# nature research

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## **Reporting Summary**

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our <u>Editorial Policies</u> and the <u>Editorial Policy Checklist</u>.

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For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.				
n/a	Confirmed			
	$\square$ The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement			
	A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly			
	The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.			
	A description of all covariates tested			
	A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons			
	A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)			
	For null hypothesis testing, the test statistic (e.g. <i>F</i> , <i>t</i> , <i>r</i> ) with confidence intervals, effect sizes, degrees of freedom and <i>P</i> value noted <i>Give P values as exact values whenever suitable.</i>			
$\boxtimes$	For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings			
	For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes			
	$\boxtimes$ Estimates of effect sizes (e.g. Cohen's $d$ , Pearson's $r$ ), indicating how they were calculated			
	Our web collection on <u>statistics for biologists</u> contains articles on many of the points above.			
Sof	ftware and code			

## Policy information about <u>availability of computer code</u>

Data collection No commercial, open source, or custom software or code was used for data collection.

Data analysis Individual software components are as follows:

 $R(v\ 4.1.2), DeMixT\ (v\ 1.2.2), ASCAT\ (v2.5.2), sequenza\ (v\ 3.0.0), Seurat\ (v\ 3.0), Monocle\ 2\ (v\ 2.14.0), CytoTRACE\ (v\ 0.3.3))$ 

Downstream analyses and statistical analyses were performed using package as follows:

rpart (v 4.1.15), survival (v 3.2.0), survminer (v 0.4.9), gprofiler2 (v 0.2.0), fgsea (v 1.14.0), DSS (v 2.42.0), ChIPseeker (v 3.11), ComBat (v

3.20.0), limma (v 3.50.1), GeneOverlap (v 1.24.0), ComplexHeatmap (v 2.4.3), RColorBrewer (v1.1.2)

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

#### Data

Policy information about <u>availability of data</u>

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data
- A description of any restrictions on data availability

The UMI counts of the hepatocellular carcinoma single cell RNA sequencing data were downloaded from the Gene Expression Omnibus (GEO) with the accession code GSE125449. The UMI counts and cell type annotations of the lung adenocarcinoma single cell RNA sequencing data were downloaded from the ArrayExpress under accessions E-MTAB-6149. The UMI counts of the colorectal adenocarcinoma single cell RNA sequencing data are available at http://crcmoonshot.org/?page\_id=189. FASTQ files of single-cell RNA sequencing data from pancreatic cancer will be publicly available on the GEO with the

according code GSE1					
accession code GSE156405. Raw read counts from the mixed cell-line study were downloaded from GEO with accession code GSE121127. Raw read counts of RNA sequencing data, clinical data, and somatic mutations from 7,054 tumor samples across 15 TCGA cancer types are available for download from the Genomic Data Commons Data Portal (https://portal.gdc.cancer.gov/). ATACseq data for TCGA samples were downloaded from					
https://science.scien	cemag.org/content/362/6413/eaav1898/tab-figures-data.				
	f ICGC-EOPC was downloaded from  tirect com/science/article/pii/S15356108183048232via%3Dibub#gs1				
https://www.sciencedirect.com/science/article/pii/S1535610818304823?via%3Dihub#gs1.  All primary METABRIC data including Affymetrix SNP 6.0 CEL files and Illumina HT 12 gene expression arrays, are available at the EGA (EGAS00000000083), and may be downloaded from https://ega-archive.org/studies/EGAS00000000083. Clinical information of METABRIC was downloaded from https://www.cbioportal.org/study/clinicalData?id=brca metabric.					
Clinical information of	f TRACERx was downloaded from				
	g/doi/full/10.1056/NEJMoa1616288#article_supplementary_material. was downloaded from https://ega-archive.org/studies/EGAS00001002247.				
RNAseq data of TRAC	ERx was downloaded from https://ega-archive.org/studies/EGAS00001003458.				
TmS values of all sam	ples and the identified intrinsic tumor signature genes for this study are available for download at https://github.com/wwylab/TmS.				
Fiold-sno	cific reporting				
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	e below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.				
X Life sciences	Behavioural & social sciences Ecological, evolutionary & environmental sciences				
or a reference copy of t	ne document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>				
Life scier	ces study design				
All studies must dis	close on these points even when the disclosure is negative.				
Sample size	No statistical methods were used to predetermine sample size. Sample size was determined by the availability of the data.				
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Data exclusions	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.				
Data exclusions Replication					
	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.				
Replication	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.  All attempts at replication were successful.				
Replication Randomization	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.  All attempts at replication were successful.  There are no experimental groups in this study.				
Replication Randomization Blinding	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.  All attempts at replication were successful.  There are no experimental groups in this study.  There are no experimental groups in this study.				
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Replication Randomization Blinding Reporting We require information	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.  All attempts at replication were successful.  There are no experimental groups in this study.				
Replication  Randomization  Blinding  Reporting  We require information  system or method list	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.  All attempts at replication were successful.  There are no experimental groups in this study.				
Replication  Randomization  Blinding  Reporting  We require information  system or method list	Samples with missing RNAseq data or DNA sequencing data were excluded from the analysis.  All attempts at replication were successful.  There are no experimental groups in this study.  There are no experimental groups in this study.  Some from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, and is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.  Methods  Methods				

Materials & experimental systems		Methods	
n/a	Involved in the study	n/a	Involved in the study
$\boxtimes$	Antibodies	$\boxtimes$	ChIP-seq
$\boxtimes$	Eukaryotic cell lines	$\boxtimes$	Flow cytometry
$\boxtimes$	Palaeontology and archaeology	$\boxtimes$	MRI-based neuroimaging
$\boxtimes$	Animals and other organisms		
	Human research participants		
$\boxtimes$	Clinical data		
$\boxtimes$	Dual use research of concern		

### Human research participants

Policy information about studies involving human research participants

Population characteristics

Demographically, the single-cell RNA sequencing datasets from three colorectal patients includes 1 male at age 45, 1 male at 37 and 1 female at age 63.

Recruitment

The three colorectal adenocarcinoma patients with single-cell RNA sequencing data were identified prior to surgery or biopsy and were asked to prospectively sign consent for participation on the IRB approved protocol LAB10-0982 after discussion of risks and benefits. Prospective consenting of patients was required due to the intent to utilize fresh tumor tissue. No known selection bias other than the fact that these patients went through surgery at MD Anderson.

Note that full information on the approval of the study protocol must also be provided in the manuscript.