

## **Supplementary Information**

### **Spatial analysis of the glioblastoma proteome reveals specific molecular signatures and markers of survival**

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Isabelle Fournier<sup>1,6\*</sup> and Michel Salzet<sup>1,6\*</sup>

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**Supplementary Table 1.** Model cross-validation from the SpiderMass recorded in the positive ion mode from 30 different tumors using the “20% randomly taken” and the “1 patient out” methods. Percentage of good classifications are given including or excluding outliers.

Model type	ion mode	n classes	n tested spectra	n passes	n failures	N outliers	Correct classification rate (%)	
							Excluding outliers	Including outliers
<b>Cross Validation by 20% randomly taken</b>								
<b>Glioma</b>	<b>Positive</b>	<b>3</b>	<b>135</b>	<b>124</b>	<b>0</b>	<b>11</b>	<b>100%</b>	<b>91.85%</b>
<b>Cross Validation by 1 patient out method</b>								
<b>Glioma</b>	<b>Positive</b>	<b>3</b>	<b>135</b>	<b>105</b>	<b>8</b>	<b>22</b>	<b>92.92%</b>	<b>77.78%</b>

**Supplementary Table 2.** Histological composition of each of the three regions identified by MALDI-MSI. The percentage of micro-extracted points in the histological areas was calculated for each molecular region (yellow-B, red-A, and blue-C).

Region	Tumor	Very dense infiltration	dense infiltration	necrosis+ tumor	MVP+ tumor	tumor+ inflammation	necrosis
yellow	45%	0%	0%	25%	5%	0%	20%
red	39.3%	14.3%	28.6%	10.7%	3.6%	3.6%	0%
blue	73.6%	5.7%	2.3%	3.4%	12.6%	1.1%	1.1%

**Supplementary Table 3.** List of the proteomics extraction points and their associated region

Extraction point	Region	Extraction point	Region	Extraction point	Region
12-1	A	48-1	B	34-2	C
12-2	A	53-1	B	34-3	C
12-3	A	9-2	B	35-1	C
12-4	A	10-1	C	35-2	C
14-1	A	10-2	C	35-3	C
14-2	A	1-1	C	36-3	C
14-3	A	1-2	C	37-1	C
20-1	A	13-1	C	38-2	C
20-2	A	13-2	C	39-1	C
20-3	A	13-3	C	39-3	C
20-4	A	15-1	C	39-4	C
21-1	A	15-2	C	4-1	C
21-2	A	15-3	C	41-1	C
25-1	A	16-1	C	41-2	C
25-2	A	16-2	C	41-3	C
25-3	A	16-3	C	42-2	C
31-1	A	17-1	C	42-3	C
31-2	A	17-2	C	43-1	C
31-3	A	17-3	C	44-1	C
32-1	A	17-4	C	44-2	C
36-1	A	19-1	C	44-3	C
36-2	A	19-2	C	45-1	C
43-2	A	2-1	C	45-2	C
46-3	A	22-1	C	46-1	C
49-3	A	22-2	C	46-2	C
49-4	A	2-3	C	47-1	C
5-1	A	24-1	C	47-2	C
5-2	A	24-2	C	48-2	C
1-3	B	24-3	C	48-3	C
2-2	B	24-4	C	49-1	C
26-1	B	25-4	C	49-2	C
27-1	B	25-5	C	50-1	C
27-2	B	26-2	C	50-2	C
27-3	B	28-1	C	50-3	C
32-2	B	28-2	C	51-1	C
32-3	B	29-1	C	51-2	C
37-2	B	29-2	C	51-3	C
37-3	B	29-3	C	53-2	C

<b>38-1</b>	<b>B</b>	<b>30-1</b>	<b>C</b>	<b>53-3</b>	<b>C</b>
<b>38-3</b>	<b>B</b>	<b>30-2</b>	<b>C</b>	<b>6-1</b>	<b>C</b>
<b>39-2</b>	<b>B</b>	<b>33-1</b>	<b>C</b>	<b>8-1</b>	<b>C</b>
<b>4-2</b>	<b>B</b>	<b>33-2</b>	<b>C</b>	<b>8-2</b>	<b>C</b>
<b>42-1</b>	<b>B</b>	<b>33-3</b>	<b>C</b>	<b>8-3</b>	<b>C</b>
<b>47-3</b>	<b>B</b>	<b>33-4</b>	<b>C</b>	<b>9-1</b>	<b>C</b>
<b>47-4</b>	<b>B</b>	<b>34-1</b>	<b>C</b>	<b>9-3</b>	<b>C</b>

**Supplementary Table 4.** Ghost proteins overexpressed in the 3 regions A B C

GROUP	protein accession	protein length (a.a.)	molecular weight (kDa)	isoelectric point	gene symbol	type	localization
C	IP_063564	113	12.39	9.26	<i>CLDN19</i>	mRNA	3'UTR
C	IP_065285	48	5.17	8.97	<i>C8B</i>	mRNA	CDS
C	IP_072691	36	4.26	8.51	<i>CD84</i>	mRNA	3'UTR
C	IP_073718	85	9.63	10.84	<i>CCDC181</i>	mRNA	CDS
C	IP_079312	388	42.34	5.71	<i>EDARADD</i>	mRNA	3'UTR
C	IP_092740	46	5.08	10.69	<i>TTN</i>	mRNA	CDS
C	IP_219633	35	3.95	5.67	<i>POSTN</i>	mRNA	3'UTR
C	IP_2277691	46	5.53	8.78	<i>RTTN</i>	mRNA	CDS
C	IP_2303139	56	6.24	6.49	<i>LOC105369735</i>	ncRNA	-
C	IP_2323408	61	6.86	8.55	<i>LOC105376126</i>	ncRNA	-
C	IP_2348387	31	3.34	7.26	<i>COL19A1</i>	mRNA	5'UTR
C	IP_235425	38	4.24	10.31	<i>HCN4</i>	mRNA	3'UTR
C	IP_2376460	57	6.73	7.36	<i>EPHA6</i>	mRNA	3'UTR
C	IP_2393992	43	5.22	11.65	<i>LOC105376830</i>	ncRNA	-
C	IP_256988	39	4.52	9.45	<i>TBX21</i>	mRNA	3'UTR
C	IP_261897	41	4.44	9.67	<i>AC108004.2</i>	ncRNA	-
C	IP_557239	90	10.19	9.53	<i>TUBB4AP1</i>	ncRNA	-
C	IP_559054	30	3.56	4.27	<i>SLITRK2</i>	mRNA	5'UTR
C	IP_563986	424	47.8	4.43	<i>KRT8P11</i>	ncRNA	-
C	IP_572422	212	24.18	4.64	<i>TUBBP1</i>	ncRNA	-
C	IP_580018	247	29.04	11.36	<i>RPL7P32</i>	ncRNA	-
C	IP_590938	74	8.75	10.05	<i>HMGB3P18</i>	ncRNA	-
C	IP_629960	30	3.57	9.69	<i>LRRC34</i>	ncRNA	-
C	IP_669601	75	8.32	10.63	<i>HIST2H2BB</i>	ncRNA	-
C	IP_671464	249	26.94	8.21	<i>TPI1P1</i>	ncRNA	-
C	IP_701463	179	20.51	9.05	<i>CTD-2541J13.2</i>	ncRNA	-
C	IP_739889	139	15.18	5.67	<i>GOLGA6L9</i>	ncRNA	-
C	IP_752010	59	6.39	10.62	<i>MIPOL1</i>	mRNA	3'UTR
C	IP_774693	75	8.68	3.92	<i>TUBAP2</i>	ncRNA	-
C	IP_774695	374	41.01	5.68	<i>TUBAP2</i>	ncRNA	-
A	IP_2390879	36	4.08	5.49	<i>LOC107985743</i>	ncRNA	-
A	IP_244732	78	8.56	7.13	<i>KIFC3</i>	mRNA	5'UTR
B	IP_156671	120	14.22	10.43	<i>SLC13A1</i>	mRNA	3'UTR
B	IP_222588	58	6.32	11.27	<i>CPB2-AS1</i>	ncRNA	-
B	IP_2389079	29	2.98	8.35	<i>MYT1L</i>	mRNA	5'UTR
B	IP_265416	32	3.81	10.78	<i>LIPG</i>	mRNA	CDS
B	IP_273562	76	8.34	11.89	<i>LGI4</i>	mRNA	5'UTR
B	IP_278725	69	7.77	10.83	<i>ZNF888</i>	mRNA	CDS

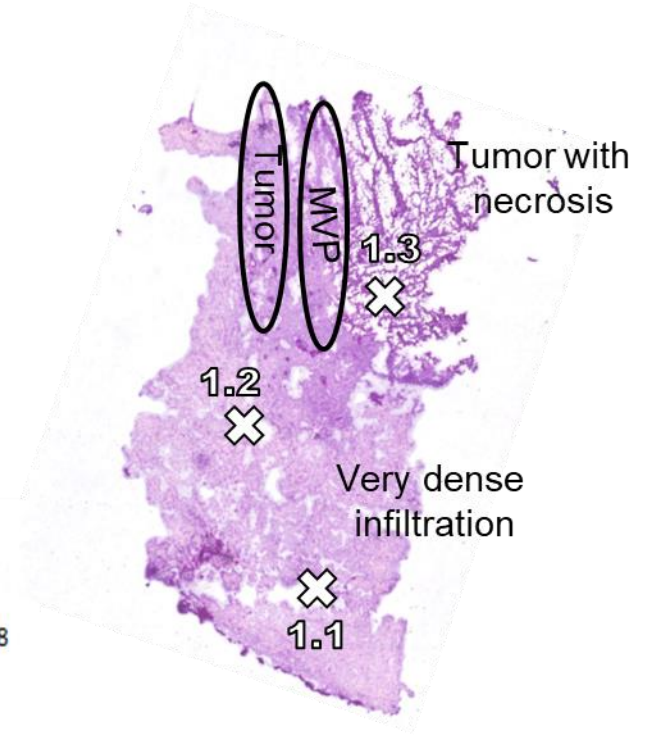
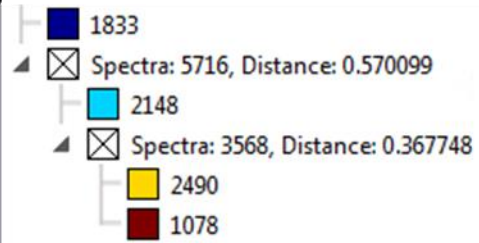
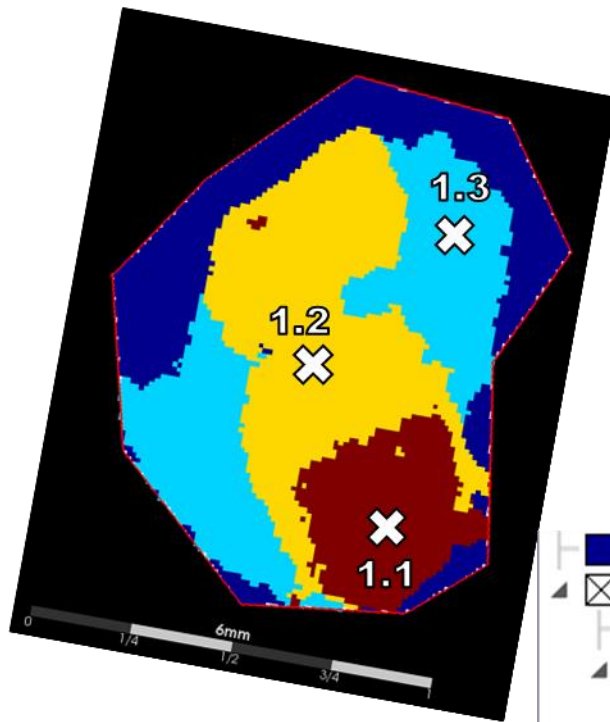
B	IP_592742	86	9.64	5.86	<i>KRT18P64</i>	ncRNA	-
B	IP_602497	38	4.24	5.65	<i>CTD-2151A2.2</i>	ncRNA	-
B	IP_672223	178	20.37	5.14	<i>GBP1P1</i>	ncRNA	-



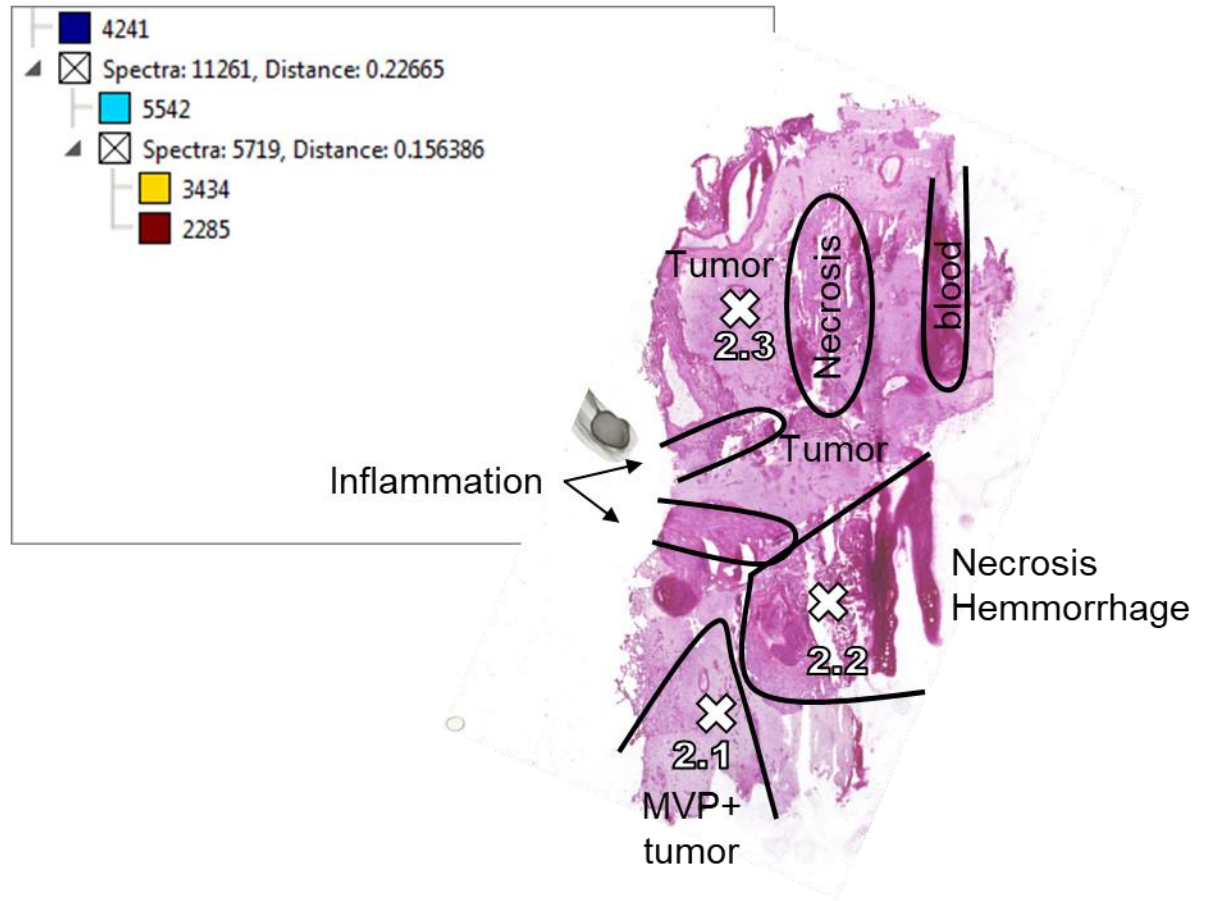
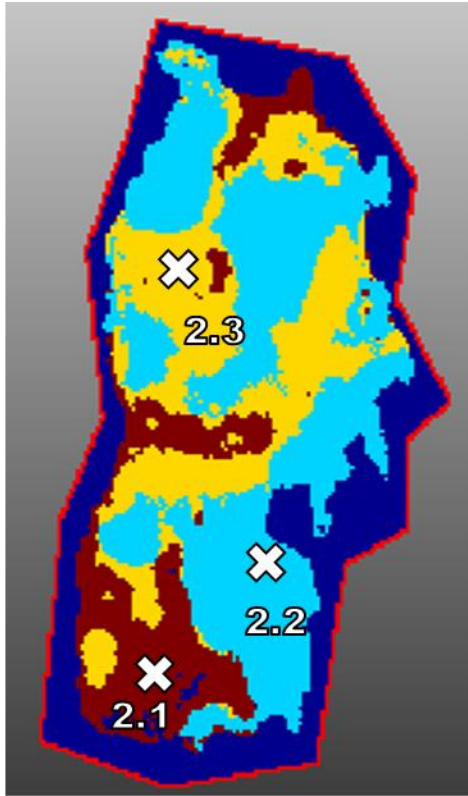


- A.** Scanned pictures after hematoxylin-eosin staining of the 46 glioblastoma tumors
- B.** Pathologist annotations of the glioblastoma samples. The pathologist annotated each sample's different regions of interest (tumors, endothelial proliferation, necrosis, infiltration, blood...).
- C.** MALDI MSI images of characteristics  $m/z$  ions for each group.
- D.** Ward clustering method gives 3 main branches with the same characteristic ions.
- E.** Principal component analysis (PCA) of each spectra reveals separation between the three regions.

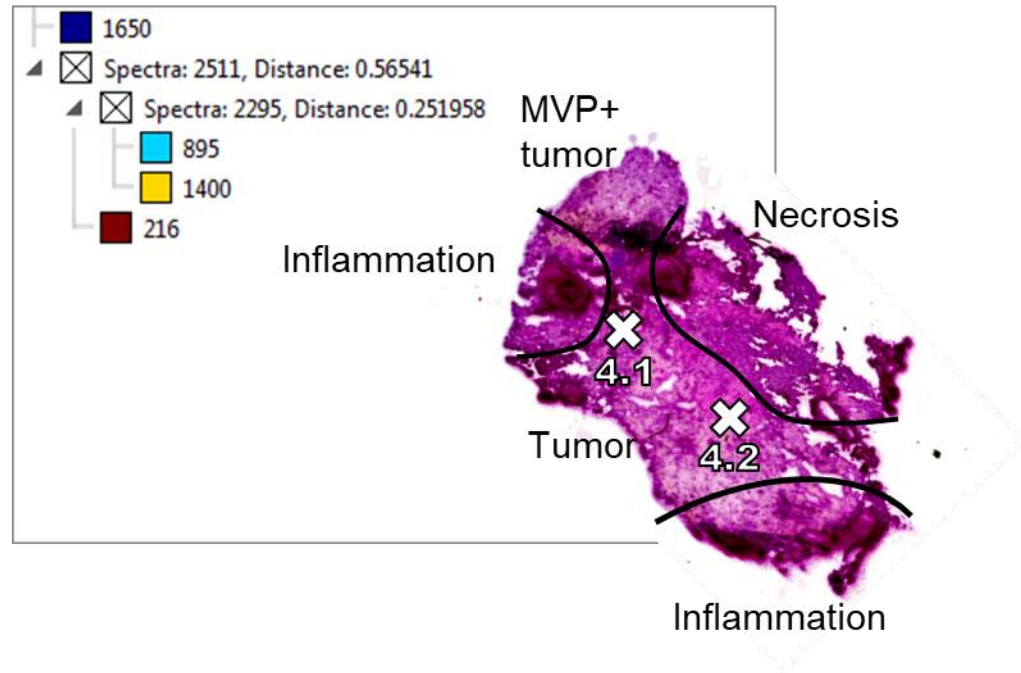
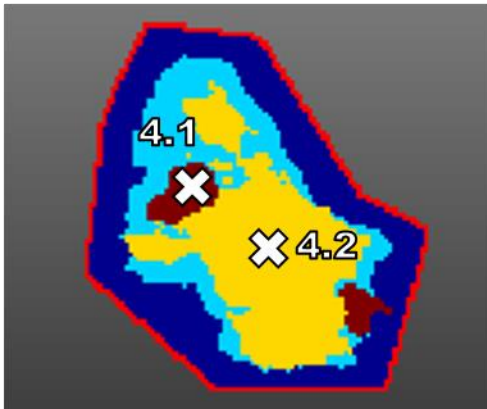
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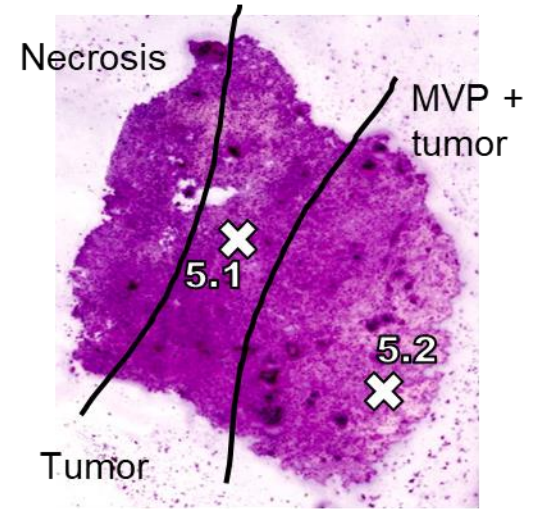
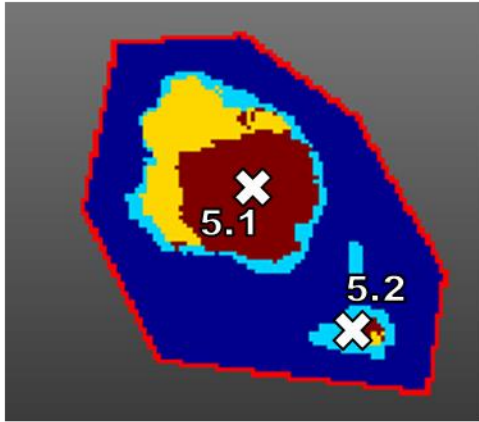
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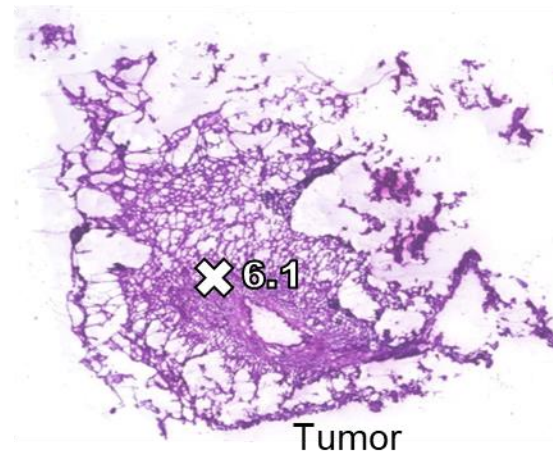
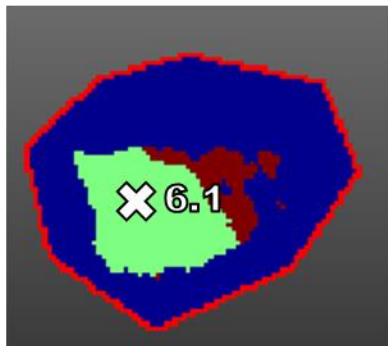
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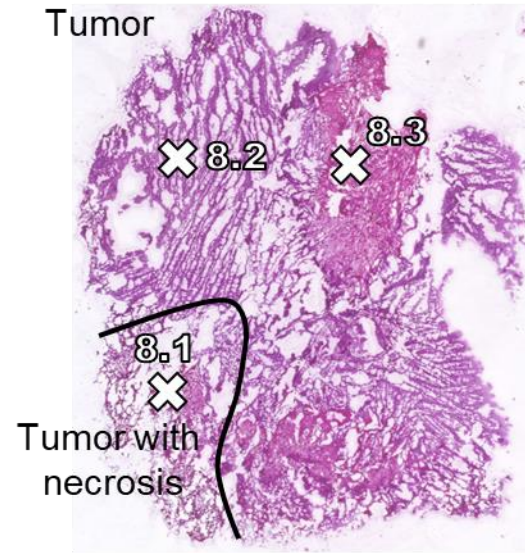
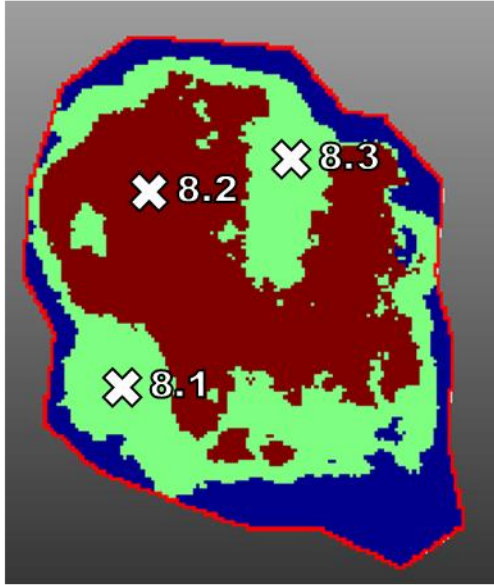
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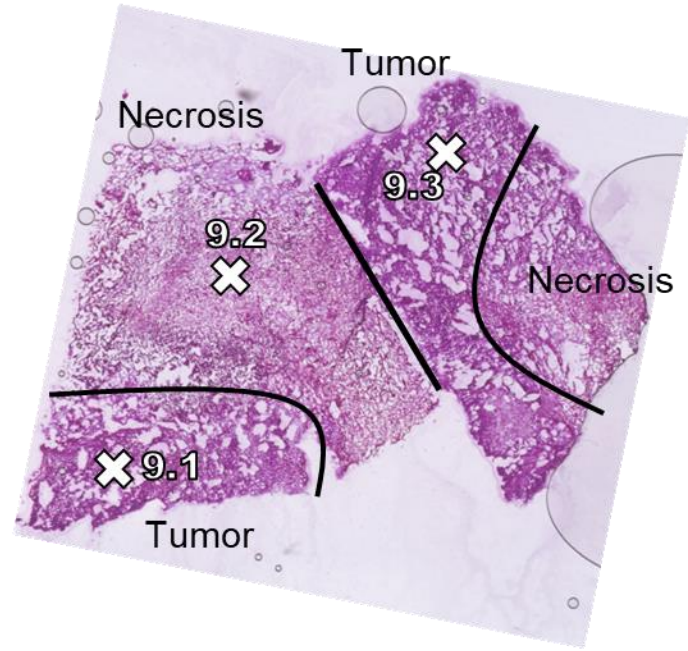
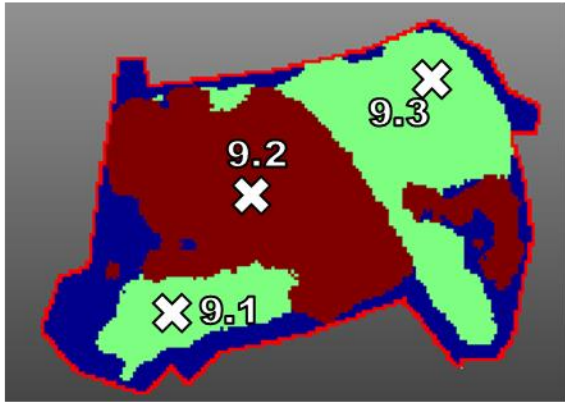
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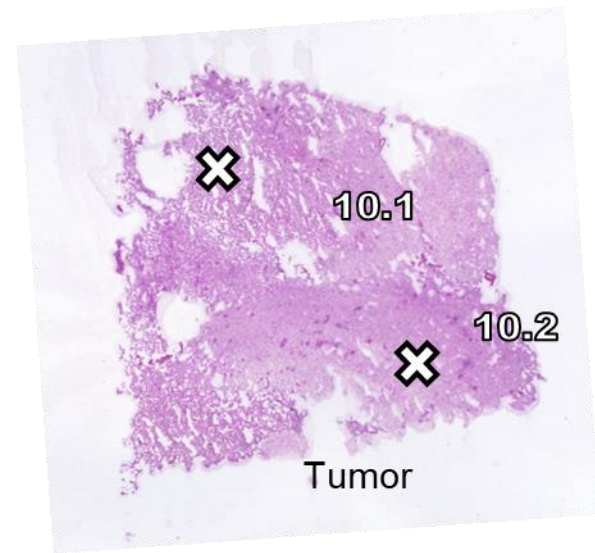
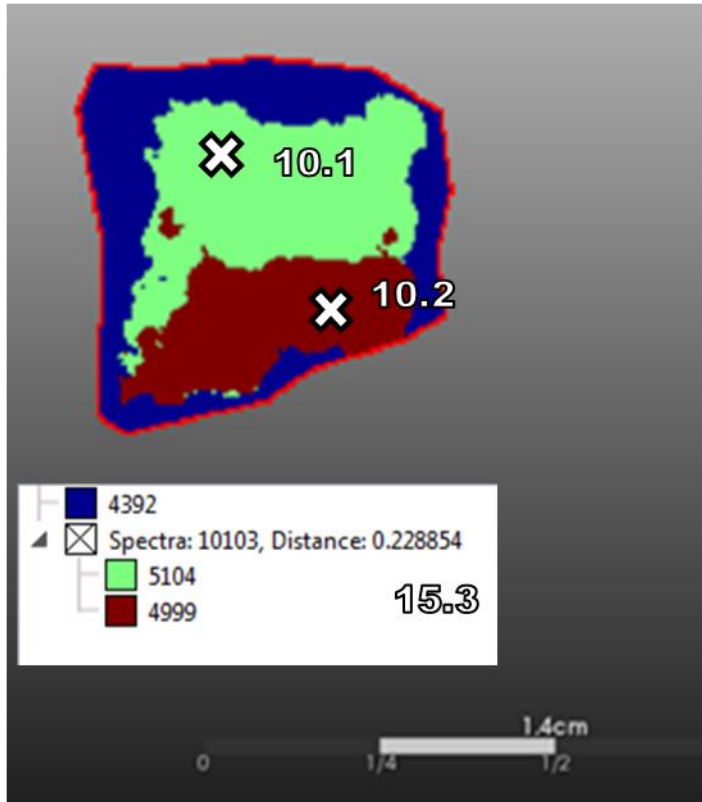
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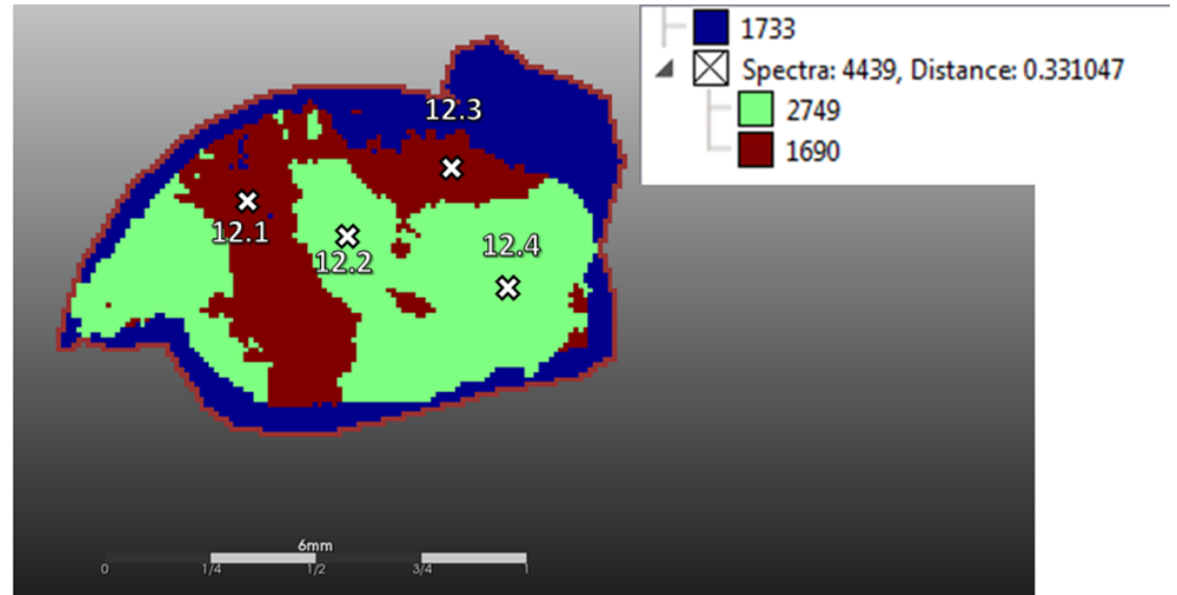
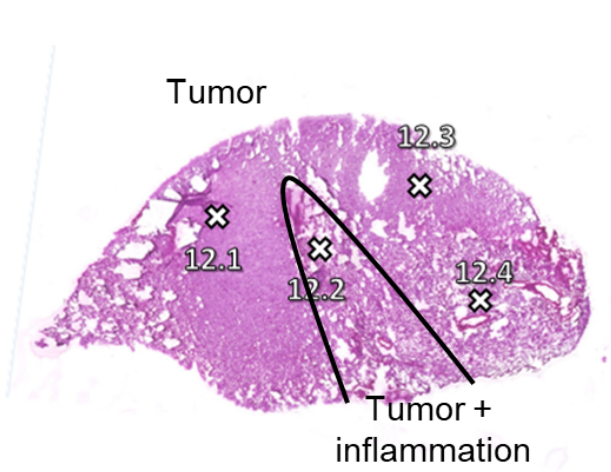


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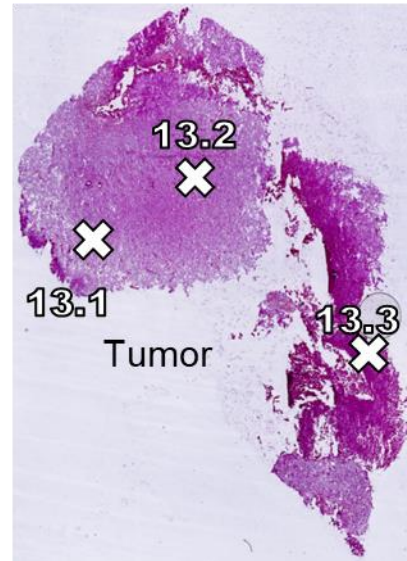
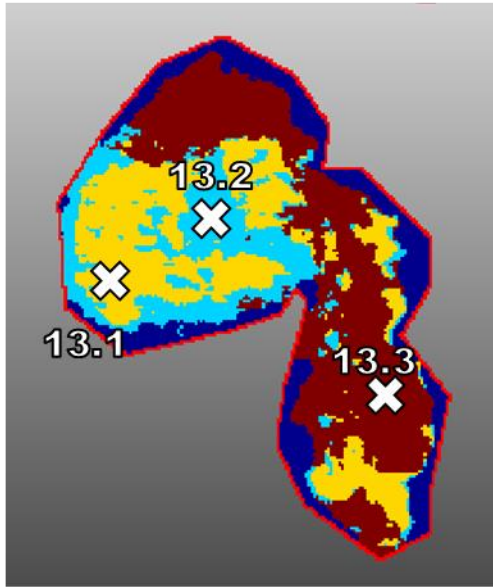




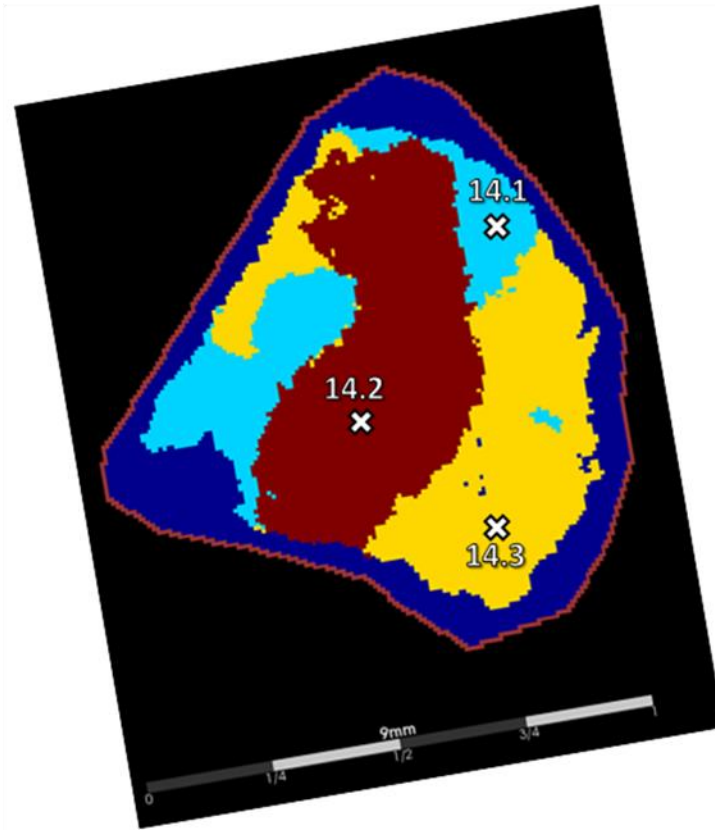
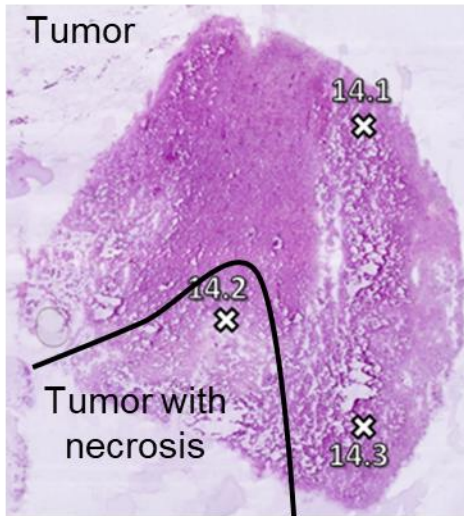
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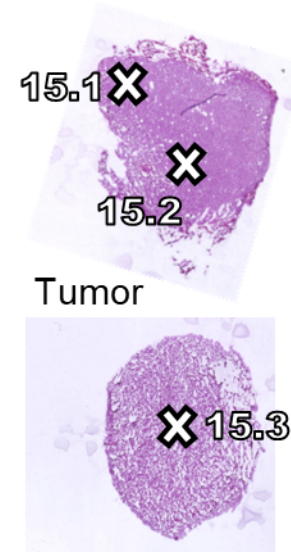
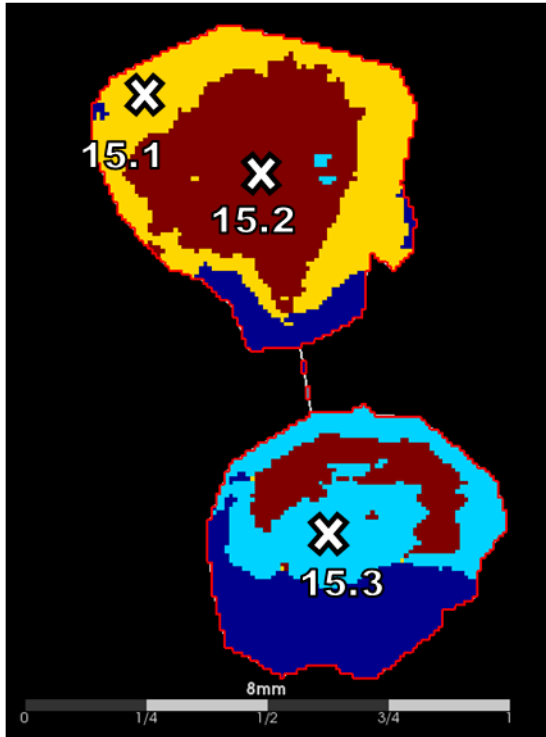
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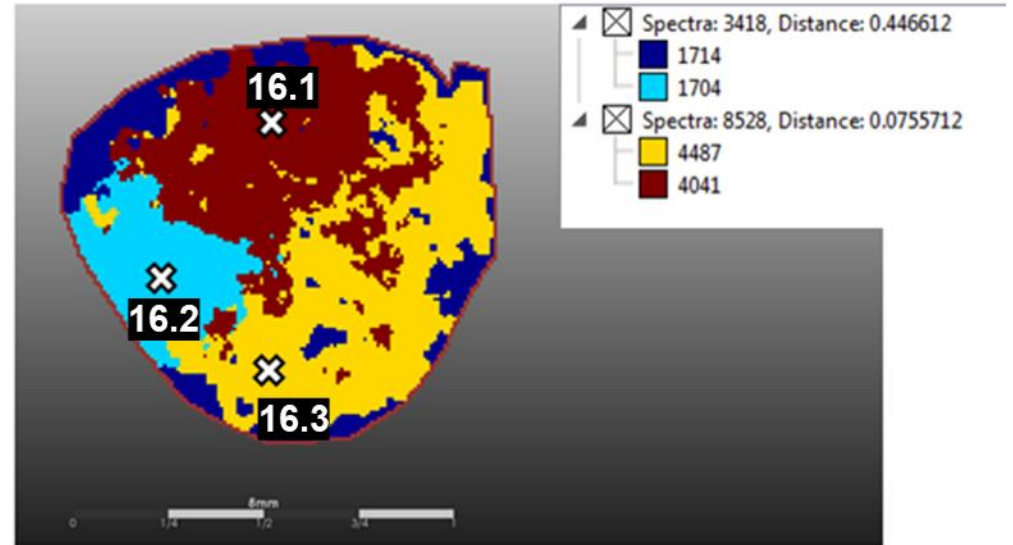
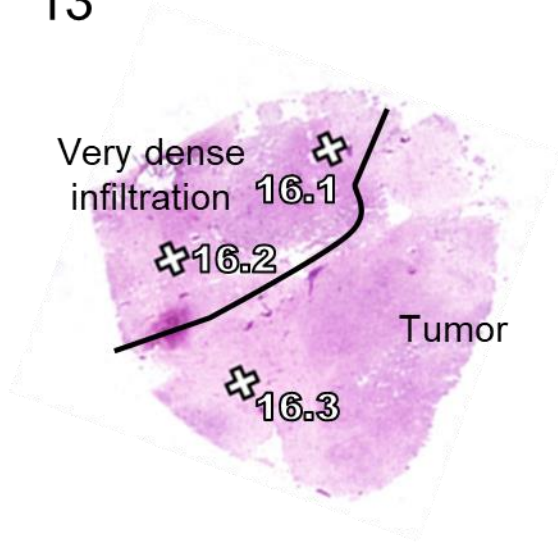
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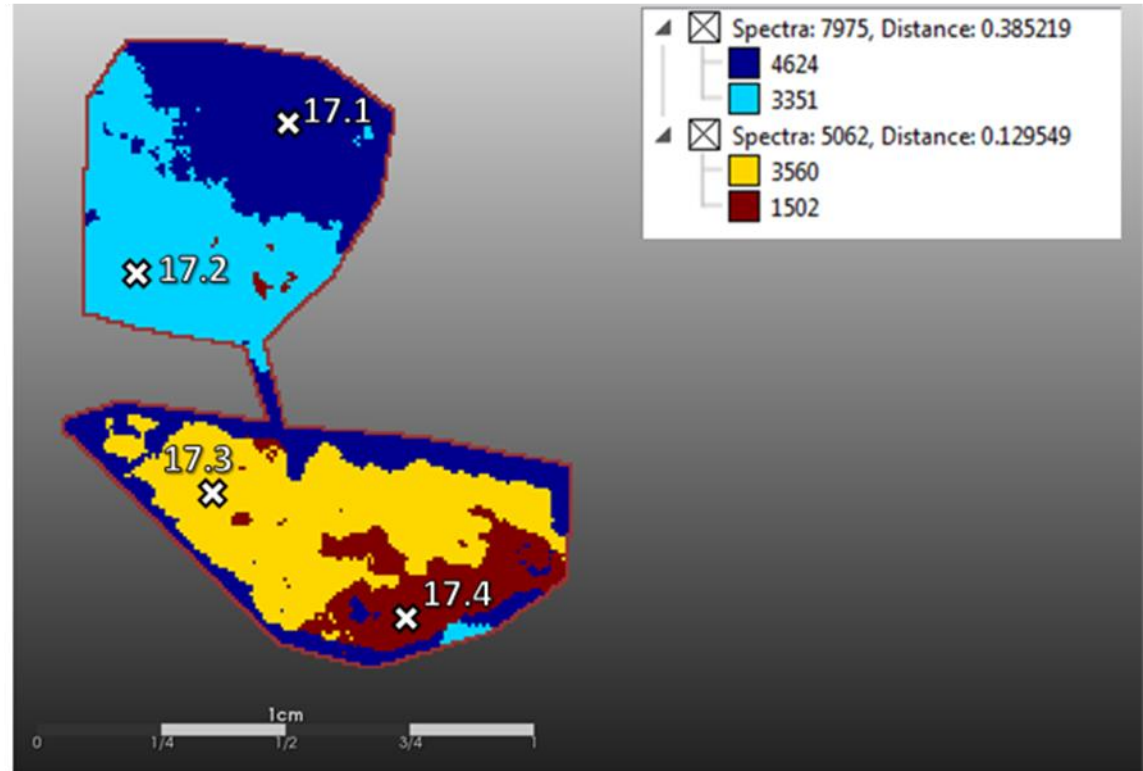
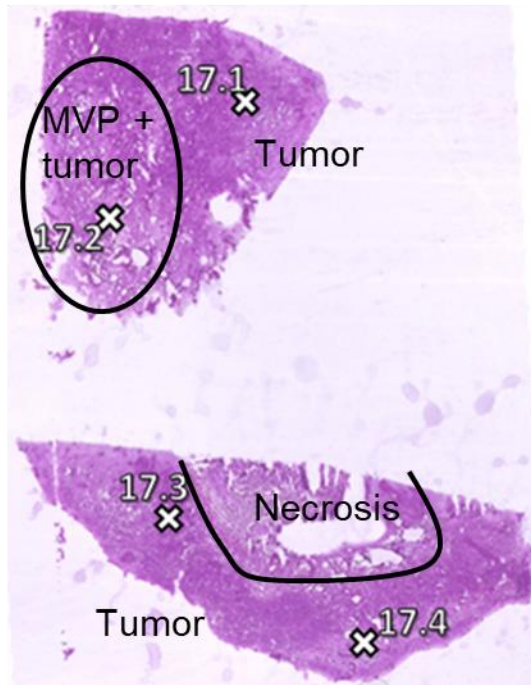
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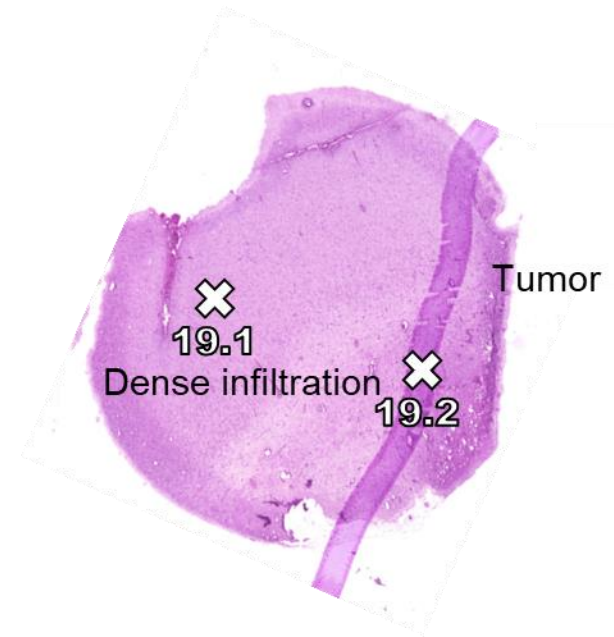
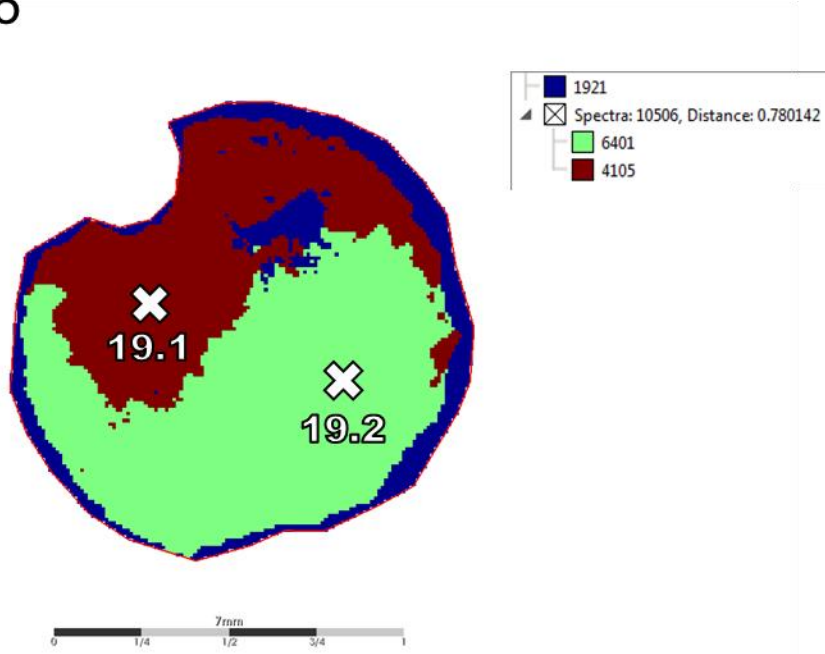
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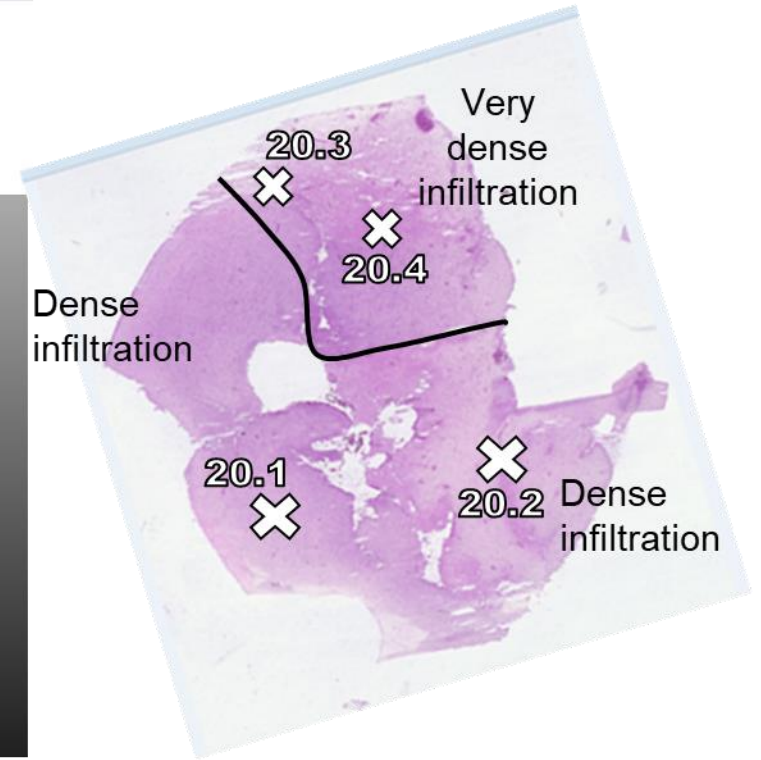
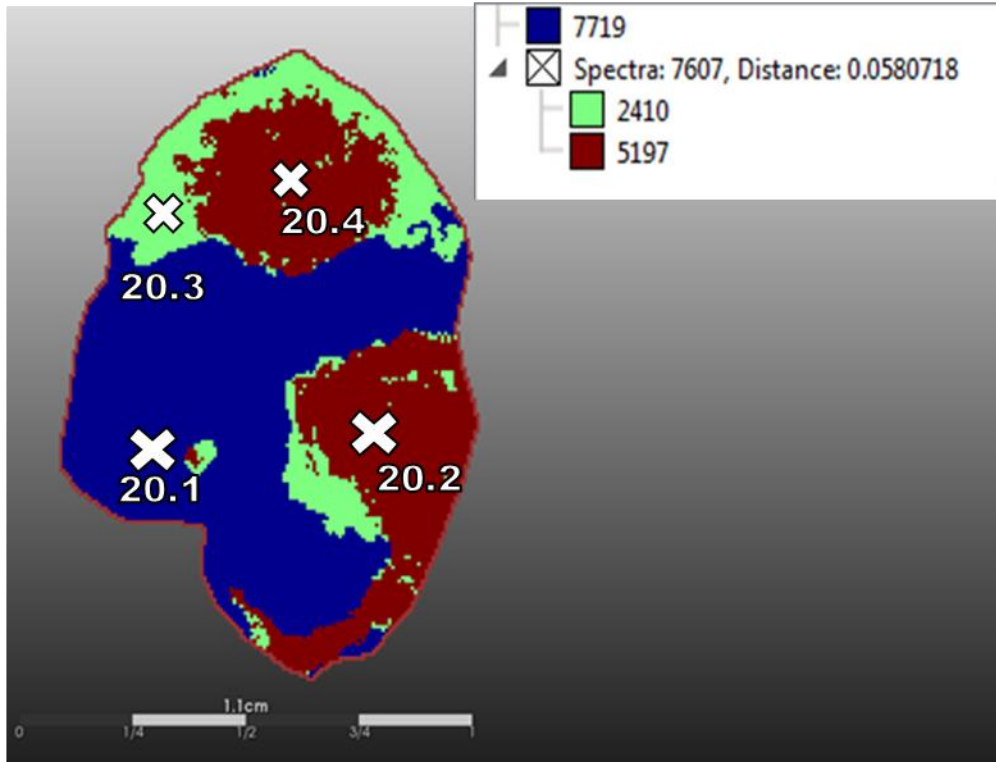
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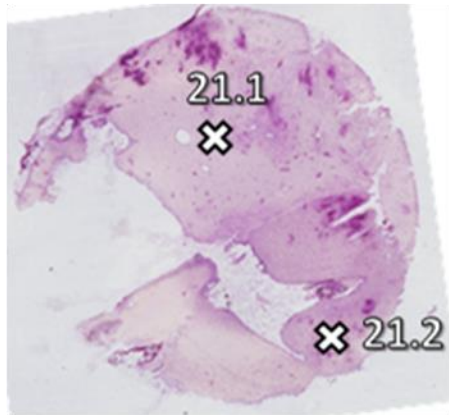


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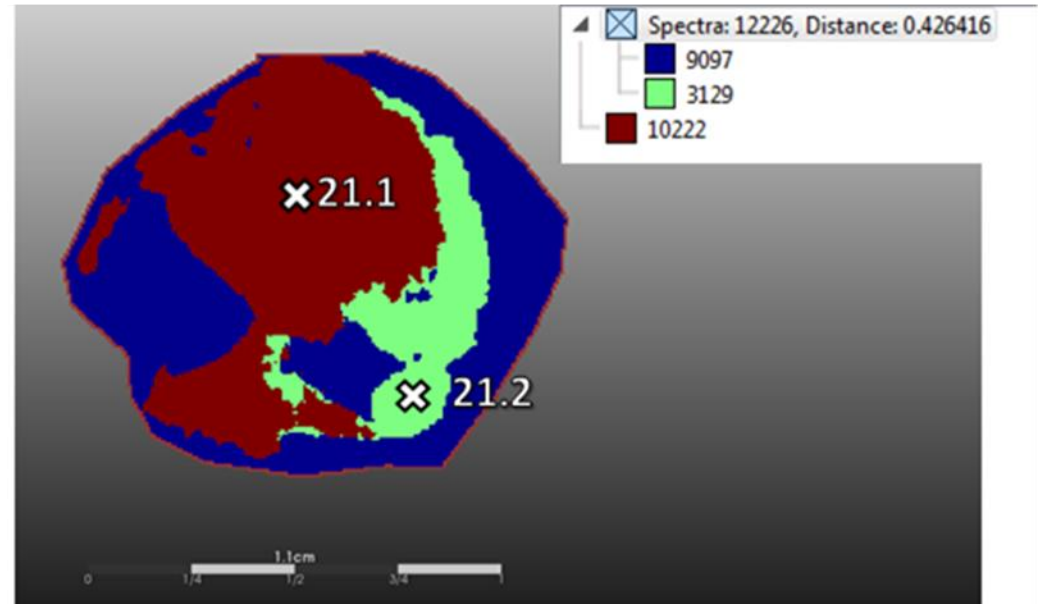




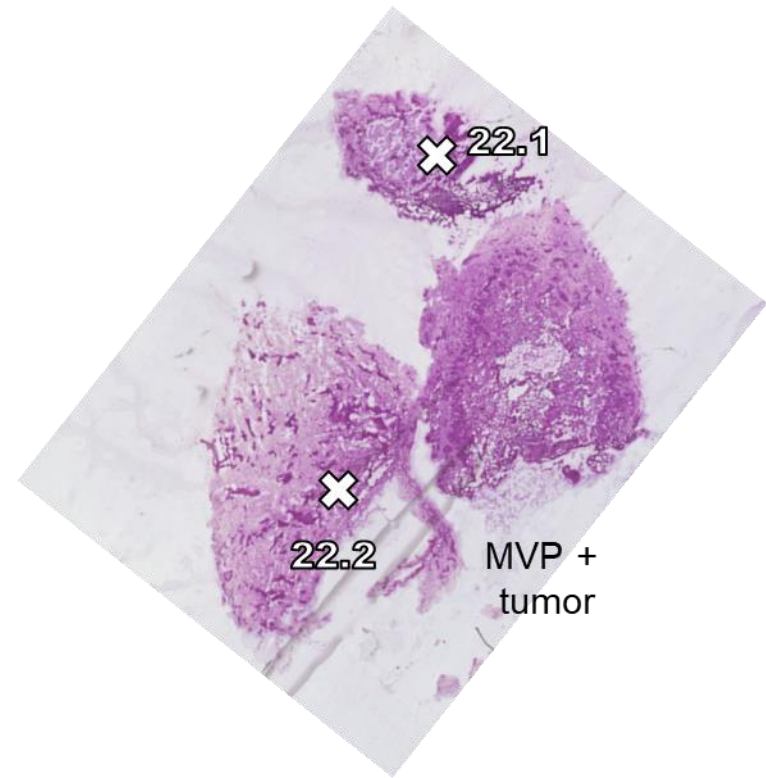
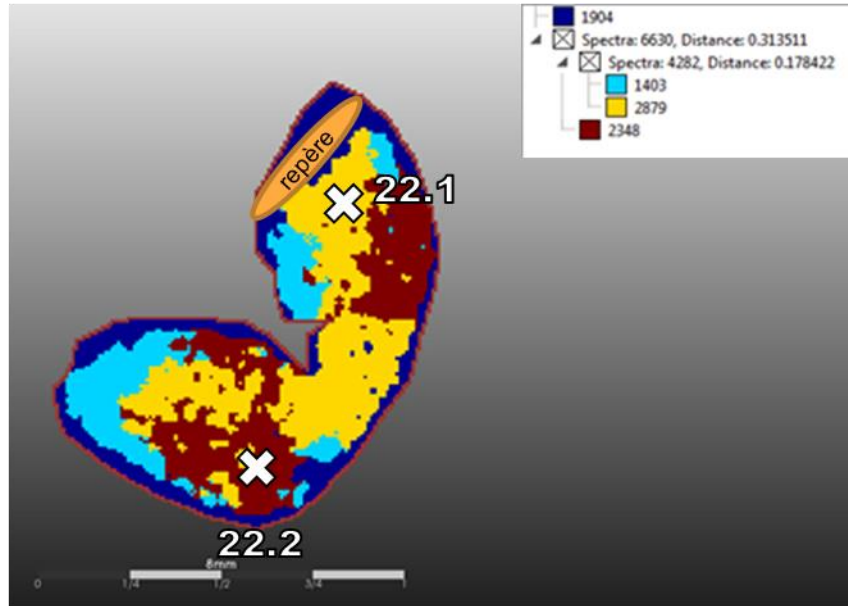
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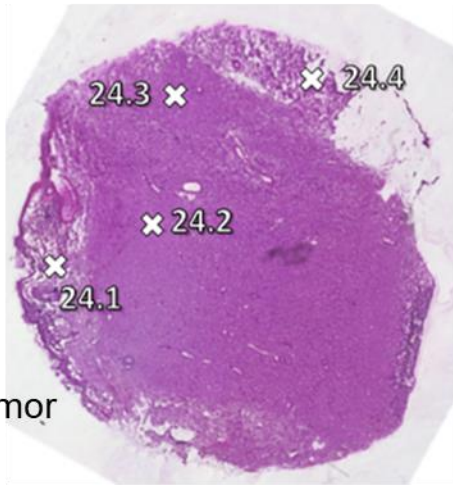
Dense infiltration



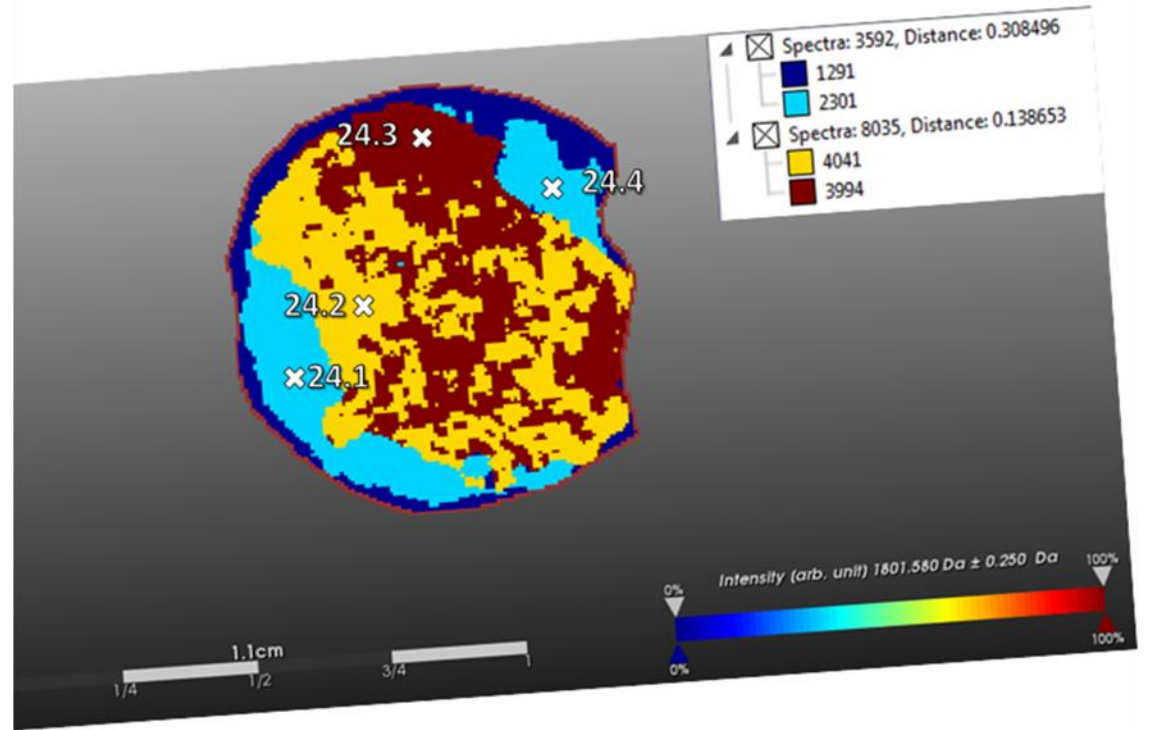
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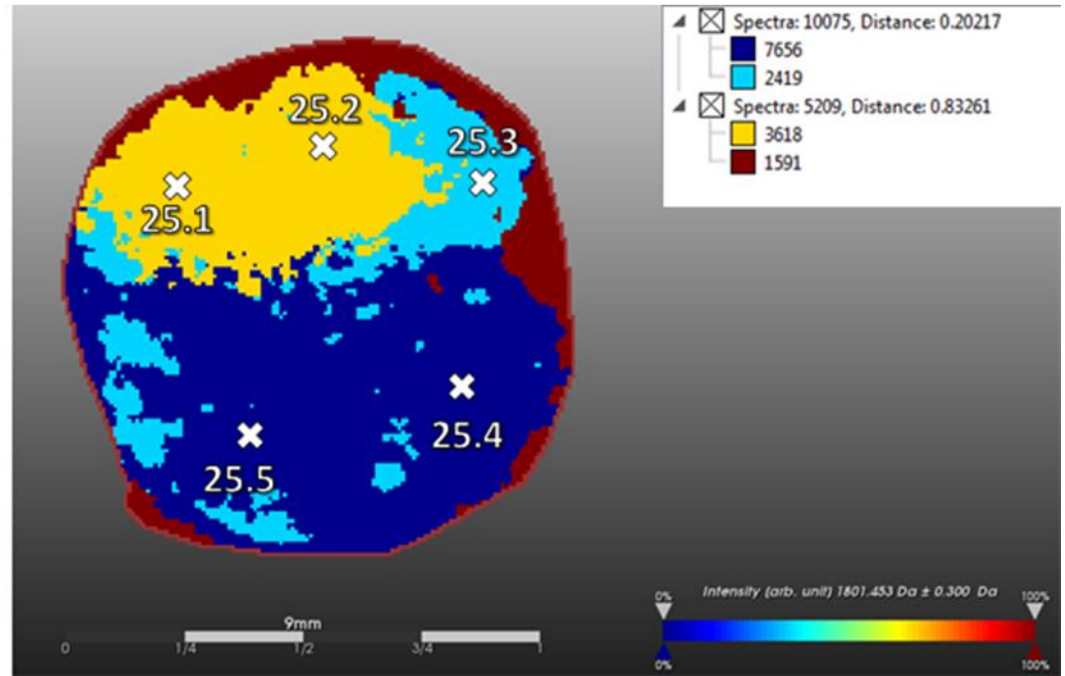
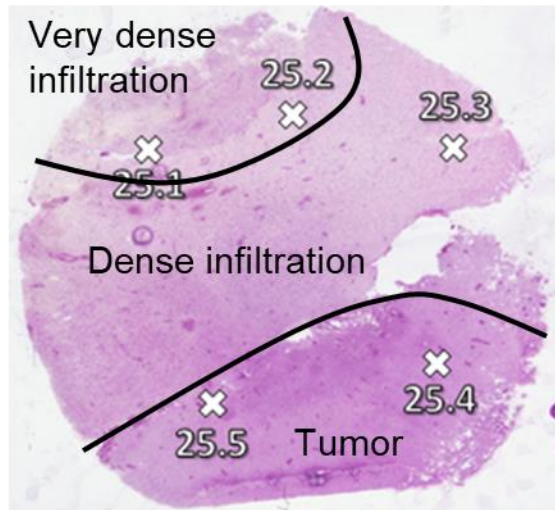
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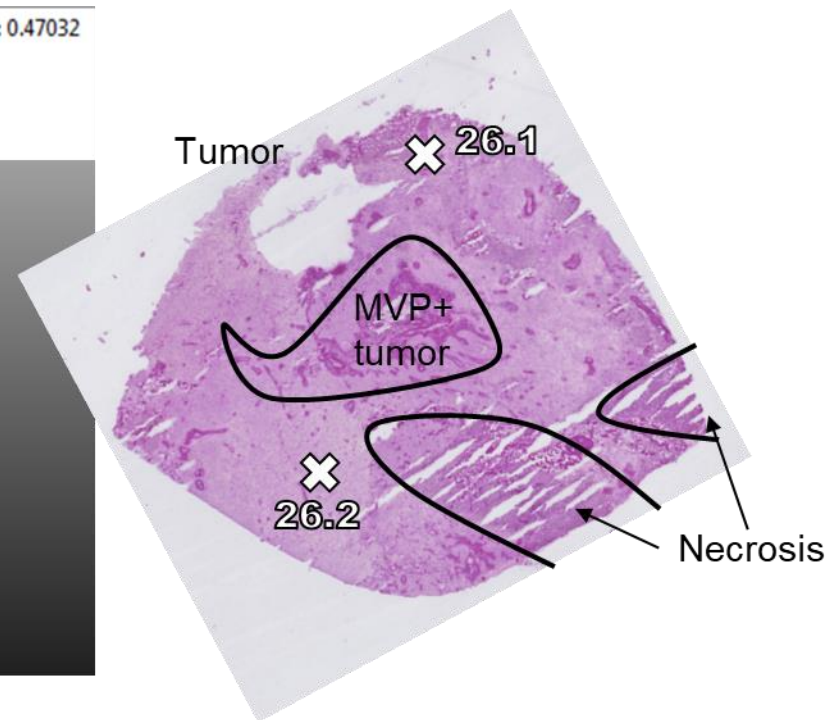
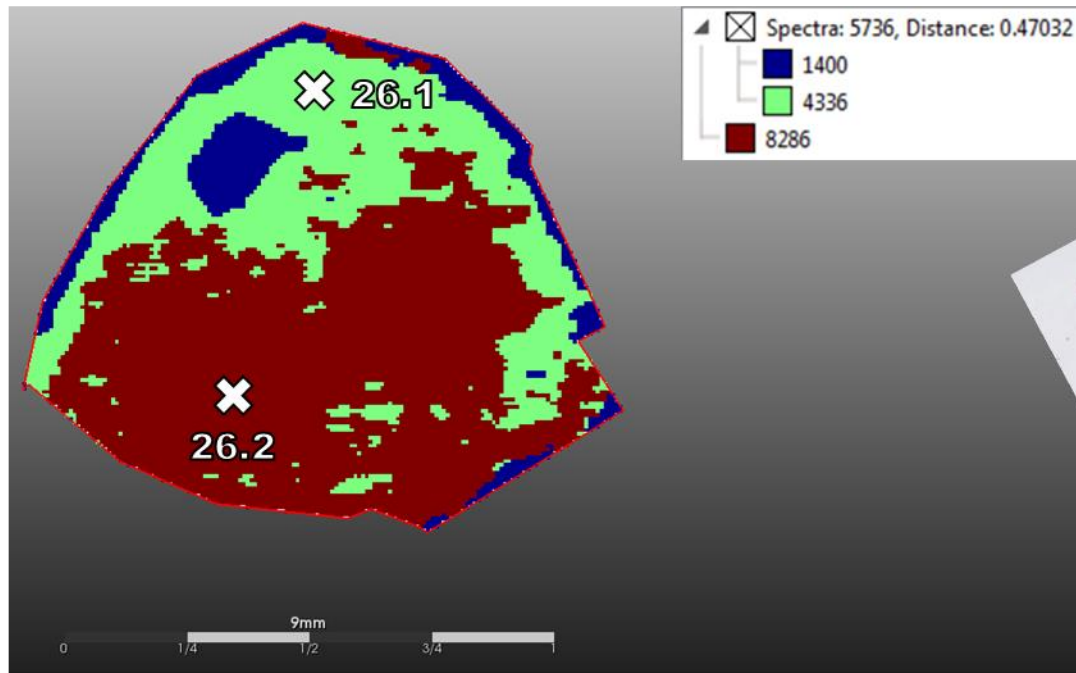
Tumor



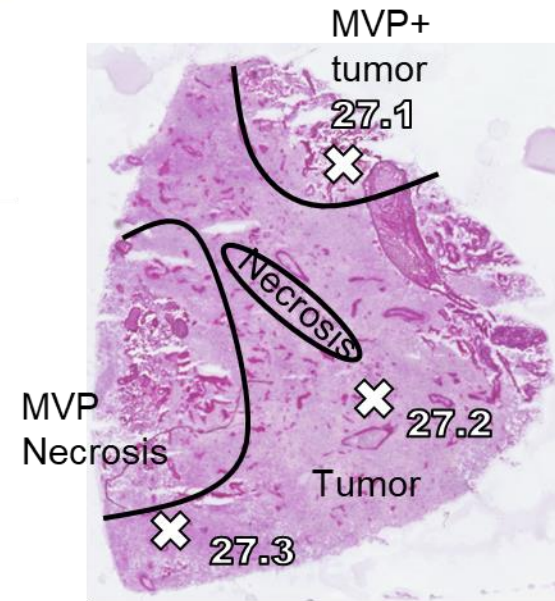
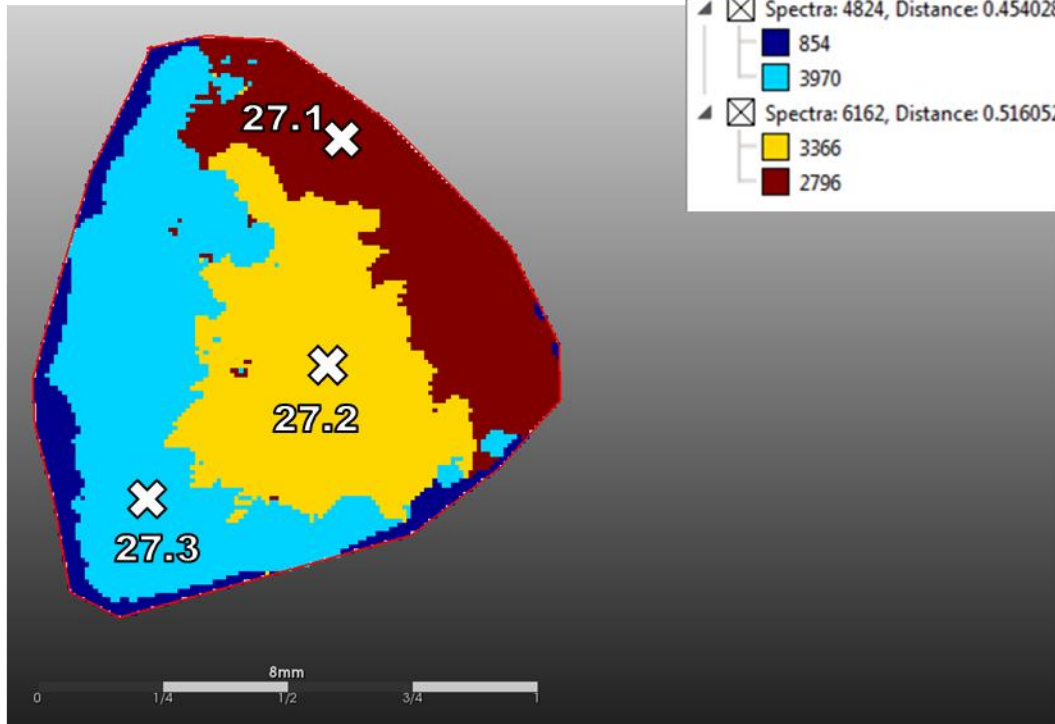
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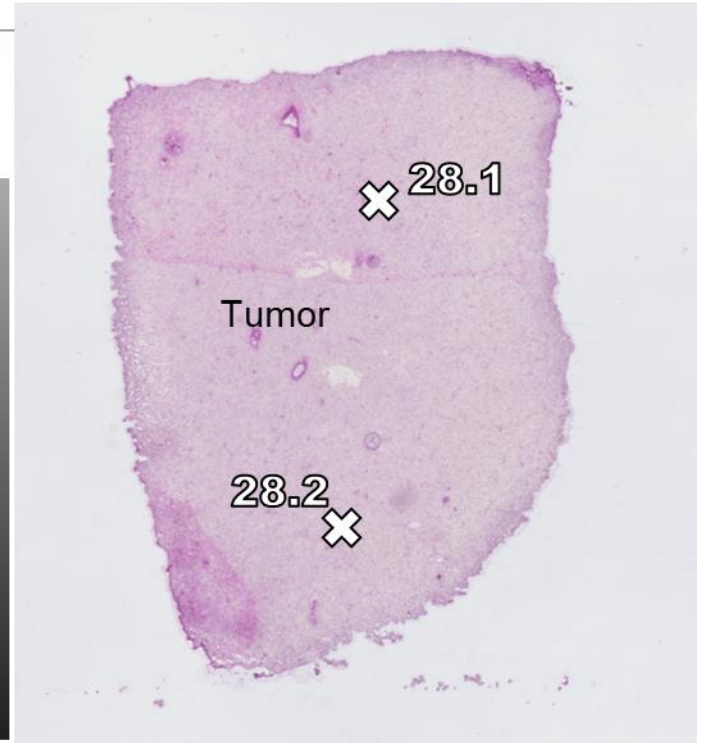
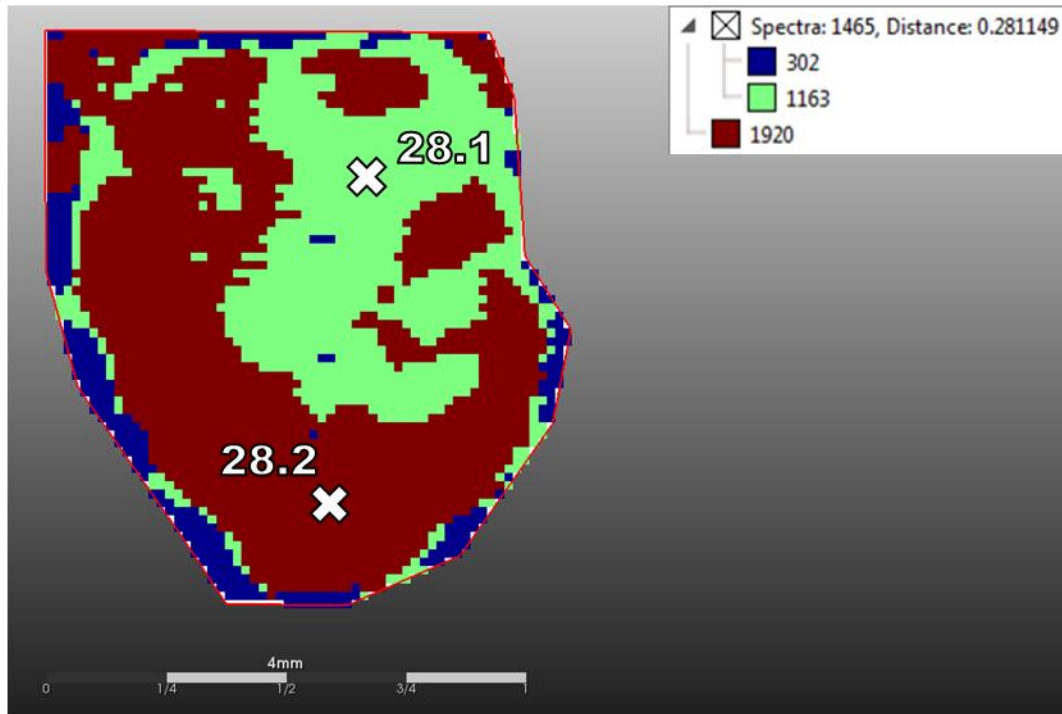
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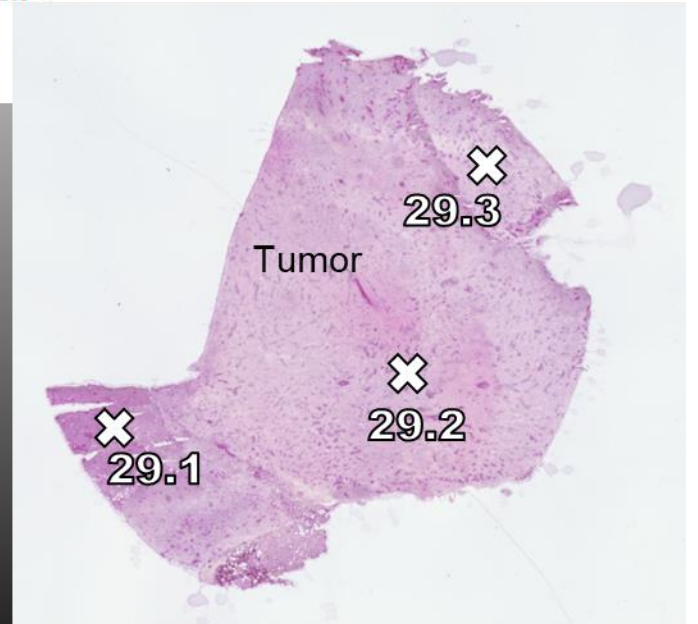
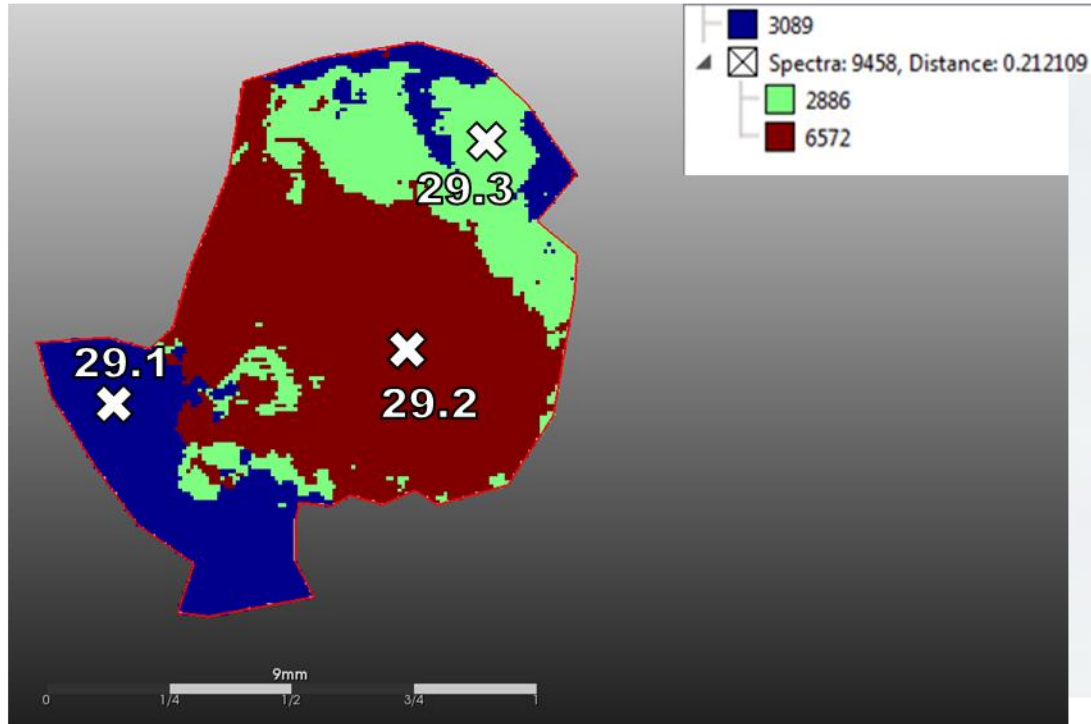
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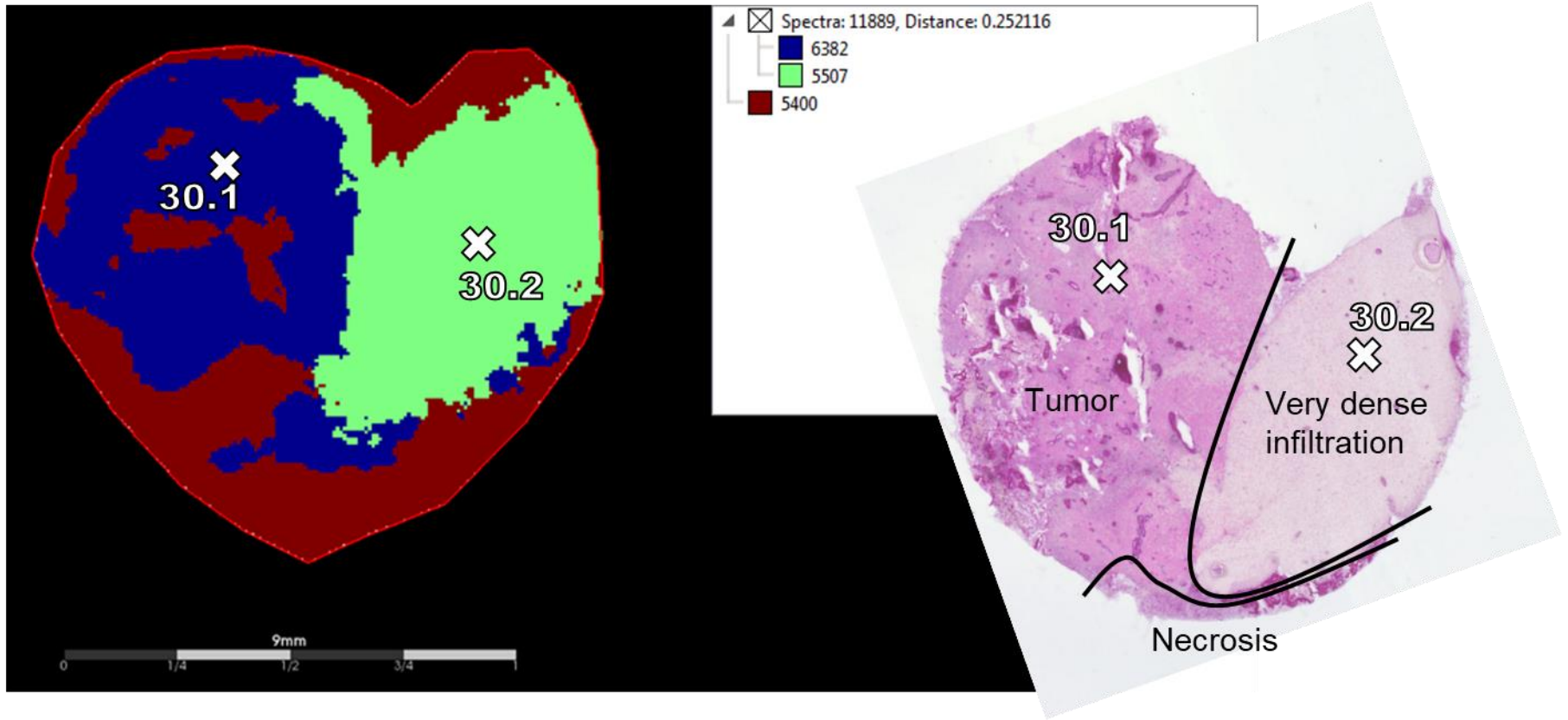


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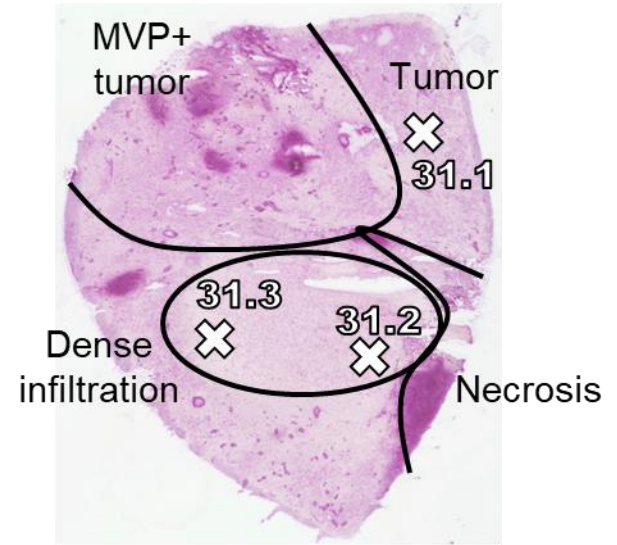
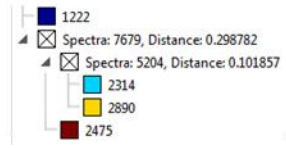
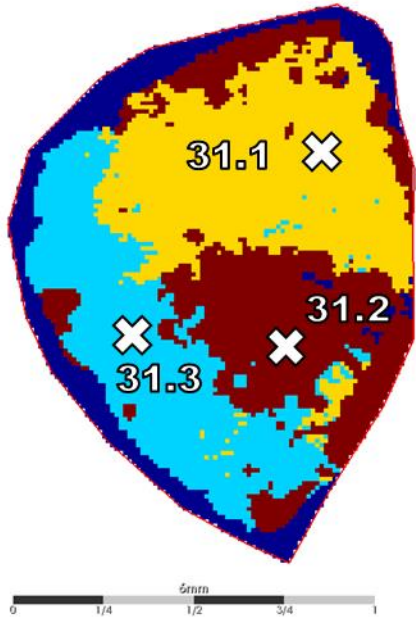




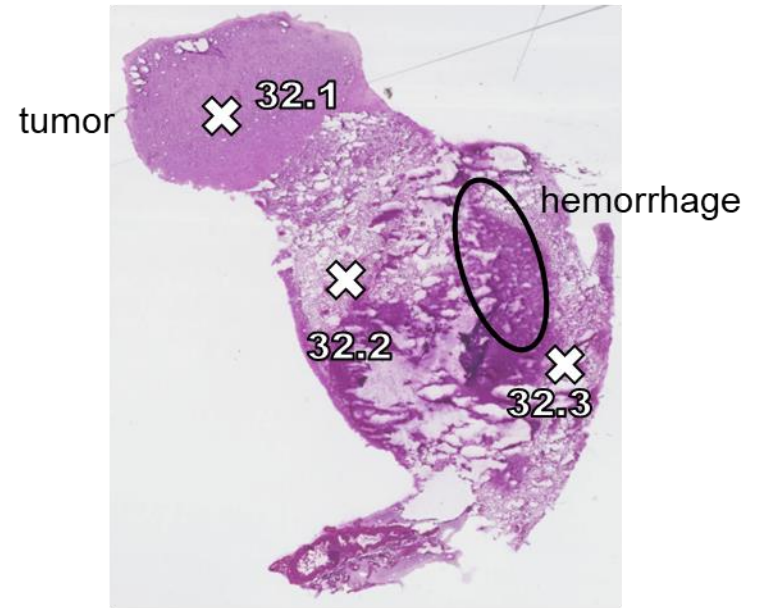
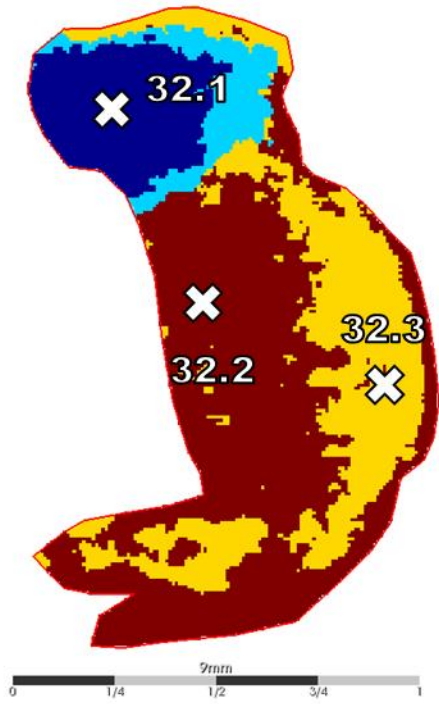
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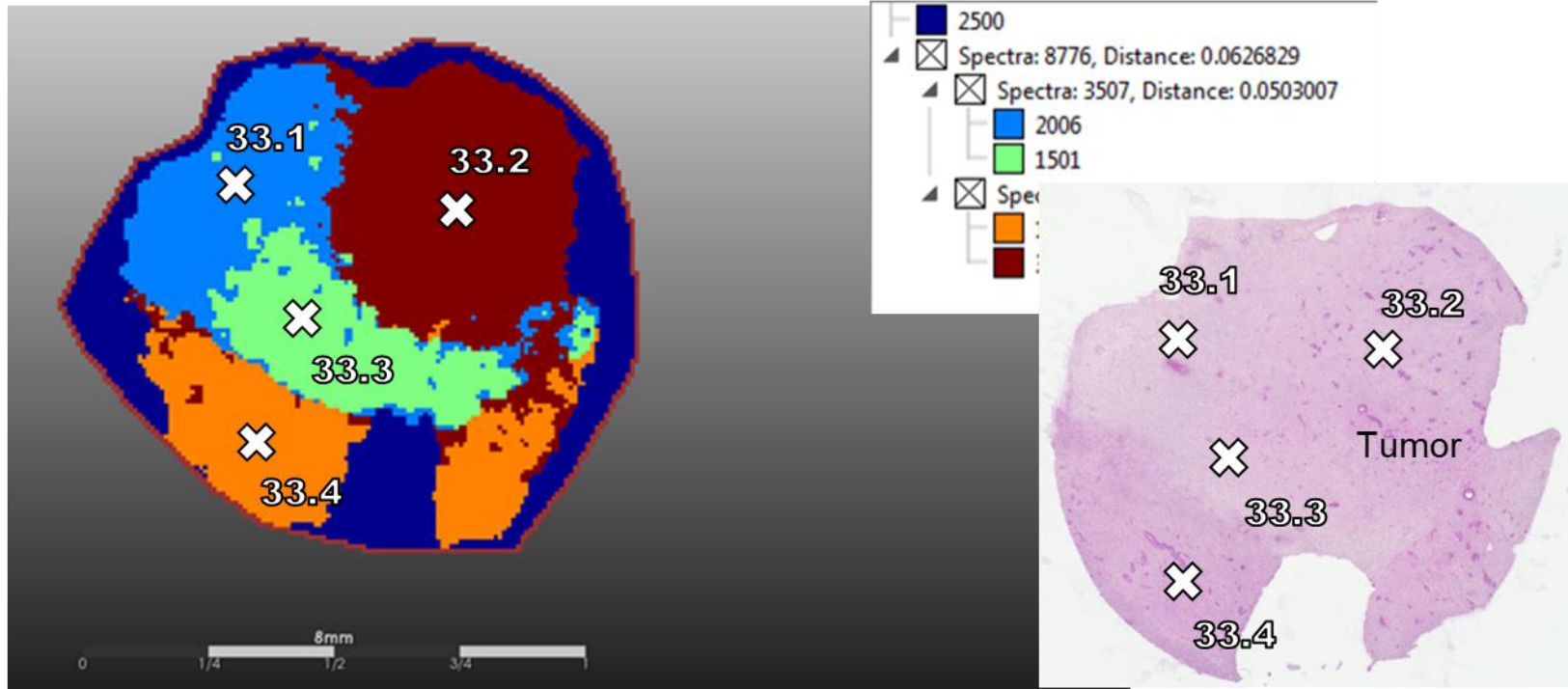


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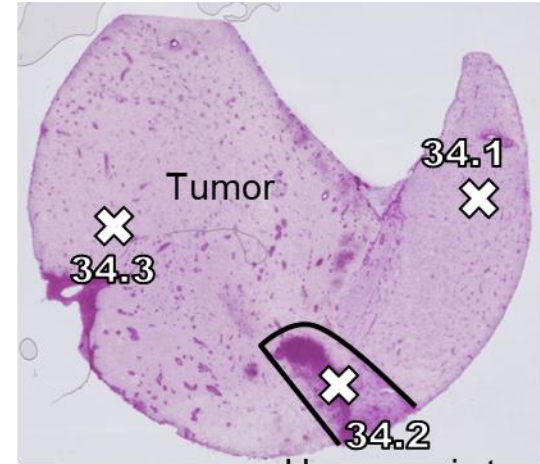
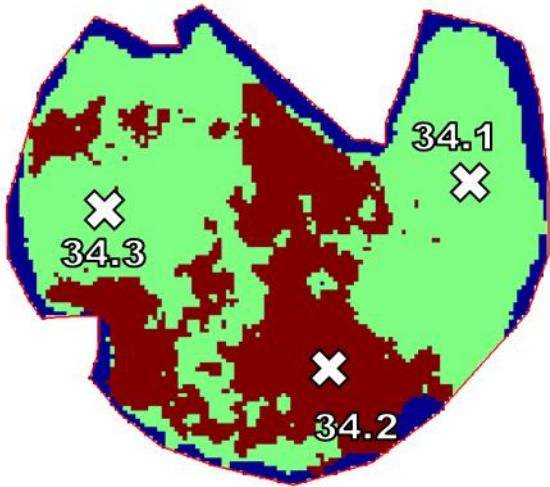


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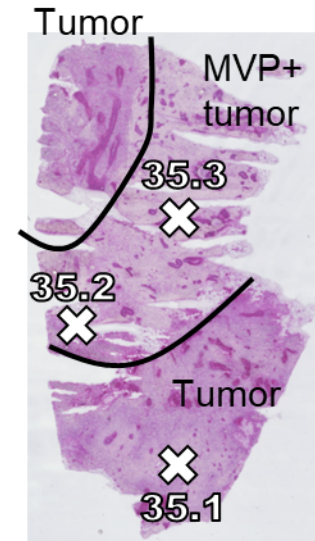
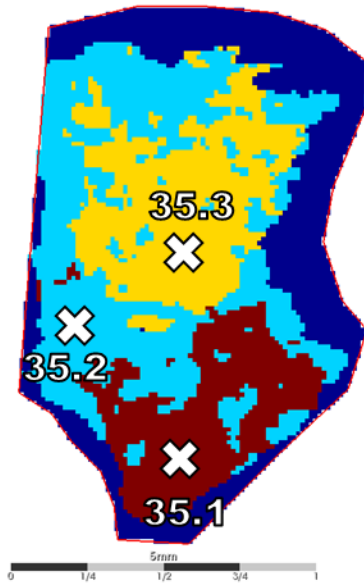
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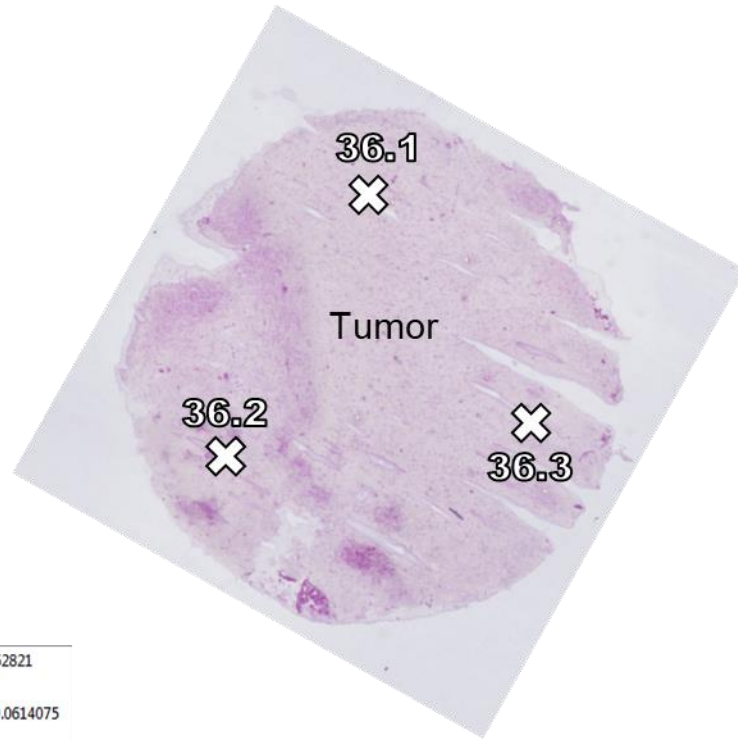
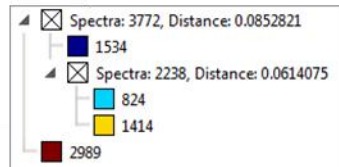
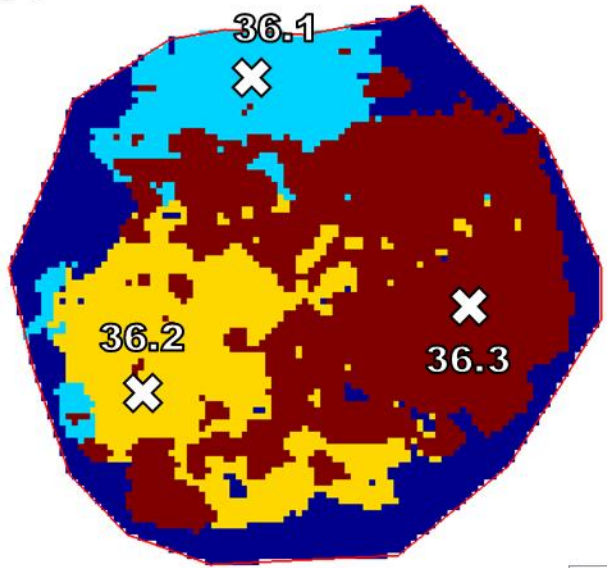
Tumor

Hemorrhagic tumor

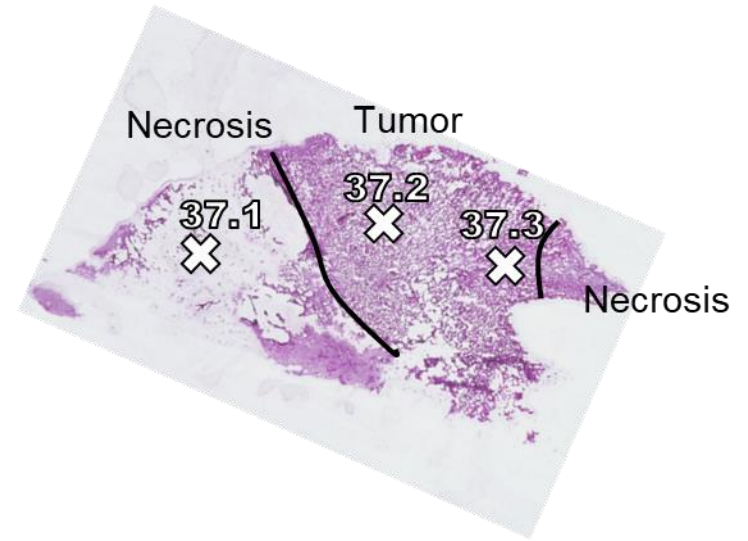
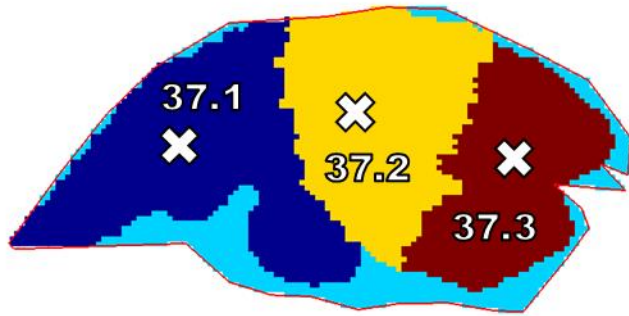
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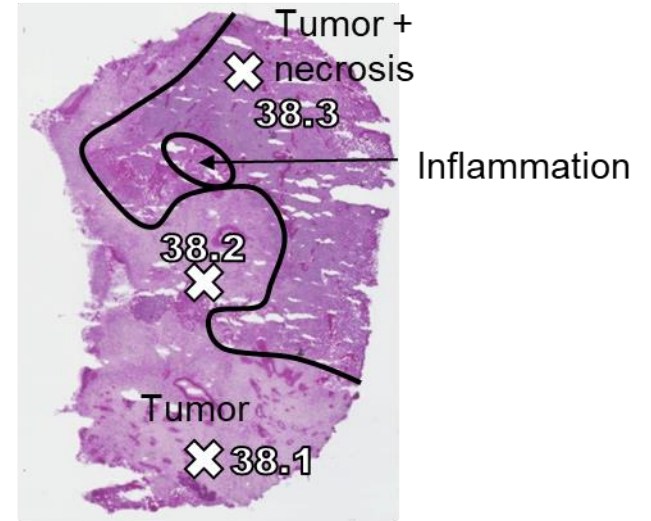
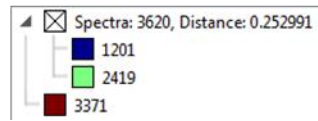
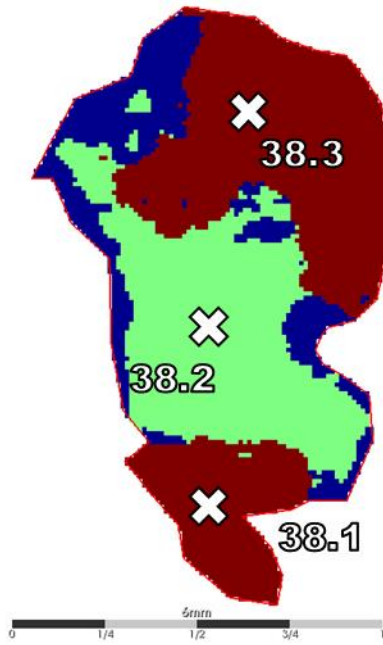


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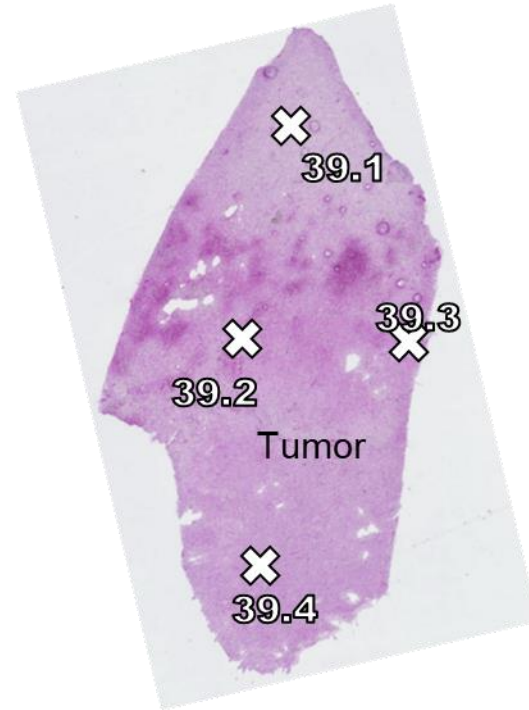
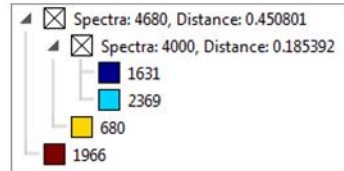
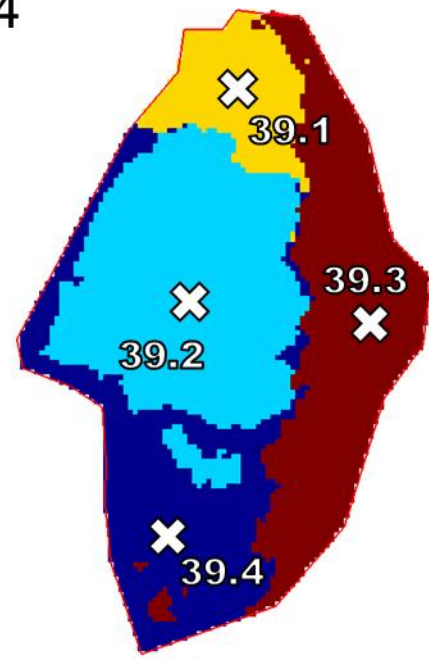




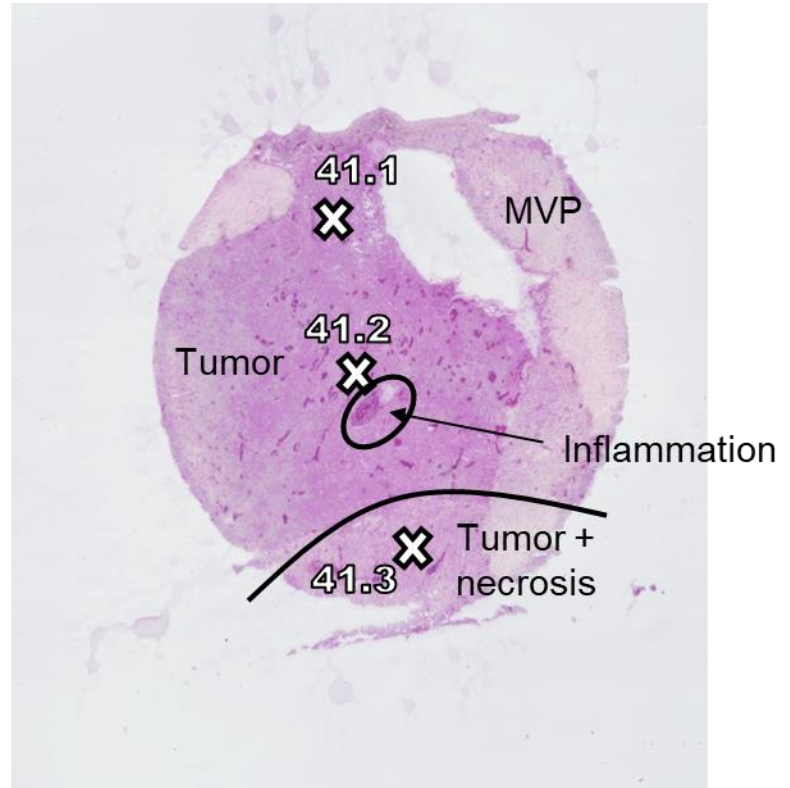
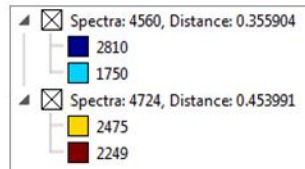
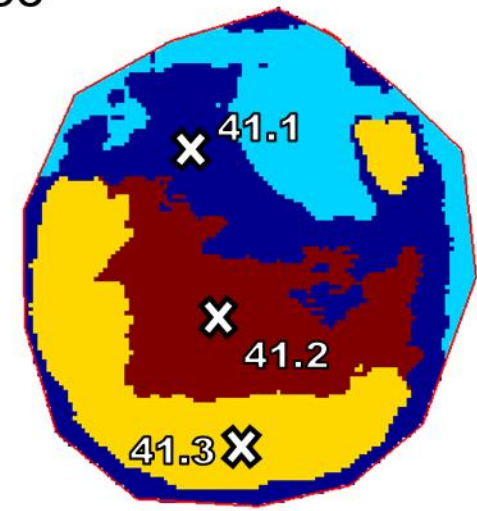
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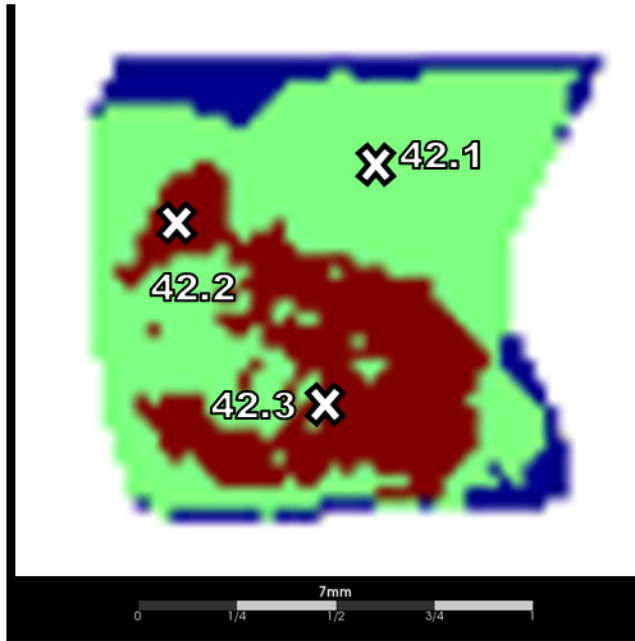
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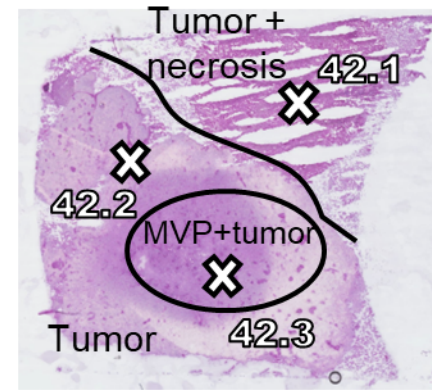
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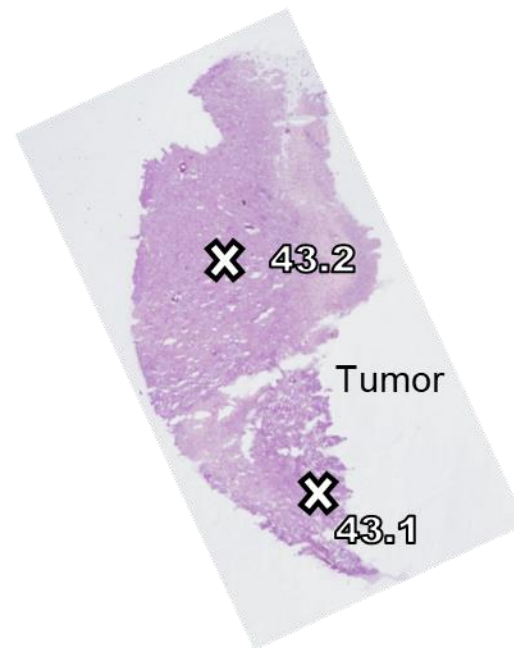
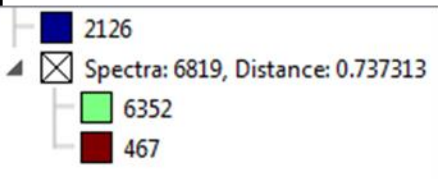
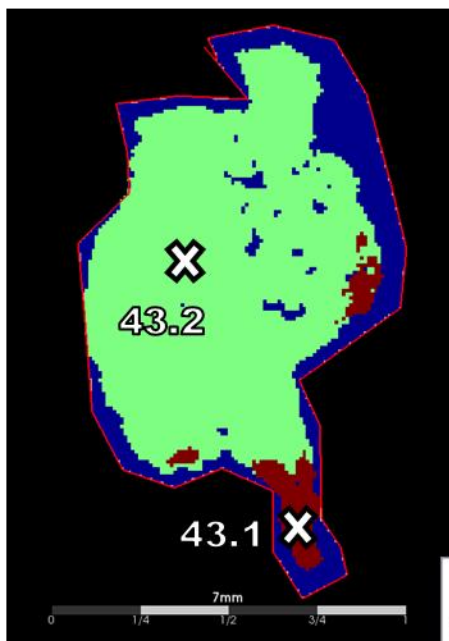
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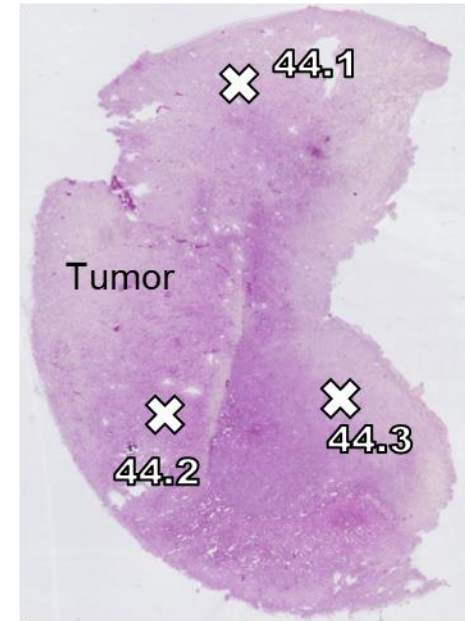
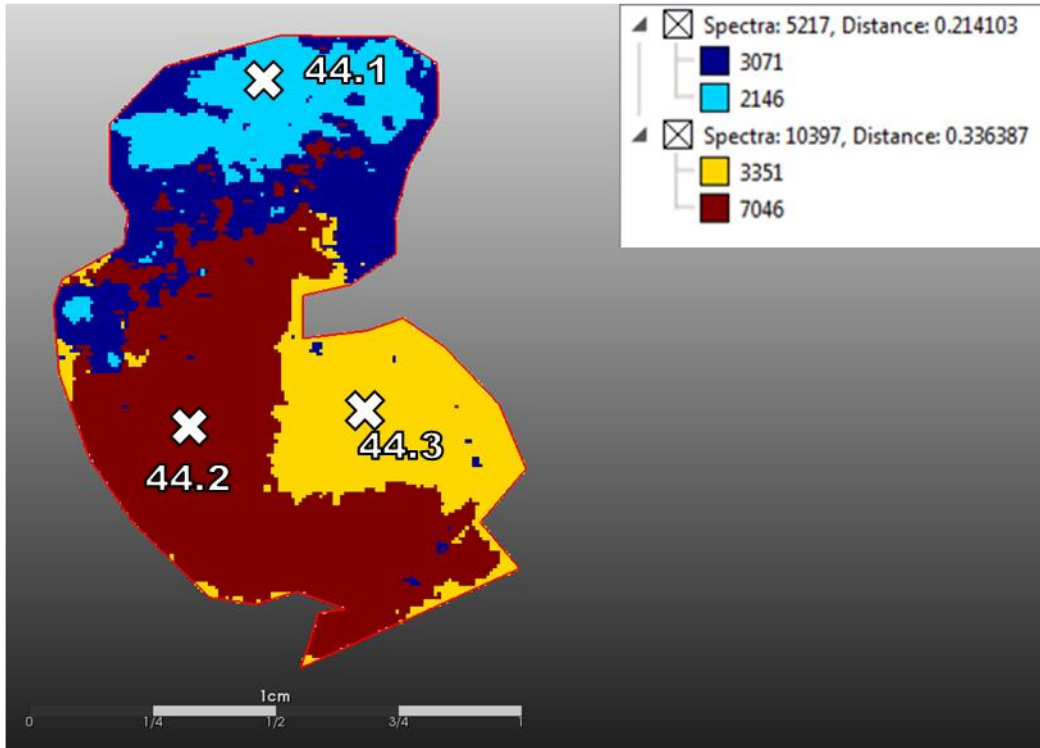


- ☒ Spectra: 7707, Distance: 0.72712
- ☒ Spectra: 5626, Distance: 0.243224
  - 2425
  - 3201
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- ☒ Spectra: 5564, Distance: 0.142983
  - 1980
  - 3584

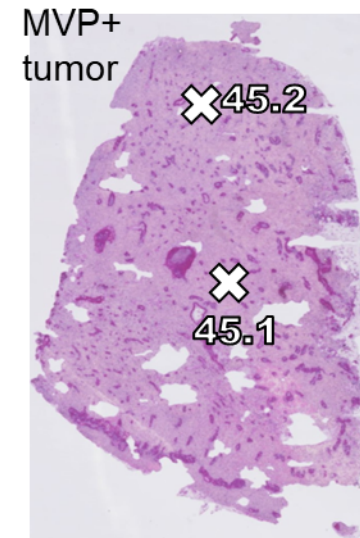
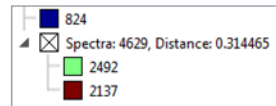
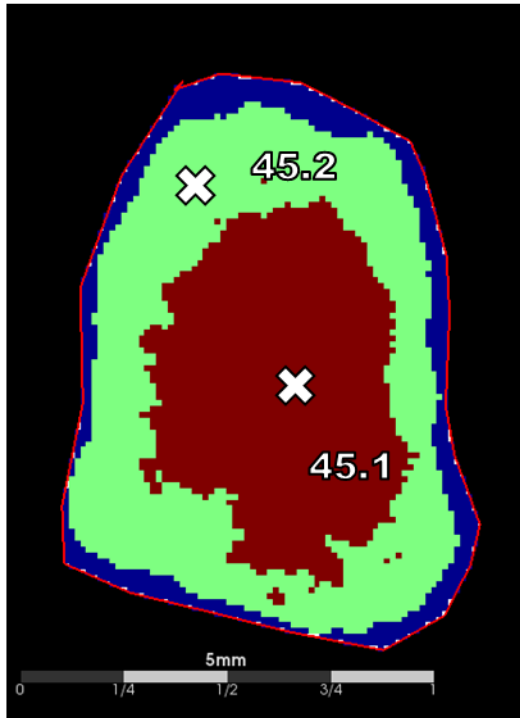


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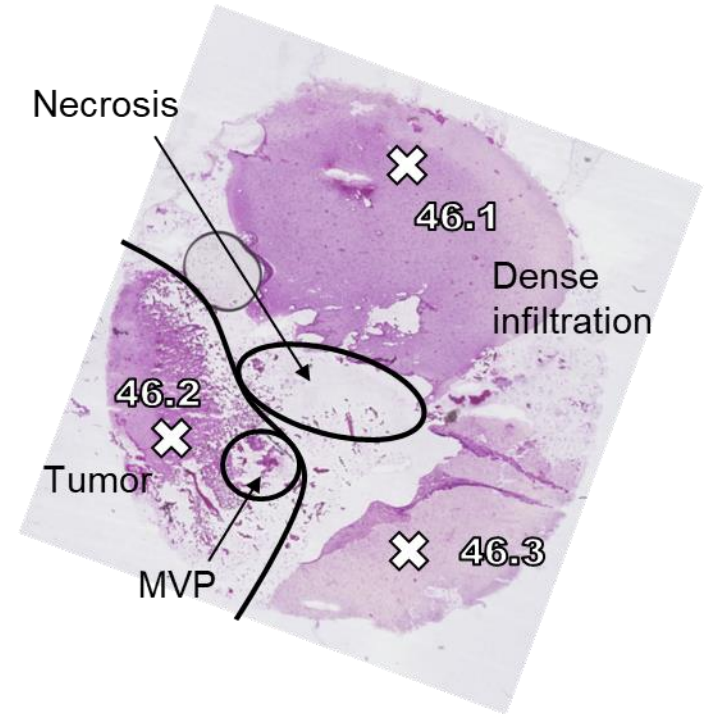
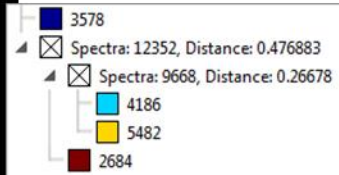
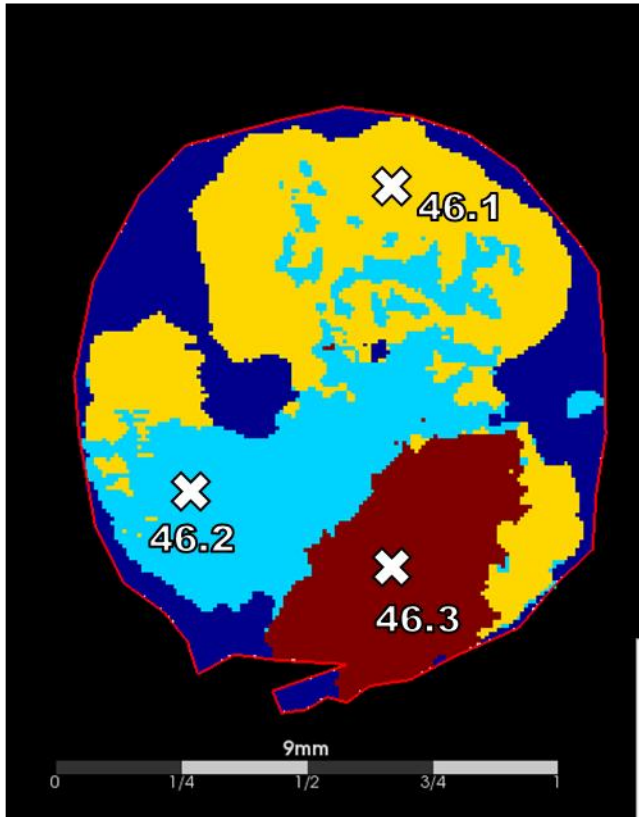




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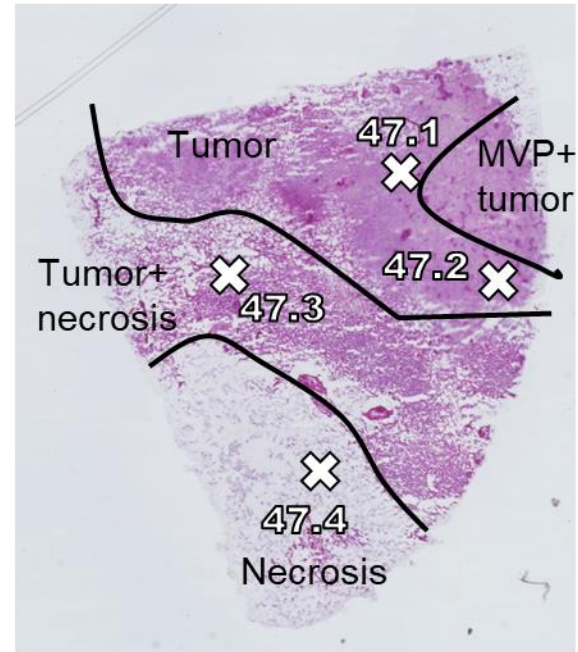
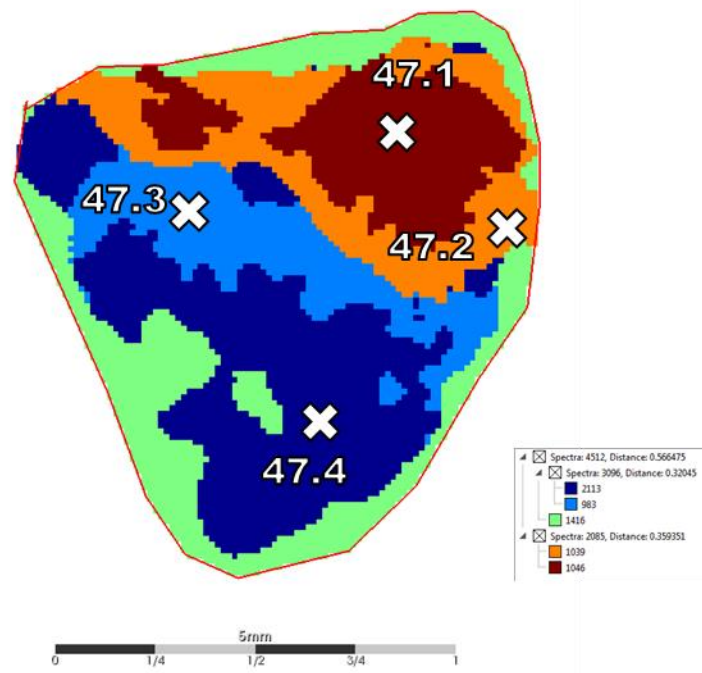


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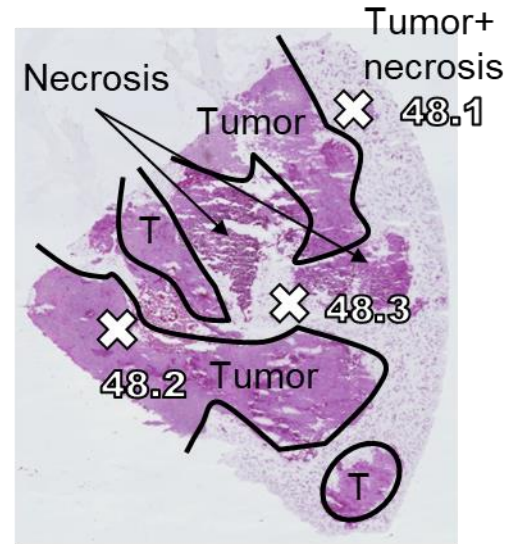
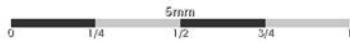
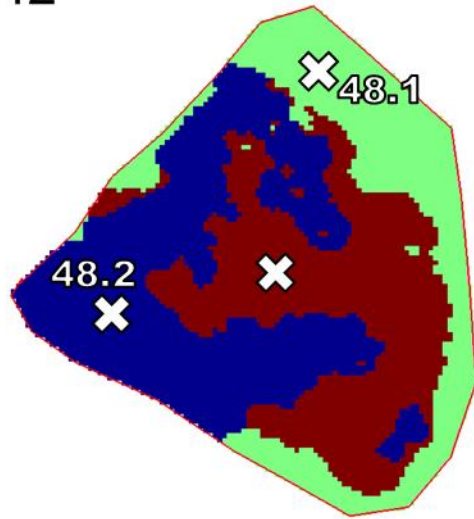




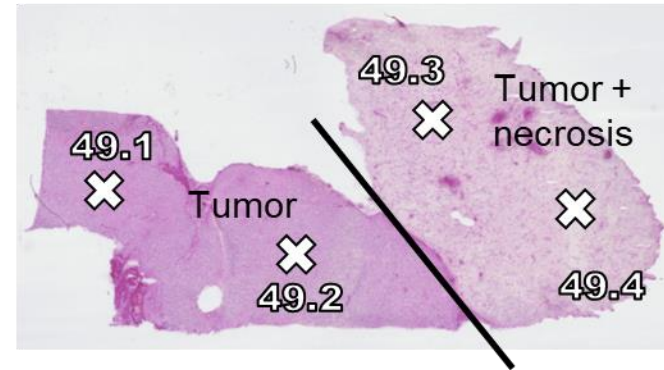
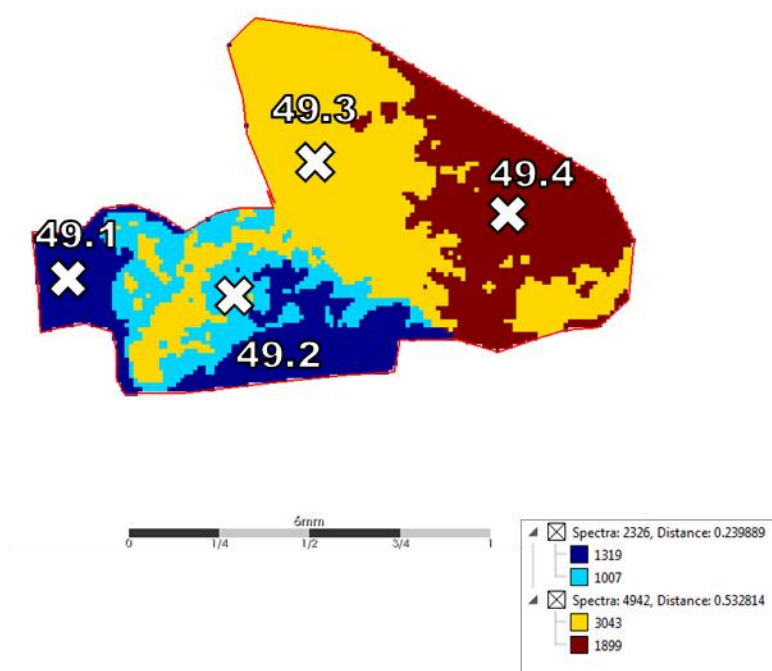
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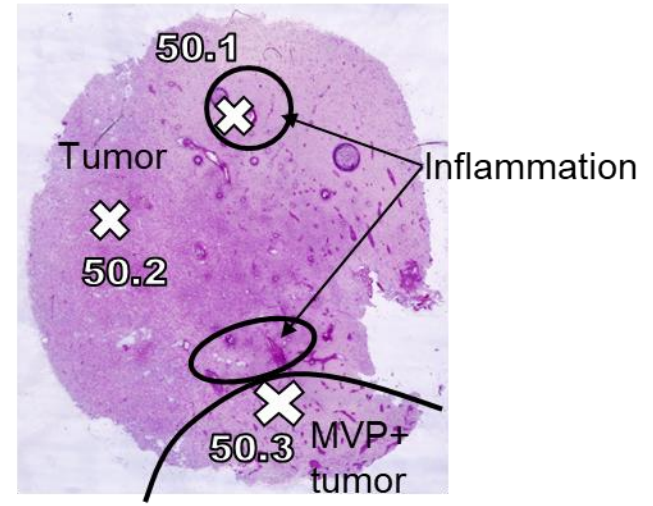
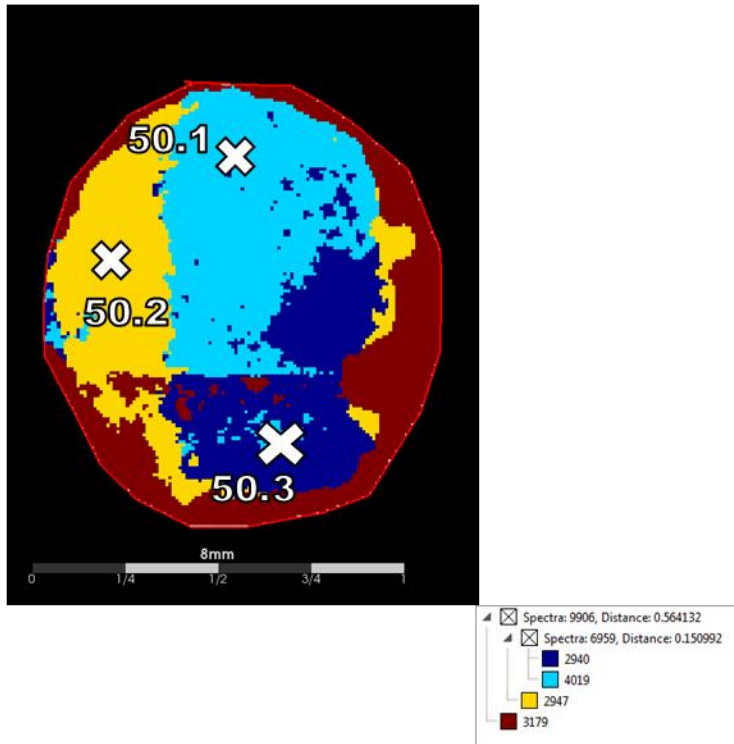


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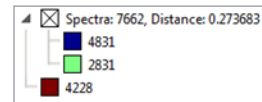
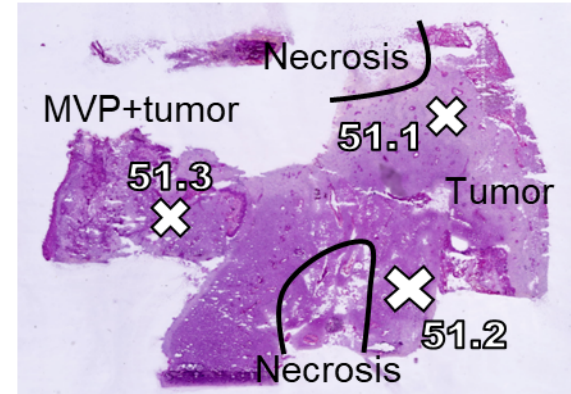
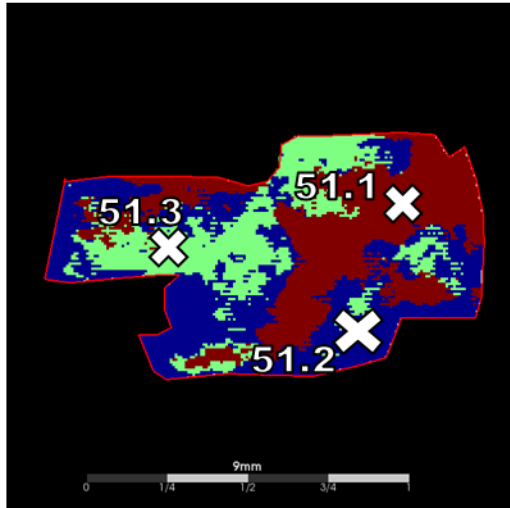


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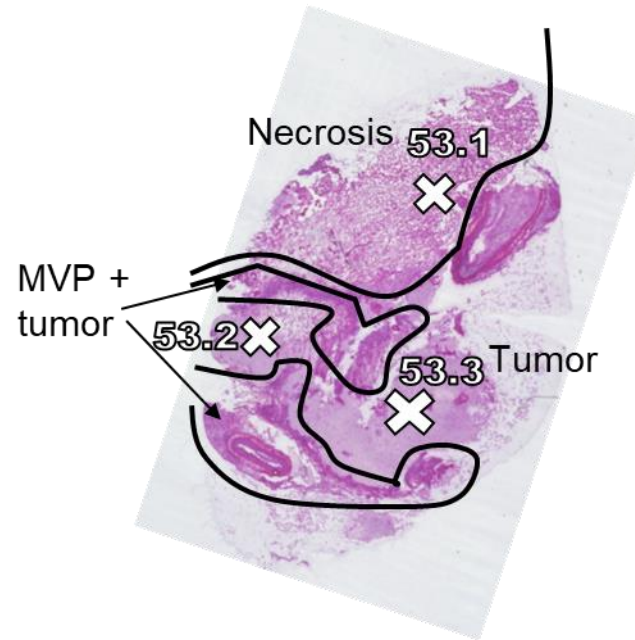
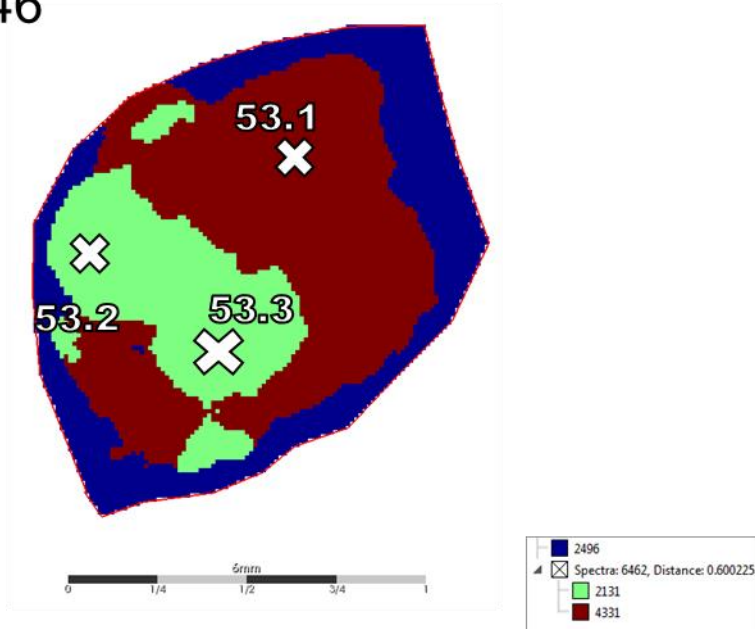




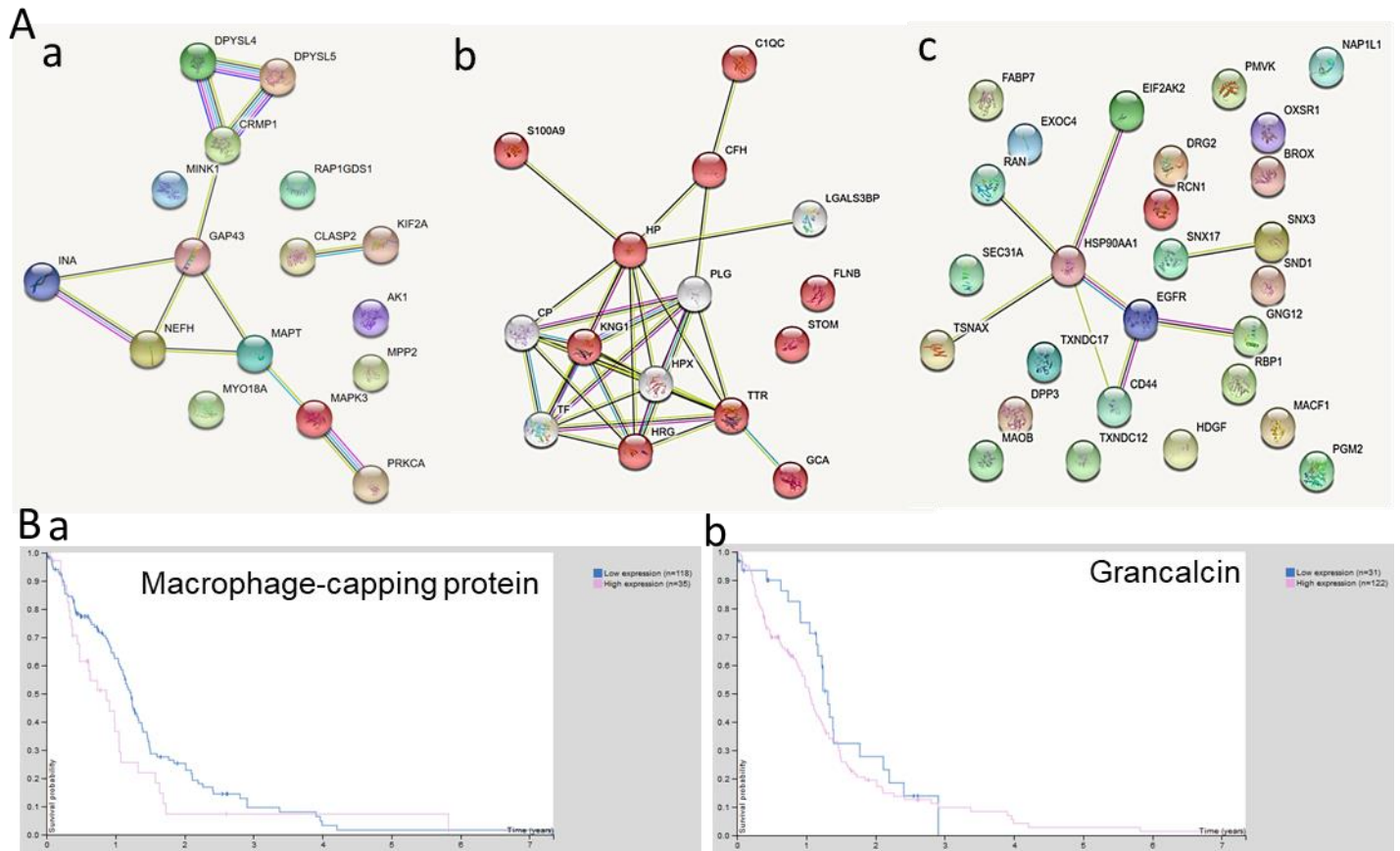
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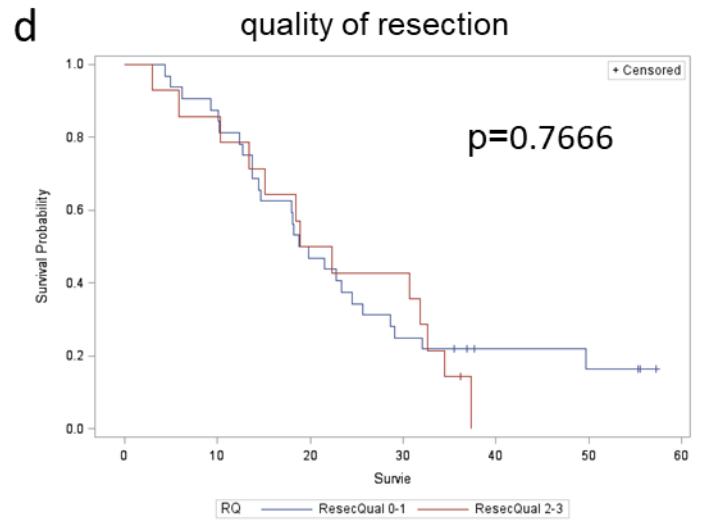
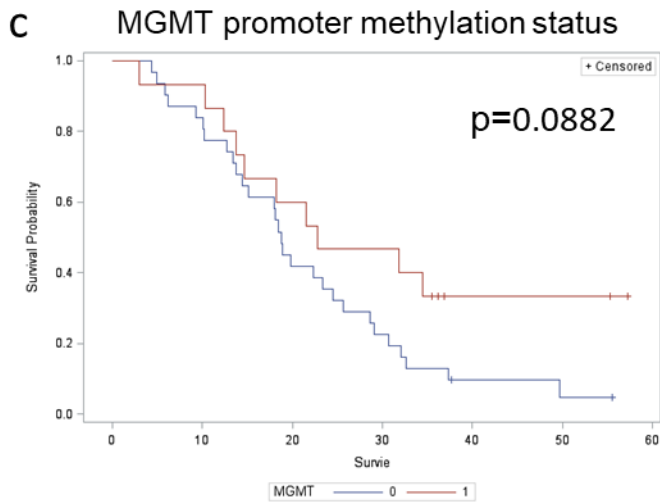
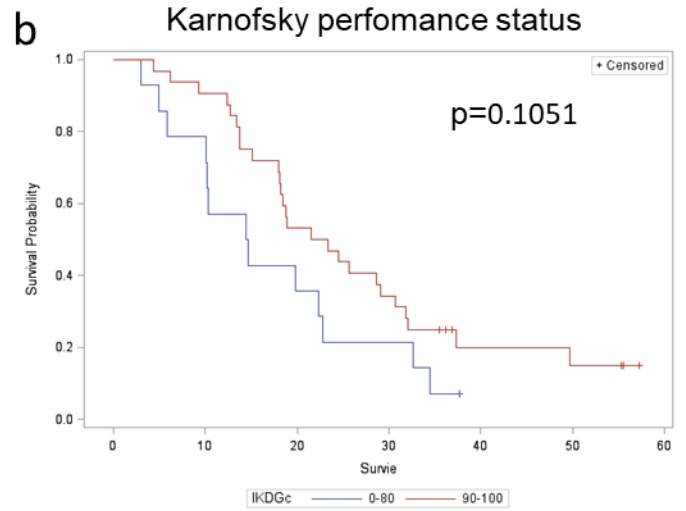
**Supplementary Figure 2:** Individual segmentation of 46 tumors by MALDI-MSI and comparison with pathologist annotations.



**Supplementary Figure 3.** String analysis of overexpressed proteins in each of the three proteomic groups and relation to patient survival **A. a)** Analysis of proteins overexpressed in group A shows involvement in axon guidance. **b)** Proteins overexpressed in group B and mainly involved in complements, coagulation cascade and, inflammation **c)** Analysis of overexpressed proteins in group C shows a network of proteins involved in Epstein Barr infection. **B.** Correlation between Grancalcin and CAPG expression and glioma patient survival according to the TGCA data. Patients were divided based on level of expression into “low” or “high” (n = 153 patients).

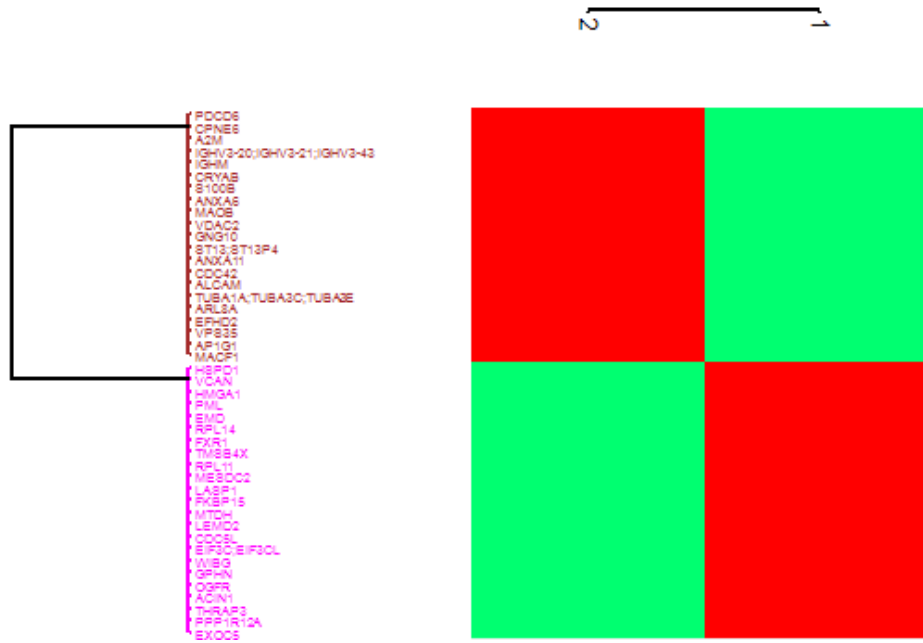
a

	Pr > Chi-Square
<b>Karnofsky indice at diagnostic 0-80 vs. 90-100</b>	0.1051
<b>MGMT</b>	0.0882
<b>Quality of resection complete, subtotal vs. partial, biopsies</b>	0.7666

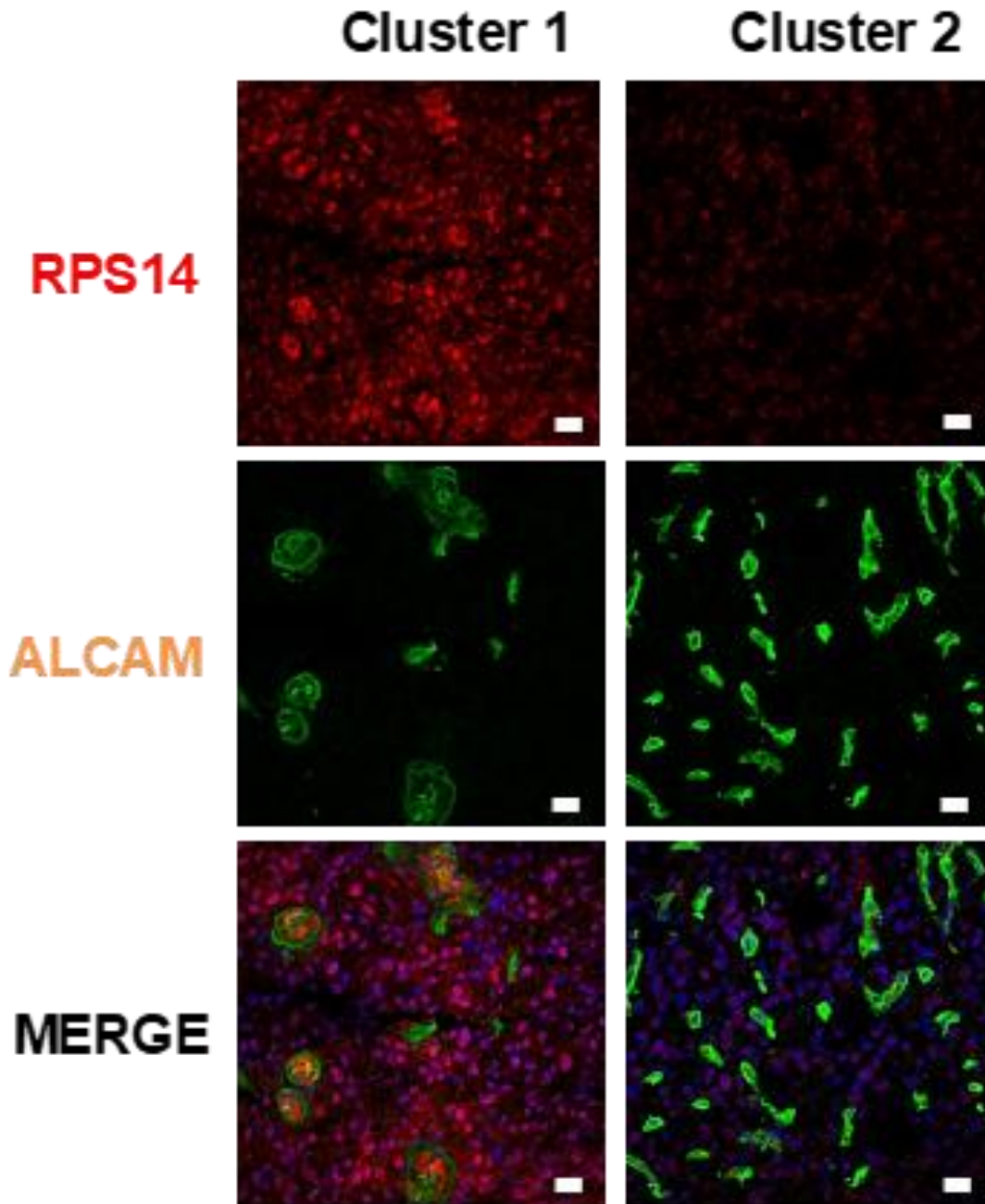


**Supplementary Figure 4.** Survival curves of all patients according to the Karnofsky performance status, MGMT promoter methylation status, and quality of resection. (a) Clinicopathological factors description ( $\chi^2$ -test). Survival curves (Kaplan-Meier Analysis) of patients according to the Karnofsky performance status (b), MGMT status (c), and resection quality (d). n = 46 patients. The lifetest procedure was used for the statistical analyses (log-rank test, Wilcoxon test).



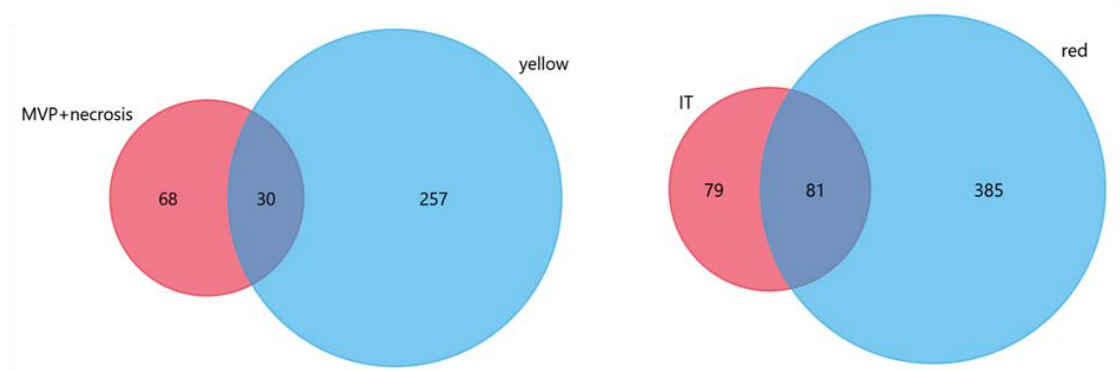


**Supplementary Figure 5.** Heatmap of the proteins significantly overexpressed in either of the 2 OS clusters (cluster 1 and cluster 2) with a  $p < 0.05$ . Heatmap obtained from the spatially resolved proteomic data which significantly showing overexpression of proteins correlating with the 2 OS clusters (cluster 1 and cluster 2) presenting identified in the patient prospective cohort with a  $p < 0.05$ . This heatmap only includes the RefProts and does not include the 5 AIProts with a  $p < 0.05$ . Statistical analyses were performed using ANOVA.

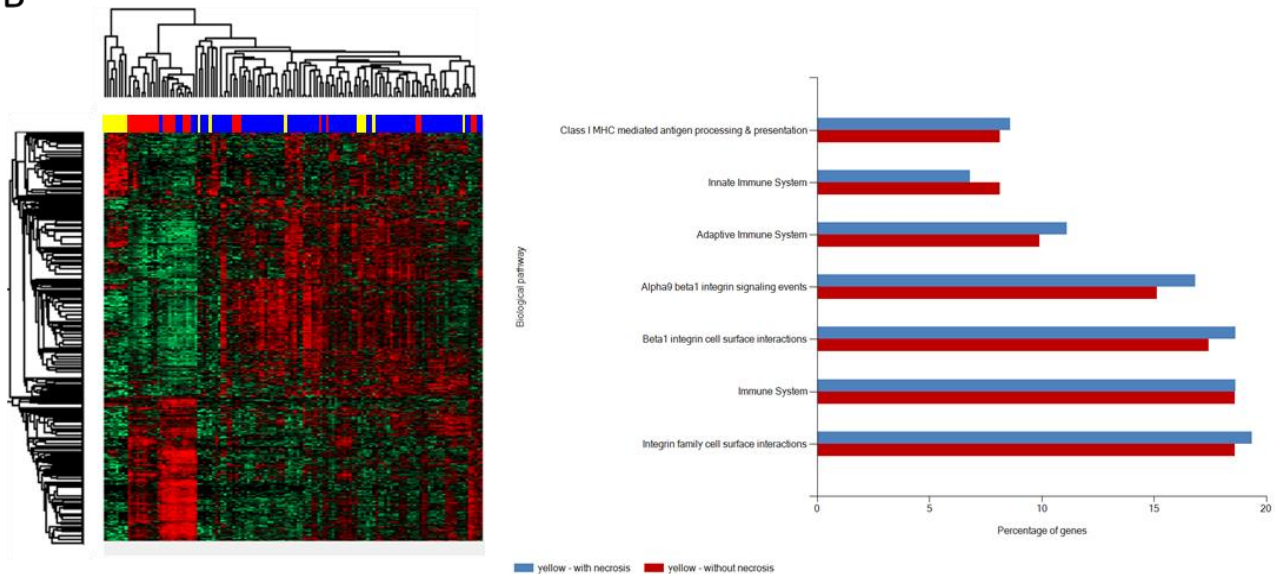


**Supplementary Figure 6.** Validation immunohistochemistry of RPS14 and ALCAM markers. Representative fluorescence images of RPS14 and ALCAM in the two OS clusters of patients. ALCAM is associated with a bad prognosis while RPS14 is related to a good prognosis. Images were acquired with a confocal microscope at 40x magnification. The experiment was repeated on representative tissues of 23 patients for the prospective cohort and on representative tissues of 50 patients for the validation cohort. For each tissue, 3 to 4 images were taken for quantification. scale bar = 20  $\mu$ m

A



B



**Supplementary Figure 7. Impact of histological features on proteomics analysis. A. Comparative analysis with the study of Lam *et al.* (2022)<sup>1</sup>.** Venn Diagrams comparing the number of common and differential proteins either in group B (yellow) vs the histological region “MVP+necrosis” of the study of Lam *et al.* or in group A (red) vs the histological region named infiltrated tumor (IT) of the study of Lam *et al.* **B. Proteomics analysis without the micro-extracted points in the necrotic region.** Heatmap of proteins with different regulation profiles as determined by label-free quantification in the three groups without the proteins coming from the necrotic region and comparative analysis of the biological pathways enriched in group B (yellow) with or without the necrotic region.

## Supplementary References

<sup>1</sup> Lam, K. H. B. et al. Topographic mapping of the glioblastoma proteome reveals a triple-axis model of intra-tumoral heterogeneity. *Nat. Commun.* 13, (2022).