

## 1 **Supplemental Information**

### 2 **Supplementary Fig. S1 (Related to Fig. 1). Gli1<sup>+</sup> cell enlargement appears differently in** 3 **different damage models.**

4 **a** The distribution of Gli1<sup>+</sup> cells were evaluated in situ in X-gal-stained sections of livers from  
5 Gli1-LacZ mice. Scale bar, 50  $\mu$ m. **b** Immunofluorescence co-staining for  $\beta$ -gal (green) with  
6 KRT19 and OPN (cholangiocyte marker, red), Alb and GS (hepatocyte marker, red), VE-CAD  
7 (endothelial marker, red) and F4/80 (macrophage marker, red) in liver sections. Slides were  
8 counterstained with DAPI (blue). Scale bar, 50  $\mu$ m. **c** Immunostaining for  $\beta$ -gal (green),  
9 EpCAM (cholangiocyte marker, red), and  $\alpha$ -SMA (activated HSC marker, red) in liver sections.  
10 Scale bar, 50  $\mu$ m. Right panels: Quantification of the percentage of Gli1<sup>+</sup> cells expressing  
11 EpCAM and  $\alpha$ -SMA. Means  $\pm$  SEM (n = 3). **d** Experimental design and representative  
12 histological images of X-gal<sup>+</sup> cells in sections of sham or PH livers. Scale bar, 50  $\mu$ m. Right  
13 panels: Quantification of the number of X-gal<sup>+</sup> cells in sham or PH livers. Means  $\pm$  SEM (n = 5).  
14 **e** Representative histological images of X-gal<sup>+</sup> cells in liver sections and the number of X-gal<sup>+</sup>  
15 liver cells from control or CCl<sub>4</sub>-treated mice. Scale bar, 50  $\mu$ m. Right panels: Quantification of  
16 the number of X-gal<sup>+</sup> cells. Means  $\pm$  SEM (n = 5). **f** Representative histological images of X-  
17 gal<sup>+</sup> cells in liver sections from control or 10 $\times$  CCl<sub>4</sub>-treated mice. Scale bar, 50  $\mu$ m. Right  
18 panels: Quantification of the number of X-gal<sup>+</sup> cells. Means  $\pm$  SEM (n = 5). \**p* < 0.05. **g, h**  
19 Representative histological images of X-gal<sup>+</sup> cells in liver sections from mice fed a normal diet  
20 and a 3-week CDE (**g**) or MCD (**h**) diet. Scale bar, 50  $\mu$ m. Right panels: Quantification of the  
21 number of X-gal<sup>+</sup> cells. Means  $\pm$  SEM (n = 5). \**p* < 0.05. \*\**p* < 0.01. **i** Immunofluorescence  
22 staining for  $\beta$ -gal (green) in liver sections from mice fed a normal diet and a 4-week DDC, CDE  
23 or MCD diet. Scale bar, 50  $\mu$ m.

### 24 **Supplementary Fig. S2 (Related to Fig. 1). Cell labeling in Gli1-CreER; Ai9 mice with Oil.**

25 **a** Schematic showing the experimental design. **b** Immunostaining for tdTomato (red) in liver  
26 sections from Gli1-CreER; Ai9 mice treated with Oil at various time points. Scale bar, 200  $\mu$ m.

27 **Supplementary Fig. S3 (Related to Fig. 1). Gli1<sup>+</sup> cells differentiate into hepatocytes**  
28 **during liver damage.**

29 **a** Schematic illustration of the experimental design. **b-e** Immunostaining for tdTomato (red) in  
30 liver sections at various time points after TAM administration. Scale bar, 100  $\mu$ m. **f**  
31 Quantification of the percentage of tdTomato<sup>+</sup> cells in liver sections. Means  $\pm$  SEM (n = 5). **g**  
32 Schematic showing the experimental design. **h, i** Immunostaining for tdTomato (red) in liver  
33 sections at various time points after DDC-induced injury. Scale bar, 100  $\mu$ m. **j** Quantification of  
34 the percentage of tdTomato<sup>+</sup> cells in liver sections. Means  $\pm$  SEM (n = 5). \* $p$  < 0.05. **k**  
35 Immunostaining for tdTomato (red) and HNF4 $\alpha$  (green) in serial liver sections after DDC-  
36 induced injury. Scale bar, 50  $\mu$ m. **l** Immunostaining for tdTomato (red) and KRT19 (green) or  
37 EpCAM in 50  $\mu$ m liver sections after 4 weeks of a normal diet or DDC diet. Scale bar, 50  $\mu$ m.  
38 **m** Quantification of the percentage of tdTomato<sup>+</sup> hepatocyte adjacent to the PV. Means  $\pm$  SEM  
39 (n = 4)

40 **Supplementary Fig. S4 (Related to Fig. 3). The cellular source of Gli1.**

41 **a** RNAscope analysis of hepatic expression of *Gli1* (red) and EpCAM (green) co-staining on 8-  
42 week-old WT mice. PV, portal vein. Scale bar, 50  $\mu$ m. **b** Immunofluorescence staining for  
43 tdTomato (red) and KRT19 (green) or Sox9 (green) in liver sections from Gli1-CreERT2; Ai9  
44 mice after TAM administration. Scale bar, 50  $\mu$ m. **c** Immunofluorescence staining for tdTomato  
45 (red) and F4/80 (green), HNF4 $\alpha$  (green), Alb (green) or VE-CAD (green) in liver sections from  
46 Gli1-CreERT2; Ai9 mice after TAM administration. Scale bar, 50  $\mu$ m. **d** Flow cytometric analysis  
47 of the percentage of tdTomato<sup>+</sup> cells among EpCAM<sup>+</sup> cells.

48 **Supplementary Fig. S5 (Related to Fig. 4). Analysis of scRNA-seq data.**

49 **a** Doublets were identified and filtered by DoubletDecon and DoubletFinder. **b** Principal  
50 component analysis (PCA) of the transcriptome data. **c-f** Violin plots showing the expression of  
51 select specific lineage-associated genes from the scRNA-seq data. **g** Signaling pathways that  
52 were significantly enriched in EpCAM<sup>+</sup>Gli1<sup>+</sup> cells using BioCarta gene sets for GSEA ( $p$  < 0.05).

53 **Supplementary Fig. S6 (Related to Fig. 5). Gli1<sup>+</sup> cells differentiate into hepatocytes**  
54 **during chronic injury.**

55 **a** Experimental design for lineage tracing of EpCAM<sup>+</sup> cells using EpCAM-CreERT2; Ai9 mice. **b**  
56 Schematic showing the experimental strategy. **c** Immunostaining for tdTomato (red) and  
57 EpCAM (green) in intestinal sections. Scale bar, 50  $\mu$ m. Right panels: Quantification of the  
58 percentage of tdTomato<sup>+</sup> cells expressing EpCAM. Means  $\pm$  SEM (n = 3). **d** Immunostaining  
59 for tdTomato (red) and EpCAM (green) in intestinal sections and liver sections. PV, portal vein.  
60 Scale bar, 50  $\mu$ m. Right panels: Quantification of the percentage of tdTomato<sup>+</sup> cells expressing  
61 EpCAM. Means  $\pm$  SEM (n = 3). **e** Immunostaining for GFP (red) in liver sections from Tmprss2-  
62 CreERT2; R26-YFP mice. Scale bar, 50  $\mu$ m. **f** Immunostaining for tdTomato (red) in liver  
63 sections from Tmprss2-DreERT2; R26-RSR-tdTomato mice treated with or without TAM. Scale  
64 bar, 50  $\mu$ m. **g** Immunostaining for tdTomato (red) and EpCAM (Green) or HNF4 $\alpha$ (Green) in  
65 tissue sections from livers after EpCAM-CreERT2; Ai9 mice received a DDC diet for 4 weeks.  
66 Scale bar, 50  $\mu$ m. **h** Quantification of the percentage of tdTomato<sup>+</sup> cells expressing HNF4 $\alpha$ .  
67 Means  $\pm$  SEM (n = 5). **i** Immunostaining for PDGFR $\alpha$  (Green), tdTomato (red) and HNF4 $\alpha$   
68 (brilliant blue), in tissue sections from livers after PDGFR $\alpha$ -CreERT2; Ai9 mice received a DDC  
69 diet for 4 weeks. Scale bar, 50  $\mu$ m. **j** Quantification of the percentage of tdTomato<sup>+</sup> cells  
70 expressing HNF4 $\alpha$ . Means  $\pm$  SEM (n = 5).

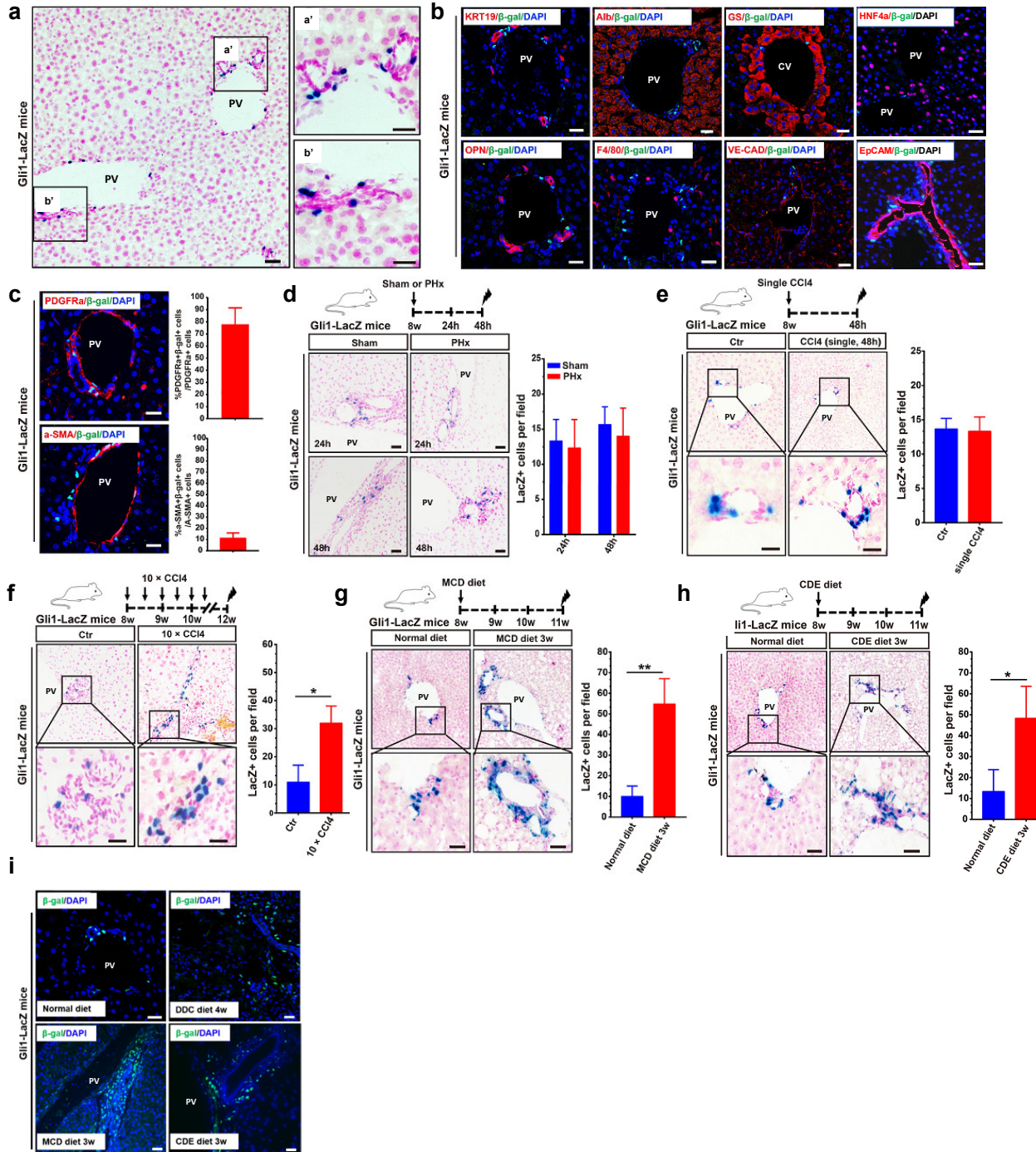
71 **Supplementary Fig. S7 (Related to Fig. 6). Organoids cultured from single EpCAM<sup>+</sup>Gli1<sup>+</sup>**  
72 **cells from Gli1-LacZ mice.**

73 **a** FACS gating strategy to isolate EpCAM<sup>+</sup> and tdTomato<sup>+</sup> cells. **b** Table showing the numbers  
74 used to calculate the % colony formation. **c** Schematic representation of organoid-development  
75 from EpCAM<sup>-</sup>Gli1<sup>-</sup>, EpCAM<sup>+</sup>Gli1<sup>-</sup>, EpCAM<sup>-</sup>Gli1<sup>+</sup> and EpCAM<sup>+</sup>Gli1<sup>+</sup> single cells from Gli1-LacZ  
76 mice sorted by FACS. **d** FACS plot showing the expression of EpCAM and Gli1 in the liver in  
77 Gli1-LacZ mice stained with CUG and EpCAM. **e** Representative images showing 5,000 sorted  
78 cells that grew into liver organoids 5 days after sorting. **f** Table showing the numbers used to

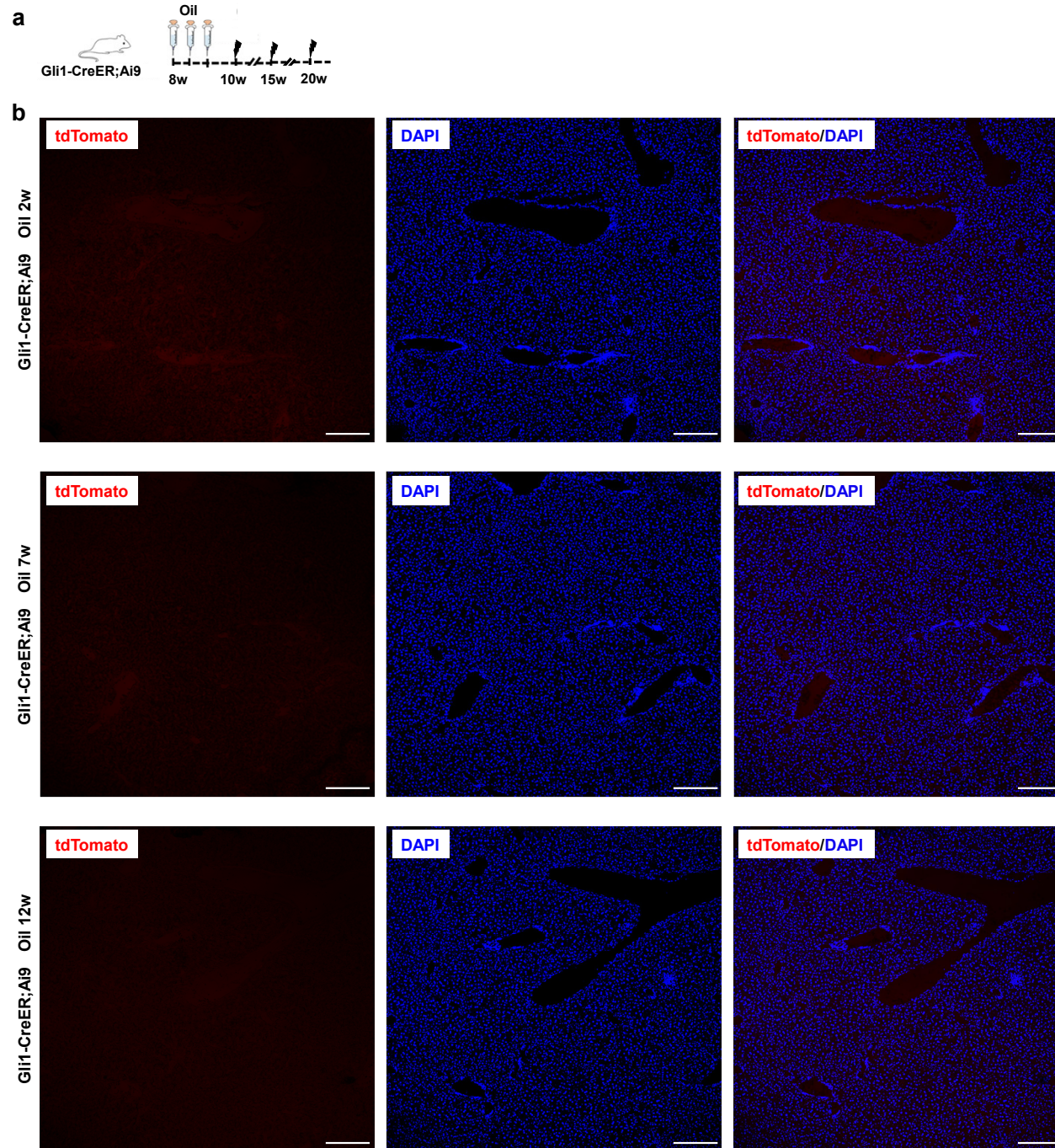
79 calculate the % colony formation for Gli1-LacZ mice. **g** Numbers of organoids formed per 5,000  
80 single cells (left) and percentage of colony formation efficiency (right). Means  $\pm$  SEM (n = 3).  
81 **\*\*** $p < 0.01$ .

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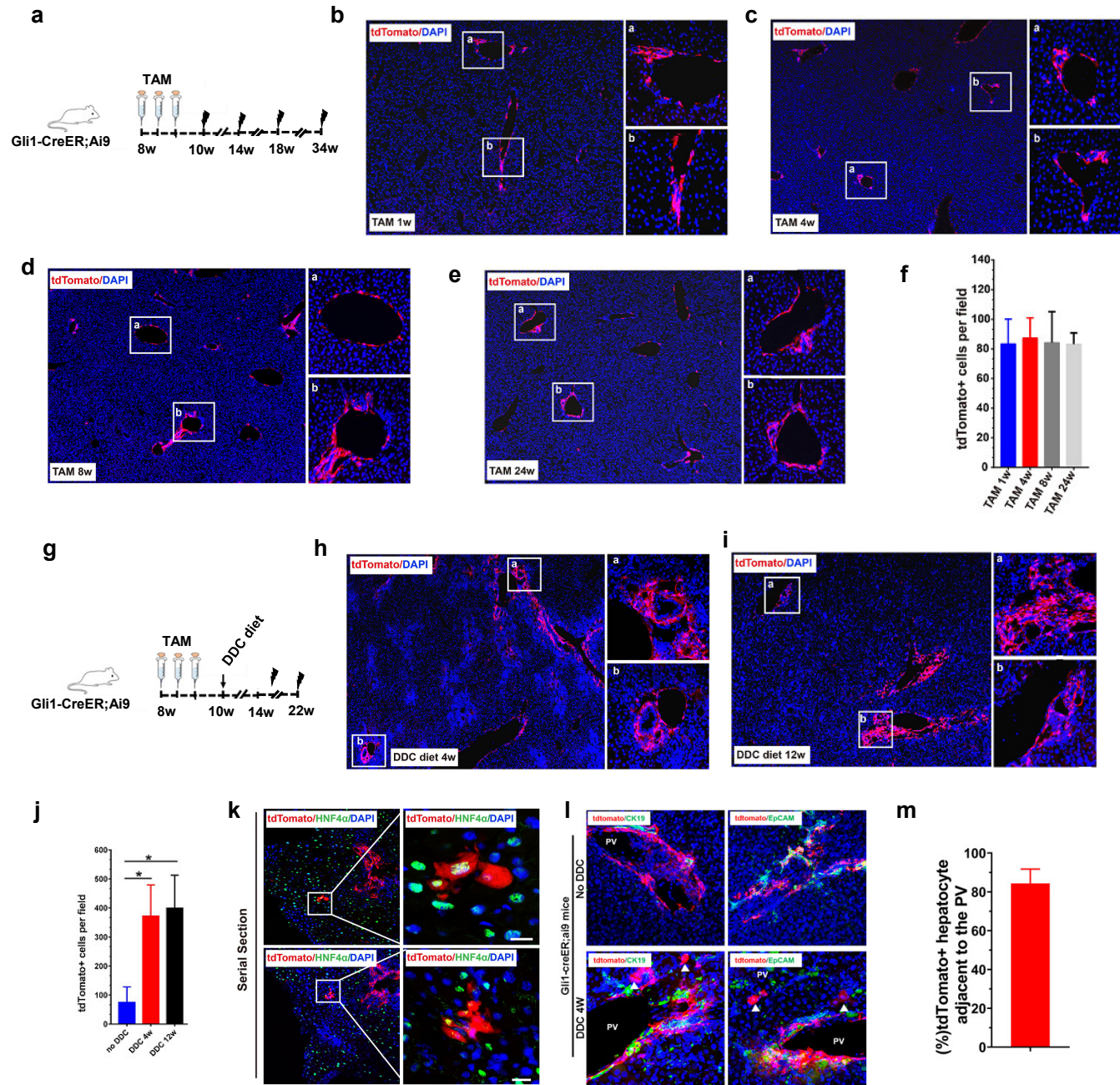
# Supplementary Fig. S1



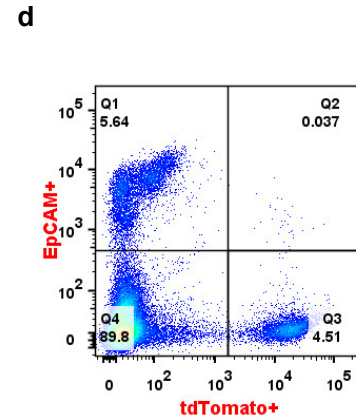
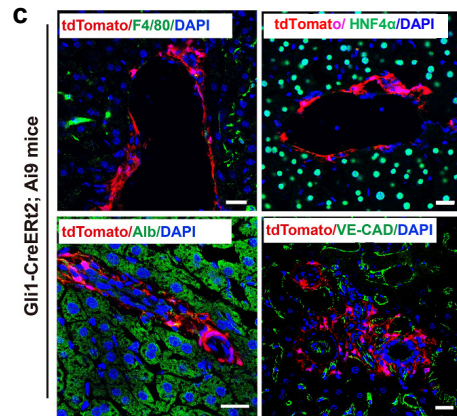
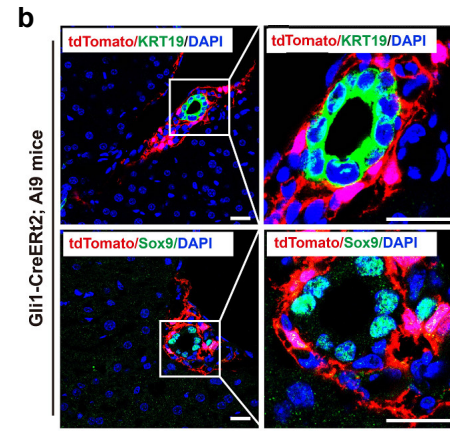
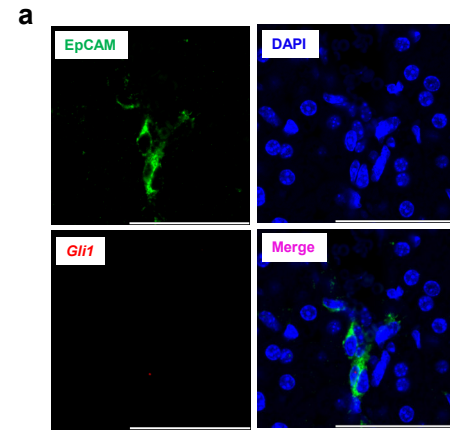
# Supplementary Fig. S2



# Supplementary Fig. S3

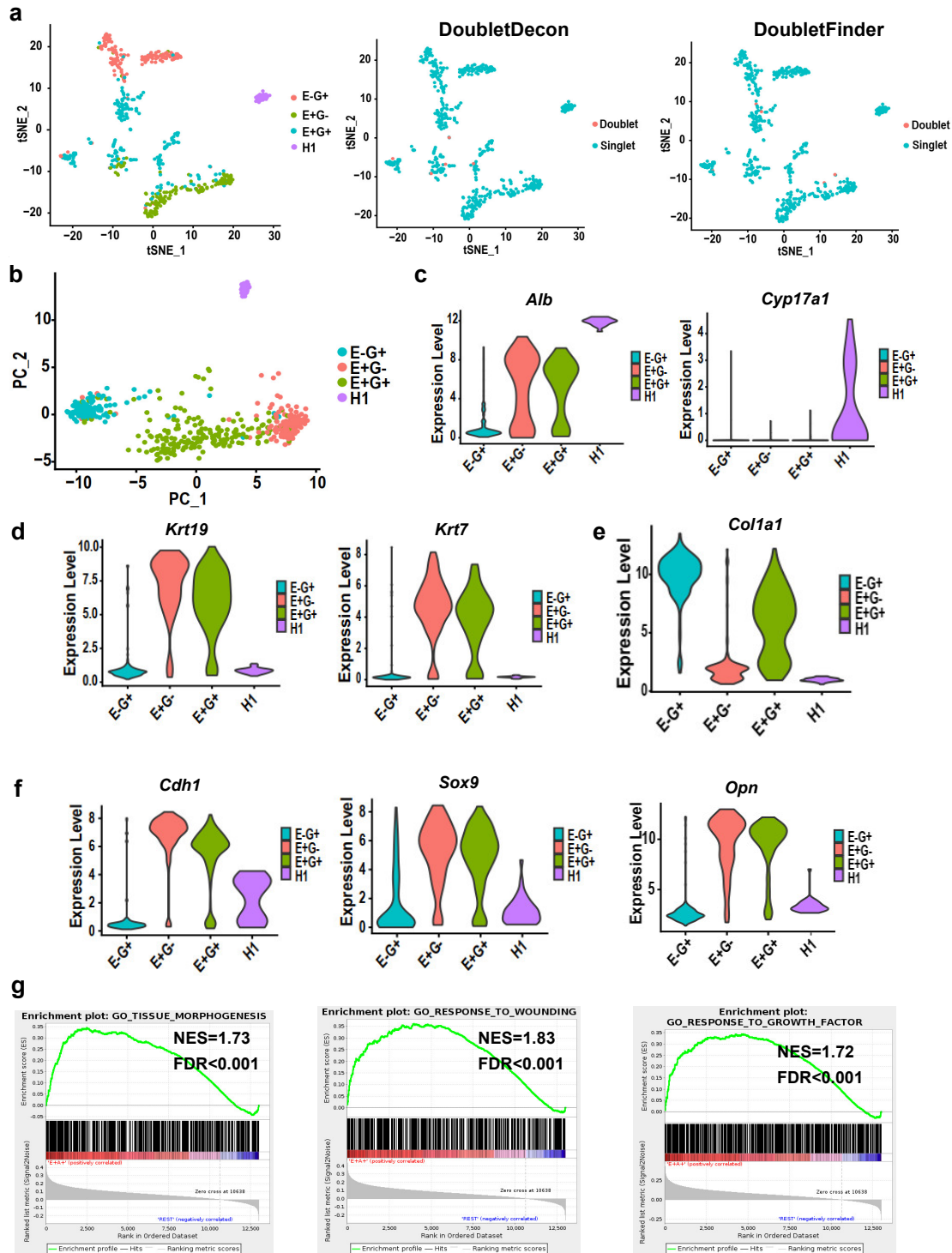


# Supplementary Fig. S4

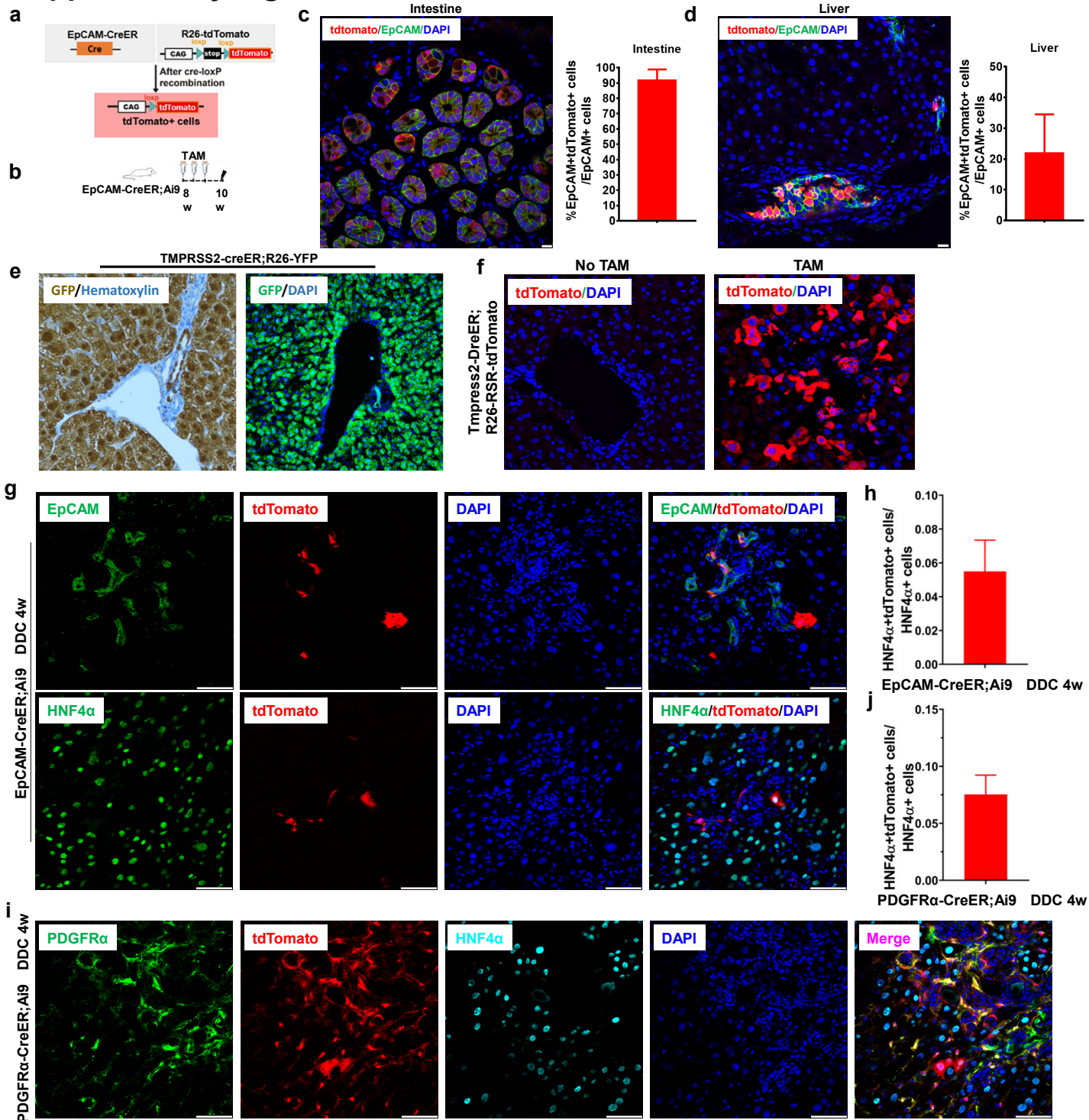




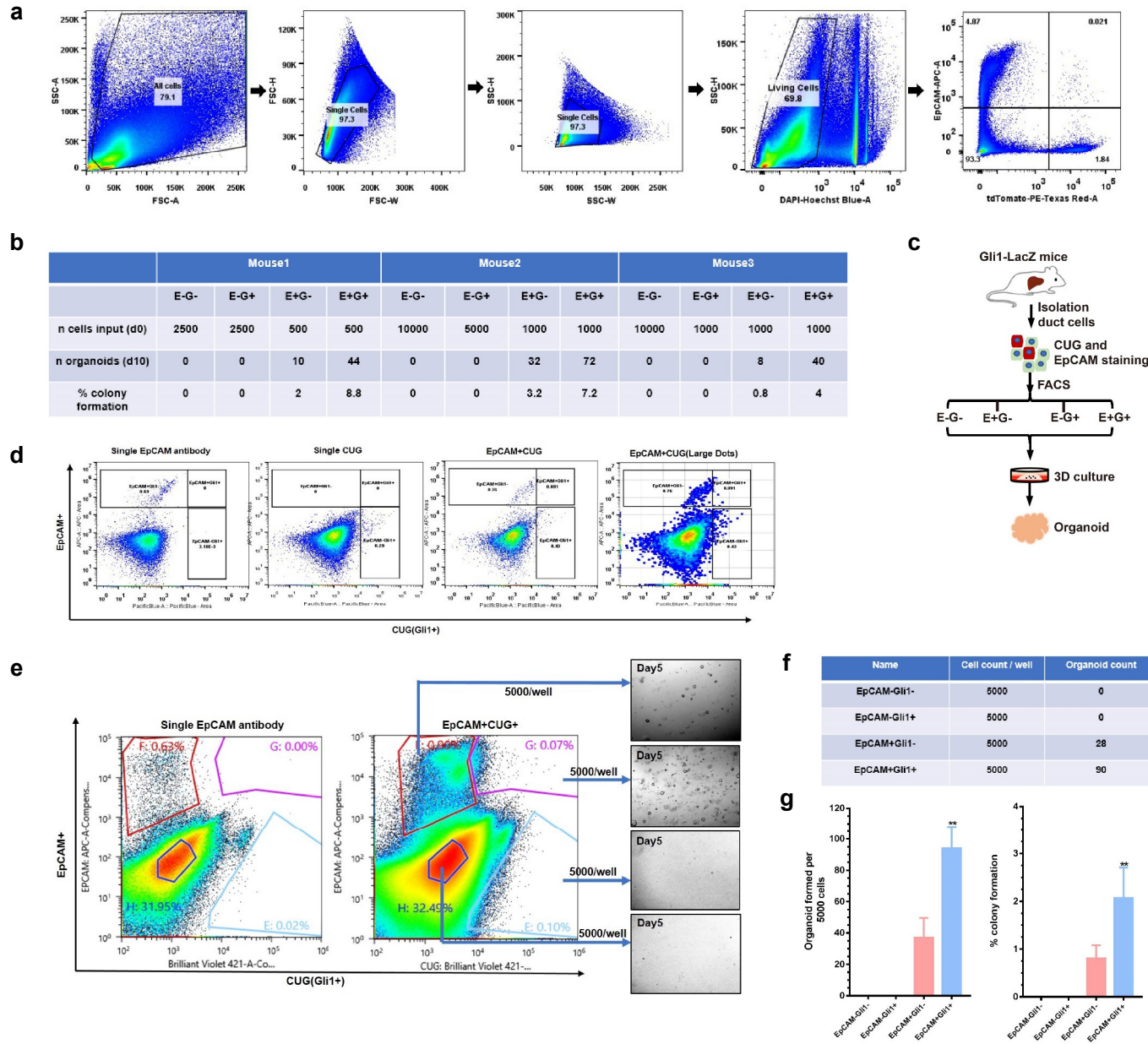
# Supplementary Fig. S5



# Supplementary Fig. S6



# Supplementary Fig. S7



83 **Supplementary Table 1: List of primary antibodies used for immunostaining**  
 84 **experiments**

<b>Primary antibody</b>	<b>Species</b>	<b>Dilution</b>	<b>Source</b>	<b>Catalog number</b>
HNF4 $\alpha$	rabbit	1/250	CST	3113S
E-cadherin	mouse	1/200	BD Biosciences	610182
CK19	rabbit	1/500	Abcam	ab52625
CK19 (Troma III)	rat	1/500	DSHB	AB_2133570
Osteopontin(OPN)	goat	1/100	R&D Systems	AF808
Sox9	rabbit	1/500	Millipore	AB5535
DsRed	rabbit	1/500	Clontech	632496
mcherry	goat	1/500	SICGEN	AB0081-200
PDGFR $\alpha$	goat	1/100	R&D Systems	AF1062
VE-CAD	goat	1/100	R&D Systems	AF1002
EpCAM	rabbit	1/250	Abcam	ab71916
GS	mouse	1/1000	BD Biosciences	610517
Alb	mouse	1/250	Sigma	SAB3500217
$\beta$ -gal	rabbit	1/250	Abcam	ab221199
$\alpha$ -SMA (Cy3-conjugated)	mouse	1/1000	Sigma	C6198
EpCAM (APC-conjugated)	rat	1/100	eBioscience	14-5791-82

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86 **Supplementary Table 2: List of primer sequences used for RT-qPCR**

<b>Gene</b>	<b>Pimer sequences (5'-3')</b>
<i>EpCAM</i>	Fw - TGTGGACATAGCTGATGTGGCTTAC Rv - CACCCTCAGGTCCATGCTCTTA
<i>Krt19</i>	Fw - GTCCTACAGATTGACAATGC Rv - CACGCTCTGGATCTGTGACA
<i>GAPDH</i>	Fw - AGGTCGGTGTGAACGGATTTG Rv - TGTAGACCATGTAGTTGAGGTCA
<i>HNF4<math>\alpha</math></i>	Fw - AGCTCGAGGCTCCGTAGTGTTT Rv - GAAAATGTGCAGGTGTTGACCA
<i>Alb</i>	Fw - GCTGAGACCTTCACCTTCCA Rv - TCTTCAGTTGCTCCGCTGTA

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