

eMaterials

Patients and controls characteristic

Patients were included in PARALS if they met definite, probable, or probable laboratory-supported ALS diagnosis according to the revised El Escorial criteria¹. Control samples were composed of 350 females and 405 males. The 755 controls were chosen randomly from the same population and matched to the cases by sex, age (± 5 years), and geographic location. Control subjects were excluded from the study if they had a first-degree relative with a neurological or psychiatric disorder.

Standard protocol approvals and patient consents

All subjects participating in the study gave written informed consent. Ethical approval was obtained from the medical ethical review board of the A.O.U Citta della Salute e della Scienza di Torino, Italy.

Whole-genome sequencing

Whole-genome sequencing was performed at The American Genome Center at the Uniformed Services University on the Walter Reed National Military Medical Center campus in Bethesda, MD, USA. Briefly, libraries were prepared using TruSeq DNA PCR-Free High Throughput Library Prep Kit (Illumina Inc.) as per the manufacturer's instructions. Sequencing was performed on an Illumina HiSeq X10 sequencer using paired-end 150 base pair reads, and the data were processed according to Genome Analysis Toolkit's (GATK) best practices (<https://software.broadinstitute.org/gatk/best-practices/>). Variant quality control was performed using the GATK variant quality score method with default filters using Genome Reference Consortium Human Build 38 as the reference.

ALS-related genes

We extracted variant information from the data for the following genes: *ALS2*, *ANXA11*, *ATXN2*, *C21orf2*, *C9orf72*, *CCNF*, *CHCHD10*, *CHMP2B*, *DAO*, *DCTN1*, *DNAJC7*, *ERBB4*, *EWSR1*, *FIG4*, *FUS*, *GLE1*, *GRN*, *HRNPA1*, *HRNPA2B1*, *HNRNPD*, *KIF5A*, *MAPT*, *MATR3*, *NEFH*, *NEK1*, *OPTN*, *PFN1*, *PRNP*, *PRPH*, *SETX*,

SIGMAR1, SOD1, SPG11, SPTLC1, SQSTM1, SS18L1, TAF15, TARDBP, TBK1, TUBA4A, UBQLN2, VAPB, and *VCP*. Prognostic genes (*UNC13A, CAMTA1*) were also extracted.

Variant annotation

Annotation was then performed using ANNOVAR v2020-06-07 (<https://annovar.openbioinformatics.org>) and KGGseq v1.0 (<http://pmglab.top/kggseq/>) using the gnomAD database (version 2.1.1) to determine minor allele frequency (MAF) in the European-derived population. The current study was focused on coding variants due to the limitations in interpreting non-coding variants.

Expansion screening

The samples were screened for *C9orf72* expansions and the microsatellite repeat in *ATXN2* using repeat-primed PCR methodology as previously described.^{2,3} A cut-off of 30 repeated expansion and the characteristic sawtooth pattern was considered pathogenic for *C9orf72*.² *ATXN2* CAG expansions were deemed intermediate if they were within the range of 30-33 repeats³. ExpansionHunter - Targeted software (version 0.3) was used to estimate repeat lengths of known, disease-causing expansions in samples undergoing whole-genome sequencing. This algorithm has been validated using experimentally confirmed samples carrying expansions⁴. The performance of whole-genome sequencing in terms of sensitivity for repeated expansion has been validated in the same ALS cohort^{5,6}.

Variant interpretation

We set a conservative MAF frequency threshold of <0.01% based on the epidemiology of ALS. We defined Loss of Function (LoF) variants when the sequence changes were predicted to be a premature stop codon, a frameshift causing insertion/deletion (indel), or a splice-site disrupting variant located in the canonical splice sites (+1 and +2, -1 and -2) that cause the premature termination codon falling < 50–55 nucleotides upstream of the 3' most exon–exon junctions. Loss of function variants was considered deleterious unless they exceeded the 0.01% MAF threshold. For all other type of variants, the combination of different sets of algorithms was

considered using recommended threshold [MutationTaster, Mendelian Clinically Applicable Pathogenicity (M-CAP), CADD, Variant Effect Scoring Tool (VEST3), Rare Exome Variant Ensemble Learner (REVEL), Meta-analytic Support Vector Machine (Meta-SVM)]⁷. These variants underwent an independent review by clinical and genetic ALS experts and were confirmed to be clinically reportable if agreed by consensus. When available, gene-specific information data was used in the classification

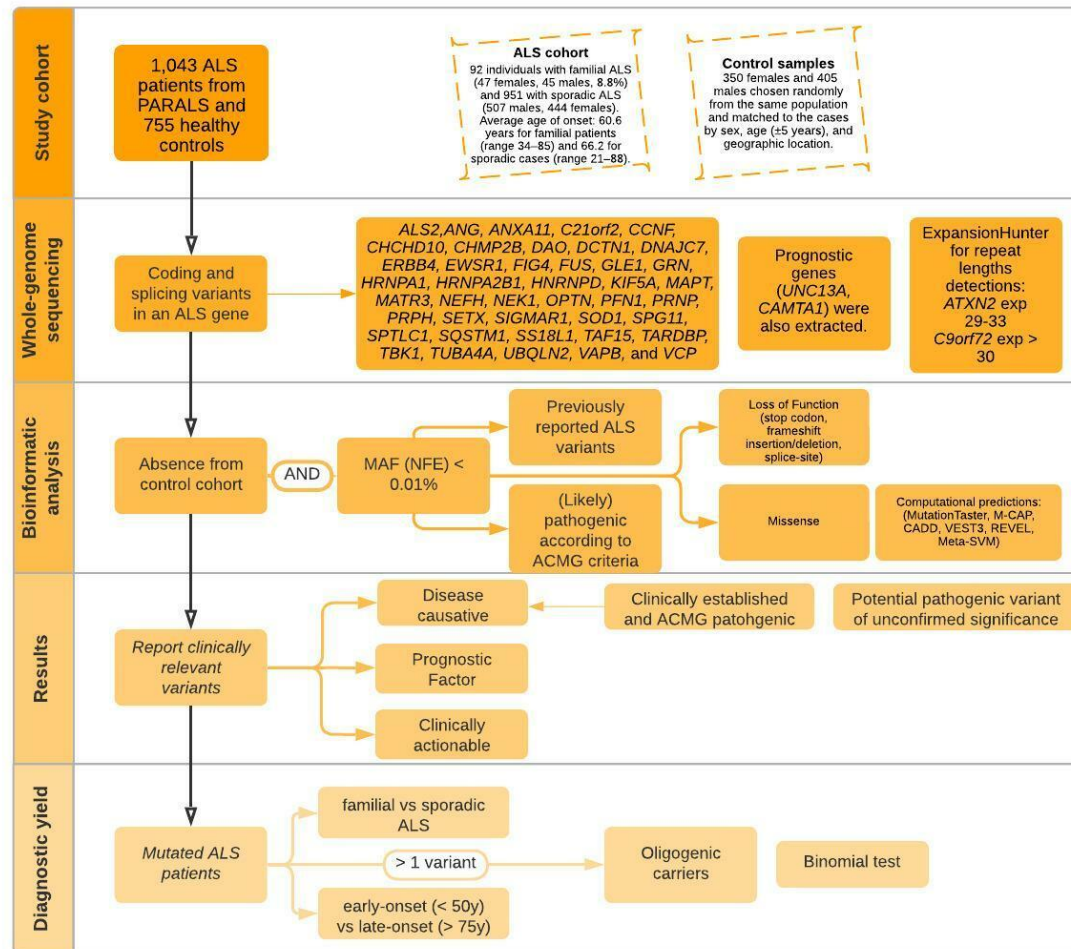
Statistical Analysis

A 2-tailed Fisher exact test was used to evaluate the genetic association between *ATXN2* CAG repeat sizes and ALS (significance set at $P < 0.05$). The burden of multiple variant carriers was assessed by a binomial test⁸. Age at disease onset and disease progression were also assessed across oligogenic and monogenic with analysis of variance (ANOVA). The analyses were performed in R (version 3.6.0).

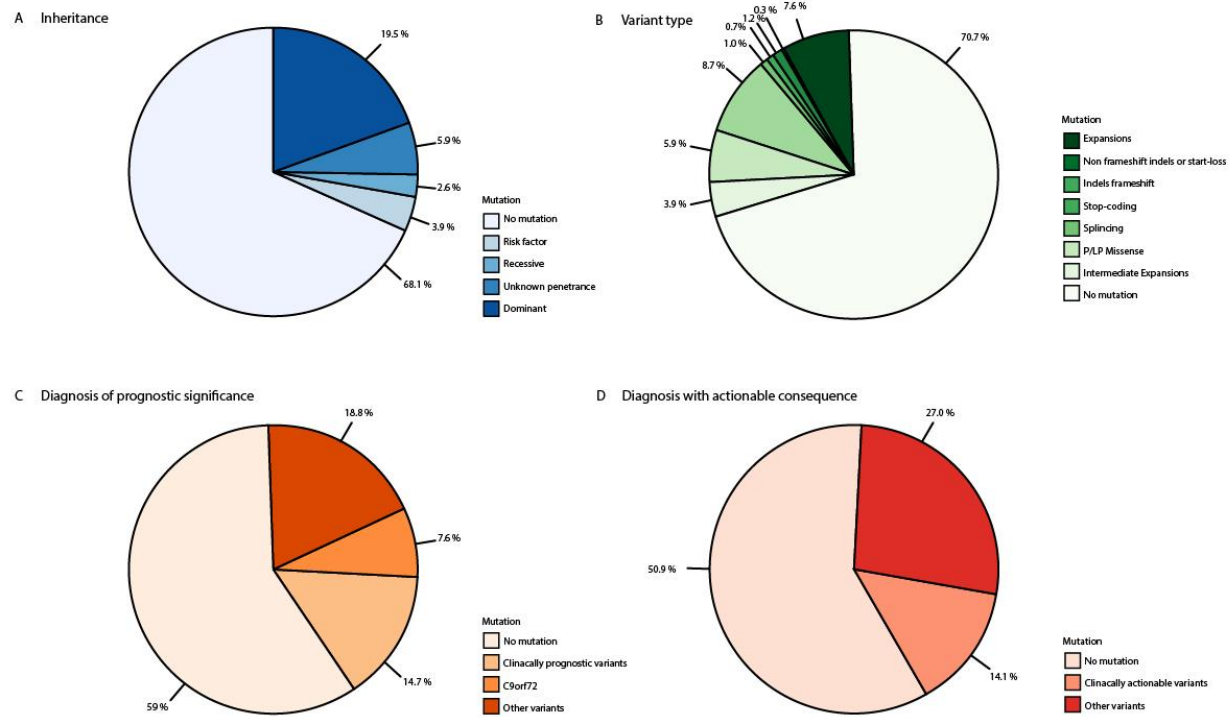
Data availability statement

The individual-level sequence data are available on dbGaP (accession number: phs001963.v1.p1) upon motivated request by interested researchers.

eFigure 1. Study Flowchart. Simplified flowchart describing the steps taken to filter and prioritize variants in ALS genes. MAF, minor allele frequency; NFE, non-Finnish Europeans; ACMG, American College of Medical Genetic.



eFigure 2. Summary of identified variants.



eTables

eTable 1 – Variants detected in our cohort previously reported in ALS cases.

Gene	Transcript	Nucleotide change	Amino acid change	Exon	Function
ALS2	NM_020919	c.4867C>T	p.L1623F	exon33	missense
ALS2	NM_020919	c.3836+1G>A	.	exon24	splicing GT donor
ALS2	NM_020919	c.3797C>G	p.P1266R	exon24	missense
ALS2	NM_020919	c.3462G>T	p.Q1154H	exon21	missense
ALS2	NM_020919	c.3394C>A	p.R1132S	exon21	missense
ALS2	NM_020919	c.3206G>A	p.G1069E	exon19	missense
ALS2	NM_020919	c.2979+1G>A	.	exon17	splicing GT donor
ALS2	NM_020919	c.2479A>T	p.T827S	exon13	missense
ALS2	NM_020919	c.1804C>T	p.R602C	exon8	missense
ALS2	NM_020919	c.1649C>T	p.P550L	exon7	missense
ALS2	NM_020919	c.1628A>G	p.D543G	exon6	missense
ALS2	NM_020919	c.1550C>G	p.A517G	exon6	missense
ALS2	NM_020919	c.1472-1G>C	.	exon6	splicing AG acceptor
ALS2	NM_020919	c.37G>A	p.G13R	exon3	missense

<i>ANXA11</i>	NM_001157	c.922C>T	p.R308X	exon8	stopgain
<i>ANXA11</i>	NM_001157	c.102G>A	p.M34I	exon3	startloss
<i>ANXA11</i>	NM_001157	c.744+2C>T	.	exon7	splicing
<i>C21ORF2</i>	NM_001271441	c.418_420del	p.G140del	exon5	non-frameshift deletion
<i>C21ORF2</i>	NM_001271441.	c.735_736insCGT	p.P245_V246insR	exon6	non-frameshift
	2	GGGGAGGGAG	GQGAWSLTGP		insertion
		CATGGAGCCT			
		CACAGGGCCC			
<i>C21ORF2</i>	NM_001271441.	c.701_702insGCC	p.V235Pfs*38	exon6	frameshift
	2	TCACAGGGCC			insertion
		CCGTGGGGAG			
		GGAGCATGGA			
		GCCTCACAGG			
		GCC			
<i>CCNF</i>	NM_001761	c.139C>T	p.L47F	exon2	missense
<i>CCNF</i>	NM_001761	c.316C>G	p.L106V	exon4	missense
<i>CCNF</i>	NM_001761	c.1043C>T	p.P348L	exon9	missense
<i>CCNF</i>	NM_001761	c.1132G>A	p.V378I	exon11	missense
<i>CHCHD1</i>	NM_213720	c.239C>T	p.P80L	exon2	missense
	0				
<i>CHMP2B</i>	NM_014043	c.74A>G	p.Q25R	exon2	missense
<i>CHMP2B</i>	NM_014043	c.85A>G	p.I29V	exon2	missense
<i>DAO</i>	NM_001917	c.34G>A	p.G12R	exon2	missense

<i>DAO</i>	NM_001917	c.992G>A	p.G331E	exon11	missense
<i>DCTN1</i>	NM_001190836	c.3575C>T	p.A1192V	exon28	missense
<i>DCTN1</i>	NM_001190836	c.3574G>A	p.A1192T	exon28	missense
<i>DCTN1</i>	NM_001190836	c.2588T>C	p.M863T	exon20	missense
<i>DCTN1</i>	NM_001190836	c.2522A>G	p.Y841C	exon20	missense
<i>DCTN1</i>	NM_001190836	c.2321C>G	p.P774R	exon18	missense
<i>DCTN1</i>	NM_001190836	c.2147C>T	p.T716M	exon17	missense
<i>DCTN1</i>	NM_001190836	c.1484G>C	p.R495P	exon12	missense
<i>DCTN1</i>	NM_001190836	c.964C>T	p.Q322*	exon8	stopgain
<i>DCTN1</i>	NM_001190836	c.884T>C	p.V295A	exon7	missense
<i>DNAJC7</i>	NM_003315	c.611G>A	p.R204Q	exon7	missense
<i>DNAJC7</i>	NM_003315	c.203G>A	p.R68Q	exon3	missense
<i>EPHA4</i>	NM_001304537	c.2060C>G	p.S687C	exon12	missense
<i>EPHA4</i>	NM_001304537	c.1702T>C	p.S568P	exon9	missense
<i>EPHA4</i>	NM_001304537	c.1058T>C	p.L353P	exon4	missense
<i>ERBB4</i>	NM_005235	c.3446G>T	p.G1149V	exon27	missense
<i>ERBB4</i>	NM_005235	c.2525G>A	p.R842Q	exon21	missense
<i>ERBB4</i>	NM_005235	c.1913G>A	p.W638*	exon16	stopgain
<i>ERBB4</i>	NM_005235	c.1912T>C	p.W638R	exon16	missense
<i>ERBB4</i>	NM_005235	c.1898G>C	p.C633S	exon16	missense
<i>ERBB4</i>	NM_005235	c.1772A>G	p.E591G	exon15	missense

<i>ERBB4</i>	NM_005235	c.586C>T	p.R196C	exon5	missense
<i>ERBB4</i>	NM_005235	c.532G>C	p.V178L	exon4	missense
<i>ERBB4</i>	NM_005235	c.308G>A	p.R103H	exon3	missense
<i>EWSR1</i>	NM_001163285	c.1447A>G	p.M483V	exon14	missense
<i>EWSR1</i>	NM_001163285	c.1775_1776insT TTTT	p.Q593Ffs*20	exon17	frameshift
<i>FIG4</i>	NM_014845	c.30C>G	p.S10R	exon1	missense
<i>FIG4</i>	NM_014845	c.52T>C	p.Y18H	exon1	missense
<i>FIG4</i>	NM_014845	c.2200G>A	p.E734K	exon20	missense
<i>FIG4</i>	NM_014845	c.2467C>T	p.Q823*	exon22	stopgain
<i>FIG4</i>	NM_014845	c.2639T>A	p.I880N	exon23	missense
<i>FIG4</i>	NM_014845	c.2650C>T	p.Q884*	exon23	stopgain
<i>FUS</i>	NM_001170634	c.785A>G	p.N262S	exon7	missense
<i>FUS</i>	NM_001170634	c.1480C>T	p.R494*	exon14	stopgain
<i>FUS</i>	NM_001170634	c.1539G>C	p.R513S	exon15	missense
<i>FUS</i>	NM_001170634	c.1552C>G	p.Q518E	exon15	missense
<i>GLE1</i>	NM_001003722	c.1771C>T	p.Q591*	exon12	stopgain
<i>GRN</i>	NM_002087	c.763C>A	p.L255M	exon8	missense
<i>GRN</i>	NM_002087	c.1562G>A	p.C521Y	exon12	missense
<i>GRN</i>	NM_002087	c.1595C>A	p.T532N	exon12	missense
<i>HNRNPA1</i>	NM_031157	c.380A>G	p.Q127R	exon4	missense

<i>HNRNPA1</i>	NM_031157	c.666C>G	p.F222L	exon6	missense
<i>HNRNPA1</i>	NM_031157	c.824G>T	p.G275V	exon8	missense
<i>HNRNPA1</i>	NM_031157	c.876C>G	p.N292K	exon8	missense
<i>HNRNPA1</i>	NM_031157	c.883G>A	p.G295R	exon8	missense
<i>KIF5A</i>	NM_004984	c.340C>T	p.R114*	exon4	stopgain
<i>KIF5A</i>	NM_004984	c.1463T>G	p.L488R	exon14	missense
<i>KIF5A</i>	NM_004984	c.2152C>T	p.R718W	exon19	missense
<i>KIF5A</i>	NM_004984	c.2263G>A	p.E755K	exon20	missense
<i>KIF5A</i>	NM_004984	c.2757del	p.K920Nfs*128	exon25	frameshift
<i>MAPT</i>	NM_016834	c.149T>C	p.I50T	exon3	missense
<i>MAPT</i>	NM_016834	c.736G>A	p.G246S	exon7	missense
<i>MAPT</i>	NM_016834	c.880T>C	p.S294P	exon9	missense
<i>MAPT</i>	NM_016834	c.913G>A	p.V305I	exon9	missense
<i>MATR3</i>	NM_018834	c.368A>G	p.D123G	exon2	missense
<i>MATR3</i>	NM_018834	c.2148+2T>+GG AC	.	exon14	splicing GT donor
<i>MATR3</i>	NM_018834	c.2371+2T>+AT A	.	exon14	splicing GT donor
<i>NEFH</i>	NM_021076	c.1036C>T	p.R346C	exon2	missense
<i>NEFH</i>	NM_021076	c.1723C>T	p.P575S	exon4	missense
<i>NEFH</i>	NM_021076	c.1783C>T	p.P595S	exon4	missense
<i>NEFH</i>	NM_021076	c.2461A>G	p.K821E	exon4	missense

<i>NEK1</i>	NM_001199399	c.3502C>T	p.H1168Y	exon32	missense
<i>NEK1</i>	NM_001199399	c.3451A>C	p.I1151L	exon32	missense
<i>NEK1</i>	NM_001199399	c.3419_3422del	p.I1140Rfs*17	exon31	frameshift
<i>NEK1</i>	NM_001199399	c.3373G>A	p.E1125K	exon31	missense
<i>NEK1</i>	NM_001199399	c.3214delA	p.T1072Lfs*20	exon30	frameshift
<i>NEK1</i>	NM_001199399	c.2816C>G	p.S939*	exon28	stopgain
<i>NEK1</i>	NM_001199399	c.2698G>T	p.D900Y	exon28	missense
<i>NEK1</i>	NM_001199399	c.2523_2526del	p.N841Kfs*53	exon26	frameshift
<i>NEK1</i>	NM_001199399	c.1226G>A	p.W409*	exon15	stopgain
<i>NEK1</i>	NM_001199399	c.1129_1132del	p.Q377Rfs*7	exon13	frameshift
<i>NEK1</i>	NM_001199399	c.781C>T	p.R261C	exon10	missense
<i>NEK1</i>	NM_001199399	c.577C>T	p.L193F	exon9	missense
<i>NEK1</i>	NM_001199399	c.464+1G>AG+	.	exon6	GT splicing donor
<i>NEK1</i>	NM_001199399	c.449C>G	p.A150G	exon7	missense
<i>NEK1</i>	NM_001199399	c.380G>A	p.R127Q	exon6	missense
<i>OPTN</i>	NM_001008212	c.247C>T	p.R83C	exon4	missense
<i>OPTN</i>	NM_001008212	c.265C>T	p.Q89*	exon4	stopgain
<i>OPTN</i>	NM_001008212	c.332T>G	p.L111R	exon4	missense
<i>OPTN</i>	NM_001008212	c.403G>T	p.E135*	exon5	stopgain
<i>OPTN</i>	NM_001008212	c.644G>A	p.R215K	exon7	missense

<i>OPTN</i>	NM_001008212	c.917_921del	p.T307Sfs*3	exon9	frameshift
<i>OPTN</i>	NM_001008212	c.1499T>C	p.L500P	exon13	missense
<i>OPTN</i>	NM_001008212	c.1634G>A	p.R545Q	exon15	missense
<i>OPTN</i>	NM_001008212	c.1639C>T	p.Q547*	exon15	stopgain
<i>OPTN</i>	NM_001008212	c.1719G>A	p.M573I	exon15	missense
<i>PFNI</i>	NM_005022	c.67G>A	p.V23M	exon1	missense
<i>PRPH</i>	NM_006262	c.487G>T	p.D163Y	exon1	missense
<i>PRPH</i>	NM_006262	c.611T>A	p.V204E	exon3	missense
<i>PRPH</i>	NM_006262	c.996+1G>A	.	exon6	GT splicing donor
<i>PRPH</i>	NM_006262	c.1024G>A	p.E342K	exon6	missense
<i>PRPH</i>	NM_006262.4	c.1222_1223insG CAG	p.S408Cfs*19	exon7	frameshift
<i>PRPH</i>	NM_006262	c.1409A>C	p.Y470S	exon9	missense
<i>SETX</i>	NM_015046	c.7240C>T	p.R2414*	exon25	stopgain
<i>SETX</i>	NM_015046	c.6848_6851del	p.T2283Kfs*32	exon22	frameshift
<i>SETX</i>	NM_015046	c.6685A>G	p.M2229V	exon21	missense
<i>SETX</i>	NM_015046	c.1427A>G	p.H476R	exon10	missense
<i>SETX</i>	NM_015046	c.1427A>C	p.H476P	exon10	missense
<i>SETX</i>	NM_015046	c.1178T>C	p.L393P	exon10	missense
<i>SETX</i>	NM_015046	c.638C>T	p.S213F	exon6	missense
<i>SETX</i>	NM_015046	c.62A>T	p.Y21F	exon3	missense

<i>SETX</i>	NM_015046	c.4A>G	p.S2G	exon3	missense
<i>SOD1</i>	NM_000454	c.16G>A	p.V6M	exon1	missense
<i>SOD1</i>	NM_000454	c.59A>G	p.N20S	exon1	missense
<i>SOD1</i>	NM_000454	c.115C>G	p.L39V	exon2	missense
<i>SOD1</i>	NM_000454	c.197A>G	p.N66S	exon3	missense
<i>SOD1</i>	NM_000454	c.217G>A	p.G73S	exon3	missense
<i>SOD1</i>	NM_000454	c.271G>A	p.D91N	exon4	missense
<i>SOD1</i>	NM_000454	c.281G>A	p.G94D	exon4	missense
<i>SOD1</i>	NM_000454	c.357+1G>+T	.	exon5	GT splicing donor
<i>SOD1</i>	NM_000454	c.409A>T	p.K137*	exon5	stopgain
<i>SOD1</i>	NM_000454	c.435G>C	p.L145F	exon5	missense
<i>SOD1</i>	NM_000454	c.435G>T	p.L145F	exon5	missense
<i>SOD1</i>	NM_000454	c.442G>A	p.G148S	exon5	missense
<i>SPAST</i>	NM_001363875	c.455T>C	p.I152T	exon2	missense
<i>SPAST</i>	NM_001363875	c.806C>T	p.S269F	exon5	missense
<i>SPAST</i>	NM_001363875	c.1526A>G	p.D509G	exon14	missense
<i>SPG11</i>	NM_025137	c.6877C>T	p.R2293W	exon38	missense
<i>SPG11</i>	NM_025137	c.6857G>C	p.R2286P	exon38	missense
<i>SPG11</i>	NM_025137	c.6094T>C	p.C2032R	exon32	missense
<i>SPG11</i>	NM_025137	c.6010T>G	p.L2004V	exon32	missense

<i>SPG11</i>	NM_025137	c.5414G>A	p.R1805H	exon30	missense
<i>SPG11</i>	NM_025137	c.3095C>T	p.P1032L	exon17	missense
<i>SPG11</i>	NM_025137	c.148C>T	p.Q50*	exon1	stopgain
<i>SQSTM1</i>	NM_003900	c.447C>G	p.D149E	exon3	missense
<i>SQSTM1</i>	NM_003900	c.775G>A	p.V259M	exon6	missense
<i>SQSTM1</i>	NM_003900	c.775G>C	p.V259L	exon6	missense
<i>SQSTM1</i>	NM_003900	c.1043C>T	p.P348L	exon7	missense
<i>SQSTM1</i>	NM_003900	c.1084G>A	p.E362K	exon7	missense
<i>SQSTM1</i>	NM_003900	c.1142C>T	p.A381V	exon7	missense
<i>SSI8L1</i>	NM_001301778	c.253A>C	p.M85L	exon7	missense
<i>TAF15</i>	NM_139215	c.116G>A	p.G39E	exon4	missense
<i>TAF15</i>	NM_139215	c.329A>G	p.Y110C	exon6	missense
<i>TAF15</i>	NM_139215	c.1088+1G>A	.	exon14	splicing GT donor
<i>TARDBP</i>	NM_007375	c.800A>G	p.N267S	exon6	missense
<i>TARDBP</i>	NM_007375	c.1144G>A	p.A382T	exon6	missense
<i>TARDBP</i>	NM_007375	c.1169A>G	p.N390S	exon6	missense
<i>TARDBP</i>	NM_007375	c.1178C>T	p.S393L	exon6	missense
<i>TBK1</i>	NM_013254	c.254T>C	p.I85T	exon4	missense
<i>TBK1</i>	NM_013254	c.454G>C	p.V152L	exon5	missense
<i>TBK1</i>	NM_013254	c.521A>G	p.Y174C	exon5	missense

<i>TBK1</i>	NM_013254	c.992+1G>A	.	exon9	splicing GT donor
<i>TBK1</i>	NM_013254	c.1343_1346del	p.I450Kfs*15	exon12	frameshift
<i>TIA1</i>	NM_022173	c.1154C>T	p.T385I	exon13	missense
<i>TIA1</i>	NM_022173	c.746A>C	p.K249T	exon10	missense
<i>TIA1</i>	NM_022173	c.95C>A	p.P32H	exon2	missense
<i>TIA1</i>	NM_022173	c.94C>A	p.P32T	exon2	missense
<i>TUBA4A</i>	NM_001278552	c.1312G>A	p.D438N	exon4	missense
<i>TUBA4A</i>	NM_001278552	c.1184_1190del	p.G395Afs*67	exon4	frameshift deletion
<i>TUBA4A</i>	NM_001278552	c.1045A>T	p.T349S	exon4	missense
<i>TUBA4A</i>	NM_001278552	c.148delG	p.A50Qfs*89	exon2	frameshift deletion
<i>UBQLN2</i>	NM_013444	c.401C>T	p.T134I	exon1	missense
<i>UBQLN2</i>	NM_013444	c.1172A>G	p.Y391C	exon1	missense
<i>UBQLN2</i>	NM_013444	c.1505G>A	p.G502D	exon1	missense
<i>VAPB</i>	NM_004738	c.332C>T	p.P111L	exon4	missense
<i>VCP</i>	NM_001354927	c.2086C>T	p.R696C	exon16	missense
<i>VCP</i>	NM_001354927	c.2002A>G	p.R668G	exon15	missense
<i>VCP</i>	NM_001354927	c.265T>C	p.Y89H	exon4	missense

eTable 2A. Oligogenic combinations in our cohort

Mutation 1	Mutation 2
<i>C9orf72</i> GGGGCC exp	<i>CHCHD10</i> :c.239C>T:p.P80L
<i>C9orf72</i> GGGGCC exp	<i>KIF5A</i> :c.T1463G:p.L488R
<i>C9orf72</i> GGGGCC exp	<i>C21orf2</i> :c.700_701ins:p.V246 Wfs*35
<i>C9orf72</i> GGGGCC exp	<i>NEK1</i> :c.G3373A:p.E1125K
<i>C9orf72</i> GGGGCC exp	<i>OPTN</i> :c.G644A:p.R215K
<i>SOD1</i> :c.59A>G:p.N20S	<i>FUS</i> :c.1542G>C:p.R514S
<i>SOD1</i> :c.115C>G:p.L39V	<i>MATR3</i> :c.2148+2->GGAC
<i>SOD1</i> :c.271G>A:p.D91N	<i>CHCHD10</i> :c.239C>T:p.P80L
<i>SOD1</i> :c.435G>C:p.L145F	<i>OPTN</i> :c.403G>T:p.E135*
<i>SOD1</i> :c.435G>C:p.L145F	<i>ERBB4</i> :c.308G>A:p.R103H
<i>TARDBP</i> :c.1144G>A:p.A382T	<i>UBQLN2</i> :c.1612G>C:p.V538L
<i>DCTN1</i> :c.C673T:p.Q225*	<i>MATR3</i> :c.2148+2->GGAC
<i>ANXA11</i> :c.C922T:p.R308X	<i>ERBB4</i> :c.A1772G:p.E591G

eTable 2B. Analysis of oligogenic frequency and effect of multiple variants on age at disease onset in our cohort

	Oligogenic cases	Monogenic cases	Binomial p-value	ANOVA p-value
Cases (% cohort)	13 (1.3%)	266 (25.6%)	0.98	-
Age at onset (SD)	59.9 (11.9)	63.9 (11.2)	-	0.1310

Supplementary References

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