
Chicken	Tsc2	Forward	TCTGTAAGCCCAAAGAATAAGCA	117
		Reverse	AGCTTATCACCCACACCA	
		probe	[FAM]CACACAGCACCTTAGCTAGAGCAC[BHQ1]	
Reprogramming vector	PB-TAD-7F	Forward	GACAACAATTTGCCGGTTCGAGG	91
		Reverse	GGCTTCTTCCACCACTTCTCCA	
		probe	[FAM]ACATGTGCAAGCTGAGGCCACT[BHQ1]	

Supplementary references

1. Katayama M, *et al.* Induced pluripotent stem cells with six reprogramming factors from Prairie Vole, which is an animal model for social behaviors. *Cell Transplant* **25**, 783-796 (2016).
2. Katayama M, *et al.* Immortalized prairie vole-derived fibroblasts (VMF-K4DTs) can be transformed into pluripotent stem cells and provide a useful tool with which to determine optimal reprogramming conditions. *J Reprod Dev* **63**, 311-318 (2017).