1	A small PPR protein interacts with PPR-SMR1 to facilitate the splicing of introns in
2	maize mitochondria

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- 16 Supplemental data
- 17 Supplemental Figure S1. Phenotypes of *spr2-1/+* ear, overexpressed ear *Spr2*-OE1, and
- 18 complemented ear *Spr2*-Com1.
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- 23 Supplemental Figure S6. The *spr2* mutant is deficient for mitochondrial *nad1*, *nad2*,
- 24 *nad4*, *nad5*, and *nad7* mature transcripts.
- 25 Supplemental Figure S7. RT-PCR analysis of the splicing of the *nad1*, *nad2*, *nad4*,
- 26 *nad5*, and *nad7* introns in the *spr2* mutants.
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- 28 overexpression lines, complemented lines, and WT.
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- 31 in Y2H and LCI assays.
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- **33** Supplemental Table S1. Alteration of the respiration rate of WT and *spr2-1*.
- 34 Supplemental Table S2. List of some splicing factors which shared target intron(s) with
- 35 SPR2.
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- 37

38 Supplemental data



- 40 Supplemental Figure S1. Phenotypes of *spr2-1/+* ear, overexpressed ear *Spr2*-OE1, and
- 41 complemented ear *Spr2*-Com1.
- 42 (A) A self-pollinated *spr2-1/+* ear segregating *spr2* and wild-type (WT) kernels at 17
- 43 days after pollination (DAP). Arrows point to *spr2-1* mutant kernels.
- 44 (B) A self-pollinated overexpressed ear *Spr2*-OE1 (*Ubi::Spr2*-HA).
- 45 (C) A self-pollinated complemented ear *Spr2*-Com1 (*spr2*+*Ubi::Spr2*-HA).
- 46



- 48 Supplemental Figure S2. Linkage analysis of *spr2-1* mutant.
- 49 TIR8 and Spr2-F1 are primers used to detect the Mu insertion of spr2-1 mutants, Spr2-F1
- 50 and *Spr2*-R1 are the primers used to detect the *Spr2* gene. N: Non-segregating. S:
- 51 Segregating.





54 Supplemental Figure S3. Expression analysis of *Spr2*.

- 55 (A) Relative expression of *Spr2* in eleven independent *Spr2* overexpression (OE)
- 56 transgenic lines compared with WT.
- 57 (B) RT-qPCR analysis on the expression of *Spr2* in major tissues and developing seeds in
- 58 maize. R: Root; Sh: Shoot; L: Leaf; Si: Silk; F: Flower; Br: Bract. DAP: days after
- 59 pollination. RNAs were normalized against maize *Actin* gene (*GRMZM2G126010*).
- 60 Values represent the mean and standard deviation of three biological replicates, \pm SD.
- 61



63 Supplemental Figure S4. Genetic complementation of maize *spr2-2* with *Spr2*.

- 64 (A) Transgenic construct for overexpressing Spr2. Ubi-F and Spr2-R1 primers were used
- 65 in genotyping. LB: left border; RB: right border; 3xHA: triple HA tag.
- 66 (B) The genotype of Spr2 in overexpressed lines Spr2-OE, complemented lines Spr2-
- 67 Com, and wild type.
- 68 (C) Seedling phenotype comparison among overexpressed lines *Spr2*-OE, complemented
- 69 lines *Spr2*-Com, and wild type. Scale bar, 5 cm.
- 70





- 73 (A) RT-PCR analyses of AOX genes in WT and spr2. RNAs were normalized against
- 74 ZmActin gene (GRMZM2G126010).
- 75 (B) RT-qPCR analyses of AOX genes in WT and spr2. RNAs were normalized against
- 76 *ZmActin* gene (*GRMZM2G126010*). Values represent the mean and standard deviation of
- 77 three biological replicates, \pm SD.
- 78



80 Supplemental Figure S6. The *spr2* mutant is deficient for mitochondrial *nad1*, *nad2*,

81 *nad4*, *nad5*, and *nad7* mature transcripts.

82 The substantial decrease of *nad1*, *nad2*, *nad4*, *nad5* and *nad7* transcripts occur in *spr2*

83 alleles. Normalization was performed against *ZmActin* gene (*GRMZM2G126010*). The

84 RNA was isolated from the same ear segregating for WT and *spr2* mutants. DAP: days

85 after pollination.





89 *nad5*, and *nad7* introns in the *spr2* mutants.

90 Normalization was performed against *ZmActin* gene (*GRMZM2G126010*). S, intron

91 spliced; U, intron unspliced; MW, molecular markers; Kbp, thousand base pair. These

92 gel images are the same as that in Figure 5B.



94 Supplemental Figure S8. Splicing efficiency analysis of all 22 mitochondrial introns in
95 overexpression lines, complemented lines, and WT.

- 96 (A) RT-qPCR analysis of splicing efficiency of all 22 mitochondrial introns in
- 97 overexpression lines *Spr2*-OE and WT.
- 98 (B) RT-qPCR analysis of splicing efficiency of all 22 mitochondrial introns in
- 99 complemented *Spr2*-Com and WT.
- 100 The RNA was isolated from the leaves of *Spr2*-OE, *Spr2*-Com, and WT. Data are means
- 101 $(\pm SE)$ of three biological replicates. Normalization was performed against *ZmActin* gene
- 102 (*GRMZM2G126010*).
- 103



- 105 Supplemental Figure S9. Negative controls in BiFC analysis.
- 106 Different combinations of the proteins were tested in *Nicotiana benthamiana* leaves and
- 107 detected under a laser confocal microscope. Scale bars, $20 \ \mu m$.
- 108



Supplemental Figure S10. PPR-SMR1 has no directly physical interaction with EMP16

- 111 in Y2H and LCI assays.
- 112 (A) Y2H assay to determine the interactions between PPR-SMR1 and EMP16. DDO
- 113 medium: SD/-Leu/-Trp medium; QDO medium: SD/-Ade/-His/-Leu/-Trp medium;
- 114 QDO/X/A: QDO medium with added the X- α -gal and AbA.
- 115 (B) Determine the interactions between PPR-SMR1 and EMP16 by LCI analysis. An
- 116 equal amount of *Agrobacteria* was infiltrated in each test.
- 117



- **Supplemental Figure S11.** SPR2 can form multimer.
- 120 (A) Y2H assay to determine SPR2 multimer. DDO medium: SD/-Leu/-Trp medium;
- 121 QDO medium: SD/-Ade/-His/-Leu/-Trp medium; QDO/X/A: QDO medium with added
- 122 the X- α -gal and AbA.
- 123 (B) Determine SPR2 multimer by LCI analysis.
- 124 (B) Determine SPR2 multimer by BiFC analysis. Scale bars, 20 μm.

	Respiration rate (nmol O ₂ min ⁻¹ g ⁻¹ fresh weight)				
	Vt	Valt	V _{cyt}	$V_{alt}/V_t(\%)$	$V_{cyt}/V_t(\%)$
WT	1348.39±11.71	562.10±19.81	908.06±98.4	41.69	67.34
emp36	214.81±9.72	142.22±14.45	10493±7.65	66.21	48.84

126 Supplemental Table S1. Alteration of the respiration rate of WT and *spr2-1*.

128 Mitochondrial total respiration rate (V_t) , the capacity of the cytochrome pathway (V_{cyt})

129 and the alternative pathway (V_{alt}) were measured using a Chlorolab II liquid-phase

130 oxygen electrode; 2 mM KCN and 2 mM SHAM were used to inhibit the activity of

131 cytochrome c oxidase and alternative oxidase, respectively. Data are mean values \pm SEs

132 from three independent biological samples.

134	Supplemental Table S2.	List of some splicing factors	which shared target intron(s) with
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135 SPR2.

Protein	Target transcripts	Reference
EMP16	nad2-int4	(Xiu et al., 2016)
PPR-SMR1	<i>nad1</i> -int1, 2, 3, 4; <i>nad2</i> -int1, 2, 3, 4; <i>nad4</i> -int1, 2, 3; <i>nad5</i> -int1, 3, 4; <i>nad7</i> -int2; <i>rps3</i> -int1	(Chen et al., 2019)
Zm-mCSF1	<i>nad2</i> -int2, 3; <i>nad5</i> -int1, 2; <i>nad7</i> -int3; <i>ccmFc</i> -int1	(Chen et al., 2019)
PPR14	<i>nad2</i> -int3; <i>nad7</i> -int1, 2	(Wang et al., 2020)
SPR2	<i>nad1</i> -int1, 2, 3, 4; <i>nad2</i> -int1, 2, 3, 4; <i>nad4</i> -int1, 3; <i>nad5</i> -int1, 2, 4; <i>nad7</i> -int1, 2	This study

Primer name	Primer sequences (5'-3')	Application	
Spr2-F1	GAAGCCAGTGGTGAGCATCGC		
Spr2-R1	GAATGGTCGGTCTGATTTTCTCTTGCC	_	
TIR8.1	CGCCTCCATTTCGTCGAATCCCCTS	Genotype <i>spr2-1</i> and	
TIR8.2	CGCCTCCATTTCGTCGAATCCSCTT	<i>spr2-2</i> mutant	
TIR8.3	SGCCTCCATTTCGTCGAATCCCKT		
TIR8.4	CGCCTCCATTTCGTCGAATCACCTC		
Spr2-F2	CACCATGAGCCTGCAGATTCCGG	Spr2 cloning into pEntry	
Spr2-R2	TGATTTCTCATCAATAGCGAAGTTTCTG	vector	
Spr2-F3	ATGAGCCTGCAGATTCCGG	RT-PCR analysis of <i>Spr2</i>	
Spr2-R3	GAATGGTCGGTCTGATTTTCTCTTGC	gene	
Spr2-F4	GGTGGACGTGGACGAGATCTTC	RT-qPCR analysis of	
Spr2-R4	CCACAGAACCCGACAGCATC	Spr2 gene	
Spr2-F5	GGATCCATGAGCCTGCAGATTCCGGT	Snr 2 HA aloning into	
Spr2-R5	CCCGGGGAATGGTCGGTCTGATTTTCTCTT	nLINTE vector for	
3xHA-pUNTF-F	CCCGGGGGGGTTAATTAACATCTTTTACCCATA	_ pUNTF vector for	
3xHA-pUNTF-R	GAGCTCTTAGCTGCACTGAGCAGCGTA		
Ubi-F	CTTGGATGATGGCATATGCAGC	Genotype <i>Spr2</i> -OE and <i>Spr2</i> -Com	
Spr2-YTH-F	GAATTCGCAAGGCAAGAGGAGGAGGACG		
Spr2-YTH-R	GGATCCGAATGGTCGGTCTGATTTTC	yeast two-hybrid (Y2H)	
Emp16-YTH-F	CGCGGAATTCACCGCCGCCGCTTCCACTT	assay	
Emp16-YTH-R	CCCGGGATCCATTAACTGTTAAATGGTCCCAG	_	
Spr2-cLUC-F	CCGGGAGCTCATGAGCCTGCAGATTCCGGT		
Spr2-cLUC-R	CGCGGAGCTCGAATGGTCGGTCTGATTTTCTCT	 Luciferase complementation imaging (LCI) assay 	
Spr2-nLUC-F	CCGGGGGATCCATGAGCCTGCAGATTCCGGT		
Spr2-nLUC-R	CGCGGTCGACGAATGGTCGGTCTGATTTTCTCT		
Emp16-nLUC-F	CCGGGGATCCATGCCGCCGGCCAACTCCA		
Emp16-nLUC-R	CGCGGTCGACATTAACTGTTAAATGGTCCCAGA	-	
Spr2-pSPYE-F	tggcgcgccactagtggatccATGAGCCTGCAGATTCCGG	Bimolecular fluorescence	
Spr2-pSPYE-R	cccgggagcggtaccctcgagGAATGGTCGGTCTGATTTTCTCT	complementation (BIFC)	
Emp16-pSPYE-F	tggcgcgccactagtggatccATGCCGCCGGCCAACTCCA	analysis	

137	Supplemental	Table S3. L	ist of the primers	used in this study.
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Emp16-pSPYE-R	cccgggagcggtaccctcgagATTAACTGTTAAATGGTCCCAGA
Zm-mCSF1-pSPYE-F	tggcgcgccactagtggatccATGCTCACCCTCCCCGGTAC
Zm-mCSF1-pSPYE-R	cccgggagcggtaccctcgagAATTACTTTTGTAATTTGGCAC
PPR14-pSPYE-F	tggcgcgccactagtggatccATGCGTCGCTACTGCCACGT
PPR14-pSPYE-R	cccgggagcggtaccctcgagTTCAAACAGTGTTTGAAATTTC
PPR-SMR1-pSPYE-F	tggcgcgccactagtggatccATGCTGCTCCGCGTTGGC
PPR-SMR1-pSPYE-R	cccgggagcggtaccctcgagCCTAGGCATGCCAAGGGATCT
400	