

Supporting Information

Perivascular Stromal Cells Instruct Glioblastoma Invasion, Proliferation, and Therapeutic Response within an Engineered Brain Perivascular Niche Model

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Figure S1. (**A**) Immunofluorescent markers for human brain microvascular endothelial cells (HBMVEC), normal human astrocytes (NHA), and human brain vascular pericytes (HBVP). Cells were fixed 48 hours after seeding. Scale bar = $50 \mu m$. (**B**) Matrigel tube formation assay for brain microvascular endothelial cells. Images were taken 16 hours after cell seeding. Scale bar = $100 \mu m$



Figure S2. Immunofluorescent staining for occludins in microvascular cultures. Scale bar = $100 \ \mu m$.







Figure S4. Quantification of total pixel intensity and total pixel intensity normalized to outgrowth area in spheroid invasion cultures. Data analyzed using one-way ANOVA with Scheffe's post-hoc; p<0.05, p<0.01, p<0.01, p<0.001, N = 9 - 11 hydrogels.



Figure S5. Quantitative comparison of proteins secreted by spheroid invasion cultures. Data analyzed using one-way ANOVA with Tukey's post-hoc or Kruskal-Wallis test with Dunn's post-hoc; p<0.05, p<0.01, p<0.01, p<0.001 compared to GBM6-only cultures; N = 5 conditioned media samples.



Figure S6. Quantification of the fraction of RFP-expressing GBM6 tumor cells that incorporate EdU in a 24-hour pulse. N = 6 hydrogels.



Figure S7. Dose-response curve for GBM12 cells treated with temozolomide for 48 hours. N = 4 - 6 hydrogels.



Figure S8. Representative images of tumor-vascular co-cultures with and without 48 hours of temozolomide treatment. cPARP staining was performed at the end of the treatment period. Scale bar = $200 \ \mu m$.



Figure S9. Calculated ratios for **A**) cPARP, **B**) EdU, and **C**) KI67 expression between TMZ-treated (TMZ+) and DMSO control (TMZ-) groups. * p<0.05, **p<0.01, and ***p<0.001 between groups.



Figure S10. A) Comparing the proximity of cPARP- vs. cPARP+ GBM12 cells to the microvascular network after temozolomide treatment. Data points represent the fraction of tumor cells that are a specified distance from the microvascular network in a single hydrogel. Data analyzed using paired-sample t-test or Wilcoxon signed-rank test;*p<0.05, **p<0.01, ***p<0.001 **B**) Comparing the fraction of cPARP- and cPARP+ GBM12 cells that are within 10 µm of the microvascular network. Data analyzed using one-way ANOVA with Tukey's post-hoc; ^p<0.05, ^^p<0.01 compared to EC, cPARP-; #p>0.05 compared to EC, cPARP+ **C**) Comparing the fraction of cPARP- and cPARP+ GBM12 cells that are greater than 50 µm from the microvascular network. Data analyzed using one-way ANOVA with Tukey's post-hoc; #p<0.05 compared to EC, cPARP+ GBM12 cells that are greater than 50 µm from the microvascular network. Data analyzed using one-way ANOVA with Tukey's post-hoc; #p<0.05 compared to EC, cPARP+ GBM12 cells that are greater than 50 µm from the microvascular network. Data analyzed using one-way ANOVA with Tukey's post-hoc; #p<0.05 compared to EC, cPARP+; N = 7 hydrogels



Figure S11. Representative images of tumor-vascular co-cultures with and without 48 hours of temozolomide treatment. EdU staining occurred after a 24-hour pulse of EdU after temozolomide treatment. Scale bar = $200 \ \mu m$.



Figure S12. Representative images of tumor-vascular co-cultures with and without 48 hours of temozolomide treatment. KI67 staining was performed at the end of the treatment period. Scale bar = $200 \ \mu m$.